

'Clustering' SIRP α into the Plasma Membrane Lipid Microdomains Is Required for Activated Monocytes and Macrophages to Mediate Effective Cell Surface Interactions with CD47

Binh Ha¹✉, Zhiyuan Lv^{1,2}✉, Zhen Bian^{1,2}, Xiugen Zhang¹, Aarti Mishra¹, Yuan Liu^{1*}

1 Program of Cellular Biology and Immunology, Center of Inflammation, Immunity and Infection, Georgia State University, Atlanta, Georgia, United States of America, **2** School of Life Sciences, Nanjing University, Nanjing, Jiangsu, China

Abstract

SIRP α , an ITIMs-containing signaling receptor, negatively regulates leukocyte responses through extracellular interactions with CD47. However, the dynamics of SIRP α -CD47 interactions on the cell surface and the governing mechanisms remain unclear. Here we report that while the purified SIRP α binds to CD47 and that SIRP α is expressed on monocytes and monocytic THP-1 or U937, these SIRP α are ineffective to mediate cell binding to immobilized CD47. However, cell binding to CD47 is significantly enhanced when monocytes transmigrating across endothelia, or being differentiated into macrophages. Cell surface labeling reveals SIRP α to be diffused on naive monocytes but highly clustered on transmigrated monocytes and macrophages. Protein crosslink and equilibrium centrifugation confirm that SIRP α in the latter cells forms oligomerized complexes resulting in increased avidity for CD47 binding. Furthermore, formation of SIRP α complexes/clusters requires the plasma membrane 'lipid rafts' and the activity of Src family kinase during macrophage differentiation. These results together suggest that 'clustering' SIRP α into plasma membrane microdomains is essential for activated monocytes and macrophages to effectively interact with CD47 and initiate intracellular signaling.

Citation: Ha B, Lv Z, Bian Z, Zhang X, Mishra A, et al. (2013) 'Clustering' SIRP α into the Plasma Membrane Lipid Microdomains Is Required for Activated Monocytes and Macrophages to Mediate Effective Cell Surface Interactions with CD47. PLoS ONE 8(10): e77615. doi:10.1371/journal.pone.0077615

Editor: Yingxiao Wang, UC San Diego, United States of America

Received: May 22, 2013; **Accepted:** September 9, 2013; **Published:** October 15, 2013

Copyright: © 2013 Ha et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by a Research Scholar Grant from the American Cancer Society (YL). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

* E-mail: yliu@gsu.edu

✉ These authors contributed equally to this work.

Introduction

Signal regulatory protein α (SIRP α) (also known as SHPS1, CD172A, or P84) is a cell surface receptor-like signaling protein predominantly expressed in myeloid leukocytes including granulocytes, monocytes, macrophages and dendritic cells [1,2]. The expression of SIRP α is also reported in other specific types of cells such as neurons [3] and podocytes in the kidney [4]. As a typical type-1 transmembrane protein, SIRP α functions tightly correlate with its structure, which contains an extracellular domain with a highly interactive IgV-like loop and a cytoplasmic domain containing two immunoreceptor tyrosine-based inhibitory motifs (ITIMs) that are essential for signaling [5]. SIRP α has been reported to be crucial for leukocyte functions, including monocyte and neutrophil (PMN) chemotactic transmigration [6,7], macrophage phagocytosis [8], macrophage fusion [9], dendritic cell antigen presentation [10],

and other possible cellular functions such as neuronal axonal growth and synapse formation [3] and kidney filtration [4]. The regulation of all these cellular functions by SIRP α has been attributed to its extracellular binding interactions with the ligand CD47, another important cell surface protein broadly expressed on most cells, and to the SIRP α ITIMs-mediated intracellular signaling. It has been suggested that the extracellular binding of SIRP α to CD47 triggers tyrosine phosphorylation in its cytoplasmic ITIMs, resulting in the association of the SH2 domain-containing protein tyrosine phosphatase (SHP-1 or 2), which consequently initiates negative signaling events leading to the inhibition of cell function [11-16]. Studies of macrophage phagocytosis indicate that ligation of macrophage SIRP α by CD47 expressed on the encountered cells prohibits macrophage phagocytosis, whereas failure of SIRP α engagement by CD47, or deficiency of SIRP α ITIM-mediated signaling, promotes macrophage engulfment of the cell

[2,8,17-22]. It has also been suggested that increased CD47 expression on cancer cells serves as a crucial mechanism for the evasion of immune clearance [23,24].

The evidence found for high affinity binding between the extracellular domains of SIRP α and CD47 is convincing. Experiments using recombinant SIRP α and CD47 extracellular domains as well as transfected cells that overexpress the full-length SIRP α or CD47 protein on the cell surface [7,25-27] have demonstrated efficient binding between the two proteins. Despite being expressed by different cell types, we previously observed that CD47 purified from RBCs, leukocytes, spleen cells, fibroblasts and epithelial cells all directly bound to SIRP α [7]. Key amino acid residues in the SIRP α extracellular IgV-like loop, which mediates binding to CD47, have also been identified [26]. However, most of these experiments confirming SIRP α -CD47 binding were performed in acellular or transfection systems; cell functions implicating the involvement of SIRP α -CD47 interactions were also observed often indirectly, primarily through the usage of inhibitory antibodies against SIRP α or CD47 or recombinant soluble proteins that compete for binding. The interaction between SIRP α and CD47 on the surface of their native cells is not very clear. Moreover, as cell interactions are often transient, the mechanisms that regulate the dynamic binding and disengagement of SIRP α and CD47 require further elucidation.

In this study, we report that peripheral monocytes, despite the abundant cell surface expression of SIRP α , do not bind CD47. We found that effective SIRP α -mediated cell binding to CD47 occurs when monocytes are activated and/or differentiated into macrophages. Our data indicate that concomitant with monocyte activation/differentiation, SIRP α is distributed into the cholesterol-rich lipid microdomains of the plasma membrane, where it forms high density protein complexes. With this "clustered" structure, SIRP α on the cell surface mediates high avidity and effective binding interactions with CD47.

Materials and Methods

Cells

Human peripheral polymorphonuclear leukocytes (PMN) and peripheral blood mononuclear cells (PBMC) were isolated from whole blood donated by healthy volunteers who provided written informed consent according to the protocol approved by the institutional review board of Georgia State University [7,28]. Monocyte leukemic cell lines THP-1 and U937 obtained from American Tissue Culture Collection (ATCC) were cultured in RPMI 1640 supplemented with 10% FBS. Additionally the THP-1 medium contained 50 μ M β -mercaptoethanol. To induce macrophage differentiation, THP-1 cells were cultured in the presence of phorbol myristate acetate (PMA, 50 nM) for 3-4 days. Human microvascular endothelial cells (HMEC-1), kindly provided by Dr. E.W. Ades (Centers for Disease Control and Prevention, Atlanta) [29], were cultured in MCDB 131 medium supplemented with 10 mM/L L-glutamine, 10 ng/ml mouse epidermal growth factor (mEGF, BD Biosciences), 1 μ g/ml hydrocortisone (Sigma) and 10% FBS. Mouse NIH 3T3 cells, which produce murine macrophage colony stimulating factor

(M-CSF), were kindly provided by Dr. David Weiss (Emory University) and maintained in DMEM with 10% FBS. The harvested medium containing M-CSF was used to induce the differentiation of mouse bone marrow cells into macrophages [30]. Mouse peritoneal macrophages (PEM) were obtained by lavaging the peritoneal area with cold PBS followed by centrifugation. CHO-SIRP α cells, which permanently express the human SIRP α that was previously generated [26], and non-transfected CHO cells were maintained in DMEM with 10% FBS.

Antibodies, recombinant proteins and other reagents

Monoclonal antibodies (mAb) SE5A5 and SE7C2, against SIRP α , and anti-CD47 mAb B6H12.2 were described previously [7,26,28]. Polyclonal rabbit antibody against the human SIRP α extracellular domain (anti-SIRP α .ex) was generated by immunizing rabbits with a purified fusion protein consisting GST and the entire extracellular domain of SIRP α (SIRP α .ex-GST) [5]. After obtaining a highly reactive serum, SIRP α -specific IgG was enriched by affinity purification using protein A/G-conjugated Sepharose, and then passed through SIRP β .ex-GST-coated glutathione-Sepharose to eliminate the cross-reactivity with the close SIRP family member SIRP β [31]. A rabbit polyclonal antibody that recognizes the SIRP α cytoplasmic terminus (anti-SIRP α .ct) was obtained from EMD Millipore and was also generated in the lab; the antibody detects SIRP α of both human and murine origins. The rat anti-mouse SIRP α extracellular domain mAb P84 was purchased from BD Biosciences. CHO cells producing the fusion protein of human CD47 extracellular domain and alkaline phosphatase (CD47-AP) had been previously generated and the resulting CD47-AP in the medium was purified, as described previously [7,26]. Recombinant human monocyte chemotactic protein 1 (MCP-1), methyl- β -cyclodextrin (M β CD), triparanol, cholesterol oxidase (CO), filipin were purchased from Sigma. Chemicals, including 3,3'-dithiobis(sulfosuccinimidyl)propionate (DTSSP), Sulfo-NHS-Biotin, and NHS-Biotin, were from Pierce.

Immunofluorescence labeling of cells

To assess SIRP α expression on the cell surface, monocytes and macrophages were blocked with 2% BSA for 30 min followed by incubation with anti-SIRP α -ex for human cells and CHO, and mAb P84 for murine cells (2 μ g/ml each) for 60 min (25°C). Cells were washed, incubated with Alexa Fluor-conjugated secondary antibodies and then analyzed by fluorescence microscopy and/or FACS. To distinguish monocytes in PBMC, the cells were co-stained with the anti-CD14 (2 μ g/ml) antibody (BD Biosciences). Lymphocytes were detected by staining with the anti-Lck (Cell Signaling) and anti-CD45R/B220 (BioLegend) antibodies, while mouse macrophages were identified by co-staining with the anti-F4/80 antibody (BioLegend). Control staining was performed using isotype-matched IgG or the corresponding secondary antibodies alone. To identify lipid rafts, cells were co-stained with the Alexa Fluor 488-conjugated cholera toxin B subunit (CT-B) (Life Technologies). For perturbation of the lipid rafts, PMA-induced THP-1 cells were treated with M β CD (10 mg/ml), filipin (0.5 μ g/ml), triparanol (5 μ M) or cholesterol oxidase (CO,

1 unit/ml) for 1 h (former two agents) or 12 h (latter two agents) under cell culture conditions prior to cell staining. For kinase inhibition, THP-1 cells were treated with genistein (100 μ g/ml) (Sigma), PP1 (35 μ M) (BioMol), the genistein analog daidzein (non-inhibitory, 100 μ g/ml) (BioMol), a mixture of MAP kinase inhibitors SB203580 (p38 inhibitor) (BioMol) and PD98059 (p42/44 MEK) (Calbiochem) (20 μ M each), the phosphatidylinositol 3 (PI3) kinase inhibitor LY294002 (25 μ M) (Calbiochem), the Bruton's tyrosine kinase (Btk) inhibitor LFM-A13 (100 μ M) (Calbiochem), the Janus protein tyrosine kinase (JAK) inhibitor (JAK inhibitor I) (100 nM) (Calbiochem), which inhibits JAK1, JAK2, and JAK3, on day 2 and again on day 3 during PMA-induced macrophage differentiation. Cells were collected on day 4 for immunofluorescence staining.

SDS-PAGE, immunoblot, dot blot, biotinylation, protein cross-linking and equilibrium centrifugation

Cells were lysed in a cold lysis buffer containing 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 1% Triton X-100, protease inhibitor cocktail (1:50 dilution, Sigma) and 3 mM PMSF. After centrifugation, the supernatant lysates were collected. For western blot (WB) analysis, lysates were applied to SDS-PAGE and transferred to nitrocellulose. After blocking with 5% non-fat milk, the membrane was incubated with the appropriate primary antibodies followed by detection using the corresponding horseradish peroxidase (HRP)-conjugated secondary antibodies and ECL. For dot blot experiments, cell lysates were adsorbed to the nitrocellulose membrane by vacuum, blocked and detected with primary and secondary antibodies. For protein biotinylation, PMN and PBMC were treated with 2 mM NHS-biotin (cell permeable) or Sulfo-NHS-biotin (non-permeable) for 60 min (25°C), followed by washing and quenching with 100 mM glycine. Following cell lysis, SIRP α in PMN and PBMC was immunoprecipitated using the anti-SIRP α .ex-conjugated Sepharose and total and biotinylated SIRP α was detected using anti-SIRP α .ct and HRP-conjugated streptavidin, respectively. To cross-link cell surface proteins, monocytes and macrophages were treated with DTSSP for 30 min (25°C). After quenching, cells were lysed and SIRP α was analyzed by SDS-PAGE and WB under non-reducing and reducing conditions (+DTT); the reducing condition reverses DTSSP crosslinking. To detect the SIRP α complexes by equilibrium centrifugation, cell lysates with or without DTSSP crosslinking were supplemented with 5% sucrose and laid on top of sucrose density gradients (5-35% for human monocytes and 15-45% for mouse macrophages) prepared in 1.2 ml thick-walled ultracentrifuge tubes (Beckman Coulter). Ultracentrifugation was carried out using a TLS-55 rotor in an Optima TL Ultracentrifuge (Beckman Coulter) at 165,000 g for 14 h (4°C). Ten fractions (100 μ l each) were collected and SIRP α was analyzed by SDS-PAGE and WB. A biotinylated protein marker ranging from 40-250 kD (BioRad) was ultracentrifuged along with the cell lysates to serve as the MW reference.

Cell adhesion assay to test SIRP α -mediated cell interaction

Purified CD47-AP (10 μ g/ml in HBSS) was immobilized by incubation in 96-well microtiter plates for 2 h (25°C). After blocking with 1% BSA, the cell suspension (1 x 10⁶) in 50 μ l blocking buffer was added into CD47-AP-coated wells, in the presence or absence of antibodies or fusion proteins (10 μ g/ml), followed by stationary incubation for 30 min (25°C). The wells were then gently washed three times and cells that adhered were determined by microscopy. To calculate cell adhesion percentile, cells were labeled with the fluorescent dye BCECF (Molecular Probes) prior to incubation with CD47-AP. The fluorescence intensities of the cells before and after washing were recorded using a fluorescence plate reader at excitation and emission wavelengths of 485 and 535 nm, respectively.

Monocyte transendothelial migration

To establish endothelial monolayers, HMEC-1 cells were cultured on polycarbonate transwells (0.33 cm², 5 μ m pore size, Corning) until confluence. To induce monocyte transmigration, freshly isolated human PBMC (5 x 10⁶, containing ~1 x 10⁶ monocytes) were added to the upper chamber of the transwells in 150 μ l RPMI 1640 followed by addition of MCP-1 (50 ng/ml) in 0.5 ml the same medium into the lower chamber (supplemental data). After incubation for 2 h (37°C), monocytes that transmigrated into the lower chamber were collected for further analysis.

SIRP α purification and examination of binding to CD47

After lysis of various cells, SIRP α in detergent soluble supernatants was incubated with anti-SIRP α .ex-conjugated Sepharose for 3 h (4°C). The Sepharose beads were washed with Tris buffer, pH 7.5, containing 1% n-octyl- β -D-glucopyranoside (OG). Elution of SIRP α was performed using 100 mM sodium acetate, pH 4.0, containing 1% OG, followed by immediate neutralization with 1.0 M Tris buffer, pH 8.0. Affinity purified SIRP α was confirmed by WB using anti-SIRP α .ex and anti-SIRP α .ct antibodies. To examine SIRP α binding to CD47, purified SIRP α was diluted in HBSS (1:10) and immobilized in 96-well microtiter plates (2 h, 4°C). After blocking, the wells were incubated with CD47-AP (1 μ g/ml) for 30 min (25°C) in the presence or absence of inhibitory antibodies. After washing, CD47 binding was detected using the AP substrate p-nitrophenylphosphate (pNPP, Sigma). In some experiments, dot blots were conducted by absorption of purified SIRP α onto nitrocellulose. After blocking, the membrane was incubated with CD47-AP (1 μ g/ml) followed by the analysis of AP activity using NBT (nitro-blue tetrazolium chloride) and BCIP (5-bromo-4-chloro-3'-indolylphosphate p-toluidine salt) (Promega). The densities of the dots were quantified using the NIH densitometry software Image J.

Results

Peripheral monocytes express ample SIRP α on the cell surface but do not effectively interact with CD47

We examined SIRP α expression in human peripheral granulocytes (PMN) and peripheral mononuclear cells (PBMC). As shown in Figure 1A, immunoblotting (WB) using polyclonal antibodies against the SIRP α extracellular domain (anti-SIRP α .ex) or the cytoplasmic tail (anti-SIRP α .ct) detected the abundant expression of SIRP α (~65-85kD) in human PMN and PBMC. Cell surface immunofluorescence labeling showed that in PBMC, SIRP α is selectively expressed on CD14⁺ monocytes but not on lymphocyte-specific protein tyrosine kinase (Lck)⁺ T cells or B220⁺ B lymphocytes (Figure 1B), confirming previous results [25]. Protein biotinylation assays to compare SIRP α distribution in monocytes and PMN showed that SIRP α in PMN was labeled with biotin only following membrane permeabilization, as shown in Figure 1C. This was in agreement with our previous finding indicating that the majority of SIRP α is stored in intracellular pools in PMN prior to chemoattractant stimulation [31]. However, when labeling PBMC, we observed strong biotinylated SIRP α under both permeable and impermeable conditions, suggesting that SIRP α was distributed on the monocyte surface. FACS analysis confirmed that the majority of SIRP α is directly distributed on the monocyte cell surface, as membrane permeabilization did not show additional labeling (Figure 1D). Similar results were obtained when detecting SIRP α expression in monocyte-like, THP-1 and U-937 cells, in which the majority of SIRP α was found on the cell surface instead of being located in intracellular pools (Figure S1).

While SIRP α is significantly expressed on the cell surface, we found that peripheral monocytes, as well as cultured THP-1 and U-937, could not effectively interact with extracellular CD47. The cell adhesion assay, depicted in Figure 2A, was used to test SIRP α -mediated cellular adhesion interactions with recombinant CD47 containing the SIRP α -binding domain (CD47-AP) [7]. As shown in Figure 2B, none of the human PBMC (~30% monocytes), CD14-enriched monocytes (>90% purity), cultured THP-1 or U-937 cells adhered to immobilized CD47-AP. In contrast, the control CHO cells with SIRP α over-expression (CHO-SIRP α), but not the mock-transfected cells, displayed direct CD47-AP adhesion (Figure 2C). Addition of either the anti-SIRP α mAb SE5A5 or the anti-CD47 mAb B6H12.2 completely eliminated CHO-SIRP α adhesion (Figure 2D), confirming the presence of the SIRP α and CD47-mediated cell surface interaction. FACS analyses revealed that CHO-SIRP α expressed significantly higher amounts of SIRP α on the cell surface than monocytes (Figure 2E), suggesting that the density of SIRP α may play an important role in SIRP α -mediated cell surface binding interactions.

Activated monocytes and macrophages mediate high avidity binding to CD47 via SIRP α

Previous reports have shown that monocyte transmigration in response to inflammatory challenge involves the interaction of SIRP α on the monocyte surface with CD47 expressed on tissue cells [6], suggesting that the activation of monocytes

might be required for SIRP α to interact with CD47. To test this, we treated freshly isolated PBMC with MCP-1, and also induced monocytes to transmigrate across HMEC-1 monolayers towards MCP-1 (depicted in Figure S2). As shown in Figure 3, direct treatment with MCP-1, a method which is known to partially activate monocytes, only slightly increased monocyte adhesion to CD47-AP. In contrast, monocytes that were induced to transmigrate across HMEC-1 monolayers exhibited remarkable binding to CD47-AP. Addition of inhibitory anti-SIRP α mAbs (SE5A5 and SE7C2) or a soluble SIRP α extracellular domain (SIRP α .ex Fc), which competitively binds to CD47, abrogated the adhesion of transmigrated monocytes to CD47-AP, confirming the SIRP α -mediated cell surface interaction (Figure 3).

Similarly, we tested if the activation/differentiation of THP-1 cells would alter SIRP α -mediated cell binding. In these experiments, THP-1 cells were treated with PMA for 3-4 days to induce macrophage phenotypic differentiation. In contrast to non-induced THP-1 cells that displayed no adhesion, PMA-induced THP-1 macrophages demonstrated readily binding to CD47 (Figure 3). Inhibition of SIRP α -CD47 extracellular interaction with mAbs and SIRP α .ex Fc completely eliminated the THP-1 macrophage adhesion (Figure 3). Further, FACS analyses of THP-1 macrophages, as well as transmigrated monocytes, showed similar labeling intensities on these cells as on non-differentiated THP-1 or naive monocytes (Figure 3C), suggesting that the enhanced adhesion to CD47 was not due to increases in SIRP α expression.

To examine if the failure of binding to CD47 by naive monocytes and non-differentiated THP-1 is due to the SIRP α incompetent binding (contrary to the competent binding on transmigrated monocytes and THP-1 macrophages), we affinity purified SIRP α from different cells and compared their direct binding. As shown in Figure 4A, SIRP α in the different cell lysates was affinity absorbed to anti-SIRP α .ex-conjugated Sepharose and, after washing, was eluted at pH 4.0 in a buffer containing 1% OG. Purified SIRP α was then immobilized onto microtiter plates (Figure 4B) or nitrocellulose (Figure 4C), followed by testing for direct CD47 binding. As shown, all purified SIRP α , including those from naive monocytes and non-differentiated THP-1, displayed effective binding to CD47, implying that the differences of cellular interactions are unlikely caused by alterations in the SIRP α structure or the modification.

SIRP α clusters on activated monocytes and macrophages

Interestingly, when conducting the cell surface labeling, we observed that the distribution of SIRP α on naive monocytes is different from that on transmigrated monocytes or macrophages. As shown in Figure 5A, SIRP α presents as a diffuse distribution pattern on freshly isolated human monocytes, but a highly punctate staining pattern on transmigrated monocytes and THP-1 macrophages. This suggests that SIRP α is clustered at the plasma membrane. Similar diffuse and punctate staining patterns of SIRP α were also observed when labeling murine peripheral monocytes and macrophages, respectively (Figure 5A). Both peritoneal

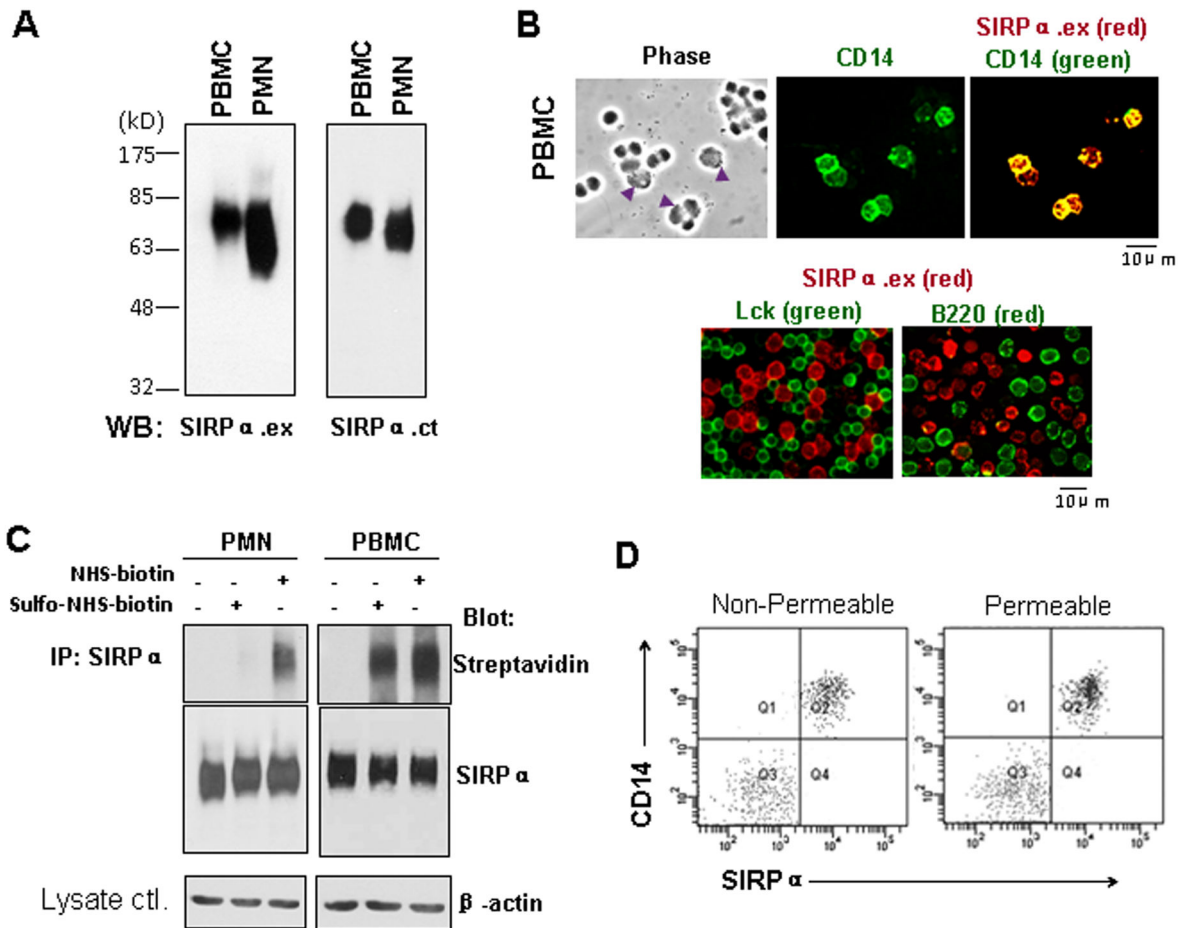


Fig. 1

Figure 1. Expression of SIRP α on monocytes. Detection of SIRP α by WB in freshly isolated human PBMC and PMN using anti-SIRP α .ex (SIRP α .ex) and anti-SIRP α .ct (SIRP α .ct) antibodies. B) Immunofluorescence staining of PBMC. Cell surface SIRP α was labeled with anti-SIRP α .ex under non-permeable conditions. Monocytes (arrow heads) were defined by anti-human CD14 antibody, and T and B lymphocytes were defined with anti-Lck and anti-B220 antibodies, respectively. C) Determination of the distribution of SIRP α in PMN and monocytes by cell biotinylation. Cells were biotinylated with either NHS-biotin (cell permeable) or sulfo-NHS-biotin (non-cell permeable), followed by the IP of SIRP α from the lysates. Biotinylated and total SIRP α were blotted by streptavidin-HRP and anti-SIRP α .ct, respectively. The same amount of protein was used for IP as confirmed by the actin blots. D) FACS analysis to determine SIRP α in monocytes labeled under cell permeable and non-permeable conditions.

doi: 10.1371/journal.pone.0077615.g001

macrophages (PEM) and bone marrow derived macrophages (BMDM) displayed highly punctate SIRP α staining on the cell surface. Given that CD47 is also expressed in monocytes and macrophages, we assessed if SIRP α clustering involves interactions with CD47 in the same membrane (*cis*-interaction). However, double staining of THP-1 macrophages and BMDM showed that CD47 displayed a mixture of diffuse and punctate pattern that did not largely overlap with the staining of SIRP α (Figure 5B).

To confirm SIRP α clustering on activated/differentiated leukocytes, we performed chemical crosslinking using a cell-impermeable crosslinker DTSSP. As shown in Figure 5C, DTSSP was unable to crosslink SIRP α on naïve monocytes, which remained to be resolved as the monomer by SDS-PAGE (70-80 kD and 110-120 kD for human and murine SIRP α , respectively). In contrast, DTSSP dramatically crosslinked SIRP α on transmigrated monocytes and macrophages, resulting in SIRP α shifting to a much larger size that was barely resolved by 8-10% polyacrylamide gel electrophoresis.

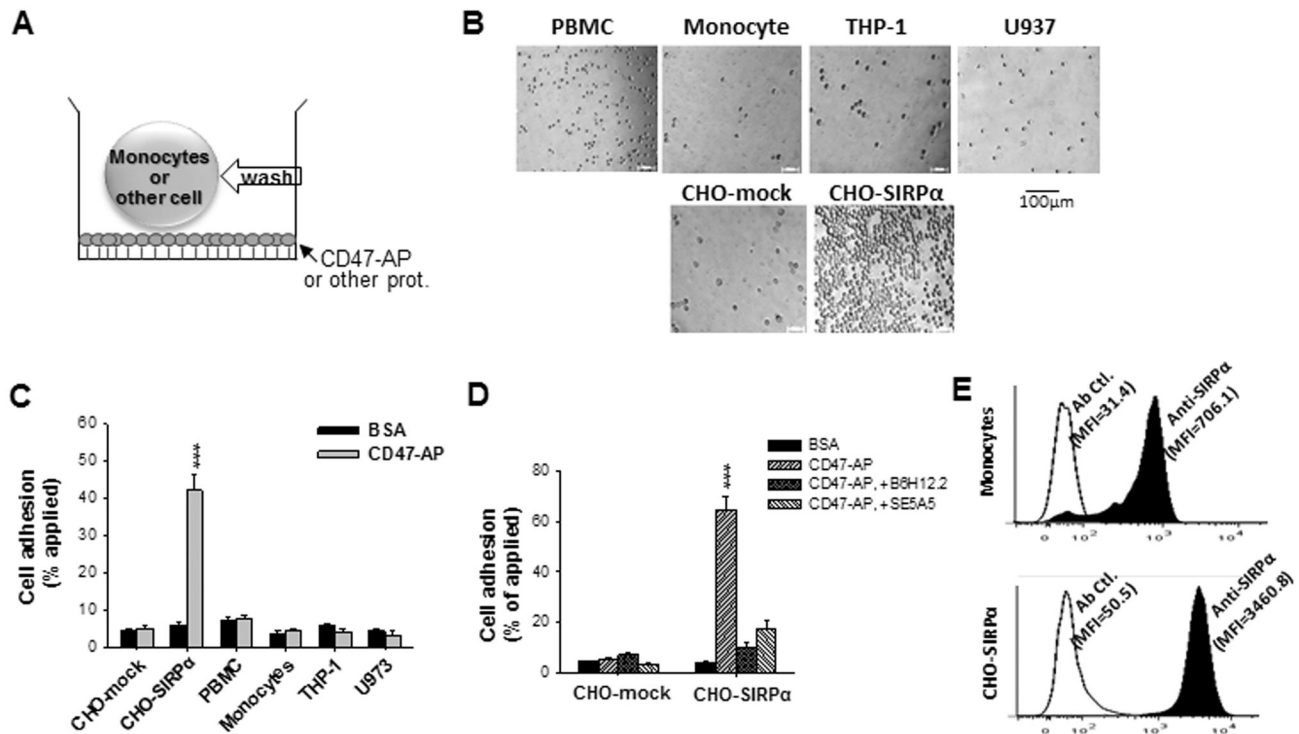


Fig. 2

Figure 2. SIRP α in naïve monocytes do not mediate cell adhesion to immobilized CD47. A) Schematic depiction of the cell adhesion assay. B) Adhesion of different cells to immobilized CD47-AP. Cell suspensions were incubated with CD47-AP or BSA (ctl.) for 30 min (25°C), followed by washing. C) Percentage of cell adhesion to CD47-AP versus BSA. D) Control adhesion performed using CHO cells with or without SIRP α transfection (CHO-mock and CHO-SIRP α , respectively), and in the presence or absence of inhibitory anti-SIRP α (SE5A5) and anti-CD47 (B6H12.2) mAbs (20 μ g/ml for each). The data are expressed as the means \pm SE and represent three independent experiments with triplicate wells per cell type. ***, $P < 0.001$. E) FACS analyses of SIRP α on the cell surfaces of peripheral monocytes and CHO-SIRP α cells.

doi: 10.1371/journal.pone.0077615.g002

Reversal of the cross-linking by DTT instantly resolved SIRP α to its monomeric size (Figure 5C). This remarkable change of SIRP α by crosslinking was consistent with the cell labeling observation, suggesting that SIRP α is clustered to large complexes on activated monocytes and macrophages. However, these SIRP α complexes remained in the detergent-soluble lysates, as dot blots without gel electrophoresis detected the protein (Figure 5C). We also investigated if SHP-1/2, the intracellular signaling molecule downstream of SIRP α , would be affected by DTSSP crosslinking. As shown, in naïve monocytes and unstimulated THP-1 cells, SHP-1 resolves as a distinctive protein of ~65kD, despite DTSSP treatment. Interestingly, SHP-1 could not be detected in transmigrated monocytes and macrophages after DTSSP cross-linking, suggesting that SHP-1 also formed protein complexes. As SHP-1 stays intracellularly and DTSSP is

membrane impermeable, the complexes of SHP-1 observed by DTSSP treatment are thus likely formed through protein interactions with certain transmembrane partners such as SIRP α . To determine the size of SIRP α complexes, lysates prepared from DTSSP-treated cells were applied to sucrose density gradients followed by equilibrium ultracentrifugation. As shown in Figure 5D, the sedimentation of crosslinking-stabilized SIRP α complexes from transmigrated monocytes and macrophages was significantly faster than the monomeric SIRP α from PBMC and the non-crosslinked macrophages. Judging by the reference protein markers, the sizes of SIRP α complexes formed in human monocytes and murine macrophages were both ≥ 250 kD (Figure S3).

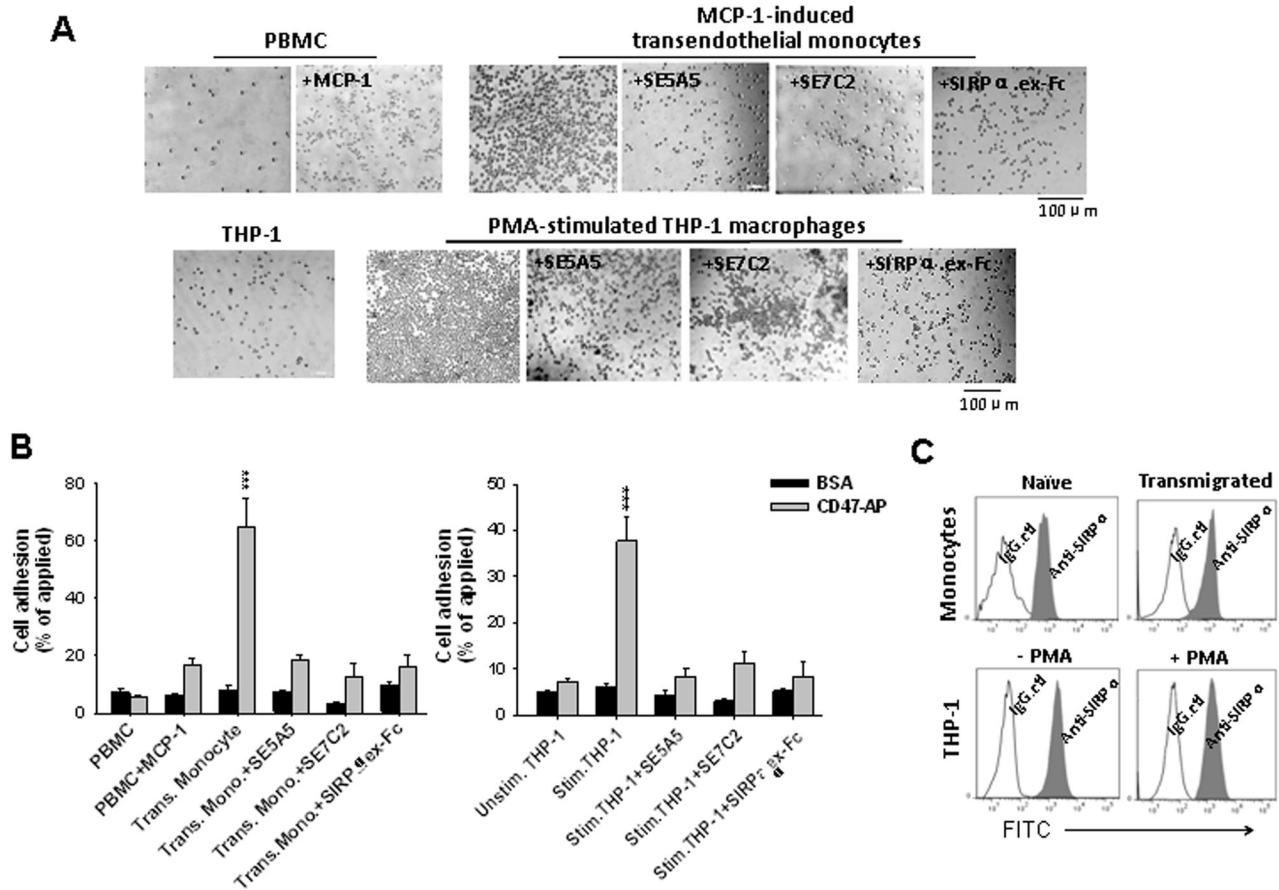


Fig. 3

Figure 3. SIRP α in activated monocytes and macrophages mediate high avidity cell adhesion to immobilized CD47. To obtain activated monocytes, freshly isolated PMBC were treated with 50 ng/ml MCP-1 (37°C, 1 h) and were also induced to transmigrate across HMEC-1 endothelial monolayers by MCP-1. To obtain macrophages, THP-1 cells were treated with PMA (50 nM) for 3-4 days. A) Cell adhesion assays. SE5A5 and SE7C2 are both inhibitory anti-SIRP α mAbs. B) Percentage of cell adhesion to CD47-AP versus BSA. ***, $P < 0.001$. C) FACS analysis of SIRP α on the surface of naïve (PBMC) and activated monocytes, unstimulated and stimulated THP-1 using anti-SIRP α .ex antibody.

doi: 10.1371/journal.pone.0077615.g003

SIRP α clustering involves cholesterol-rich membrane rafts and actin filaments

As the cholesterol-rich lipid rafts in the plasma membrane play a critical role in the assembly of important cell surface structures, we investigated the role of lipid rafts in clustering SIRP α on activated monocytes and macrophages. As shown in Figure 6A, brief (30 min) treatments of THP-1 macrophages with M β CD, a widely used cholesterol-depleting/lipid raft perturbation reagent, disassembled the SIRP α clusters on the cell surface. Likewise, inhibition of cholesterol biosynthesis with triparanol or modifying cholesterol with cholesterol oxidase (CO) also dispersed the SIRP α complexes. Treatment with

filipin, which binds cholesterol but does not necessarily perturb the lipid rafts [32,33], however, did not disassociate SIRP α clusters. The concomitant labeling of cell surface ganglioside GM1, a characteristic lipid raft marker, with cholera toxin B subunit (CT-B) revealed that the majority of SIRP α co-localized within the lipid rafts (~55-65%, calculated by the ZEISS microscope software ZEN) (Figure 6B). Furthermore, the loss of SIRP α clusters correlated with a reduction in the strength of THP-1 macrophage binding to CD47. As shown in Figure 6C, treating PMA-differentiated THP-1 with M β CD, triparanol and CO, but not filipin, resulted in the attenuation of SIRP α -mediated cell binding to CD47-AP. As the actin cytoskeleton

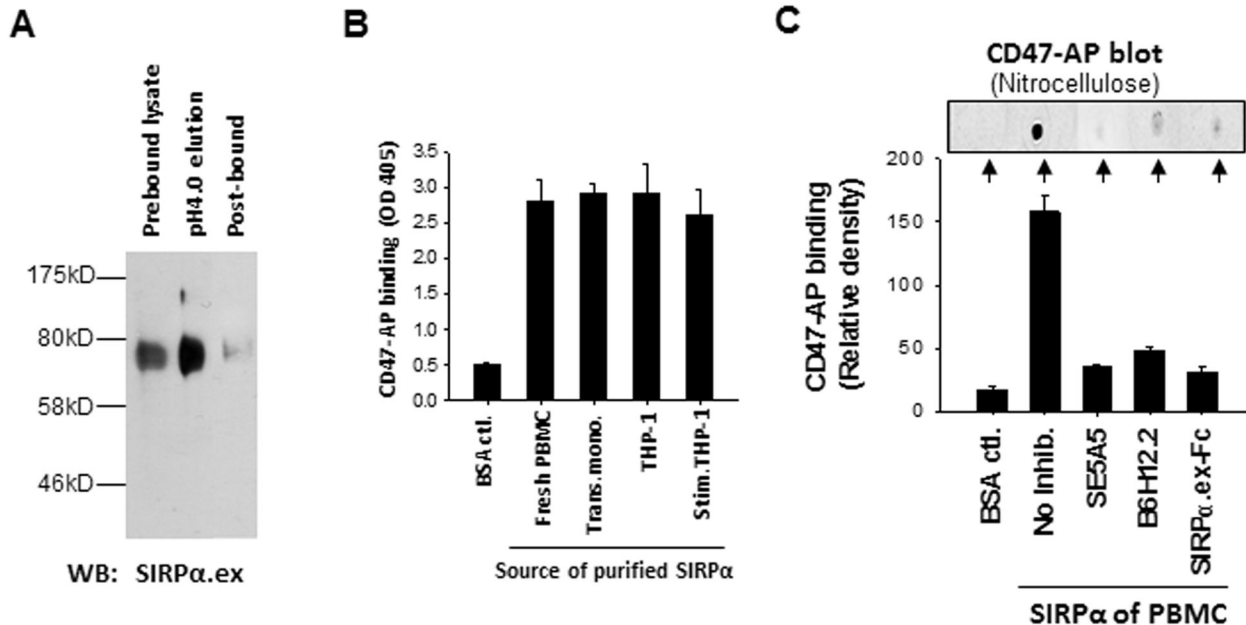


Fig. 4

Figure 4. Purified SIRP α binding to CD47. SIRP α , affinity purified from naïve and activated monocytes and unstimulated and stimulated THP-1, was tested for direct binding to CD47-AP. A) Affinity purification of SIRP α . SIRP α in different cell lysates was absorbed by anti-SIRP α .ex-conjugated Sepharose and eluted at pH 4.0 in the presence of 1% OG. B) Binding of purified SIRP α from various sources to CD47-AP. Purified SIRP α was immobilized in 96-well plates following a 1:10 dilution of OG. Binding to CD47 was tested by incubation with CD47-AP (2 μ g/ml, 30 min), followed by washing and detection of AP activity. The figure inset shows similar amounts of immobilized SIRP α in the wells as detected by WB (anti-SIRP α .ct). C) Binding of SIRP α purified from PBMC to CD47-AP in the presence of inhibitory anti-SIRP α (SE5A5) and anti-CD47 (B6H12.2) mAbs, or SIRP α .ex Fc. Purified SIRP α samples were absorbed onto 96-well plates (lower panel) or nitrocellulose membrane (upper panel). Following blocking with BSA and incubation with CD47-AP, the binding was assessed by AP activity.

doi: 10.1371/journal.pone.0077615.g004

has been well recognized as a dynamic regulator of lipid raft stability, we also treated cells with cytochalasin D to disrupt actin filaments. As shown, brief treatment with cytochalasin D (30 min) largely diminished SIRP α clusters on THP-1 macrophages. Altogether, these results suggest that cholesterol-rich lipid rafts are critically involved in clustering and stabilization of SIRP α complexes during monocyte activation and/or differentiation into macrophages.

Involvement of Src family tyrosine kinase in the formation of SIRP α clusters

To investigate the potential mechanisms that control the SIRP α complex formation, inhibitors targeting different signaling pathways were tested during PMA-induced THP-1 differentiation into macrophages. As shown in Figure 7A, inhibiting the Src family tyrosine kinases with PP1 and

genistein significantly blocked SIRP α clustering on the cell surface. On the contrary, treating THP-1 with daidzein, a genistein analog that lacks tyrosine kinase inhibition, had no effect. Inhibition of other signaling pathways including PI3 kinase (LY294002), MAP kinases such as p38 MAP (SB 203580) and MAP kinase kinase (PD 98059), Janus kinases (JAK1-3) and Bruton's tyrosine kinase (Btk) (LFM-A13) did not affect the formation of SIRP α complexes. Consistent with the pattern of SIRP α distribution on the cell surface, cell adhesion assays demonstrated that the treatment with genistein and PP1, but not the other inhibitors, eliminated THP-1 macrophage binding to CD47 (Figure 7B).

Discussion

This study was initiated following the observation that peripheral monocytes express abundant SIRP α on the cell

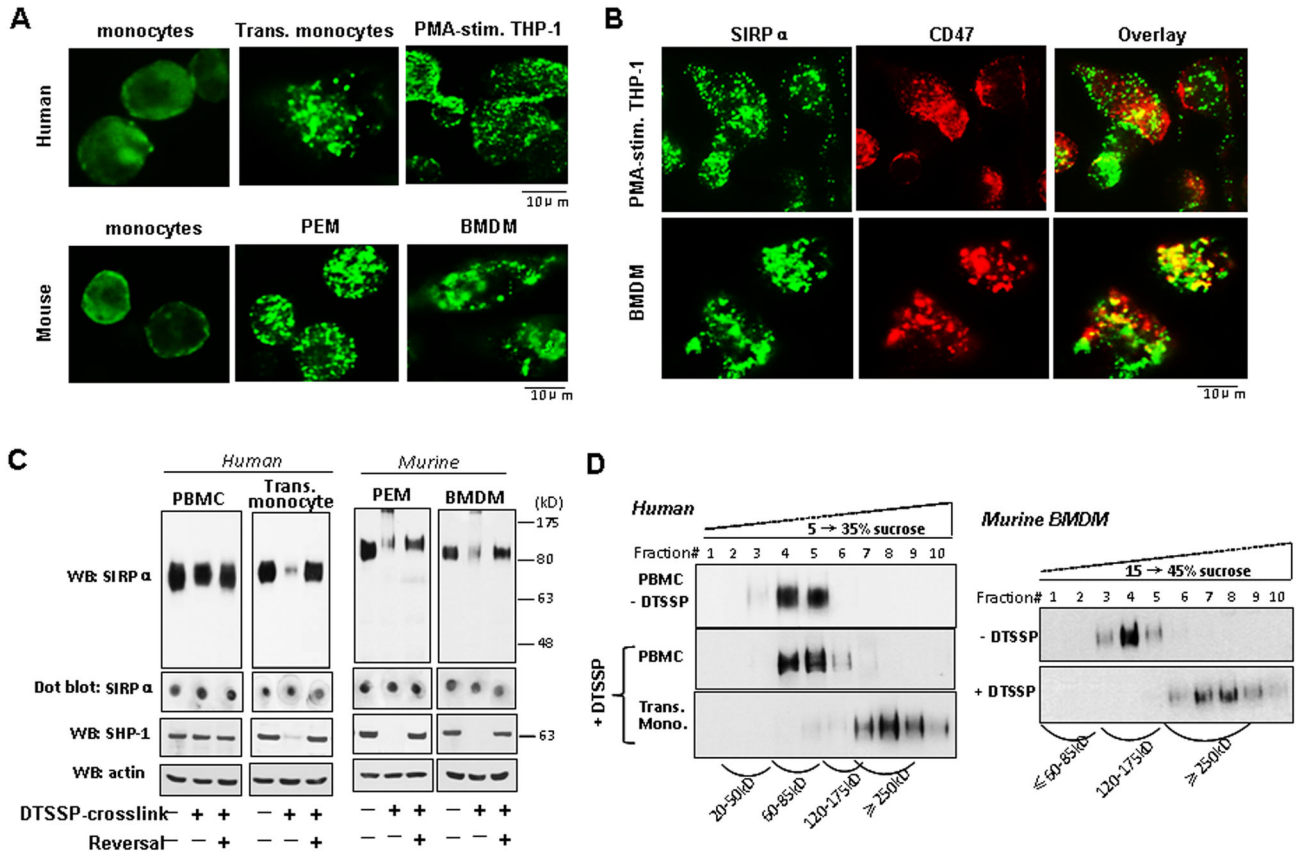


Fig. 5

Figure 5. SIRP α on activated monocytes and macrophages forms clusters/complexes. A) Assessing SIRP α on naïve PBMC, transmigrated monocytes, PMA-induced THP-1 and murine macrophages by immunofluorescence staining using anti-SIRP α .ex and mAb P84, an antibody specific for the murine SIRP α extracellular domain. B) Staining of SIRP α and CD47 on PMA-stimulated THP-1 using anti-SIRP α .ex and mAb B6H12.2. C) DTSSP crosslinking to assess protein complexes consisting SIRP α . Different cells were treated with DTSSP (25°C, 30 min) prior to lysis in detergent-containing buffer. Lysate samples were analyzed by SDS-PAGE and WB under non-reducing or reducing (+DTT) conditions. The latter reverses the DTSSP crosslinking. Note: murine SIRP α appears to be 110-120 kD, larger than the human counterpart. Dot blot detection of SIRP α was performed by directly absorbing cell lysates onto nitrocellulose, prior to antibody blotting. D) Equilibrium centrifugation analysis of SIRP α complexes. After DTSSP crosslinking, cell lysates were applied to ultracentrifugation in sucrose density gradients. Following fractionation, SIRP α localization was analyzed by WB and compared to the sedimentation of a reference protein marker.

doi: 10.1371/journal.pone.0077615.g005

surface but do not mediate cell binding interactions with the extracellular ligand CD47. Unlike PMN, which store the majority of SIRP α in intracellular granules to be released only after inflammatory stimulation [31], naïve monocytes lack intracellular SIRP α pools but display a majority of SIRP α on the cell surface. Despite ample SIRP α on the surface, peripheral monocytes maintain low adhesive interactions with CD47 or CD47-expressing cells, e.g., RBCs. In fact, no monocyte aggregation with RBCs under physiological conditions has ever been observed in the peripheral blood stream, even in the microvasculature in which the blood flow is slow. In this study,

more definitive evidence confirming the inefficient binding of monocytes to CD47 was obtained by testing peripheral monocytes, or monocytic THP-1 and U937, for direct adhesion/ binding to purified recombinant CD47 *in vitro*.

While naïve monocytes must avoid adhesive interactions to maintain peripheral blood homeostasis under resting conditions, activated monocytes and monocyte-differentiated macrophages require constant interaction with the surrounding cells when migrating through endothelial monolayers and tissues during inflammation or for immune surveillance. Studies have shown that SIRP α and CD47-mediated cell surface

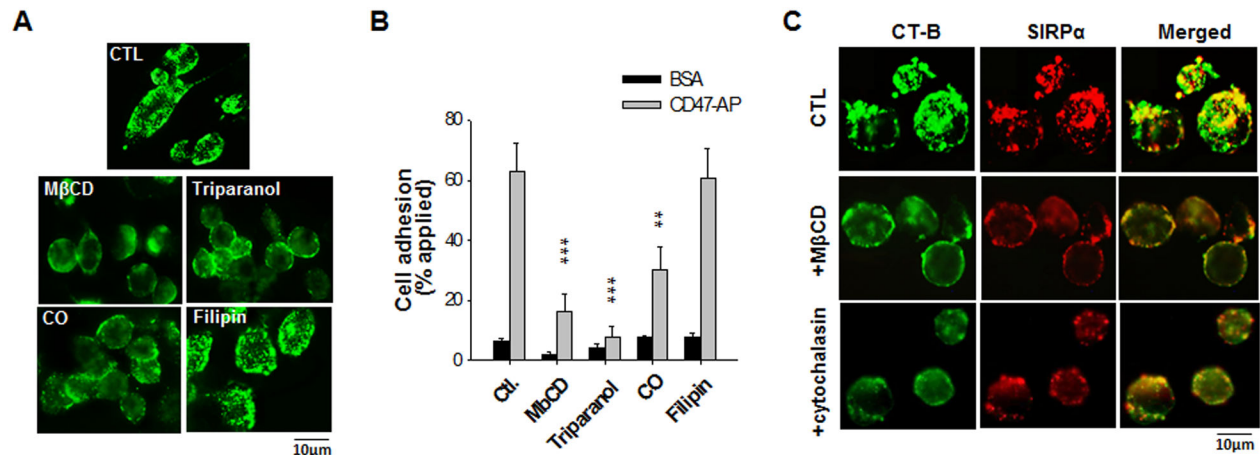


Fig. 6

Figure 6. Clustering of SIRP α on cell surfaces requires cholesterol-rich lipid rafts. PMA-stimulated THP-1 cells were treated with lipid raft/cholesterol perturbation/modification agents, including M β CD (10 mg/ml), triparanol (5 μ M), cholesterol oxidase (CO) (1 unit/ml) and filipin (0.5 μ g/ml), prior to A) assessment of SIRP α on the cell surface by immunofluorescence staining, B) analysis of SIRP α localization in lipid rafts by co-staining for raft marker GM1 by cholera toxin B subunit (CT-B), C) assay of SIRP α -mediated cell adhesion to immobilized CD47-AP. **, $P < 0.01$. ***, $P < 0.001$. The effect of the actin filament disruption agent cytochalasin D on lipid rafts and SIRP α distribution was also tested (shown in C).

doi: 10.1371/journal.pone.0077615.g006

interactions play a critical role in monocyte chemotactic transmigration and macrophage recognition of phagocytic targets. Inhibition of the SIRP α -CD47 interaction delays monocyte transmigration [6] and also perturbs macrophages to elicit intracellular signaling, which normally restrains macrophage phagocytosis of the surrounding cells [2,17-22,34]. Consistently, our cell adhesion assays showed that monocytes that had been activated and had transmigrated through the endothelial monolayers or that had differentiated into macrophages displayed significant direct binding to CD47. The striking difference between the activated monocytes/macrophages and the naïve monocytes in CD47 binding, and the lack of information on the mechanism(s) by which cell binding is so dynamically controlled, motivated the studies presented here.

Our investigations led to the following conclusions: first, the levels of SIRP α in the plasma membrane did not differ considerably between naïve and activated monocytes or un-induced and induced THP-1 macrophages. Second, by isolating individual SIRP α proteins from both naïve and activated monocytes and macrophages, we excluded the possibility that a conformational change or modifications in the SIRP α extracellular domain as the likely mechanism conferring CD47 binding. Third, there is a difference in the distribution of SIRP α on cell surfaces between naïve and activated monocytes or macrophages. When activated by cytokines and during transmigration, monocytes experience a redistribution of

SIRP α , from being dispersed in the plasma membrane to cluster into multi-stranded protein complexes. This 'clustered' SIRP α distribution confers high avidity binding to CD47 at the cell surface, enabling monocytes transmigration across endothelial monolayers. Similarly in macrophages, the cell surface bundling of SIRP α allows high avidity interactions with CD47 displayed on the encountered cells, leading to strong ITIM-mediated signaling and inhibition of phagocytosis (Figure 7C). In addition, our studies also found that SIRP α complexes are stabilized by cholesterol-rich 'lipid rafts' in the plasma membrane. Disruption of the lipid rafts by cholesterol-depletion or raft perturbation caused disassembly of SIRP α clusters on the cell surface, concomitant with reduced cell binding to CD47. Intriguingly, instead of resistance to non-ionic detergent, we found that the SIRP α complex-associated 'lipid rafts' are sensitive to Triton X-100 solubility and only partially co-localize with flotillin, a lipid raft marker [35], as analyzed by WB (Figure S3). Perhaps, the increased sensitivity to detergent indicate that these lipid rafts consisting of SIRP α complexes are rather "soft" and may contain relatively low levels of saturated lipids, thus providing a high degree of lipid and protein binding dynamics. In fact, similar 'soft' rafts have been reported to associate with other important receptors on the cell surface. These include the T cell antigen receptor (TCR) [36], immunoglobulin E (IgE) receptor [37], some Fc receptors [38], and ion channels [39], etc. The mechanisms involved in

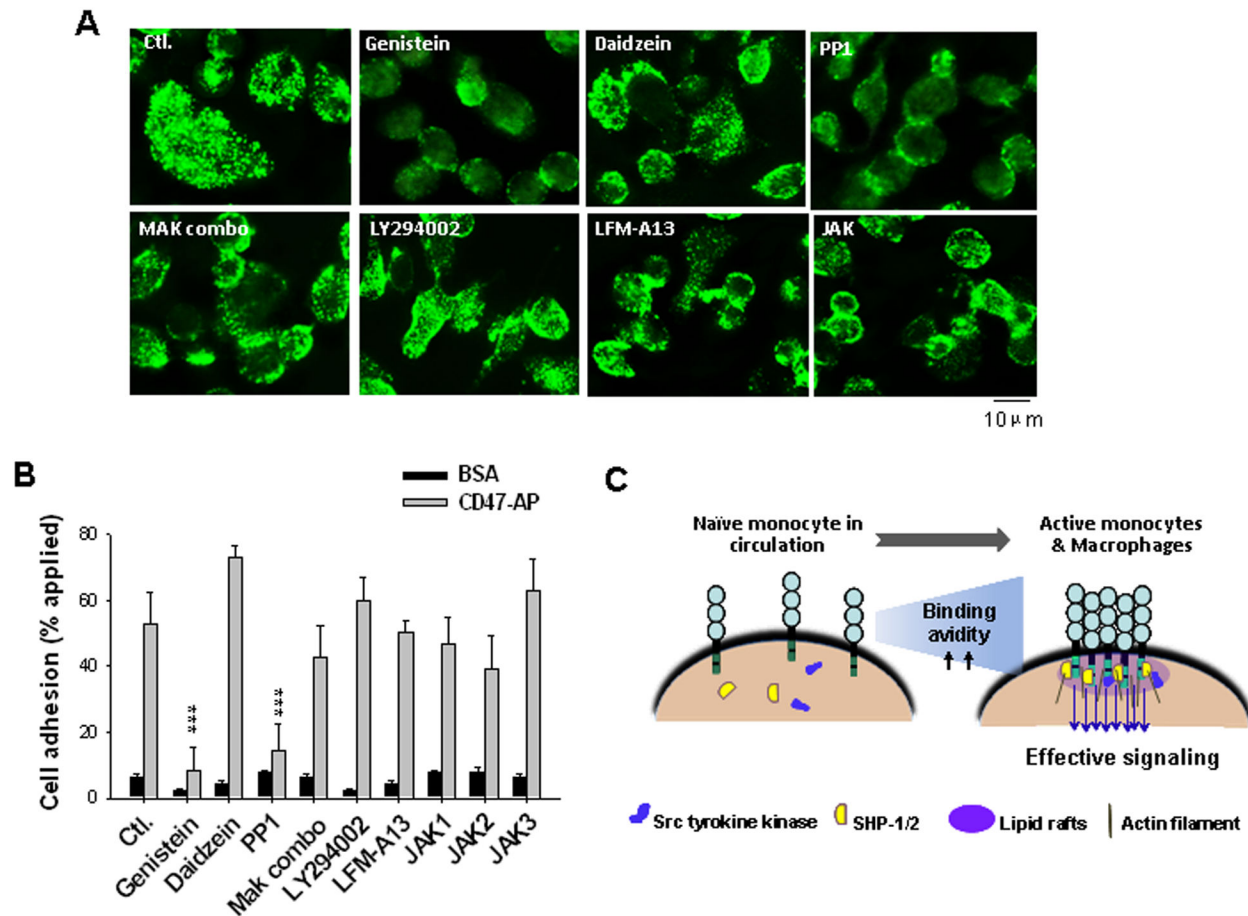


Fig. 7

Figure 7. Role of the Src family tyrosine kinase in SIRP α clustering. THP-1 cells were induced to differentiate into macrophages by treating with PMA for 5 days. From day 3, various pharmacological inhibitors were added into the cell culture, including the Src tyrosine kinase inhibitors genistein (100 μ g/ml) and PP1 (35 μ M), the genistein analog daidzein (non-inhibitory, 100 μ g/ml), a mixture of MAP kinase inhibitors (MAP combo) SB203580 and PD98059 (20 μ M each), the PI3 kinase inhibitor LY294002 (25 μ M), the Btk inhibitor LFM-A13 (100 μ M), the JAK inhibitor (JAK inhibitor I, 100 nM). A) Immunofluorescence staining of SIRP α on the THP-1 surface by anti-SIRP α .ex. B) Assay of SIRP α -mediated cell adhesion to immobilized CD47-AP. ***, $P < 0.001$. C) A model of SIRP α distribution and complex formation on monocytes and macrophages.

doi: 10.1371/journal.pone.0077615.g007

formation of this particular raft, or other types of membrane microdomains, remain unresolved.

We further explored if the formation of SIRP α complexes on activated monocytes and macrophages is controlled by specific signal transduction mechanisms. Our studies showed that inhibition of the Src family tyrosine kinase by PP1 and genistein prevented the clustering of SIRP α during THP-1 differentiation into macrophages, suggesting that Src kinase-mediated tyrosine phosphorylation plays a key role in bundling SIRP α into lipid rafts. One of the likely targets for Src kinase appears to be SIRP α itself, as its essential cytoplasmic ITIMs contain

four tyrosine residues. Consistent with this postulation, the study by Johansen et al. [40] observed that in adhesive macrophages, SIRP α exists as a tyrosine phosphorylated protein, and the status of SIRP α modification is independent of extracellular CD47 ligation. Studies in other cell types, including kidney podocytes and melanoma cells, suggested that SIRP α maintains tyrosine phosphorylation in the cytoplasmic domain and association with SHP-(1/2); cell surface CD47 ligation, on the other hand, triggers SIRP α dephosphorylation and releases SHP proteins [4,41]. Perhaps the formation of SIRP α complexes in the lipid rafts of activated

monocytes and macrophages, which is a result of cytokine stimulation, requires SIRP α cytoplasmic ITIM phosphorylation and subsequent association with SHP (Figure 7C). Indeed, the cell surface crosslinking of activated monocytes and macrophages impeded not only SIRP α but also SHP-1 from its monomeric MW, suggesting that SHP-1 is also formed protein complexes. As the crosslinker is membrane-impermeable, the formation of SHP-1 complexes must be a result of intermolecular interactions with transmembrane proteins such as SIRP α (Figure 7C). It is possible that well-assembled, multi-stranded SIRP α -SHP complexes in the lipid rafts facilitate high avidity and stable interactions with CD47 in activated monocytes or macrophages migrating in tissues; these structures also allow prompt and robust intracellular signals that prevent monocytes/macrophages from attacking encountered the self cells.

Supporting Information

Figure S1. Immunofluorescence staining of SIRP α in THP-1 and U937. SIRP α in THP-1 and U937 cells were labeled by anti-SIRP α .ex antibody under non-membrane permeable and membrane-permeable conditions. Labeling with isotype-matched IgG was performed as controls. Labeled cells were analyzed by FACS. (PDF)

Figure S2. Schematic depiction of monocyte transmigration. HMEC-1 cells were cultured on the transwell filter (with 5 μ m pore size) until confluency. For monocyte transmigration, freshly isolated PBMC were added to the upper

chamber of the setup and MCP-1 (50ng/ml) was added into the lower chamber. Following incubation at 37°C (2h), monocytes that transmigrated across the endothelial monolayer into the lower chamber were harvested and used for cell adhesion assays and other experiments. (PDF)

Figure S3. SIRP α -containing complexes are sensitive to Triton X-100 and other detergents. A) Peripheral PBMC, transmigrated monocytes, and BM-derived macrophages (BMDM) were lysed in cold lysis buffer containing 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 1% Triton X-100, protease inhibitors. After centrifugation, the supernatants were separated and labeled as soluble fractions (S). The pellets were further lysed in lysis buffer containing 0.5% SDS and designated as insoluble fractions (I). Soluble and insoluble fractions were analyzed by SDS-PAGE and WB by anti-SIRP α .ct and anti-Flotillin-1 antibodies. B) Same experiments were performed as in A except using Brij 58 as the detergent. (PDF)

Acknowledgements

The authors wish to thank Dr. Jill Leslie Littrell for critical reading and revision of the manuscript.

Author Contributions

Conceived and designed the experiments: YL. Performed the experiments: BH ZL ZB XZ AM. Analyzed the data: BH ZL ZB. Wrote the manuscript: YL.

References

- Barclay AN, Brown MH (2006) The SIRP family of receptors and immune regulation. *Nat Rev Immunol* 6: 457-464. doi:10.1038/nri1859. PubMed: 16691243.
- Matozaki T, Murata Y, Okazawa H, Ohnishi H (2009) Functions and molecular mechanisms of the CD47-SIRP α signalling pathway. *Trends Cell Biol* 19: 72-80. doi:10.1016/j.tcb.2008.12.001. PubMed: 19144521.
- Ohnishi H, Kaneko Y, Okazawa H, Miyashita M, Sato R et al. (2005) Differential localization of Src homology 2 domain-containing protein tyrosine phosphatase substrate-1 and CD47 and its molecular mechanisms in cultured hippocampal neurons. *J Neurosci* 25: 2702-2711. doi:10.1523/JNEUROSCI.5173-04.2005. PubMed: 15758180.
- Kurihara H, Harita Y, Ichimura K, Hattori S, Sakai T (2010) SIRP- α -CD47 system functions as an intercellular signal in the renal glomerulus. *Am J Physiol Renal Physiol* 299: F517-F527. doi:10.1152/ajprenal.00571.2009. PubMed: 20554646.
- Kharitonov A, Chen Z, Sures I, Wang H, Schilling J et al. (1997) A family of proteins that inhibit signalling through tyrosine kinase receptors. *Nature* 386: 181-186. doi:10.1038/386181a0. PubMed: 9062191.
- de Vries HE, Hendriks JJ, Honing H, De Lavellette CR, van der Pol SM et al. (2002) Signal-regulatory protein α -CD47 interactions are required for the transmigration of monocytes across cerebral endothelium. *J Immunol* 168: 5832-5839. PubMed: 12023387.
- Liu Y, Bühring HJ, Zen K, Burst SL, Schnell FJ et al. (2002) Signal regulatory protein (SIRP α), a cellular ligand for CD47, regulates neutrophil transmigration. *J Biol Chem* 277: 10028-10036. doi:10.1074/jbc.M109720200. PubMed: 11792697.
- Oldenborg PA, Zheleznyak A, Fang YF, Lagenaur CF, Gresham HD et al. (2000) Role of CD47 as a marker of self on red blood cells. *Science* 288: 2051-2054. doi:10.1126/science.288.5473.2051. PubMed: 10856220.
- Han X, Sterling H, Chen Y, Saginario C, Brown EJ et al. (2000) CD47, a ligand for the macrophage fusion receptor, participates in macrophage multinucleation. *J Biol Chem* 275: 37984-37992. doi:10.1074/jbc.M002334200. PubMed: 10964914.
- Okajo J, Kaneko Y, Murata Y, Tomizawa T, Okuzawa C et al. (2007) Regulation by Src homology 2 domain-containing protein tyrosine phosphatase substrate-1 of α -galactosylceramide-induced antimetastatic activity and Th1 and Th2 responses of NKT cells. *J Immunol* 178: 6164-6172. PubMed: 17475843.
- Fujioka Y, Matozaki T, Noguchi T, Iwamoto A, Yamao T et al. (1996) A novel membrane glycoprotein, SHPS-1, that binds the SH2-domain-containing protein tyrosine phosphatase SHP-2 in response to mitogens and cell adhesion. *Mol Cell Biol* 16: 6887-6899. PubMed: 8943344.
- Takada T, Matozaki T, Takeda H, Fukunaga K, Noguchi T et al. (1998) Roles of the complex formation of SHPS-1 with SHP-2 in insulin-stimulated mitogen-activated protein kinase activation. *J Biol Chem* 273: 9234-9242. doi:10.1074/jbc.273.15.9234. PubMed: 9535915.
- Kontaridis MI, Liu X, Zhang L, Bennett AM (2001) SHP-2 complex formation with the SHP-2 substrate-1 during C2C12 myogenesis. *J Cell Sci* 114: 2187-2198. PubMed: 11493654.
- Oh ES, Gu H, Saxton TM, Timms JF, Hausdorff S et al. (1999) Regulation of early events in integrin signaling by protein tyrosine phosphatase SHP-2. *Mol Cell Biol* 19: 3205-3215. PubMed: 10082587.
- Ochi F, Matozaki T, Noguchi T, Fujioka Y, Yamao T et al. (1997) Epidermal growth factor stimulates the tyrosine phosphorylation of SHPS-1 and association of SHPS-1 with SHP-2, a SH2 domain-containing protein tyrosine phosphatase. *Biochem Biophys Res Commun* 239: 483-487. doi:10.1006/bbrc.1997.7489. PubMed: 9344856.

16. Veillette A, Thibaudedeau E, Latour S (1998) High expression of inhibitory receptor SHPS-1 and its association with protein-tyrosine phosphatase SHP-1 in macrophages. *J Biol Chem* 273: 22719-22728. doi:10.1074/jbc.273.35.22719. PubMed: 9712903.
17. Ikeda H, Okazawa H, Ohnishi H, Murata Y, Oldenborg PA et al. (2006) Mutational analysis of the mechanism of negative regulation by SRC homology 2 domain-containing protein tyrosine phosphatase substrate-1 of phagocytosis in macrophages. *J Immunol* 177: 3123-3132. PubMed: 16920950.
18. Takizawa H, Manz MG (2007) Macrophage tolerance: CD47-SIRP α -mediated signals matter. *Nat Immunol* 8: 1287-1289. doi: 10.1038/ni1207-1287. PubMed: 18026079.
19. Kinchen JM, Ravichandran KS (2008) Phagocytic signaling: you can touch, but you can't eat. *Curr Biol* 18: R521-R524. doi:10.1016/j.cub.2008.04.058. PubMed: 18579095.
20. Ide K, Wang H, Tahara H, Liu J, Wang X et al. (2007) Role for CD47-SIRP α signaling in xenograft rejection by macrophages. *Proc Natl Acad Sci U S A* 104: 5062-5066. doi:10.1073/pnas.0609661104. PubMed: 17360380.
21. Mamtani M, Sharma M, Amin M, Amin A, Jawahirani A et al. (2007) Erythrophagocytosis in sickle cell anemia: statistical evidence for a biological phenomenon. *Med Hypotheses* 68: 1065-1070. doi:10.1016/j.mehy.2006.09.044. PubMed: 17097820.
22. Olsson M, Nilsson A, Oldenborg PA (2007) Dose-dependent inhibitory effect of CD47 in macrophage uptake of IgG-opsonized murine erythrocytes. *Biochem Biophys Res Commun* 352: 193-197. doi: 10.1016/j.bbrc.2006.11.002. PubMed: 17112468.
23. Willingham SB, Volkmer JP, Gentles AJ, Sahoo D, Dalerba P et al. (2012) The CD47-signal regulatory protein α (SIRP α) interaction is a therapeutic target for human solid tumors. *Proc Natl Acad Sci U S A* 109: 6662-6667. doi:10.1073/pnas.1121623109. PubMed: 22451913.
24. Jaiswal S, Jamieson CH, Pang WW, Park CY, Chao MP et al. (2009) CD47 is upregulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. *Cell* 138: 271-285. doi:10.1016/j.cell.2009.05.046. PubMed: 19632178.
25. Seiffert M, Cant C, Chen Z, Rappold I, Brugger W et al. (1999) Human signal-regulatory protein is expressed on normal, but not on subsets of leukemic myeloid cells and mediates cellular adhesion involving its counterreceptor CD47. *Blood* 94: 3633-3643. PubMed: 10572074.
26. Liu Y, Tong Q, Zhou Y, Lee HW, Yang JJ et al. (2007) Functional elements on SIRP α IgV domain mediate cell surface binding to CD47. *J Mol Biol* 365: 680-693. doi:10.1016/j.jmb.2006.09.079. PubMed: 17070842.
27. Vernon-Wilson EF, Kee WJ, Willis AC, Barclay AN, Simmons DL et al. (2000) CD47 is a ligand for rat macrophage membrane signal regulatory protein SIRP (OX41) and human SIRP α 1. *Eur J Immunol* 30: 2130-2137. doi:10.1002/1521-4141(2000)30:8. PubMed: 10940903.
28. Liu Y, Merlin D, Burst SL, Pochet M, Madara JL et al. (2001) The role of CD47 in neutrophil transmigration. Increased rate of migration correlates with increased cell surface expression of CD47. *J Biol Chem* 276: 40156-40166. PubMed: 11479293.
29. Ades EW, Candal FJ, Swerlick RA, George VG, Summers S et al. (1992) HMEC-1: establishment of an immortalized human microvascular endothelial cell line. *J Invest Dermatol* 99: 683-690. doi: 10.1111/1523-1747.ep12613748. PubMed: 1361507.
30. Roussel MF, Sherr CJ (1989) Mouse NIH 3T3 cells expressing human colony-stimulating factor 1 (CSF-1) receptors overgrow in serum-free medium containing human CSF-1 as their only growth factor. *Proc Natl Acad Sci U S A* 86: 7924-7927. doi:10.1073/pnas.86.20.7924. PubMed: 2554296.
31. Liu Y, Soto I, Tong Q, Chin A, Bühring HJ et al. (2005) SIRP β 1 is expressed as a disulfide-linked homodimer in leukocytes and positively regulates neutrophil transepithelial migration. *J Biol Chem* 280: 36132-36140. doi:10.1074/jbc.M506419200. PubMed: 16081415.
32. Patschan S, Li H, Brodsky S, Sullivan D, De Angelis DA et al. (2006) Probing lipid rafts with proximity imaging: actions of proatherogenic stimuli. *Am J Physiol Heart Circ Physiol* 290: H2210-H2219. doi: 10.1152/ajpheart.01112.2005. PubMed: 16361365.
33. Riff JD, Callahan JW, Sherman PM (2005) Cholesterol-enriched membrane microdomains are required for inducing host cell cytoskeleton rearrangements in response to attaching-effacing *Escherichia coli*. *Infect Immun* 73: 7113-7125. doi:10.1128/IAI.73.11.7113-7125.2005. PubMed: 16239505.
34. Oldenborg PA, Zheleznyak A, Fang YF, Lagenaur CF, Gresham HD et al. (2000) Role of CD47 as a marker of self on red blood cells [see comments]. *Science* 288: 2051-2054. doi:10.1126/science.288.5473.2051. PubMed: 10856220.
35. Morrow IC, Parton RG (2005) Flotillins and the PHB domain protein family: rafts, worms and anaesthetics. *Traffic* 6: 725-740. doi:10.1111/j.1600-0854.2005.00318.x. PubMed: 16101677.
36. Janes PW, Ley SC, Magee AI (1999) Aggregation of lipid rafts accompanies signaling via the T cell antigen receptor. *J Cell Biol* 147: 447-461. doi:10.1083/jcb.147.2.447. PubMed: 10525547.
37. Field KA, Holowka D, Baird B (1997) Compartmentalized activation of the high affinity immunoglobulin E receptor within membrane domains. *J Biol Chem* 272: 4276-4280. doi:10.1074/jbc.272.7.4276. PubMed: 9020144.
38. Brown DA, London E (1998) Functions of lipid rafts in biological membranes. *Annu Rev Cell Dev Biol* 14: 111-136. doi:10.1146/annurev.cellbio.14.1.111. PubMed: 9891780.
39. Dart C (2010) Lipid microdomains and the regulation of ion channel function. *J Physiol* 588: 3169-3178. doi:10.1113/jphysiol.2010.191585. PubMed: 20519314.
40. Johansen ML, Brown EJ (2007) Dual regulation of SIRP α phosphorylation by integrins and CD47. *J Biol Chem* 282: 24219-24230. doi:10.1074/jbc.M701565200. PubMed: 17584740.
41. Motegi S, Okazawa H, Ohnishi H, Sato R, Kaneko Y et al. (2003) Role of the CD47-SHPS-1 system in regulation of cell migration. *EMBO J* 22: 2634-2644. doi:10.1093/emboj/cdg278. PubMed: 12773380.