

Lumbar disc herniation: Is there an association between histological and magnetic resonance imaging findings?

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ABSTRACT

Background: Although validated radiological scoring systems and histological scoring system of surgically removed degenerated disc are used in assessment of progression of intervertebral disc degeneration, there have not been many studies that integrate these two aspects of assessments. The data available in this respect are very limited. This clinical study was designed to find the correlation between quantitative radiological score (Pfirmann grading system and Modic changes [MC]) and quantitative histological degeneration score (HDS).

Materials and Methods: A cohort of 77 patients (45 males, 32 females; mean age of 38 years [range 18–58 years]) who presented with complaints of discogenic pain or radiculopathy at single level were assessed radiologically. They were graded according to the radiological pattern. The surgically excised disc specimen was graded according to HDS. The degree of radiological changes were correlated with the degree of histological changes.

Results: Though the overall HDS (0–15) did not show statistically significant correlation with Pfirmann grading system, there were positive association found between mucoid degeneration, chondrocyte proliferation with the Pfirmann grading and mucoid degeneration, which were statistically significant. Female sex also had a higher association with instability pattern.

Conclusion: The study shows that the Pfirmann grading system, MCs and HDS can reliably be used as scoring systems for assessing lumbar disc degeneration. The radiological assessment can be used as a noninvasive tool to assess the probable change in content rather than the microstructure of a disc undergoing degeneration.

Key words: Histological degeneration score, Modic changes, Pfirmann grading, surgical discectomy, lumbar disc herniation **MeSH terms:** Interventional disc, disc, herniated, IV Disc displacement, magnetic resonance imaging

INTRODUCTION

ow back pain is a common clinical problem affecting millions of people. Intervertebral disc (IVD) degeneration is attributed as a common cause of low back pain. However, patients with varying degrees of IVD degeneration exist and their clinical picture is variable. In addition, there is a section of patients who have predominant radicular symptoms in their legs and

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tolerable or minimal back pain. The association between disc degeneration and clinical symptom profile of the patient is not clear. Age related changes in the IVDs are also similar to disc degeneration except for the diffusion delay across the endplates in aged discs as proved by Rajasekaran *et al.*,¹ change in the type of proteoglycans, decrease in type 2 collagen networks, endplate fractures, fissures in the annulus have all been seen in degenerate IVDs.²⁻⁴ Magnetic resonance imaging (MRI) is currently used to assess the degeneration of IVDs.⁵ MRI provides excellent anatomic detail of spinal tissues, but fails to provide the type of information that permits a definitive diagnosis in many patients with back pain.⁵ Anatomic imaging of the spine is

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How to cite this article: Majeed SA, Seshadrinath NA, Binoy KR, Raji L. Lumbar disc herniation: Is there an association between histological and magnetic resonance imaging findings?. Indian J Orthop 2016;50:234-42. highly accurate in the evaluation of non mechanical causes of back pain and less beneficial in the evaluation of back pain, that is, due to mechanical causes.⁵ MRI helps in noting the signal changes in the discs, which have resulted from changes in the biochemical alterations in the disc while micro structural changes are less appreciated. MRI also helps to note the changes in endplates of the vertebra known as Modic changes (MC).⁶ It can structurally classify a disc herniation to bulge, protrusion, extrusion and sequestration. Pfirmann⁷ has graded the disc degeneration seen in T2 sagittal sequence of fast spin echo MRI into five grades. A normal well hydrated disc with a clear demarcation between nucleus pulposus and the annulus is graded as Grade 1 whereas a completely collapsed disc with absent nucleo-annular differentiation is considered as Grade 5. However, this grading is an arbitrary grading based on MRI characteristics alone. Though the histological changes in composition were taken into account while making the scoring system, it is also not clear whether the structural histopathological changes in the disc will be reflected in the routine MRI T2 protocols. MCs⁶ have been graded into type 1–3 depending on the signal intensity in T1 and T2 sequences.

Boos et al.⁸ categorized the histopathological changes in the degenerating IVDs in cadavers. They found that chondrocyte proliferation, granular changes, tears or cleft formation, mucoid degeneration, rim lesions and edge neovascularization to be the common findings in the degenerated IVDs. Based on this study, they developed a histological degeneration score (HDS) for the pathological grading of the degenerated discs. Weiler et al.⁹ in his study validated this in surgically removed specimens in Caucasian patients undergoing discectomies and found that the first 4 variables proposed by Boos et al.⁸ remained true for the surgically removed specimens [Table 1].

We studied the histological changes in herniated disc tissue and to find out if there is any kind of correlation between this and MRI findings of disc degeneration and end plate changes.

MATERIALS AND METHODS

77 consecutive patients undergoing lumbar discectomies for lumbar disc herniation (LDH) were studied for HDS. All patients were of Indian origin. Ethical clearance from Institutional Ethics Committee was obtained and all patients gave valid informed consent. All patients underwent plain radiographs of lumbosacral spine with flexion and extension views and MRI assessment with 1.5 Tesla machine. Our criteria for inclusion were patients who had single level LDH, who had failed a period of atleast 2 months of conservative care. We excluded chronic low back pain patients, patients

Table 1: Modified parameters collected for the histologic assessment of disc degeneration and scoring (Boos et al. 2002) Cı

| assessment of this degeneration | | | | | | | |
|--|--|--|--|--|--|--|--|
| Criteria | Grading | | | | | | |
| Cell density (chondrocyte | 0=no proliferation | | | | | | |
| proliferation): Multiple | 1=increased cell density | | | | | | |
| chondrocytes growing in small | 2=connection of two chondrocytes | | | | | | |
| rounded groups or clusters sharply demarcated by a rim of territorial matrix | 3=small size clones (several chondrocytes grouped | | | | | | |
| | together, 3-7 cells) | | | | | | |
| | 4=moderate size clones (8-15 cells) | | | | | | |
| | 5=huge clones (>15 cells) | | | | | | |
| Structural alterations (tears and | 0=absent | | | | | | |
| clefts): Concentric tears following | 1=rarely present | | | | | | |
| the collagen fiber bundle | 2=present in intermediate | | | | | | |
| orientation in the annulus fibrosus or radiating defects extending | amounts between 1 and 3 | | | | | | |
| from the nucleus pulposus to the | 3=abundantly present | | | | | | |
| outer annulus lamellae parallel or oblique to the end-plate (clefts) | 4=scar/tissue defects | | | | | | |
| Granular changes: | 0=absent | | | | | | |
| Eosinophilic-staining amorphous | 1=rarely present | | | | | | |
| granules within the fibrocartilage matrix | 2=present in intermediate amounts between 1 and 3 | | | | | | |
| | 3=abundantly present | | | | | | |
| Mucous degeneration: | 0=absent | | | | | | |
| Cystic, oval or irregular areas | 1=rarely present | | | | | | |
| with intense deposition of | 2=present in intermediate | | | | | | |
| acid mucopolysaccharides (i.e., sulfated glycosaminoglycans) | amounts between 1 and 3 | | | | | | |
| staining dark blue with alcian-PAS | 3=abundantly present | | | | | | |
| HDS | 0-15 points | | | | | | |

HDS=Histologic degeneration score, PAS=Periodic acid-Schiff

with multilevel spinal canal stenosis and patients with evidence of higher grades of instability (spondylolisthesis of Graded 2 or above) in plain radiographs. Thus, we had 14 patients with Grade 1 instability at a single level in the cohort of 77 patients as evidenced in flexion extension lateral views of lumbosacral spine.¹⁰ (>3.5 mm of anterior translation).

The Pfirmann grading⁷ of the concerned disc was noted according to the criteria suggested by Pfirmann by two experienced radiologists. The presence of MCs⁷ in the endplates was also noted. MRI assessment of the grade of disc protrusion was also done.

During surgical discectomy, the entire removed disc fragment was collected and stored in 10% formaldehyde, decalcified for 72 h, fixed in paraffin and thin sections of $2-4 \mu m$ was made. At least 5 sections were made from each specimen and analyzed by 2 experienced pathologists. Hematoxylin and eosin staining were performed and HDS was assessed. Alcian blue staining in acidic pH¹¹ was done to assess the mucoid changes. As per Boos et al. criteria, the HDS of the discectomy specimen was calculated taking into account four parameters: (1) Chondrocyte proliferation, (2) Tears or clefts, (3) Granular changes and (4) Mucoid changes [Figures 1-4]. The maximum HDS was 15. The CD34 immunohistochemistry was used to identify blood vessels in the inner annulus/nucleus and S100 immunohistochemistry was used to identify Schwann cells in the inner annulus/ nucleus¹² [Figure 5a and b]. A histomorphological distinction between annular and nuclear disc tissue was performed by the use of light microscopic criteria particularly under polarized light, allowing the evaluation of the organization of the collagen network. Three kinds of tissue were identified in the discectomy specimens. (a) Annulus fibrosus: Fibrocartilage that includes large crimped collagen fibers organized in lamellae, with elongated fibroblast like cells lying between lamellae. (b) Nucleus pulposus: Abundant irregular matrix is containing clefts, with a sparse population of rounded cells. (c) Cartilage endplate: Typical hyaline cartilage, with an amorphous matrix and with chondrocyte-like cells exhibiting clearly visible perinuclear halo.

Alcian blue stains¹⁰ the acidic mucopolysaccharide matrix blue. It is suggested that there is a relative increase in the more acidic keratin sulfate in degenerated discs.¹¹ The



Figure 1: Histopathological slide showing chondrocyte proliferation (black arrow) seen in herniated disc material. Haematoxylin and Eosin stain. ×100 magnification



Figure 3: Photomicrograph of hyaline cartilage seen in herniated disc specimen showing chondrocytes and cartilage matrix

intensity of the alcian blue staining of the specimen is graded from 0 to 3 (0 being absent alcian blue staining and 3 being intense homogenous blue staining).

The following parameters were statistically analyzed: Sex versus type of disc herniation, sex versus instability, sex versus MCs, MCs versus HDS, Pfirmann grades versus HDS, Pfirmann grades versus alcian blue, MC versus HDS and chondrocyte proliferation versus MCs.

The Chi-square test was used for the analysis (SPSS software, Ver. 16.0, by SPSS Inc., IBM, Armonk, New York, USA). P < 0.05 was taken as significant.

RESULTS

There were 45 males and 32 females in this series. The mean age was 38 years (range 18-58 years). Forty eight



Figure 2: Granular changes, tears and clefts marked by black arrow. ×100 magnification. H and E stains



Figure 4: (a-d) Alcian blue staining of Herniated nucleus pulposus showing different grades

discs were protruded, 23 were extruded and six were sequestrated in the entire series. Sex wise distribution of the type of herniation is shown in Table 2 and Figure 6. There was no association between the type of disc herniation and sex (P = 0.198).

Sex versus instability

We looked at whether there was any association between sex and instability seen in plain radiographs 34.4% of females showed radiographic evidence of instability while only 6.7% of males showed instability. Thus, female sex has higher incidence of instability (P = 0.002, odds ratio 7.333, [1.845–29.146]) [Table 2 and Figure 7].

MC versus sex; MC versus Pfirmann Grades; MC versus histological degeneration score; MC versus chondrocyte proliferation

MC (either 1 or 2) was present in 28.2% of females and 26.7% of males. No significant association was found between sex and MCs (P = 0.917) [Table 2 and Figure 8]. About 27.27% of cases showed MC. No significant association was found between Pfirmann grades and MC. The analysis of variance was done to compare mean HDS between the three Modic groups. No statistically significant difference was seen in mean HDS of different Modic groups [Table 3 and Figure 9].

However, following findings were notable. Among cases that showed no MC, 15 out of 56 had evidence of

hyaline cartilage and in the set with MC, 16 out of 21 had the presence of hyaline cartilage. Fisher two-tailed *t*-test was done and this was found to be statistically significant (P = 0.0002). We categorized the degree of chondrocyte proliferation, low grade being Grade 0–3 and high grade being 4 and 5 and compared with the presence or absence of MCs, there was a statistically significant association (P = 0.02) [Table 4 and Figure 10]. No other parameter used in the HDS had a significant association between MCs. There was no association between the presence of CD34 and MCs, in our study (P = 0.989).

Pfirmann grades versus sex, Pfirmann grades versus histological degeneration score, Pfirmann grades versus alcian blue score

The predominant grading of Pfirmann seen in the group was Grade 3 amounting to 51.1% in males and 53.1% in females. There was no significant association between grades of Pfirmann and sex [Table 5 and Figure 11]. Again when grades of Pfirmann were compared with HDS, there was no association (P = 0.123) [Table 6 and Figure 12]. This shows that a single Pfirmann grade can comprise a wide varying degree of histological changes. The MRI grading of disc degeneration by Pfirmann does not completely reflect the pathological changes going on in the herniated discs. Pfirmann *et al.*⁷ have rightly commented in their paper that this grading system can reflect the changes in chemical composition (histologically this being



Figure 5: Immunohistochemistry showing CD34 positivity (a and b) and S100 positivity (c)



Figure 6: Bar diagram showing the sex wise distribution of various types of disc herniation



Figure 7: Bar diagram showing the relationship between sex and instability

| Sex | Туре | of herniatio | on (%) | Ins | stability (% | 6) | MC (%) | | | |
|--------------------|----------------|--------------|---------------|----------------|--------------|---------------|----------------|----------|--------------|--|
| | Protrusion | Extrusion | Sequestration | Yes | No | | 0 | 1 | 2 | |
| Female | 23 (71.9) | 6 (18.8) | 3 (9.4) | 11 (34.4) | 21 (65.6) | | 23 (71.9) | 6 (18.8) | 3 (9.4) | |
| Male | 25 (55.6) | 17 (37.8) | 3 (6.7) | 3 (6.7) | 42 (93.3) | | 33 (73.3) | 7 (15.6) | 5 (11.1) | |
| | χ^2 | Df | Р | χ^2 | df | Р | χ^2 | Df | Р | |
| Pearson Chi-square | 3.242 (<5.991) | 2 | 0.198 (>0.05) | 9.652 (>3.841) | 1 | 0.002 (<0.05) | 0.173 (<5.991) | 2 | 0.917 (>0.05 | |

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Odds Ratio=7.333 (1.845-29.146) 95% CI. MC=Modic changes

the amount of proteoglycans) rather than the alteration in micro structure.

There was a statistically significant association between the alcian blue scoring and the Pfirmann grading (P = 0.000). There is an increased association between alcian blue grades with Pfirmann 2 and 3 while alcian blue scores are found to decrease in Pfirmann 4. The graphs show a positive trend in Pfirmann 2 and 3 and a negative trend in Pfirmann 4 [Table 6 and Figure 13].

Pfirmann grades versus MC

No significant association was found between varying grades of Pfirmann and MC on analysis with Chi-square test (P = 0.416) [Table 6 and Figure 14].

DISCUSSION

Degenerated intervertebral discs show increased chondrocyte proliferation into clones as opposed to the scanty presence of discal cells in a normal disc.¹³ Discal cells are metabolically less active cells¹³ which depend on diffusion of oxygen and solutes across endplates for their nutrition.¹⁴ The changed biomechanical environment during degeneration might result in loss of apoptotic mechanism and result in cell proliferation.¹⁵ The relative proportion of acidic keratin sulfate to chondroitin sulfate increases in disc degeneration¹¹ possibly due to secretion of altered form of glycosaminoglycans by the chondrocytes. Large chain aggrecan molecules become fragmented resulting in decreased water imbibing capacity.

MCs have been suggested to be due to endplate micro fractures (type 1) and their replacement by fibro-fatty tissue (type 2). Albert and Manniche^{16,17} proposed two theories for their occurrence: An infective possibility where the endplate changes are due to anaerobic bacterial invasion through the ruptured disc and a mechanical theory where endplate changes are due to micro fractures due to compressive loads in the vertebrae. Shan et al.,¹⁸ proved that presence of MCs in adjacent endplates of herniated discs are associated with cartilaginous herniations that are less resorbed than the nucleus rich herniations, Presence of MCs in LDH may indicate a subset of patients who are less likely to benefit by conservative care. Our study also

Table 3: Frequency table MC versus HDS

| Tan | C J. I. | reque | itcy table w | IC VEIS | susi | IDO | | | | | | |
|---------------|---------|-------|--------------|---------|------|--------|-------|-------|-------|--|--|--|
| MC | n | Minii | num HDS | Maxi | mun | n HDS | Mean | HDS | SD | | | |
| 0 | 44 | | 3 | | 14 | | 8.7 | 2.502 | | | | |
| 1 | 10 | | 6 | | 13 | | 9.9 | 90 | 2.079 | | | |
| 2 | 7 | | 6 | | 12 | | 9.0 | 2.449 | | | | |
| ANOVA | | | | | | | | | | | | |
| HDS | | | Sum of sq | uares | df | Mean s | quare | F | Р | | | |
| Betw | een g | roups | 11.679 | 9 | 2 | 5.83 | 39 | 0.984 | 0.380 | | | |
| Within groups | | | 344.05 | 9 | 58 | 5.93 | 32 | | | | | |
| Total | | | 355.73 | 8 | 60 | | | | | | | |

HDS=Histologic degeneration score, SD=Standard deviation, MC=Modic changes, ANOVA= Analysis of variance

| Chondrocyte | | | MC | | | | | | |
|---------------|--------|-------------------|----------|---------------|--|--|--|--|--|
| proliferation | | No | | Yes | | | | | |
| Low | | 45 | | 11 | | | | | |
| High | | 11 | 11 | | | | | | |
| | | Chi-so | juare te | sts | | | | | |
| | | χ ² | df | Р | | | | | |
| Pearson Chi- | square | 6.026538 (>3.841) | 1 | 0.014 (<0.05) | | | | | |
| | | Risk estimate | | | | | | | |
| | Valu | 16 | 95 | 95% CI | | | | | |
| | | Low | er | Upper | | | | | |
| OR | 3.71 | 9 1.26 | 2 | 10.959 | | | | | |

| OR | 3.719 | 1.262 | 10 |
|----------|---------|-------|----|
| 00 011 1 | 0.0 5.1 | | |

OR=Odds ratio. CI=Confidence interval. MC=Modic changes

| Sex | | Pfirmann grade | | | | | | | | | | |
|--------------------|--------|----------------|-------------|---------------|--|--|--|--|--|--|--|--|
| | 2 | | 3 | | | | | | | | | |
| Female | 8 (25 | 5) | 17 (53.1) | | | | | | | | | |
| Male | 11 (24 | .4) | 23 (51.1) | 11 (24.4) | | | | | | | | |
| | | C | hi-square t | ests | | | | | | | | |
| | | χ ² | df | Р | | | | | | | | |
| Pearson Chi-square | | 0.070 (<5.991 |) 2 | 0.966 (>0.05) | | | | | | | | |
| Pearson Chi- | • | |) 2 | 0.966 | | | | | | | | |

No significant association with Chi-square test

had significantly higher proportion of hyaline cartilage and chondrocyte proliferation in discs with MCs.

Presence of blood vessels was seen in 56% of cases in outer annulus while in 20% of cases they were seen extending to inner annulus and nucleus. Uncontained disc herniations had higher presence of blood vessels which was in concurrence with Yasuma et al.¹⁹ In a healthy adult disc, only the outermost regions have sparse blood vessels.²⁰ The tear in the annulus, which precedes disc prolapse, could be acting like an angiogenic stimulator for the blood vessels.²¹

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Figure 8: Bar diagram showing the distribution of MC in both sexes



Figure 9: A diagram showing histological degeneration score distribution in various MC



Figure 10: Bar diagram showing the relationship between chondrocyte proliferation and presence of MC

Koike *et al.*,²² also identified the presence of vascular proliferation and macrophage infiltration in herniated tissue and suggested that the purpose and mechanism of angiogenesis could be different from the small number of blood vessels seen in the outer annulus during aging.

S100 positivity as a marker of Schwann cells²³ was present only in cases, which had blood vessels. Hence, the presence of blood vessels might help in neuronal migration into the



Figure 11: Bar diagram showing the distribution of Pfirmann grades in both sexes



Figure 12: Graph showing distribution of histological degeneration score in various Pfirmann grades



Figure 13: Graph showing the relation between Pfirmann grades and alcian blue scores (note the inverse association between Pfirmann grade 4 and alcian blue scores)

herniated nucleus. Studies that have used glial fibrillary acid protein, calcitonin gene related peptide and substance P staining have been able to demonstrate varying types of nerve fibers in the inner annulus and nucleus of degenerated discs.²⁴⁻²⁸

The nucleus pulposus contains water (87%),¹⁹ high molecular weight aggrecan and collagen. The annulus contains more of type 2 collagen, elastin and sparse long chain proteoglycans.^{29,30} The quantity of high

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|--|---|
|--|---|

| Pfirmann grade | HDS out of 15 | | | | | | | | | | | Alcian blue scoring | | | | MC | | | |
|--------------------|---------------|------|-------|------|-----|---|----|----|------|--------|-------|---------------------|----------|-----------|----|----|----------------|----|---------------|
| | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 1 | 2 | 3 | 4 | 0 | 1 | 2 |
| 2 | 1 | 0 | 2 | 3 | 1 | 4 | 3 | 2 | 2 | 0 | 1 | 0 | 2 | 5 | 10 | 2 | 15 | 1 | 3 |
| 3 | 0 | 2 | 2 | 4 | 4 | 4 | 4 | 6 | 11 | 1 | 0 | 2 | 5 | 17 | 15 | 3 | 29 | 7 | 4 |
| 4 | 0 | 0 | 1 | 1 | 1 | 0 | 7 | 1 | 3 | 3 | 1 | 0 | 13 | 4 | 1 | 0 | 12 | 5 | 1 |
| | | | χ² | | | c | lf | | | Ρ | | | ; | χ² | df | Ρ | χ^2 | df | Р |
| Pearson Chi-square | 29 | .814 | + (<3 | 3.92 | 24) | 2 | 2 | | 0.12 | 23 (>0 | 0.05) | | 29.408 (| (>12.592) | 6 | 0 | 3.930 (<9.488) | 4 | 0.416 (>0.05) |

Table 6 · Frequency table Pfirmann grade versus HDS alcian blue score and MC

HDS=Histologic degeneration score, MC=Modic Changes



Figure 14: Bar diagram showing distribution of Pfirmann grades and MC

molecular weight proteoglycans and water decrease centrifugally in a disc. As degeneration starts, the water and proteoglycan content of the nucleus decreases, nucleus fissures and extends into the annulus. Later the degradation of type 2 collagen results in fissuring of the lamellar layers of annulus through which nucleus is prone to prolapse. These events lead to altered mechanical properties of the IVD.^{3,4,27-30} As the discs degenerate, the differentiation between the nucleus and annulus is lost to varying degrees.

The composition of a disc herniation is clinically important. Nucleus pulposus has high proteoglycan content and only a loose collagen network, therefore when it escapes from the pressurized confines of a loaded disc it swells rapidly in tissue fluid, before shrinking again as much of its proteoglycans leach out. This probably explains the typical "resorption" of many disc herniations, leading to spontaneous resolution of symptoms. Herniated annulus also can swell, but by a lesser amount and its high collagen content leads to rapid invasion by blood vessels and inflammatory cells, leading to some tissue resorption. Less is known about the significance of hyaline cartilage within a herniation. Articular cartilage (a type of hyaline cartilage) does not swell greatly, even when physically disrupted because its collagen type 2 fibrils form a dense three-dimensional network.³¹ A lack of swelling restricts proteoglycan loss and subsequent resorption. This could possibly explain why a "hard" disc herniation containing hualine cartilage can give rise to more severe and persisting signs of nerve root compression.^{32,33}

Prior to herniation, the intervertebral disc goes through phases of degeneration.³⁴ Pfirmann grading of disc degeneration in MRI is based on the nucleo annular differentiation seen in T2 sequencing. This reflects the amount of water imbibed in the two compartments of the disc. It would be worthwhile to know whether the Pfirmann grading has a correlation with the actual proteoglycan content in the disc material. Our results show that in Pfirmann Grades 2 and 3, there is a positive correlation between glycoprotein content and Pfirmann grade. However, higher Grades 4 and 5 show an inverse correlation to the glycoprotein content. Thus, it is possible to postulate that the lack of nucleo-annular differentiation seen in advance Pfirmann grades is due to poor water content consequent to the lack of adequate glycoprotein in the degenerate discs. Pfirmann 4 and 5 have more annular content in the herniation as the amount of glycoprotein is less. Patients with more annular content and cartilaginous content could have a more protracted period of pain while a herniation with predominant nucleus pulposus content can expect to become better in a shorter period of time.³³ This postulate needs to be further studied.

Discal cells in degenerated discs secrete catabolic enzymes like metalloproteinase 3 which break down aggrecan molecules in smaller chains which are inferior in their capacity to imbibe water. In addition, it is postulated that increase in metalloproteinase 3 can positively regulate metalloproteinase 1, which are responsible for lysis of collagen fibers.35-37

The histological changes in the degenerative disc are well known and have been extensively described in the literature. However, a well-validated histological grading system for IVD degeneration is currently lacking. Since many translational studies are using histological changes as the primary outcome variable, a comprehensive and validated histological grading for IVD degeneration is essential to the field of IVD research. In IVD regeneration research, we need to look into the histology of the disc degeneration to bring in clues that might help in their regeneration.

Again, female sex has a higher propensity for instability as shown in our series. It is also an established fact that degenerative spondylolisthesis is much more common in females than males. The role of sex hormones in discal degeneration needs to be studied further.

CONCLUSION

Lumbar disc herniation comprises a spectrum of histomorphological changes in the disc tissue. The herniated disc contains varying amounts of nucleus, annulus and hyaline cartilage. Present MRI grading of disc degeneration correlates only with the quantitative change in glycoproteins in the disc tissue and does not show the microstructural alterations in a degenerated disc. Presence of modic changes in the endplates adjacent to the herniated disc level suggests significant presence of hyaline cartilage in the prolapsed disc tissue.

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Conflicts of interest

There are no conflicts of interest.

REFERENCES

- 1. Rajasekaran S, Babu JN, Arun R, Armstrong BR, Shetty AP, Murugan S. ISSLS prize winner: A study of diffusion in human lumbar discs: A serial magnetic resonance imaging study documenting the influence of the endplate on diffusion in normal and degenerate discs. Spine (Phila Pa 1976) 2004;29:2654-67.
- 2. Adams MA, Roughley PJ. What is intervertebral disc degeneration, and what causes it? Spine (Phila Pa 1976) 2006;31:2151-61.
- 3. Antoniou J, Goudsouzian NM, Heathfield TF, Winterbottom N, Steffen T, Poole AR, *et al.* The human lumbar endplate. Evidence of changes in biosynthesis and denaturation of the extracellular matrix with growth, maturation, aging, and degeneration. Spine (Phila Pa 1976) 1996;21:1153-61.
- 4. Gruber HE, Hanley EN Jr. Recent advances in disc cell biology. Spine (Phila Pa 1976) 2003;28:186-93.
- Haughton V. Medical imaging of intervertebral disc degeneration: Current status of imaging. Spine (Phila Pa 1976) 2004;29:2751-6.
- 6. Modic MT, Steinberg PM, Ross JS, Masaryk TJ, Carter JR. Degenerative disk disease: Assessment of changes in vertebral body marrow with MR imaging. Radiology 1988;166 (1 Pt 1):193-9.
- 7. Pfirrmann CW, Metzdorf A, Zanetti M, Hodler J, Boos N. Magnetic resonance classification of lumbar intervertebral disc degeneration. Spine (Phila Pa 1976) 2001;26:1873-8.
- 8. Boos N, Weissbach S, Rohrbach H, Weiler C, Spratt KF, Nerlich AG. Classification of age-related changes in lumbar intervertebral discs: 2002 Volvo Award in basic science. Spine (Phila Pa 1976) 2002;27:2631-44.

- 9. Weiler C, Lopez-Ramos M, Mayer HM, Korge A, Siepe CJ, Wuertz K, *et al.* Histological analysis of surgical lumbar intervertebral disc tissue provides evidence for an association between disc degeneration and increased body mass index. BMC Res Notes 2011;4:497.
- Boden SD, Wiesel SW. Lumbosacral segmental motion in normal individuals. Have we been measuring instability properly? Spine (Phila Pa 1976) 1990;15:571-6.
- 11. Taylor JR, Scott JE, Cribb AM, Bosworth TR. Human intervertebral disc acid glycosaminoglycans. J Anat 1992;180 (Pt 1):137-41.
- 12. Johnson WE, Evans H, Menage J, Eisenstein SM, El Haj A, Roberts S. Immunohistochemical detection of Schwann cells in innervated and vascularized human intervertebral discs. Spine (Phila Pa 1976) 2001;26:2550-7.
- 13. Trout JJ, Buckwalter JA, Moore KC. Ultrastructure of the human intervertebral disc: II. Cells of the nucleus pulposus. Anat Rec 1982;204:307-14.
- 14. Holm SH. Nutrition of the intervertebral disc. In: Weinstein JN, Wiesel SW, editors. The Lumbar Spine. Philadelphia, PA: W.B. Saunders; 1990. p. 244-60.
- 15. Ariga K, Miyamoto S, Nakase T, Okuda S, Meng W, Yonenobu K, *et al.* The relationship between apoptosis of endplate chondrocytes and aging and degeneration of the intervertebral disc. Spine (Phila Pa 1976) 2001;26:2414-20.
- 16. Albert HB, Manniche C. Modic changes following lumbar disc herniation. Eur Spine J 2007;16:977-82.
- 17. Albert HB, Kjaer P, Jensen TS, Sorensen JS, Bendix T, Manniche C. Modic changes, possible causes and relation to low back pain. Med Hypotheses 2008;70:361-8.
- Shan Z, Fan S, Xie Q, Suyou L, Liu J, Wang C, *et al.* Spontaneous resorption of lumbar disc herniation is less likely when Modic changes are present. Spine (Phila Pa 1976) 2014;39:736-44.
- 19. Yasuma T, Arai K, Yamauchi Y. The histology of lumbar intervertebral disc herniation. The significance of small blood vessels in the extruded tissue. Spine (Phila Pa 1976) 1993;18:1761-5.
- 20. Urban JP, Roberts S. Degeneration of the intervertebral disc. Arthritis Res Ther 2003;5:120-30.
- 21. Crock HV, Goldwasser M, Yoshizawa H. Vascular anatomy related to the intervertebral disc. In: Ghosh P, editor. Biology of the Intervertebral Disc. Boca Raton: CRC Press; 1991. p. 109-33.
- 22. Koike Y, Uzuki M, Kokubun S, Sawai T. Angiogenesis and inflammatory cell infiltration in lumbar disc herniation. Spine (Phila Pa 1976) 2003;28:1928-33.
- 23. Freemont AJ, Peacock TE, Goupille P, Hoyland JA, O'Brien J, Jayson MI. Nerve ingrowth into diseased intervertebral disc in chronic back pain. Lancet 1997;350:178-81.
- 24. García-Cosamalón J, del Valle ME, Calavia MG, García-Suárez O, López-Muñiz A, Otero J, *et al.* Intervertebral disc, sensory nerves and neurotrophins: Who is who in discogenic pain? J Anat 2010;217:1-15.
- 25. Palmgren T, Grönblad M, Virri J, Kääpä E, Karaharju E. An immunohistochemical study of nerve structures in the anulus fibrosus of human normal lumbar intervertebral discs. Spine (Phila Pa 1976) 1999;24:2075-9.
- 26. Fagan A, Moore R, Vernon Roberts B, Blumbergs P, Fraser R. ISSLS prize winner: The innervation of the intervertebral disc: A quantitative analysis. Spine (Phila Pa 1976) 2003;28:2570-6.
- 27. Dolan P, Adams MA, Hutton WC. The short-term effects of

chymopapain on intervertebral discs. J Bone Joint Surg Br 1987;69:422-8.

- 28. Freemont AJ. The cellular pathobiology of the degenerate intervertebral disc and discogenic back pain. Rheumatology (Oxford) 2009;48:5-10.
- 29. Buckwalter JA. Aging and degeneration of the human intervertebral disc. Spine (Phila Pa 1976) 1995;20:1307-14.
- 30. Freemont TJ, LeMaitre C, Watkins A, Hoyland JA. Degeneration of intervertebral discs: Current understanding of cellular and molecular events, and implications for novel therapies. Expert Rev Mol Med 2001;2001:1-10.
- 31. Summers GC, Merrill A, Sharif M, Adams MA. Swelling of articular cartilage depends on the integrity of adjacent cartilage and bone. Biorheology 2008;45:365-74.
- 32. Autio RA, Karppinen J, Niinimäki J, Ojala R, Kurunlahti M, Haapea M, *et al.* Determinants of spontaneous resorption of intervertebral disc herniations. Spine (Phila Pa 1976) 2006;31:1247-52.

- 33. Willburger RE, Ehiosun UK, Kuhnen C, Krämer J, Schmid G. Clinical symptoms in lumbar disc herniations and their correlation to the histological composition of the extruded disc material. Spine (Phila Pa 1976) 2004;29:1655-61.
- 34. Lama P, Le Maitre CL, Dolan P, Tarlton JF, Harding IJ, Adams MA. Do intervertebral discs degenerate before they herniate, or after? Bone Joint J 2013;95-B: 1127-33.
- 35. Goupille P, Jayson MI, Valat JP, Freemont AJ. Matrix metalloproteinases: The clue to intervertebral disc degeneration? Spine (Phila Pa 1976) 1998;23:1612-26.
- 36. Kanemoto M, Hukuda S, Komiya Y, Katsuura A, Nishioka J. Immunohistochemical study of matrix metalloproteinase-3 and tissue inhibitor of metalloproteinase-1 human intervertebral discs. Spine (Phila Pa 1976) 1996;21:1-8.
- 37. Roberts S, Caterson B, Menage J, Evans EH, Jaffray DC, Eisenstein SM. Matrix metalloproteinases and aggrecanase: Their role in disorders of the human intervertebral disc. Spine (Phila Pa 1976) 2000;25:3005-13.

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