

## Status of *Anaplasma* spp. infection in domestic ruminants from Iran: A systematic review with meta-analysis

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### ABSTRACT

*Anaplasma* species are tick-borne pathogens that are obligatory intracellular of ruminants and other mammals. In this investigation, we systematically reviewed the distribution of anaplasmosis among domestic ruminants in Iran. Five and four English and Persian databases were studied, respectively, based on keywords and throughout 17 years (2001–2017). Thirty-eight articles were included in this systematic review and meta-analysis. Totally, 5093 cattle, 1958 sheep, and 1232 goats corresponding to prevalence of *Anaplasma* infection from different areas of Iran were examined. The total prevalence of *Anaplasma* infection was estimated to be 34% (95% CI 27%, 41%) in domestic ruminants. Based on our data, Khozestan (54%) and Khorasan Razavi (46%) provinces were the most prevalent areas in Iran and Kerman (3%) and Hamedan (1%) provinces are the lowest. The highest prevalence of *Anaplasma* spp. infection was belonged to *A. ovis* (44%) and the lowest to *A. phagocytophilum* (1%) with a significant difference among them ( $p < .001$ ). In addition, the most common diagnostic tests were PCR (54%), microscopy (35%) and ELISA (7%) assays. The high prevalence of ovine and bovine anaplasmosis in Iran, confirms the stability situations of animal anaplasmosis in the studied regions particularly northeastern and southwestern parts of the country. Our data offer valuable and encouraging information as regards the current situation of anaplasmosis in domestic livestock in Iran, which might be useful for active and passive surveillance and preventing plans.

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## 1. Introduction

*Anaplasma* belongs to the family of Anaplasmataceae (Order Rickettsiales), Gram-negative and infects red blood cells that vertebrates are its main hosts and reservoirs. Anaplasmosis is an important bacterial infection in human and animal health (Bah, 2016). It is mainly transmitted by a number of species of hard *Ixodes* ticks. According to the many previous reports, *Ixodes*, *Dermacentor*, *Rhipicephalus* and *Amblyomma* genera are the main species that transmit the *Anaplasma* spp. in different districts of the world (Rymaszewska and Grenda, 2008). To date, six *Anaplasma* spp. are recognized in domestic animals (Rymaszewska and Grenda, 2008). Five species of them include, *A. marginale*, *A. centrale*, *A. phagocytophilum*, *A. bovis* and *A. ovis* were identified in Iranian ruminants (Aktas et al., 2011). Anaplasmosis, causes important economic losses to animal breeders. Clinical manifestation such as anemia, fever, weight loss, breathlessness, jaundice, abortion and finally death are common in ruminants with anaplasmosis infections (Kaewmongkol et al., 2017). Diagnosis of anaplasmosis in animals is often based on microscopically examinations of thin blood smears with Giemsa staining. Also, several conventional diagnostic tools vary from low to high sensitivity, such as Enzyme-Linked Immunosorbent Assay (ELISA) and Polymerase Chain Reaction (PCR) techniques, were used for determining the prevalence and differentiating the *Anaplasma* spp. (Khaki et al., 2015; Rodriguez-Morales et al., 2019).

The current study is designed to review the studies that have been conducted on the prevalence of *Anaplasma* infections in domesticated ruminants from different parts of Iran. Despite different studies about *Anaplasma* spp. prevalence among domestic animals in Iran, there is not any comprehensive information. According to the effect of anaplasmosis on the economy and public health, more epidemiological studies are recommended. Based on our research, there is no documented review about the prevalence of anaplasmosis among livestock in Iran.

## 2. Materials and methods

### 2.1. Searching approach

Nine most valuable databases in medicine and veterinary sciences in English and Persian languages, including, Science Direct, Scopus, Web of Science, PubMed, Medical Subject Headings (MeSH/mh), Google Scholar, Magiran, Barakat (formerly Iranmedex), Elm net, and Scientific Information Database (SID), were selected between 2001 and 2017. To explore the articles, some key words such as: *Anaplasma* spp., anaplasmosis, *Anaplasma phagocytophilum*, *Anaplasma marginale*, *Anaplasma ovis*, *Anaplasma bovis*, *Anaplasma centrale*, livestock, domestic herbivores, cattle, sheep and goat and "Iran" alone or in combination were used. To avoid the risk of selection bias in this study, the inclusion criteria were clearly classified and studied. The stages of the study plan are briefly explained in Fig. 1.

### 2.2. Paper selection

All studies were independently screened and eligibility was determined by two reviewers (MH and MS) with the agreement between reviewers of 94% using Kappa index and a third opinion (MF) resolved the disagreements.

### 2.3. Statistical analysis

The quality of meta-analysis was evaluated with STROBE scale. The score under 7.75 considered poor quality, between 7.76 and 15.5 low, between 15.6 and 23.5 moderate and more than 23.6 high quality (Von Elm et al., 2007). The mean of scores for the STROBE scale was obtained 19.43 which showed that the quality of these studies was moderate to high. The prevalence of *Anaplasma* spp. infection in each study was collected and according to binomial distribution, standard error ( $SE = \sqrt{\frac{p \cdot q}{n}}$ ) for each study was calculated and the inverse of SE for each study considered as the weight of that study. The effect size (ES) for each study and pooled outcome revealed as a forest plot [reported as ES with a 95% confidence interval (95%CI)].

Cochran's heterogeneity statistics based on chi-square test Q-test ( $p < .1$  as heterogeneities) and the I-squared statistic were used to evaluate the percentage of variation through studies with the value of 25% (low), 50% (moderate), and 75% (high) of heterogeneity. The mean of scores for the STROBE scale was obtained 19.43 which showed that the quality of these studies was moderate to high.

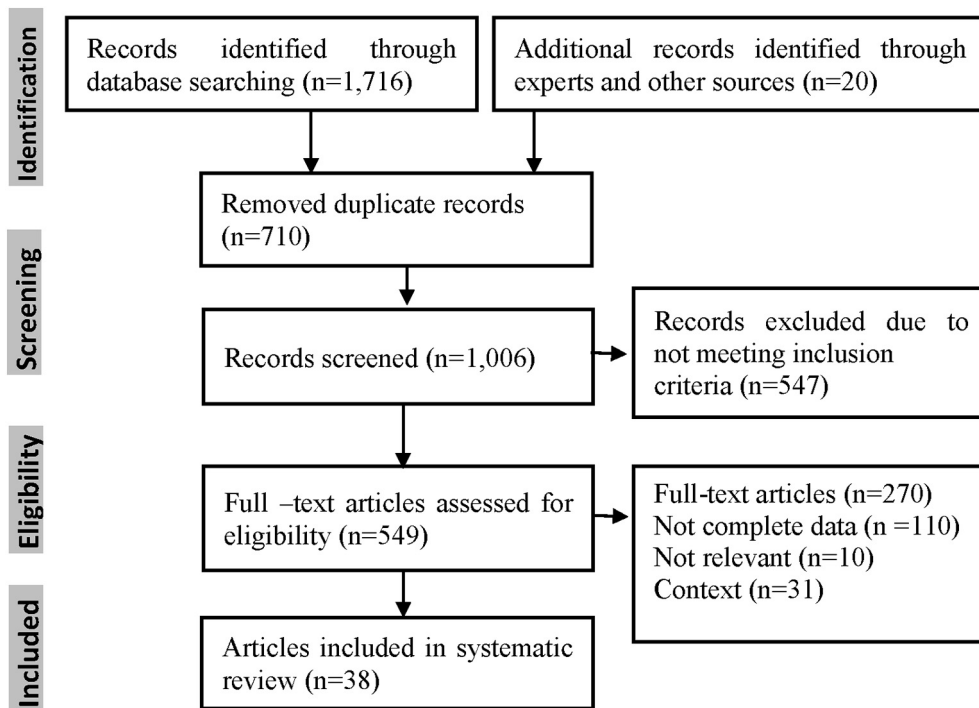


Fig. 1. PRISMA flowchart describing the study design process.

At present heterogeneity, random effects model (DerSimonian and Kacker, 2007) and otherwise applied fixed effect model (Mantel Haenszel) were used to compute overall effect size. Subgroup analyses were performed to investigate potential sources of heterogeneity from different types of animal, *Anaplasma* spp., laboratory methods and study area. Egger's test was used to evaluate publication bias. All statistical analyses were done with the Statistical Software Package (Stata) version 11.1.

### 3. Results

The process of study selection is shown in Fig. 1. Among all databases; a total of 38 articles published during 17 years (2001–2017) were selected to be included in this systematic review and meta-analysis (Table 1). All articles were cross-sectionally designated and evaluated the prevalence of *Anaplasma* infection in domestic herbivorous including cattle, sheep and goat in different districts of Iran. Totally, 5093 cattle, 1958 sheep, and 1232 goats were examined. The Overall prevalence of *Anaplasma* infection based on a random effect meta-analysis was estimated to be 34% (95% CI, 27–41%,  $I^2 = 99.24\%$ ,  $p < .001$ ), which indicated a substantial heterogeneity among studies. The results of subgroup analysis indicated the prevalence of *Anaplasma* infection among sheep 39.0% (95% CI, 20.0–58.0%), cattle 24.0% (95% CI, 16.0–31.0%) and goats 39.0% (95% CI, 21.0–57.0%), which the differences was not statistically significant among them ( $p = .14$ ) (Table 2, Fig. 2).

The maximum prevalence of *Anaplasma* spp. was in Ahvaz (54%, 95% CI: 36%–72%) and the minimum occurred in Hamedan (1%, 95% CI, 0.1%–72%) (Table 2, Fig. 3). Additionally, the most common diagnostic tests were PCR (54%), microscopy (35%) and ELISA (7%) assays (Fig. 4). In total, the highest prevalence of *Anaplasma* spp. infection was belonged to *A. ovis* (44%) and the lowest to *A. phagocytophilum* (1%) with a significant difference among them ( $p < .001$ ) (Table 2, Fig. 5). For assessing publication bias of studies, we used Egger's test. The results showed that there was no essentially publication bias in included studies of this meta-analysis ( $t = -46$ ,  $p = .651$ ).

### 4. Discussion

Anaplasmosis is common among farm animals and, in Iran, overall prevalence of infection is 34%, and the prevalence in sheep, cattle and goat were 39, 24 and 39%, respectively (Fig. 2). In comparison to the other studies in neighboring countries, anaplasmosis in Iran is more prevalent than that found in Russia, Pakistan, Turkey and Iraq (Rar et al., 2010; Atif et al., 2012; Aktas et al., 2011; Ameen et al., 2012). The diseases is widely distributed throughout the world including tropical and sub-tropical areas of Asia, South, Central and North America, Europe, Africa and Australia with a prevalence ranging of 1 to 100% (Noaman et al., 2009; Sainz et al., 2015; Pokorn et al., 2016).

Around Iran; Middle East countries, including Jordan, Egypt, United Arab Emirates, Iraq, Qatar, Cyprus, and Israel are prevalent areas for *Anaplasma* spp. infections (Ameen et al., 2012; Jafarbakloo et al., 2014; Razzaq et al., 2015; Kaewmongkol et al., 2017).

**Table 1**  
Characteristics of studies included in systematic review and meta-analysis.

Author	Year of publication	Type of animal	No. of examined	No. of positive	Anaplasma spp.	Laboratory method	Place
(Noaman and Kachouei, 2001)	2001	cattle	3269	546	<i>A.marginale</i>	Microscopic	Isfahan
(Razmi et al., 2006)	2006	cattle	160	31	<i>A.marginale</i>	Microscopic	Mashhad
	2006	sheep	391	314	<i>A.ovis</i>	Microscopic	Mashhad
	2006	goat	385	150	<i>A.ovis</i>	Microscopic	Mashhad
(Noaman and Shayan, 2009, Noaman et al., 2009)	2009	cattle	150	2	<i>A. phagocytophilum</i>	Nested-PCR	Isfahan
	2009	cattle	150	58	<i>A.marginale</i>	PCR-RFLP	Isfahan
	2009	cattle	150	75	<i>A.marginale</i>	Microscopic	Isfahan
(Ahmadi-Hamedani et al., 2009)	2009	goat	193	123	<i>A.ovis</i>	PCR-RFLP	Gonbad& Mashhad
	2009	goat	193	43	<i>Anaplasma spp.</i>	Microscopic	Gonbad& Mashhad
(Noaman and Shayan, 2010a, 2010b, 2010c)	2010	cattle	150	58	<i>A.marginale</i>	PCR-RFLP	Isfahan
(Noaman and Shayan, 2010a, 2010b, 2010c)	2010	cattle	150	91	<i>Anaplasma spp.</i>	Microscopic	Isfahan
(Noaman and Shayan, 2010a, 2010b, 2010c)	2010	cattle	150	4	<i>A.ovis</i>	Nested-PCR	Isfahan
(Ahmadi-hamedani et al., 2012)	2011	goat	193	123	<i>A.ovis</i>	PCR	Gonbad& Mashhad
	2011	goat	193	43	<i>Anaplasma spp.</i>	Microscopic	Gonbad& Mashhad
(Salehzadeh et al., 2011)	2011	cattle	200	6	<i>A.marginale</i>	Microscopic	kerman
(Jalali et al., 2013; Jalali et al., 2016)	2012	sheep	119	40	<i>Anaplasma spp.</i>	Microscopic	Ahvaz
	2012	sheep	119	104	<i>Anaplasma spp.</i>	PCR	Ahvaz
	2012	sheep	104	52	<i>A.marginale</i>	PCR-RFLP	Ahvaz
Noaman et al.	2012	sheep	150	50	<i>A.ovis</i>	PCR-RFLP	Isfahan
Noaman et al.	2013	cattle	150	75	<i>A.marginale</i>	Microscopic	Isfahan
Noaman et al.	2013	cattle	150	10	<i>A.marginale</i>	PCR-RFLP	Isfahan
Noaman et al.	2013	sheep	150	50	<i>A.ovis</i>	Microscopic	Isfahan
Noaman et al.	2013	sheep	150	10	<i>A.ovis</i>	PCR-RFLP	Isfahan
Ahmadi-Hamedani et al.	2013	goat	84	47	<i>A.ovis</i>	PCR	Gonbad& Mashhad
Hosseini-Vasoukolaei et al.	2014	cattle	9	2	<i>Anaplasma spp.</i>	PCR	Mazandaran
Hosseini-Vasoukolaei et al.	2014	sheep	65	28	<i>Anaplasma spp.</i>	PCR	Mazandaran
Hosseini-Vasoukolaei et al.	2014	goat	4	1	<i>Anaplasma spp.</i>	PCR	Mazandaran
Khaki et al.	2015	sheep	109	35	<i>Anaplasma spp.</i>	Microscopic	Ahvaz
Khaki et al.	2015	sheep	109	94	<i>A.ovis</i>	PCR-RFLP	Ahvaz
Khaki et al.	2015	sheep	109	50	<i>A.ovis</i>	PCR-RFLP	Ahvaz
Khezri et al.	2015	cattle	105	8	<i>Anaplasma spp.</i>	ELISA	Kurdistan
Khezri et al.	2015	sheep	77	5	<i>Anaplasma spp.</i>	ELISA	Kurdistan
Noaman et al.	2016	cattle	100	0	<i>Anaplasma spp.</i>	Nested-PCR	West Azarbaijan
Noaman et al.	2016	sheep	100	5	<i>A.ovis</i>	Nested-PCR	West Azarbaijan
Jalali et al.	2016	goat	104	30	<i>A.ovis</i>	Microscopic	Ahvaz
Jalali et al.	2016	goat	104	68	<i>A.ovis</i>	PCR-RFLP	Ahvaz
Yousefi et al.	2017	sheep	206	1	<i>A. phagocytophilum</i>	Nested-PCR	Hamedan
Yousefi et al.	2017	goat	164	3	<i>A. phagocytophilum</i>	Nested-PCR	Hamedan

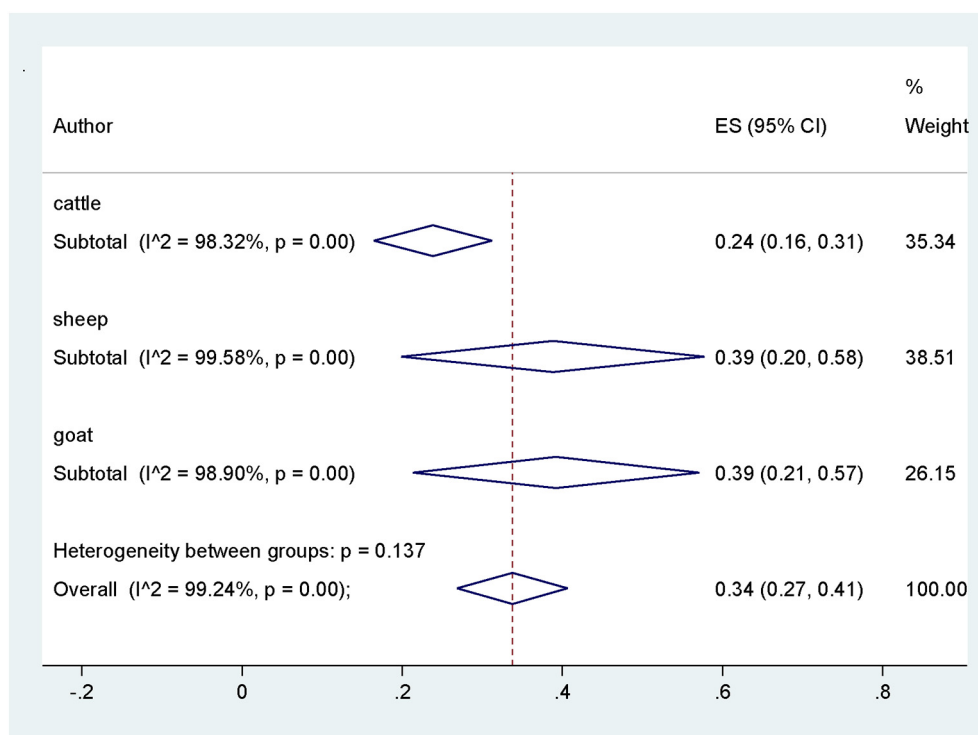
The prevalence of *Anaplasma* spp. infection among herbivores in different years in the same region of Russia, the northern Iranian neighbor, varies from 1% to 0.5% (Rar et al., 2010). In a similar study, in Pakistan, the southeast Iranian neighbor, 1050 blood samples from livestock farms were microscopically examined and revealed that 21.14% of samples were positive for blood parasites that *Anaplasma* with the prevalence of 5.81%, was the most prevalent hemoparasite (Atif et al., 2012).

Different countries from Africa, including Uganda, Kenya, Morocco, Ghana and Tanzania reported bovine anaplasmosis outbreaks during 2003 to 2019 (Bah, 2016; Sisson et al., 2017; Byaruhanga et al., 2018. Ringo et al., 2018). In a study conducted by Ait Lbacha et al., 71% (303/422) of small ruminants in Morocco were infected by *Anaplasma* spp. using PCR technique (Ait Lbacha et al., 2017).

Anaplasmosis in countries of Latin America and in Caribbean Islands exception of desert areas and certain mountain ranges is enzootic (Rodriguez-Morales et al., 2019). Human granulocytic anaplasmosis is uncommon in Europe, but it's the most prevalent tick-borne infection in animals (Sainz et al., 2015; Pokorn et al., 2016). It's reported that the prevalences of the *Ixodes ricinus* as vector have been increasing in most of the states of the USA (Ringo et al., 2018). In general, types of grazing system in the

**Table 2**Subgroup meta-analysis of the prevalence of *Anaplasma* spp. according to the type of animal, *Anaplasma* spp., detection method and place.

Characteristics	Factors	N	EF (95%CI)	I-square (%)	P-value
Type of animals	Cattle	12	0.24(0.16, 0.31)	98.0	0.14
	Sheep	14	0.39(0.20, 0.58)	99.0	
	Goat	10	0.39(0.21, 0.57)	99.0	
<i>Anaplasma</i> spp.	<i>marginale</i>	9	0.30(0.20, 0.39)	97.9	P < .001
	<i>Ovis</i>	14	0.44(0.26, 0.61)	99.3	
	<i>phagocytophilum</i>	3	0.01(0.0, 0.02)	93.2	
	<i>Anaplasma</i> spp.	11	0.33(0.16, 0.51)	98.3	
Method	Microscopy	14	0.35(0.23, 0.47)	99.1	P < .001
	Nested-PCR	5	0.02(0.001, 0.03)	41.0	
	PCR-RFLP	10	0.43(0.25, 0.62)	98.7	
	PCR	6	0.54(0.36, 0.72)	93.63	
	ELISA	2	0.07(0.03, 0.11)	99.2	
Place	Isfahan	12	0.28(0.19, 0.36)	98.2	P < .001
	Mashhad	3	0.46(0.10, 0.83)	97.6	
	Gonbad and Mashhad	5	0.45(0.25, 0.66)	97.6	
	Kerman	2	0.03(0.01, 0.06)	90.3	
	Ahvaz	8	0.54(0.36, 0.72)	97.5	
	West Azerbaijan	2	0.05(0.02, 0.11)	91.8	
	Mazandaran	3	0.37(0.24, 0.50)	84.2	
	Kurdistan	2	0.07(0.03, 0.11)	89.7	
	Hamedan	2	0.01(0.001, 0.02)	98.7	

**Fig. 2.** Pooled prevalence of *Anaplasma* spp. infection according to the type of animals.

husbandry of the livestock, climate conditions of the area, flock size, strain of *Anaplasma*, abundance of the tick as vector are some variables that significantly affects on the prevalence of infection (Rosso et al., 2017). Despite the importance of the disease in the livestock industry, there are still several areas in Iran, which none study has been performed on the *Anaplasma* spp. infection among livestock and its vectors in those areas.

Anaplasmosis as one of the most important endemic disease in many regions of Iran and the prevalence of infection is seasonally different which, increasing in spring and summer in the northern (Mazandaran, Mashhad, Gonabad and West Azerbaijan), western (Hamedan, Ahvaz and Kurdistan) and central (Isfahan, Kerman) parts of the country (Razmi et al., 2006; Ahmadi-Hamedani et al., 2009; Ahmadi-hamedani et al., 2012; Ahmadi-hamedani et al., 2014; Hosseini-Vasoukolaei et al., 2014;

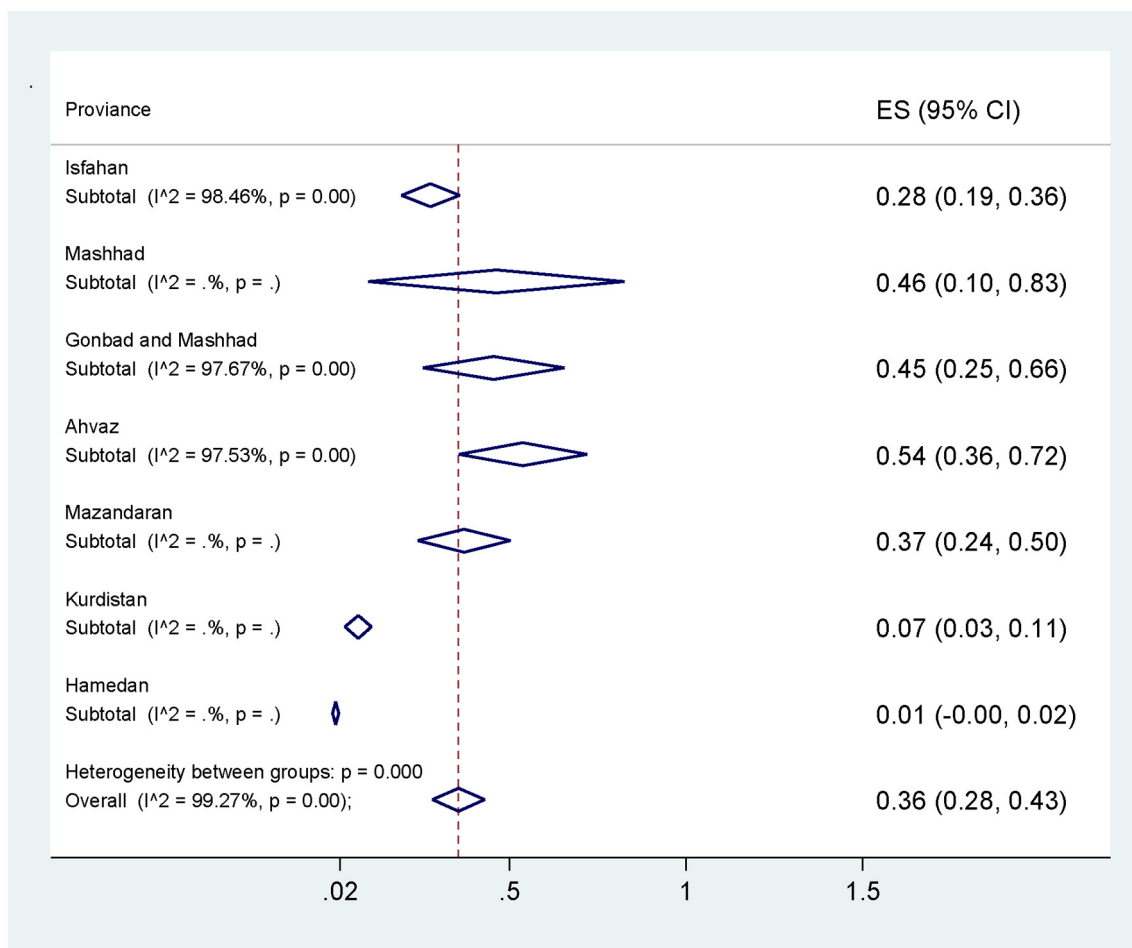


Fig. 3. Pooled prevalence of *Anaplasma* spp. according to the place of study.

Noaman and Bastani, 2016), (Jalali et al., 2013; Khaki et al., 2015; Jalali et al., 2016; Yousefi et al., 2017), (Noaman and Kachouei, 2001; Noaman and Shayan, 2009; Noaman et al., 2009; Noaman and Shayan, 2010a, 2010b, 2010c; Salehzadeh et al., 2011; Noaman, 2012; Noaman, 2013) (see Table 1). In Iran, because of the biodiversity of tick, variety of ecological and climate situations, anaplasmosis is highly prevalent. The domestic industry has a long history in Iran and cattle, sheep and goat are the most important livestock. Also, some livestock and products are exported to different parts of the world (Jalali et al., 2013; Razaq et al., 2015). Ticks have key roles in the transmission of many infectious diseases such as viral, parasitic and also bacterial diseases. Ticks are usually present in tropical and also subtropical regions. Nowadays, because of the alteration of the land use patterns, and changes in climate; the rate of tick-borne diseases have been significantly increased and spreading to new zones (Rodriguez-Morales et al., 2019). Based on many published reports, *Ixodes*, *Dermacentor*, *Rhipicephalus* and *Amblyomma* are the most important vectors for *Anaplasma* spp. in different districts of Iran (Noaman, 2012). Anaplasmosis can cause respiratory distress, enlarge the prescapular lymph nodes reduction of milk production, body weight, abortion and maybe death (Stuen et al., 2003).

In the current study, the pooled prevalence rate of infection is estimated to be 34% in Iran. Additionally, the prevalence rate of different geographical zones demonstrated that there are five zones in Iran with different prevalence rates: 54% in Ahvaz (Khozestan Province) (Jalali et al., 2013; Khaki et al., 2015), 46% in Mashhad (Khorasan Razavi Province) (Razmi et al., 2006), 45% in Gonbad and Mashhad (Khorasan Razavi Province) (Ahmadi-hamedani et al., 2012), 37% in Mazandaran Province (Hosseini-Vasoukolaei et al., 2014), 28% in Isfahan Province (Noaman and Shayan, 2010a, 2010b, 2010c) and 3% and 1% in Kerman and Hamedan provinces (Salehzadeh et al., 2011) (Fig. 3). Accordingly, the highest mean of prevalence rate of infection was in southwestern and northeastern parts of Iran (Fig. 3). In Turkey, Iranian western neighbor, *Anaplasma* spp. infection was reported that 9% (35/389) of bovine were positive using PCR method in 2011 (Aktas et al., 2011).

The western provinces of Iran such as West Azarbaijan and Kurdistan have similar weather, the height and environmental conditions to Turkey. Also in a similar study in Iraq, from 184 cattle, 44 sheep, 59 goats and 70 ibex, 4, 2, 3 and 1 cases were positive for anaplasmosis, respectively (Ameen et al., 2012). Our findings are in agreement with different studies on sheep and goats in



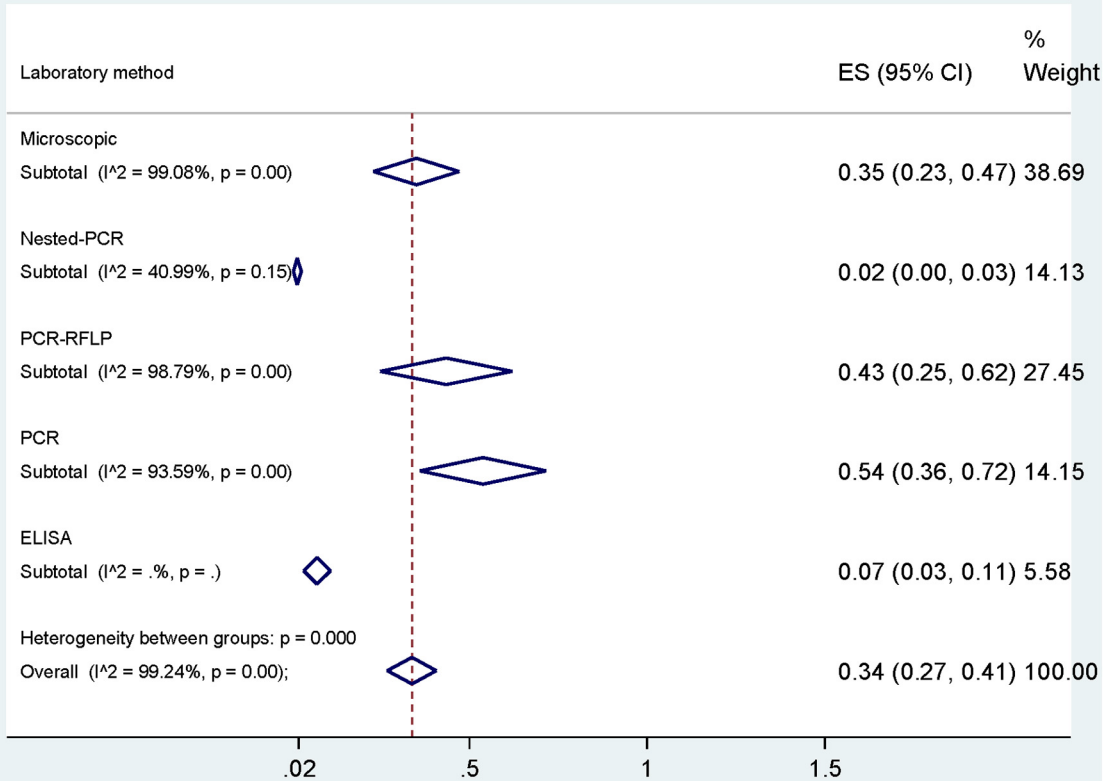


Fig. 4. Pooled prevalence of *Anaplasma* spp. according to the laboratory methods.

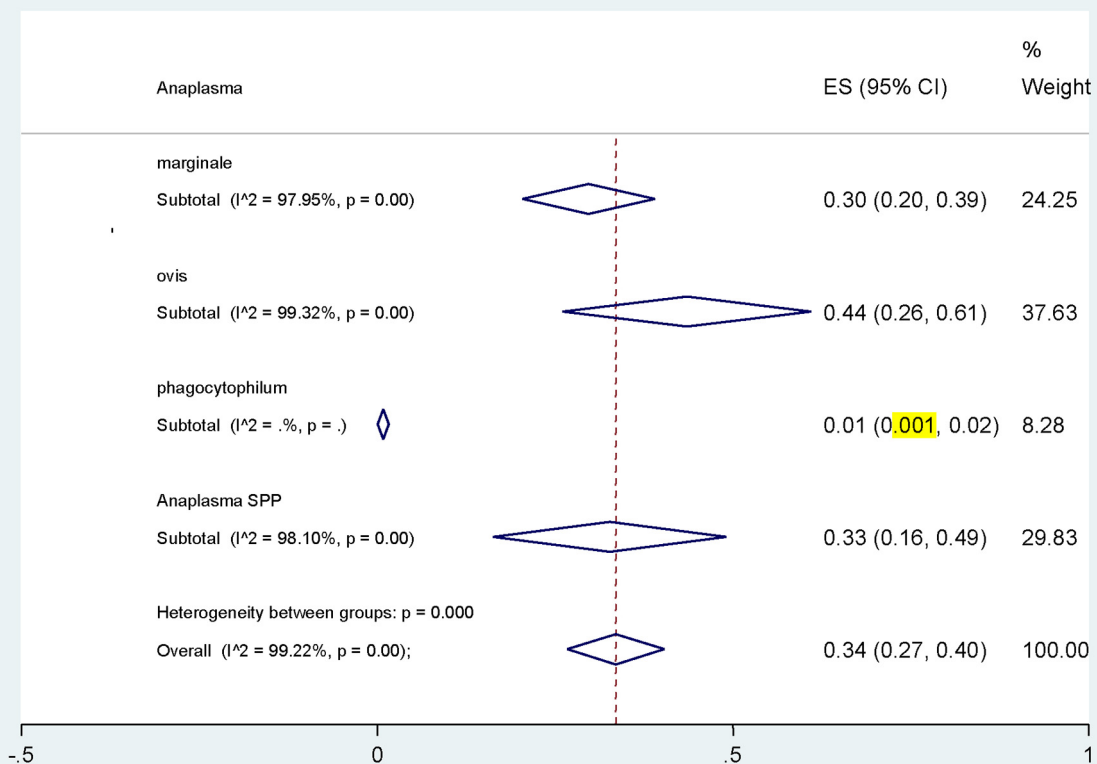


Fig. 5. Pooled prevalence of *Anaplasma* spp. according to the species of *Anaplasma*.

Pakistan in 2014 which showed 9 positive samples of 210 horses, with PCR-RFLP method, and the prevalence rate of 16% is recorded by PCR method (Razzaq et al., 2015). The prevalence of anaplasmosis in Pakistan, the southeast Iranian neighbor, is almost close to the prevalence of Khorasan Razavi, eastern Iran. In addition, in Borderline of Iran-Afghanistan, in June 2013 to May 2014, molecular studies were done on 53 samples, which the *Anaplasma*'s DNA was found in 14 samples (26.4%) out of the 53 specimens. It is a concerning prevalence of anaplasmosis among animals in Afghanistan, as the eastern neighbor of Iran (Jafarbakloo et al., 2014).

In addition, positive rate of infection in studies using PCR-RFLP method (43%) was significantly higher than other laboratory tests ( $p < .001$ ) (Fig. 4 and Table. 2). The highest infection was in Ahvaz (54%) (Jalali et al., 2013; Khaki et al., 2015; Jalali et al., 2016), and Mashhad (46%) (Razmi et al., 2006) and the lowest rate was in Hamedan (1%) (Yousefi et al., 2017) and Kerman (3%) (Salehzadeh et al., 2011), with a significant difference between them (Fig. 3 and Table. 2). The most prevalence of infection among *Anaplasma* spp. was belonged to *A. ovis* with 44% and *A. marginale* with 30% of infection rates and the lowest prevalence to *A. phagocytophilum* with 1% with significant differences among them (Fig. 4 and Table. 2). One of the most important causes of evaluation of the *Anaplasma* spp. infection is the pathogenicity of this parasite to humans and the likelihood of its transmission from animal to human. *A. phagocytophilum* is one of the most pathogenic species that is seriously posing a risk for humans, particularly pregnant women (Dhand et al., 2007).

## 5. Conclusion

The high occurrence of ovine and bovine anaplasmosis in Iran, confirms the stability situations of animal anaplasmosis in the studied regions, particularly northeastern and southwestern provinces of the country and may be a warning for animal welfare and health. In brief, our data offer valuable and encouraging information as regards the current situation of anaplasmosis in domestic livestock in Iran, which might be useful for active and passive surveillance and preventing plans. Further investigation and monitoring will be needed to expand the surveillance and control policies, such as vaccination and improvement the traditional diagnostic tools and assessment the pesticide resistance in ticks to reduce the mortality and morbidity of anaplasmosis among livestock and consequently decrease the risk of outbreaks and economic failure and public health hazard in Iran.

## Declaration of Competing Interest

All authors declare that they have no conflicts of interest.

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## References

- Ahmadi-Hamedani, M., Khaki, Z., Rahbari, S., Kazemi, B., Bandehpour, M., 2009. Molecular identification of anaplasmosis in goats using a new PCR-RFLP method. *Iran. J. Vet. Res.* 10 (4), 367–372.
- Ahmadi-hamedani, M., Khaki, Z., Rahbari, S., Ahmadi-hamedani, M.A., 2012. Hematological profiles of goats naturally infected with *Anaplasma ovis* in north and North-east Iran. *Comp. Clin. Pathol.* 21 (6), 1179–1182.
- Ahmadi-hamedani, M., Ahmadi-hamedani, M., Fathi, E., Sani, R.N., 2014. Comparison of selected biochemical parameters between naturally infected and non-infected goats with *Anaplasma ovis*. *Comp. Clin. Pathol.* 23 (4), 989–992.
- Ait Lbacha, H., Alali, S., Zouagui, Z., El Mamoun, L., Rhalem, A., Petit, E., Haddad, N., Gandoin, C., Boulouis, H.J., Maillard, R., 2017. High prevalence of *Anaplasma* spp. in small ruminants in Morocco. *Transbound. Emerg. Dis.* 64 (1), 250–263.
- Aktas, M., Altay, K., Dumanli, N., 2011. Molecular detection and identification of *Anaplasma* and *Ehrlichia* species in cattle from Turkey. *Ticks Tick-Borne Dis.* 2 (1), 62–65.
- Ameen, K., Abdullah, B., Abdul-Razaq, R., 2012. Seroprevalence of *Babesia bigemina* and *Anaplasma marginale* in domestic animals in Erbil, Iraq. *Iraqi J. Vet. Sci.* 26 (Suppl. 3), 109–114.
- Atif, F., Khan, M., Iqbal, H., Roheen, T., 2012. Prevalence of tick-borne diseases in Punjab (Pakistan) and hematological profile of *Anaplasma marginale* infection in indigenous and crossbred cattle. *Pakistan J. Sci.* 64, 11–15.
- Bah, M., 2016. Effects of arthropod Ectoparasite infestations on livestock productivity in three districts in southern Ghana. PhD diss. 2016. University of Ghana available on. <http://ugspace.ug.edu.gh/handle/123456789/22955>.
- Byaruhanga, C., Collins, N.E., Knobel, D.L., Khumalo, Z.T., Chaisi, M.E., Oosthuizen, M.C., 2018. Molecular detection and phylogenetic analysis of *Anaplasma marginale* and *Anaplasma Centrale* amongst transhumant cattle in North-Eastern Uganda. *Ticks Tick-Borne Dis.* 9 (3), 580–588.
- DerSimonian, R., Kacker, R., 2007. Random-effects model for meta-analysis of clinical trials: an update. *Contemp. Clin. Trial* 28 (2), 105–114.
- Dhand, A., Nadelman, R.B., Aguero-Rosenfeld, M., Haddad, F.A., Stokes, D.P., Horowitz, H.W., 2007. Human granulocytic anaplasmosis during pregnancy: case series and literature review. *Clin. Infect. Dis.* 45 (5), 589–593.
- Hosseini-Vasoukolaei, N., Oshaghi, M.A., Shayan, P., Vatandoost, H., Babamahmoudi, F., Yaghoobi-Ershadi, M.R., Telmadarraiy, Z., Mohtarami, F., 2014. *Anaplasma* infection in ticks, livestock and human in Ghaemshahr, Mazandaran Province, Iran. *J. Arthropod. Borne Dis.* 8 (2), 204–211.
- Jafarbakloo, A., Bakhshi, H., Faghihi, F., Telmadarraiy, Z., Khazeni, A., Oshaghi, M.A., Ramzgouyan, M.R., Sedaghat, M.M., 2014. Molecular detection of *Anaplasma* and *Ehrlichia* infection in ticks in borderline of Iran-Afghanistan. *J. Biomed. Sci. Eng.* 7 (11), 919–926.
- Jalali, S., Khaki, Z., Kazemi, B., Bandehpour, M., Rahbari, S., Razi Jalali, M., Yasini, S., 2013. Molecular detection and identification of *Anaplasma* species in sheep from Ahvaz, Iran. *Iran. J. Vet. Res.* 14 (1), 50–56.
- Jalali, S.M., Bahrami, S., Rasooli, A., Hasanvand, S., 2016. Evaluation of oxidant/antioxidant status, trace mineral levels, and erythrocyte osmotic fragility in goats naturally infected with *Anaplasma ovis*. *Trop. Anim. Health Prod.* 48 (6), 1175–1181.



- Kaewmongkol, G., Lukkana, N., Yangtara, S., Kaewmongkol, S., Thengchaisri, N., Sirinarumit, T., Jittapalpong, S., Fenwick, S.G., 2017. Association of Ehrlichia canis, Hemotropic mycoplasma spp. and Anaplasma platys and severe anemia in dogs in Thailand. *Vet. Microbiol.* 201, 195–200.
- Khaki, Z., Jalali, S.M., Kazemi, B., Razi Jalali, M., Yasini, S.P., 2015. A study of hematological changes in sheep naturally infected with Anaplasma spp. and Theileria ovis: molecular diagnosis. *Iran. J. Vet. Med.* 9 (1), 19–26.
- Noaman, V., 2012. Identification of hard ticks collected from sheep naturally infected with Anaplasma ovis in Isfahan province, Central Iran. *Comp. Clin. Pathol.* 21 (3), 367–369.
- Noaman, V., 2013. Discrimination between Anaplasma marginale and Anaplasma ovis by PCR-RFLP. *World Appl. Sci. J.* 21 (2), 190–195.
- Noaman, V., Bastani, D., 2016. Molecular study on infection rates of Anaplasma ovis and Anaplasma marginale in sheep and cattle in West-Azerbaijan province, Iran. *Vet. Res. Forum* 7 (2), 163–167.
- Noaman, V.S. Arabzadeh, Kachouei, B., 2001. A study on anaplasmosis in cattle of Flavarjan city, Isfahan Province (1995–2000). *Pajouhesh-va-Sazandegi* 14 (3), 10–12.
- Noaman, V., Shayan, P., 2009. Molecular detection of Anaplasma phagocytophilum in carrier cattle of Iran—first documented report. *Iran. J. Microbiol.* 1 (2), 37–42.
- Noaman, V., Shayan, P., 2010a. Comparison of microscopy and PCR-RFLP for detection of Anaplasma marginale in carrier cattle. *Iran. J. Microbiol.* 2 (2), 89–94.
- Noaman, V., Shayan, P., 2010b. Molecular detection of Anaplasma bovis in cattle from central part of Iran. *Vet. Res. Forum* 1, 117–122.
- Noaman, V., Shayan, P., 2010c. A new PCR-RFLP method for detection of Anaplasma marginale based on 16S rRNA. *Vet. Res. Commun.* 34 (1), 43–50.
- Noaman, V., Shayan, P., Amininia, N., 2009. Molecular diagnostic of Anaplasma marginale in carrier cattle. *Iran. J. Parasitol.* 4 (1), 26–33.
- Pokorn, M., Zupanc, T.A., Strle, F., 2016. Pediatric human granulocytic anaplasmosis is rare in Europe. *Pediatr. Infect. Dis. J.* 35 (3), 358–359.
- Rar, V.A., Livanova, N.N., Panov, V.V., Doroschenko, E.K., Pukhovskaya, N.M., Vysochina, N.P., Ivanov, L.L., 2010. Genetic diversity of Anaplasma and Ehrlichia in the Asian part of Russia. *Ticks Tick-Borne Dis.* 1 (1), 57–65.
- Razmi, G.R., Dastjerdi, K., Hossieni, H., Naghibi, A., Barati, F., Aslani, M., 2006. An epidemiological study on Anaplasma infection in cattle, sheep, and goats in Mashhad Suburb, Khorasan Province, Iran. *Ann. N. Y. Acad. Sci.* 1078 (1), 479–481.
- Razzaq, F., Khosa, T., Ahmad, S., Hussain, M., Saeed, Z., Khan, M., Shaikh, R., Ali, M., Iqbal, F., 2015. Prevalence of Anaplasma phagocytophilum in horses from Southern Punjab (Pakistan). *Trop. Biomed.* 32, 233–239.
- Ringo, A.E., Moumouni, P.F.A., Lee, S.-H., Liu, M., Khamis, Y.H., Gao, Y., Guo, H., Zheng, W., Elstratiou, A., Galon, E.M., 2018. Molecular detection and characterization of tick-borne protozoan and rickettsial pathogens isolated from cattle on Pemba Island, Tanzania. *Ticks Tick-Borne Dis.* 9 (6), 1437–1445.
- Rodriguez-Morales, A.J., Bonilla-Aldana, D.K., Idarraga-Bedoya, S.E., Garcia-Bustos, J.J., Cardona-Ospina, J.A., Faccini-Martínez, Á.A., 2019. Epidemiology of zoonotic tick-borne diseases in Latin America: Are we just seeing the tip of the iceberg? *F1000Research* 7, 1988. <https://doi.org/10.12688/f1000research.17649.2>.
- Rosso, F., Tagliapietra, V., Baráková, I., Derdáková, M., Konečný, A., Hauffe, H.C., Rizzoli, A., 2017. Prevalence and genetic variability of Anaplasma phagocytophilum in wild rodents from the Italian alps. *Parasit. Vectors* 10 (1), 293.
- Rymaszevska, A., Grenda, S., 2008. Bacteria of the genus Anaplasma—characteristics of Anaplasma and their vectors: a review. *Vet. Med.* 53 (11), 573–584.
- Sainz, Á., Roura, X., Miró, G., Estrada-Peña, A., Kohn, B., Harrus, S., Solano-Gallego, L., 2015. Guideline for veterinary practitioners on canine ehrlichiosis and anaplasmosis in Europe. *Parasit. Vectors* 8 (1), 75.
- Salehzadeh, S., Fathi, S., Dehaghi, M.M., Asl, E.N., Nezhad, H.A., 2011. Survey of Theileria annulata and Anaplasma marginale in cattle in Kerman area, southeast of Iran. *Scientia Parasitologica.* 12 (2), 61–66.
- Sisson, D., Hufschmid, J., Jolles, A., Beechler, B., Jabbar, A., 2017. Molecular characterization of anaplasma species from african buffalo (Syncerus caffer) in kruger national park, South Africa. *Ticks Tick-Borne Dis.* 8 (3), 400–406.
- Stuen, S., Bergström, K., Petrovec, M., Van de Pol, I., Schouls, L.M., 2003. Differences in clinical manifestations and hematological and serological responses after experimental infection with genetic variants of Anaplasma phagocytophilum in sheep. *Clin. Diagn. Lab. Immunol.* 10 (4), 692–695.
- Von Elm, E., Altman, D.G., Egger, M., Pocock, S.J., Gøtzsche, P.C., Vandenbroucke, J.P., 2007. The Strengthening of Reporting of Observational Studies in Epidemiology (STROBE) statement: guidelines for reporting observational studies. *Ann. Intern. Med.* 147 (8), 573–577.
- Yousefi, A., Rahbari, S., Shayan, P., Sadeghi-dehkordi, Z., Bahonar, A., 2017. Molecular evidence of Anaplasma phagocytophilum: an emerging tick-borne pathogen in domesticated small ruminant of Iran; first report. *Comp. Clin. Pathol.* 26 (3), 637–642.