### HBFP: a new repository for human body fluid proteome

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#### Abstract

Body fluid proteome has been intensively studied as a primary source for disease biomarker discovery. Using advanced proteomics technologies, early research success has resulted in increasingly accumulated proteins detected in different body fluids, among which many are promising biomarkers. However, despite a handful of small-scale and specific data resources, current research is clearly lacking effort compiling published body fluid proteins into a centralized and sustainable repository that can provide users with systematic analytic tools. In this study, we developed a new database of human body fluid proteome (HBFP) that focuses on experimentally validated proteome in 17 types of human body fluids. The current database archives 11 827 unique proteins reported by 164 scientific publications, with a maximal false discovery rate of 0.01 on both the peptide and protein levels since 2001, and enables users to query, analyze and download protein entries with respect to each body fluid. Three unique features of this new system include the following: (i) the protein annotation page includes detailed abundance information based on relative qualitative measures of peptides reported in the original references, (ii) a new score is calculated on each reported protein to indicate the discovery confidence and (iii) HBFP catalogs 7354 proteins with at least two non-nested uniquely mapping peptides of nine amino acids according to the Human Proteome Project Data Interpretation Guidelines, while the remaining 4473 proteins have more than two unique peptides without given sequence information. As an important resource for human protein secretome, we anticipate that this new HBFP database can be a powerful tool that facilitates research in clinical proteomics and biomarker discovery.

Database URL: https://bmbl.bmi.osumc.edu/HBFP/

#### Background

Human body fluids are thought to be rich resources of diseaseassociated proteins that are secreted or leaked from pathological tissues across the body, many of which are commonly obtainable through non-invasive procedures (1, 2). Driven by these factors, research interests have soared a few decades ago toward biomarker discovery by examining body fluid proteomes. It is highly plausible that empowered by innovative high-throughput technologies, modern proteomic studies have successfully identified a large number of proteins in various body fluids such as plasma, serum, saliva and urine (3).

With great effort by a few large consortiums, several community-based proteomic databases have been developed in the past decades. For example, in 2002, the international Human Proteome Organization initiated the Human Plasma Proteome Project and reported human plasma and serum protein constituents in its online databases (4). Another similar database, named Plasma Proteome Database, archived more than 10 000 proteins detected in human blood (5). Additionally, the Proteomics Identifications database (6) and Human Plasma PeptideAtlas (7) report a total of 3509 highconfidence plasma proteins. More recently, the extracellular vesicles community also reports new proteins identified in exosomes in multiple different resources including blood and breast milk, e.g. in ExoCarta (8). Additionally, the global Human Proteome Project (HPP) announces a set of mass spectrometry (MS) data interpretation guidelines that are presented to the broader research community (9).

Our team has recently conducted a systematical assessment of human proteome identified using quantitative proteomics tools such as MS and computational predictive models, as documented in a recent review article (10). To expand this effort, we developed a new human body fluid proteome (HBFP) database to organize 11 827 unique proteins reported in 164 scientific articles since 2001, which has a maximal false discovery rate (FDR) of 0.01 on both the peptide and protein levels. Until today, this database

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stores information about proteins from 17 types of body fluids including plasma/serum, saliva, urine, cerebrospinal fluid (CSF), seminal fluid (SF), amniotic fluid, tear fluid, bronchoalveolar lavage fluid (BALF), milk, synovial fluid, nipple aspirate fluid, cervical-vaginal fluid, pleural effusion, sputum, exhaled breath condensate, pancreatic juice and sweat.

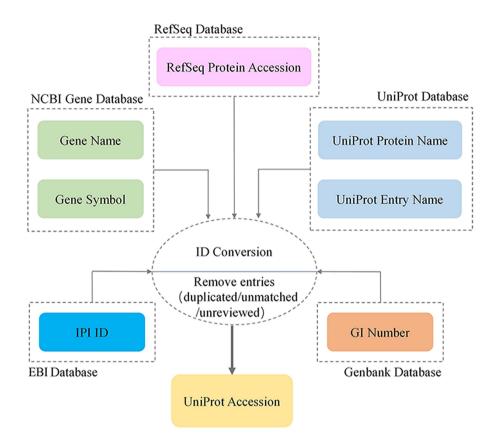
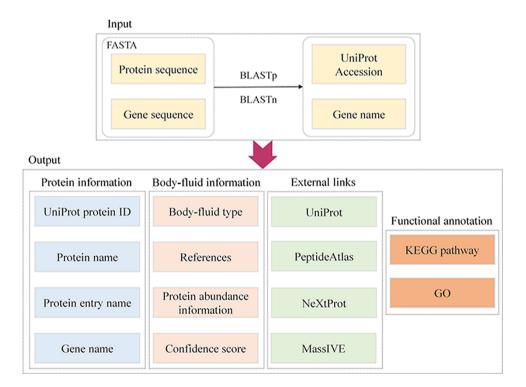


Figure 1. Workflow of protein identifier conversion.



For each protein entry, description about protein secretion information, literature source, abundances, confidence and functional annotation is provided. This database system also provides users easy access to data visualization and download and functional analysis based on Gene Ontology (GO) and pathways.

#### **Database content and design**

#### **Protein entries**

We have manually collected proteins reported in 17 types of body fluids by carefully reviewing 164 scientific references published since 2001 based on a PubMed search with FDR  $\leq$ 1% on both the peptide and protein levels.

In the HBFP database, each protein is assigned with a unique identifier of UniProtKB/Swiss-Prot accession (UniProt release 2020\_06) (11). Since different identifiers have been mixed used in the referenced studies, we first used conversion tools at BioDBnet (https://biodbnet-abcc.ncifcrf.gov/)

(12) and UniProt (https://www.UniProt.org/) to confidently convert different identifiers to UniProt accession numbers. The common identifiers involved in this study include International Protein Index ID [hosted at European Bioinformatics Institute (EBI) (closed in 2011)], GI number (from Genbank database), RefSeq protein accession (from RefSeq database), Gene name/symbol (from NCBI Gene database) and UniProt protein/entry name (from UniProt database). The ID conversion process is shown in Figure 1. During the conversion, poorly curated proteins with ambiguous identifiers were eliminated. For examples, many International Protein Index ID links to unclearly described instances that cannot be mapped to a UniProt entry are excluded.

#### **Database utilities**

The interface of the HBFP database is constructed by PHP, while the database system is based on MySQL. The main contents of the current database include query and browse pages described as follows.

Table 1. Overall statistics

		Sta	ntistics	
Body fluid types		Number of protein entries	Number of references	References
1	Plasma/serum	5790	38	(18–55)
2	Saliva	2758	21	(19, 56-75)
3	Urine	7330	23	(19, 76–97)
4	CSF	4364	12	(19, 90, 98-107)
5	SF	4084	5	(108–112)
6	Amniotic fluid	3025	6	(19, 113–117)
7	Tear fluid (TF)	1882	11	(118–128)
8	BALF	3434	6	(41, 129–133)
9	Milk	2457	14	(134–147)
10	Synovial fluid	1637	7	(148–154)
11	Nipple aspirate fluid	1734	5	(155–159)
12	Cervical-vaginal fluid	949	4	(160–163)
13	Pleural effusion	1519	3	(164–166)
14	Sputum	1809	3	(167–169)
15	Exhaled breath condensate	351	5	(170–174)
16	Pancreatic juice	702	4	(175–178)
17	Sweat	1244	3	(179–181)
Total (non-redundant)		11 827	164	

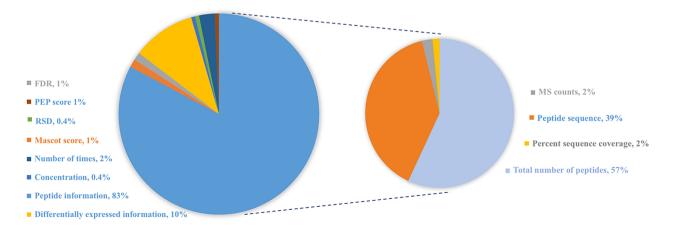


Figure 3. Distribution of protein abundance methods in HBFP database based on a number of original quantitative analysis methods from the original literature studies. Note that the sum of protein abundance is not 100% since not all of the literature studies provide quantitative analysis information.

#### Querying page

As one of the most important functions, the querying page allows users to search for body fluid proteins based on different types of input including protein ID, gene name, and protein or gene sequence. When given a FASTA input, BLASTp or BLASTn is used to translate sequence input to the best-match protein entry. The top hit (the highest bit score) from the BLAST search is considered the best match of the query. Figure 2 illustrates the workflow and content of querying page.

The annotation of each protein contains the following information:

- Protein ID/name/entry name
- Gene name
- Associated body fluid type along with indicated discovery confidence (explained in the next section)
- References and protein abundance information where the protein is reported
- External links to public databases including UniProt, PeptideAtlas and NeXtProt (13), MassIVE (14)
- Functional annotation based on the KEGG pathway (15) and GO (16)

#### Browsing page

This page provides an overview list of proteins associated with 17 types of body fluids and links to view and download selected proteins.

#### **Database highlights**

#### Data statistics

When determining the inclusion of reported proteins, we applied the following criteria for credibility of the MS

evidence. First, for papers that issued peptide sequence details, we remapped all those peptide sequences to neXtProt (release 2021-02-15) using the neXtProt peptide uniqueness checker to remove unreliable matches (17). Specifically, we applied guideline #15 of HPP Guidelines 2.1 (9) to include proteins that contain at least two non-nested uniquely mapping peptides of nine amino acids into the HBFP database. According to this criterion, 7354 proteins were confirmed confidently. Another 4473 proteins were also included as they were not explicitly provided with peptide sequence information but have more than two unique peptides.

The overall statistics about the protein entries and references in terms of each body fluid are summarized in Table 1. The current HBFP database contains 11827 distinct proteins from 17 types of body fluids. Note that urine exceeds all other body fluids in terms of protein counts while blood is at the second rank. All data are made publicly available in the HBFP and via links at https://bmbl.bmi.osumc.edu/ HBFP/.

#### Protein abundance

In order to provide users experimental evidence from the original study, this database also displays relatively abundant information from the corresponding literature studies. General proteomics approaches using MS identify proteins by matching identified peptides against predefined protein sequence databases. The qualitative measures of protein reported in the original reference include the following: (i) peptide information: most of cited studies provide explicit information about peptide sequence, the total number of peptides, MS counts or the percent sequence coverage; (ii) differential expression information including fold change (positive value demonstrates up-regulated expression and negative

А	С	Query:P58340 less		
Protein ID List (less than 50)	Select Conditions Query with UniProt Protein ID List	UniProt Protein ID Protein Name	P58340 (FASTA) Myeloid leukemia factor 1	
Gene Name List (less than 50)	P58340	UniProt Entry Name	MLF1_HUMAN Body fluid (the total) p value	
Protein		Gene Name	MLF1	
Sequence(FASTA)		Related Fluid Type Information	bronchoalveolar lavage fluid (6 papers in total) [ confidence score: 0.965 seminal fluid (5 papers in total) [ confidence score: 0.97	
Sequence(FASTA)	Query Example Clear	Reference (* means manual remap)	(bronchoalveolar lavage fluid) ← Body fluid	
В		Link to PubMed/	<ul> <li>Sim SY., Choi YR., Lee JH. et al. (2019) In-depth proteomic analysis of huma bronchoalveolar lavage fluid toward the biomarker discovery for lung cancer Proteomics. Clinical applications, 13, e1900028[Loa2,IBA0=25.5211523073577]</li> </ul>	
Protein ID List (less than 50)	Select Conditions		(seminal fluid) Protein abundance	
Gene Name List (less than 50) Protein	Sequence MUST be provided in FASTA format: >sp[P58340] MLF1_HUMAN Myeloid leukemia factor 1 OS=Homo sapiens OX=9606 GN=MLF1 PE=1 SV=1	$*$ means manual remap $\longrightarrow$ .	<ul> <li>Wang G., Guo Y., Tao Z. et al. (2013)In-depth proteomic analysis of the humar sperm reveals complex protein compositions. Journal of Proteomics, 79, 114-122.(11 unique peptides)</li> </ul>	
Sequence(FASTA)	MFRMLNSSFEDDPFFSESILAHRENMRQMIRSFSEPF0 RDLLSISDGRGRAHNRRGHNDG	Other Related Resourses	UniProt   PeptideAtlas   neXtProt   MassIVE	
Gene Sequence(FASTA)	EDSLTHTDVSSFQTMDQMVSNMRNYMQKLERNFGQ SVDPNGHSFCSSSVMTYSKIGDEP PKVFQASTQTRRAPGGIKETRKAMRDSDSGLEKMAIG		Link to Uniprot / PeptideAtlas / neXtProt / MassIVE	
	Query Example Clear D	KEGG Pathway ID		
		hsa:4291(ko:K15622)		
		Related GO Information		
		GO ID		
		ciliary basal body [GO:0036064]		
		cilium [GO:0005929]		

Figure 4. Example of query response with input as 'P58340' in the protein ID and protein sequence box.

value indicates down-regulated expression), up- or downregulated expression in case vs. control or (normalized) spectral counts and (iii) other statistical information including FDR, relative standard deviation and the number of times across different samples or experiments, as shown in Figure 3.

#### Confidence score

In the HBFP database, to evaluate the confidence level of each discovered protein in each body fluid, a new statistical measure is calculated based on Guideline # 9 of HPP guidelines 2.1 for the combined datasets. It is a well-known phenomenon that when taking N datasets with a substantial FDR and piling them all together, the overall FDR increases with the number of datasets. For example, for plasma, there are 38 papers with plasma protein lists, each with a substantial FDR ( $\leq 1\%$ ). It is probably a conservative estimate to suppose that the FDR of such a combined result is  $1\% + 0.5\% \times (N \text{ datasets}-1)$  (9). It means that 50% of the correct identifications overlap and none of the incorrect ones does, so the resulting FDR is added in a 0.5% increment. Meanwhile, the confidence level of protein in the combined datasets is also reduced. Otherwise, considering the overlap of the true positives, the larger the number of datasets in which a protein is associated with a specific fluid, the more reliable this protein is. In the end, a confidence score *C* is calculated as follows:

$$C_{i,j} = A_i + 0.5\% \times (M_j - 1)$$
(1)

$$A_i = 1 - FDR_i \tag{2}$$

$$FDR_i = 1\% + 0.5\% \times (N_i - 1) \tag{3}$$

View By 3		L Download All Download the interest or all proteins		1 to 50 of 2457 >>		
plasma/serum (5790)	2 🔳	Protein ID	Protein N	ame	(	Confidence Score
saliva (2758)		P01833	Polymeric i	mmunoglobulin receptor	0	.99
urine (7330)		P47710	Alpha-S1-c	asein [Cleaved into: Casoxin-D]	0	99
cerebrospinal fluid (4364)		Q13410	Butyrophilir	n subfamily 1 member A1	0	.99
seminal fluid (4084)		P05814	Beta-caseir		0	.985
		P10451	Osteopontii	1	0	.98
amniotic fluid (3025)		P00709	Alpha-lacta	lbumin	0	.975
tear fluid (1882)		P07498	Kappa-case	in	0	.975
bronchoalveolar lavage fluid (3434)		P10909	Clusterin		0	.975
milk (2457)		P15941	Mucin-1		0	.975
		P19835	Bile salt-ac	tivated lipase	0	.975
synovial fluid (1637)		P22897	Macrophag	e mannose receptor 1	0	.975
nipple aspirate fluid (1734)		Q6WN34	Chordin-like	e protein 2	0	.975
cervico vaginal fluid (949)		Q99541	4 A	В		C
pleural effusion (1519)		P00738	1 ProteinId	ProteinName		onfidence Score
sputum (1809)		P01024	2 P01833 3 P47710	Polymeric immunoglobulin rece Alpha-S1-casein [Cleaved into		0.99 0.99
exhaled breath		P01591	4 Q13410 5 P05814	Butyrophilin subfamily 1 memb Beta-casein	ber Al	0.99
condensate (351)		P01876	6 P10451	Osteopontin		0. 98
pancreatic juice (702)		P02788	7 P00709 8 P07498	Alpha-lactalbumin Kappa-casein		0.975
sweat (1244)		POC0L5	9 Q6WN34	Chordin-like protein 2		0.975
			10 Q99541 11 P10909	Perilipin-2 Clusterin		0.975
			12 P15941	Mucin-1		0.975
		_	13 P19835	Bile salt-activated lipase		0.975
			14 P22897	Macrophage mannose receptor 1		0.975
			15 P01024	Complement C3		0.97
	16 Q08431 Lactadherin			0.97		
17 P00738 18 P01591 19 P01876				Haptoglobin		0.97
			Immunoglobulin J chain		0.97	
				Immunoglobulin heavy constant	alpha 1	0.97
			20 P02788	Lactotransferrin		0.97
			21 POCOL5	Complement C4-B		0.97

Figure 5. Download illustration where user can choose the body fluid name and download the proteins of interest or all proteins.

where  $N_i$  is the number of relevant literature studies (datasets) of a specific fluid *i*;  $FDR_i$  represents the overall FDR of multiple datasets in body fluid *i*;  $A_i$  means the confidence level of proteins in the combined datasets of body fluid *i* and  $M_j$  refers to the number of literature studies in which a protein *j* is identified in body fluid *i*.

For example, there are 38 literature studies related to blood in the HBFP, so  $N_i = 38$ ,  $FDR_i = 0.195$  and  $A_i = 0.805$ . The protein O14791 is identified in blood by 19 independent studies, i.e.  $M_j = 19$ . As a result, the calculated  $C_{i,j}$  score for O14791 in blood is 0.895. Meanwhile, protein Q9UJV9 only is identified in one paper for blood, so  $M_j = 1$  and  $C_{i,j} =$  $A_i = 0.805$ . It means that protein Q9UJV9 maintains only the confidence level in the combined datasets of blood. Specifically, protein P01833 is identified in milk by 14 studies, and a total of 14 literature studies on milk are included in the HBFP, so protein P01833 maintains the original confidence level, i.e. 0.99. The larger the *C* score, the higher the confidence that a protein reported in that fluid will be. Note that this score can only be compared within the same type of body fluid.

#### **Database applications**

Data access

The website can be accessed through https://bmbl.bmi. osumc.edu/HBFP/.

#### Query

All proteins can be easily accessed by searching protein ID, gene name, protein sequence (FASTA) or gene sequence (FASTA) (<50 items per query) (Figure 4A and B as an example). A BLAST (182) is performed locally to find the best match when the sequence FASTA format is given. For each protein, detailed information is displayed (Figure 4C).

Users can connect directly to the PubMed or Google Scholar to view the original study through the provided links. Four databases (UniProt, PeptideAtlas, NeXtProt and MassIVE) are cross-linked for additional protein annotation, while the KEGG pathway and GO are focused on the functional aspects (Figure 4D).

#### Download

HBFP allows users to browse the entire protein list in each body fluid, where the proteins are ordered based on descending confidence scores. Users can check and download all entries of the selected body fluid type in one go, as shown in Figure 5.

## Demo of comparative analysis using the HBFP database

#### Body fluid analysis

Many proteins in the HBFP database have a broad distribution in terms of body fluid types. An internal comparative analysis across different fluids can provide further information regarding the specificity of a proposed marker protein. Of 11 827 identified proteins, 66.8% are identified in at least two body fluids (Figure 6). A total of 93 proteins (0.79%) are shared among all analyzed body fluids, which may indicate that these proteins are essential for various life activities (Table 2).

#### Venn diagram and GO annotation

To take a closer look at this comparison, we focused on five body fluids that have the most protein counts, including blood, urine, CSF, SF) and BALF. An interesting discovery is that urine shares large numbers of common proteins with other fluids (Figure 7). A total of 4109, 3212, 2990 and 2950 proteins overlapped between the plasma and the other

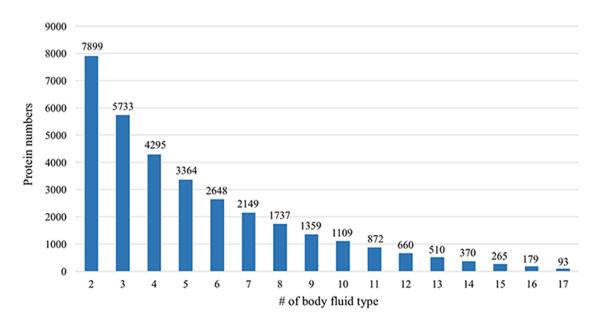


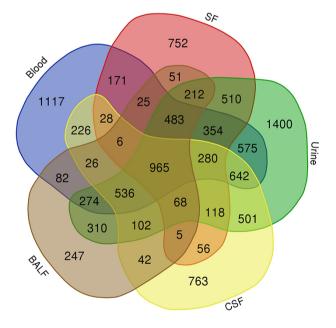
Figure 6. Comparative analysis across different body fluids. Seven thousand eight hundred and ninety-nine (7899) proteins are presented in at least two body fluids and 5733 proteins existed in at least three body fluids. Only 93 proteins exist in all 17 body fluids.

 Table 2. List of 93 proteins shared among all 17 body fluids

	UniProt accession number	Protein name	Gene name
1	P11021	Endoplasmic reticulum chaperone BiP	HSPA5
2	P55072	Transitional endoplasmic reticulum ATPase	VCP
3	P13647	Keratin, type II cytoskeletal 5	KRT5
4	O00299	Chloride intracellular channel protein 1	CLIC1
5	P02787	Serotransferrin	TF
6	P22314	Ubiquitin-like modifier-activating enzyme 1	UBA1
7	P13645	Keratin, type I cytoskeletal 10	KRT10
8	P02533	Keratin, type I cytoskeletal 14	KRT14
9 10	P07237 P06576	Protein disulfide-isomerase	P4HB ATP5F1B
10	P30041	ATP synthase subunit beta, mitochondrial Peroxiredoxin-6	PRDX6
11	P63104	14-3-3 protein zeta/delta	YWHAZ
12	P62258	14-3-3 protein epsilon	YWHAE
13	P14923	Junction plakoglobin	JUP
15	P04040	Catalase	CAT
16	P01834	Immunoglobulin kappa constant	IGKC
17	P06702	Protein S100-A9	S100A9
18	P52209	6-Phosphogluconate dehydrogenase, decarboxylating	PGD
19	P18669	Phosphoglycerate mutase 1	PGAM1
20	P14618	Pyruvate kinase PKM	PKM
21	P61981	14-3-3 protein gamma	YWHAG
22	P07384	Calpain-1 catalytic subunit	CAPN1
23	P50395	Rab GDP dissociation inhibitor beta	GDI2
24	Q00610	Clathrin heavy chain 1	CLTC
25	P26641	Elongation factor 1-gamma	EEF1G
26	P32119	Peroxiredoxin-2	PRDX2
27	P19971	Thymidine phosphorylase	TYMP
28	P26038	Moesin	MSN
29	P40121	Macrophage-capping protein	CAPG
30	P35754	Glutaredoxin-1	GLRX
31	P01009	Alpha-1-antitrypsin	SERPINA1
32	P01860	Immunoglobulin heavy constant gamma 3	IGHG3
33	P06753	Tropomyosin alpha-3 chain	TPM3
34	P68871	Hemoglobin subunit beta	HBB
35	P62805	Histone H4	H4C1
36	P30086	Phosphatidylethanolamine-binding protein 1	PEBP1
37	P35579	Myosin-9	MYH9 A2M
38 39	P01023	Alpha-2-macroglobulin Peroxiredoxin-1	PRDX1
39 40	Q06830 P02042	Hemoglobin subunit delta	HBD
40	P07737	Profilin-1	PFN1
42	P80188	Neutrophil gelatinase-associated lipocalin	LCN2
43	P02679	Fibrinogen gamma chain	FGG
44	P40925	Malate dehydrogenase, cytoplasmic	MDH1
45	P08758	Annexin A5	ANXA5
46	P46940	Ras GTPase-activating-like protein IQGAP1	IQGAP1
47	P01833	Polymeric immunoglobulin receptor	PIGR
48	P31949	Protein S100-A11	\$100A11
49	P04792	Heat shock protein beta-1	HSPB1
50	P07339	Cathepsin D	CTSD
51	P01857	Immunoglobulin heavy constant gamma 1	IGHG1
52	P06733	Alpha-enolase	ENO1
53	P23284	Peptidyl-prolyl cis-trans isomerase B	PPIB
54	P02647	Apolipoprotein A-I	APOA1
55	O43707	Alpha-actinin-4	ACTN4
56	P30740	Leukocyte elastase inhibitor	SERPINB1
57	Q16610	Extracellular matrix protein 1	ECM1
58	P60709	Actin, cytoplasmic 1	ACTB
59	P15924	Desmoplakin	DSP
60	P62937	Peptidyl-prolyl cis-trans isomerase A	PPIA
61	P17931	Galectin-3	LGALS3
62	P00491	Purine nucleoside phosphorylase	PNP
63	P04080	Cystatin-B	CSTB
64	P02788	Lactotransferrin	LTF
65	P13639	Elongation factor 2	EEF2
66	P35527	Keratin, type I cytoskeletal 9	KRT9
67	P06396 P59998	Gelsolin Actin-related protein 2/3 complex subunit 4	GSN ARPC4
68			

Table 2. (Continued)

	UniProt accession number	Protein name	Gene name
69	P25311	Zinc-alpha-2-glycoprotein	AZGP1
70	P02768	Albumin	ALB
71	P61160	Actin-related protein 2	ACTR2
72	P04406	Glyceraldehyde-3-phosphate dehydrogenase	GAPDH
73	P60174	Triosephosphate isomerase	TPI1
74	P18206	Vinculin	VCL
75	P08670	Vimentin	VIM
76	P10599	Thioredoxin	TXN
77	P11142	Heat shock cognate 71 kDa protein	HSPA8
78	P01011	Alpha-1-antichymotrypsin	SERPINA3
79	P04075	Fructose-bisphosphate aldolase A	ALDOA
80	P04264	Keratin, type II cytoskeletal 1	KRT1
81	P37837	Transaldolase	TALDO1
82	P35908	Keratin, type II cytoskeletal 2 epidermal	KRT2
83	P02545	Prelamin-A/C	LMNA
84	P69905	Hemoglobin subunit alpha	HBA1
85	P07900	Heat shock protein HSP 90-alpha	HSP90AA1
86	P29401	Transketolase	TKT
87	P00558	Phosphoglycerate kinase 1	PGK1
88	P00338	L-lactate dehydrogenase A chain	LDHA
89	P01861	Immunoglobulin heavy constant gamma 4	IGHG4
90	P05109	Protein S100-A8	S100A8
91	P04083	Annexin A1	ANXA1
92	P01024	Complement C3	C3
93	P09211	Glutathione S-transferase P	GSTP1



**Figure 7.** Venn diagram showing the common proteins among five body fluids (blood, urine, CSF, SF and BALF) that have the most number of proteins in the HBFP.

four body fluids (blood, CSF, SF and BALF, respectively). There are 965 proteins commonly detected in all five body fluids. The functional analysis using the BiNGO tool (183) in Cytoscape (184), reflecting information about cellular localization, molecular function and biological process of these proteins (Figure 8).

#### Conclusions

The new HBFP database developed in this study represents the first of its kind as a comprehensive reference resource of HBFP. All data are available through an open-access userfriendly Web platform. All protein entries were manually curated, which can be easily traced back to the original literature. Users can query and download proteins of interest to verify discovery in their own study or conduct an *in silico* analysis on human secretomes. We currently schedule a regular update every 6 months. The future plan is to include computationally identified proteins using statistical and machine learning approaches (185–191). In the past decade, many computational studies have revealed unique strengths in overcoming challenges in profiling-based proteomics research in terms of discovering new protein bioavailability and functions. Those computationally predicted proteins can serve as a secondary resource for biomarker discovery. In summary, by providing a wealth of information and functional analysis, we believe the HBFP database can be an excellent tool for the research community to explore human proteome in various body fluids.

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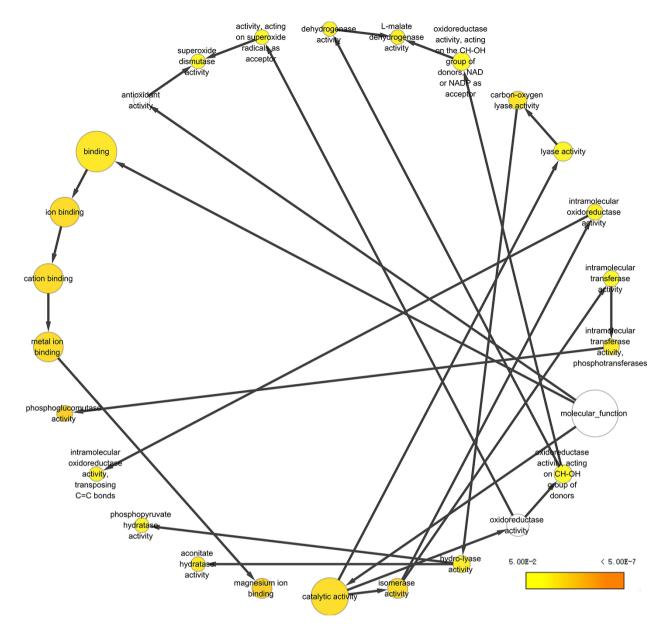


Figure 8. Example of GO annotation based on the 965 proteins common in five body fluids.

#### **Conflict of interest**

The authors declare that they have no competing interests.

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