

Check for updates

OPEN ACCESS

EDITED BY Francesco Schettini, Institut de Recerca Biomèdica August Pi i Sunyer (IDIBAPS), Spain

REVIEWED BY

Yutian Zou, Sun Yat-sen University Cancer Center (SYSUCC), China Qifeng Yang, Shandong University, China

*CORRESPONDENCE Jin Yang 1473106133@qq.com

[†]These authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Breast Cancer, a section of the journal Frontiers in Oncology

RECEIVED 04 June 2022 ACCEPTED 16 August 2022 PUBLISHED 08 September 2022

CITATION

Zhang M, Zhang J, Liu N, Wang B, Zhou Y and Yang J (2022) VISTA is associated with immune infiltration and predicts favorable prognosis in TNBC. *Front. Oncol.* 12:961374. doi: 10.3389/fonc.2022.961374

COPYRIGHT

© 2022 Zhang, Zhang, Liu, Wang, Zhou and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

VISTA is associated with immune infiltration and predicts favorable prognosis in TNBC

Mi Zhang[†], Juan Zhang[†], Na Liu, Biyuan Wang, Yan Zhou and Jin Yang^{*}

Department of Medical Oncology, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an, China

Background: V-domain Ig-containing suppressor of T cell activation (VISTA), a critical immune checkpoint protein, can regulate the immune system. Nevertheless, little information is available on the expression level of VISTA and its clinical significance as well. The immunological and prognostic role of VISTA in triple-negative breast cancer (TNBC) still remains unclear.

Methods: The clinical significance and expression of VISTA in TNBC were examined using RNA sequencing and clinical data. Cancer single-cell state atlas (CancerSEA), gene set enrichment analyses (GSEA), single sample GSEA, ESTIMATE algorithm, immunohistochemistry (IHC) were utilized to assess the functions of VISTA.

Results: VISTA was down-regulated and closely associated with good prognosis in TNBC. The expression of VISTA was higher in Immunity-H group and immunomodulatory (IM) subtype. The level of VISTA expression in TNBC gradually increased with the degree of stromal tumor infiltrating lymphocytes (sTILs) infiltration. In addition, the high expression of VISTA was strongly linked to higher proportion of CD8 (+) T cell and M1 macrophages.

Conclusion: VISTA was remarkably correlated with a favorable prognosis and high immune infiltration in patients with TNBC.

KEYWORDS

VISTA, triple-negative breast cancer (TNBC), prognosis, immune infiltration, M1 macrophages

Introduction

Breast cancer (BC) is the most common cancer in women worldwide (1). Triple-negative breast cancer (TNBC) is histologically defined by the absence of the expression of estrogen receptor, progesterone receptor, and epidermal growth factor receptor-2 (HER-2). TNBC is a highly aggressive subtype of BC with a high incidence of local recurrence and metastasis. It accounts for approximately 15% of all types of BC (2). Therefore, in the last decades, endocrine therapy and anti-HER-2 therapy were not recommended, chemotherapy was the only available systemic treatment option for TNBC patients.

In recent years, immunotherapies based on immune checkpoint-blocking antibodies have achieved some success. TNBC is the most immunogenic subtype of BC. The introduction of immunotherapy into the treatment of TNBC has produced survival benefits. Programmed cell death ligand 1 (PD-L1) blocking antibody (atezolizumab) has been approved by European Medicines Agency and Food and Drug Administration for PD-L1 positive metastatic TNBC (3), because the first-line treatment with atezolizumab plus nabpaclitaxel can improve survival. Nevertheless, the response rate to immune checkpoint inhibitors (ICIs) is low. In the United States, less than 13% of patients with cancer benefit from the immunotherapy (4). In metastatic TNBC, the objective response rates (ORR) of ICIs, as a monotherapy, do not exceed 24% (5). Additionally, some cancer patients with initial response to ICIs would develop acquired resistance later (6). Despite receiving continued therapy, about one-quarter to one-third of metastatic melanoma patients who have objective response to ICIs therapies eventually relapsed (7). Based on above reasons, the sustained benefits of immunotherapy have been limited to a small number of patients. Hence, there is an urgent need to explore new targets.

As a type I immunoglobulin membrane protein, V-domain Ig-containing suppressor of T cell activation (VISTA) exerts immunosuppressive activities on T cells and plays a crucial role in the regulation of antitumor immunity (8-11). Relevant studies have shown that VISTA inhibits T cell activation through T cell-extrinsic and intrinsic mechanisms. On the one hand, VISTA expressed on antigen presenting cells (APCs) may act as a ligand that binds to inhibitory receptors. On the other hand, VISTA expressed on T cells may act as a receptor that transmits inhibitory signals (12). Although both VISTA and programmed cell death receptor 1 (PD-1) are immune checkpoint proteins that inhibit T cell activation, VISTA and PD-L1/PD-1 pathways independently control tumor-specific Tcell responses (13). Blocking VISTA and PD-L1 in murine tumor models can achieve synergistic therapeutic efficacy and enhance antitumor responses. Besides, the combined inhibition of cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) and VISTA is more efficacious, and could increase CD8/Treg and

Tcon/Treg ratios in the tumor microenvironment (TME) (14). In conclusion, more and more evidence indicated that VISTA is a new emerging target in cancer immunotherapy.

Many studies have proved that VISTA is a potential prognostic factor, while their conclusions are controversial. Villarroel et al. revealed that the high level of VISTA measured in the tumor area was significantly associated with better 5-year overall survival (OS) in non-small-cell lung cancer (NSCLC) (15). Coincidentally, Loeser et al. found that VISTA positive tumors showed a major survival advantage in early-stage esophageal adenocarcinoma (pT1/2) compared with VISTA negative tumors (16). In contrast, Kuklinski et al. suggested that high VISTA expression was remarkably correlated with worse disease-specific survival (DSS) in primary cutaneous melanoma (17). To date, there are few studies on VISTA in TNBC. The clinical significance of VISTA, and its potential role and mechanism in tumor immune infiltration in patients with TNBC still remain elusive.

In this study, multiple databases were used to examine the expression, clinical correlates, and prognostic significance of VISTA in TNBC patients. Moreover, gene set enrichment analyses (GSEA), single sample GSEA, ESTIMATE algorithm, immunohistochemistry (IHC) and multiplex immunofluorescence (mIF) were utilized to explore the potential biological functions of VISTA. Our study aimed to shed light on the role of VISTA in prognosis and tumor immune infiltration of TNBC.

Materials and methods

Data acquisition

TNBC and para-carcinoma tissue datasets were acquired from The Cancer Genome Atlas (TCGA) (https://portal.gdc. cancer.gov/). A total of 11 TNBC samples from TCGA were included in our study. RNA sequencing and clinical data were obtained from 359 TNBC patients collected at the Fudan University Shanghai Cancer Center (FUSCC) (FUSCC cohort) (18). The clinical data included age, gender, histology, tumor T stage, N stage and stromal tumor infiltrating lymphocytes (sTILs). The RNA sequencing data of 131 TNBC patients from GSE83937 (https://www.ncbi.nlm.nih.gov/geo/) were adopted to verify the main findings from the FUSCC cohort.

Kaplan-Meier plotter

The Kaplan-Meier plotter database (http://kmplot.com/ analysis) was used to evaluate the prognostic value of VISTA expression in TNBC.

Breast cancer integrative platform

The BCIP (http://www.omicsnet.org/bcancer/) is an integrative platform. The Molecular Taxonomy of Breast

Cancer International Consortium (METABRIC) database acquired from the BCIP was used to analyze the relationship between VISTA expression and OS in TNBC.

GSEA

In order to investigate biological function and potential signaling pathway of VISTA expression level in tumor tissues, GSEA was carried out using TPM of RNA sequencing data from the FUSCC cohort.

CancerSEA

CancerSEA (http://biocc.hrbmu.edu.cn/CancerSEA/home. jsp) involves 14 functional states of 41900 cancer single cells from 25 tumor types (19). In this work, it was used to evaluate the functional state of VISTA in BC.

CIBERSORT

CIBERSORT (20) is an algorithm to determine the possible proportion of immune cells in a sample based on gene expression profile.

Landscape of immune infiltration status in TNBC

The single sample GSEA was performed to quantify the relative abundance of each cell infiltration in the TNBC tumor immune microenvironment (TIME). The enrichment scores of all samples in two cohorts (FUSCC and Gene Expression Omnibus (GEO) cohorts) were calculated by ssGSEA analysis. Subsequently, immune characteristic clustering was evaluated using R package and sparcl. All samples were clustered into two subgroups: Immunity-H group with an abundance of immune associated sets. ESTIMATE algorithm was used to calculate the tumor purity and immune scores.

TIMER2.0

TIMER2.0 (http://timer.cistrome.org/) (21) was utilized to investigate the correlations between VISTA expression and tumor purity, CD8 (+) T cell and M1 macrophage infiltration in basal-like BC.

IHC and mIF

The clinical data and tumor pathological tissue samples of 16 patients with TNBC diagnosed in the First Affiliated Hospital of Xi'an Jiaotong University from October 2015 to October 2020 were retrospectively analyzed. According to standard IHC

protocols, formalin-fixed paraffin-embedded (FFPE) tissue samples were immunohistochemically stained using VISTA antibody (D1L2G; dilution 1:200, Cell Signaling Technology, USA). The IHC scores of 0 to 4 were considered to be low expression of VISTA, while the scores of 5 to 12 were considered to be high expression of VISTA. Some markers were detected by mIF on PerkinElmer Mantra Quantitative Pathology Workstation/Quantitative Pathology Analysis Platform. The antibodies used included anti-panCK, anti-HLA-DR and anti-CD68. Previous studies used CD68 and HLA-DR for M1 identification (22–25). Hence, in this study, CD68 and HLA-DR double-positive cells were defined as M1 macrophages.

Statistical Analysis

Survival analysis was performed using Kaplan–Meier method. Kruskal-Wallis and Wilcoxon tests were conducted to compare the expression difference of VISTA. Spearman's or Pearson's test was carried out for correlation analysis. All statistical analyses were completed by R (version 3.6.3) and R packages (https://www.r-project.org/). P < 0.05 was regarded to be statistically significant. All statistical tests were two-sided.

Results

Clinical significance of VISTA in TNBC

To begin with, the data were downloaded from TCGA database to investigate the differences of the VISTA mRNA levels between paired tumor and normal tissues in TNBC. The results revealed that the VISTA levels in tumor tissues were significantly down-regulated (P = 0.0004; Figure 1A) compared with the adjacent normal tissues. Afterwards, the Kaplan-Meier Plotter database and METABRIC database were used to elucidate the relationship between VISTA expression and survival of TNBC patients. The results indicated that the high mRNA expression level of VISTA was significantly associated with better relapse-free survival (RFS) (hazard ratio (HR) = 0.63(0.46-0.86), log-rank P = 0.0033 for 225373_at; Figure 1B) and OS (HR = 0.58, P = 0.0015; Figure 1C). Furthermore, the correlation between VISTA expression and the clinical parameters of TNBC was examined using FUSCC cohort. The results illustrated that younger patients (≤ 60 years) expressed remarkably higher VISTA level than patients over 60 years of age (P = 0.0045), while VISTA expression did not differ significantly according to histology, tumor stage and nodal stage (Figures 1D-1G). Collectively, the above results demonstrated that VISTA may serve as a potential biomarker for predicting survival in TNBC.



based on TCGA database. (B) Survival curve using the Kaplan-Meier plotter is shown for RFS. (C) Kaplan-Meier survival curve of OS in the METABRIC database. Association of VISTA expression with clinicopathological characteristics, including age (D), histology (E), tumor stage (F) and nodal stage (G). IDC, invasive ductal carcinoma.

Functional characteristics of VISTA in TNBC

After determining the prognostic value of VISTA in TNBC, GSEA was further performed using the TPM of RNA sequencing data from the FUSCC cohort to evaluate biological function and potential signaling pathway of VISTA. The results of Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses indicated that multiple immune-related pathways were mainly enriched in VISTA high expression phenotype of TNBC, including chemokine signaling pathway (normalized enrichment score (NES) =2.52, P < 0.001; Figure 2A), B cell receptor (BCR) signaling pathway (NES=2.47, P < 0.001; Figure 2B), natural killer (NK) cell-mediated cytotoxicity (NES=2.42, P < 0.001; Figure 2C), T cell receptor (TCR) signaling pathway (NES=2.41, P < 0.001; Figure 2D) and cytokine-cytokine receptor interaction (CCI) pathways (NES=2.39, P < 0.001; Figure 2E), etc. The 63 positively correlated KEGG pathways (false discovery rate (FDR) q-value < 0.05, NES > 1) were shown in Supplementary Table 1. Moreover, CancerSEA was used to assess the functional state of VISTA in BC. The results showed that VISTA expression exhibited a positive correlation with inflammation (Correlation = 0.69, P < 0.001), differentiation (Correlation = 0.60, P < 0.001), angiogenesis (Correlation = 0.43, P < 0.05), quiescence (Correlation = 0.38, P < 0.05) and metastasis (Correlation = 0.37, P < 0.05) in BC (Supplementary Figure 1). The CIBERSORT algorithm was further used to calculate the distribution of 22 immune cells in each TNBC sample. The proportions of tumor immune infiltrating cell (TIIC) and the correlations between immune cells in FUSCC and GEO cohorts were illustrated in Figures 2F, G, respectively.

VISTA expression is correlated with immune infiltration in TNBC

Based on single sample GSEA algorithm, the enrichment scores of TNBC patients in FUSCC and GEO cohorts were calculated. The samples were classified into two subgroups with different immunity by unsupervised clustering. The Immunity_H group (FUSCC, 117 samples; GEO, 94 samples) was characterized with a high expression level of immunerelated sets, and Immunity_L group (FUSCC, 242 samples; GEO, 37 samples) was performed with a low expression level of immune-related sets. At the same time, the immune scores (ESTIMATEScore, ImmuneScore, StromalScore) and tumor purity between two subgroups were assessed to confirm our group assignment with ESTIMATE algorithm. After comparison, it indicated that the Immunity_H group obtained a significantly higher immune scores and a remarkably lower tumor purity than Immunity_L group. Above all, the immunity in Immunity_H group was stronger than that in Immunity_L group (Figures 3A, C). In order to further explore the immunity of VISTA, the expression level of VISTA between Immunity_H and Immunity_L group was compared. The results showed that the expression level of VISTA in Immunity_H group was significantly higher than that in Immunity_L group (Figures 3B, D). Moreover, as the degree of sTILs infiltration increased, VISTA expression level gradually increased in TNBC (Figure 3E). Additionally, VISTA expression level was notably higher in the immunomodulatory (IM) subtype compared with the basal-like immune-suppressed (BLIS) subtype (Figure 3F).

In additional, the immune cell infiltration differences between VISTA high and low groups were analyzed. The



results demonstrated that B cell memory (P < 0.001), T cell CD8 + (P < 0.001), T cell CD4 memory resting (P = 0.003), T cell CD4 memory activated (P < 0.001), T cell regulatory (Treg) (P = 0.015), monocytes (P < 0.001) and macrophage M1 (P < 0.001) exhibited a higher expression in the VISTA-high group, whereas B cells naive (P = 0.006), T cell follicular helper (P < 0.001), T cell gamma delta (P = 0.004), NK cells resting (P = 0.011), macrophage M0 (P < 0.001), mast cells resting (P = 0.013) and mast cells activated (P = 0.024) had a higher expression in the VISTA-low group in patients from the FUSCC cohort (Figure 4A). In order to ensure the accuracy of the results from the FUSCC cohort, the same analysis on the GEO cohort was performed. In agreement with the above results, T cell CD8+ (P = 0.004), T cell CD4 memory activated (P < 0.001) and macrophage M1 (P = 0.002) were upregulated, while macrophage M0 (P = 0.010) and mast cells activated (P <0.001) were downregulated in the VISTA-high group (Figure 4B). What's more, it was discovered VISTA expression

exhibited a positive correlation with M1 macrophage and CD8 (+) T cell (Figure 4C) infiltration in basal-like BC using TIMER2.0.

VISTA expression and the tumor immune microenvironment

The clinical samples in our hospital were collected. After obtaining institutional ethics committee approval and written informed consent, 16 TNBC patients were enrolled (Supplementary Table 2). The correlation between VISTA and immune cell infiltration was examined by IHC and mIF. The results reflected that the expression of VISTA in stromal cells (68.75%, 11/16) was higher than that in tumor cells (25%, 4/16). Only one case was positive for stromal cells and tumor cells, and one case was negative for both (Figures 5A, B). Most importantly, the high protein level of VISTA in stromal cells



was associated with higher infiltration of M1 macrophages in TNBC (P < 0.0001) (Figures 5C, D).

Discussion

TNBC is the most malignant subtype of BC, with limited treatment strategies, high incidence of tumor mutation burden (TMB) and tumor-infiltrating lymphocyte (TIL) rates. Phase III IMpassion130 study showed that atezolizumab combined with nab-paclitaxel improved the progression free survival (PFS) in patients with metastatic TNBC (3). Recent meta-analysis revealed that in metastatic BC, the pooled ORR of ICIs was

19% (95% CI = 12-27%), and TNBC had a relatively higher ORR (23%) than other BC subtypes (26). Although immunotherapy has achieved some success, only a few patients will benefit from it. Therefore, new immunotherapeutic targets are needed.

VISTA, a novel negative immune checkpoint, is a membrane protein with molecular weight of 55000 to 65000 daltons. It is highly conserved across different species, which suggested a conserved functional role (9). Previous study have found that TGF- β -induced Smad3 activation led to an increase in VISTA expression (27). In addition, hypoxia-inducible factor (HIF)-1 α binding to a conserved hypoxia response element in the VISTA promoter could also upregulate VISTA on myeloid cells (28). The true functional binding partner(s) for VISTA is



under investigation. At present, the potential ligands for VISTA include V-set and immunoglobulin domain 3 (VSIG-3), galectin-9 and so on (29). The engagement of VSIG-3 with VISTA can inhibit the proliferation of T cells and reduce their cytokine and chemokine release (30). Several studies have demonstrated that VISTA blockage has antitumour activity

both *in vitro* and *in vivo*. VISTA inhibition was observed to decelerate tumour growth and increase survival rates in mouse models (13). At present, there are some clinical trials on VISTA. NCT02671955 and NCT02812875 are phase I clinical trials. India CTRI/2017/12/011026 is a phase II open label randomized trial, and its results show that the clinical benefit rate of CA-170



(inhibit PD-L1/PD-L2/VISTA) is 59.5% (31). The role of VISTA in controlling T cell activation is different from the PD-1/PD-L1 pathway (13). VISTA is increased after ipilimumab therapy in patients with prostate cancer (32) and anti-PD-1 therapy in patients with metastatic melanoma (33). This implied negative immune checkpoint regulation by VISTA is an important potential mechanism of acquired immunotherapy resistance. Moreover, Liju Zong et al. demonstrated a positive association between the levels of VISTA protein and PD-1/PD-L1 in breast cancer specimens (34). We found that the expression of VISTA was also positively correlated with the expression of PD-1 (Supplementary Figure 2A) and PD-L1 (Supplementary Figure 2B) in basal-like BC using TIMER2.0. The expression, prognostic significance and function of VISTA have been demonstrated in multiple tumor types, including melanoma, pleural mesothelioma, NSCLC, BC, pancreatic cancer, colorectal cancer, ovarian and endometrial cancer, and so on (31). However, the prognostic value and immunological role of VISTA in TNBC have not been extensively studied.

In this study, the expression of VISTA in paired TNBC tissues and matched normal tissues was analyzed from TCGA. The results indicated that VISTA was lowly expressed in TNBC tissues. Besides, it was found that VISTA was more highly expressed in younger patients with TNBC. Most importantly,

survival curves revealed that VISTA was a good prognostic factor in TNBC, which was coincident with a previous report (35). A recent meta-analysis has also found that the high expression of VISTA was associated with significantly better OS (P < 0.001) (36). The above results suggested that VISTA could serve as a good prognostic factor for TNBC patients.

Subsequently, GSEA was performed using RNA sequencing data from the FUSCC cohort to evaluate the biological significance of VISTA in TNBC. The findings proved that among the top 10 statistically enriched KEGG pathways in TNBC patients with higher expression of VISTA, 5 pathways, including chemokine signaling pathway, BCR signaling pathway, NK cell-mediated cytotoxicity, TCR signaling pathway and CCI pathways, were related to immune function. In the relatively early stage of malignant tumor process, chemokines can induce lymphocyte infiltration, which may improve antitumor activity (37). For example, when CCR5 is expressed in T cells, it can enhance the antitumor response (38). BCR signaling plays a vital role at multiple checkpoints of B cell biology. BCR is necessary for B cells to correctly stimulate immune response (39). NK cells play a critical role in tumor immunosurveillance and tumor clearance. An 11-year follow-up survey conducted by Imai K illustrated that high NK cell cytotoxic activity was associated with reduced cancer risk (40). Tumor-infiltrating NK cells confer a positive prognostic value in colorectal carcinom, gastric carcinoma and squamous cell lung cancer (41). The recognition of cancer antigens by the TCR leads T cell activation, which inhibits tumor progression (42). CCI is closely related to immune reactions. Collectively, these results indicated that VISTA was closely related to immune infiltration in TNBC patients.

The Immunity_H group had a stronger immune signal. sTILs are the major immune defense against cancer cells, and associated with better prognosis in BC (43). A study conducted by Yi-Zhou Jiang classified TNBC into four transcriptome-based subtypes (1): luminal androgen receptor (2), IM, (3) BLIS, and (4) mesenchymal-like. Among them, IM subtype was characterized by high immune cell signaling. BLIS subtype was characterized by the down-regulation of immune response genes (18). In our study, it was revealed that the expression of VISTA was higher in the Immunity_H group and IM subtype. Moreover, higher expression of VISTA was remarkably associated with higher levels of sTILs. These findings suggested that VISTA expression was positively correlated with immunity in TNBC patients.

As we know, TIICs play a very vital role in tumor progression. CD8 (+) T cells have strong tumor killing ability. A study, involving 12,439 patients with BC, confirmed that CD8 (+) T cells were associated with good prognosis (44). Besides, several studies demonstrated that the expression of VISTA was positively correlated with CD8 (+) T cell infiltration. Xin-Lin He et al. retrospectively analyzed the medical records of 2440 patients. The results showed that the high expression of VISTA in solid tumours was strongly linked to better prognosis and high numbers of CD8 (+) TILs (P < 0.001) (36). Ming Zhang et al. also revealed there was a statistically significant positive association between VISTA and CD8 + mRNA expression in hepatocellular carcinoma, and VISTA+/CD8 + patients had a better OS (45). Similarly, our study found that CD8 (+) T cells exhibited a positive correlation with VISTA expression, which may partly explained why TNBC patients with high VISTA expression had a good prognosis.

Tumor associated macrophages (TAMs) are a major component of the TME. M0 macrophages are highly plastic. They can change their phenotype under the influence of environmental signals (46). Depending on the microenvironment, macrophages can mainly polarize into two functional phenotypes: classically activated macrophages (M1) and alternatively activated macrophages (M2) (47). M1 macrophages exert anti-tumor activity by releasing pro-inflammatory cytokines. The high density of M1 macrophages is associated with good prognosis in a variety of human malignancies, including NSCLC, HCC, ovarian and gastric cancers (48, 49). Besides, in the colorectal cancer, microsatellite instabilityhigh tumors had higher densities of M1 macrophages in tumor stroma. M1 polarization of tumor stromal macrophages was related to lower cancer specific mortality (50). In our study, compared with VISTA-low group, VISTA-high group experienced significantly higher proportions of macrophage M1, as well as remarkable lower proportions of macrophage M0. The results suggested that VISTAhigh group had a favorable immune microenvironment. IHC and mIF were further performed to validate the primary results that were revealed by bioinformatics analysis. The findings revealed VISTA level in stromal cells was positively correlated with M1 macrophages in TNBC, which was consistent with the above conclusion.

However, some potential limitations of our study should be noted. Our analysis showed that there was an association between the infiltration of CD8 (+) T cell and M1 macrophages and the expression of VISTA in TNBC, but the specific mechanism was unclear. Further studies are needed to verify the expression and function of VISTA *in vivo* and *in vitro*.

Conclusion

In conclusion, our analysis suggested that VISTA was a good prognostic factor, and its expression was positively correlated with immunity in TNBC. Most importantly, the high level of VISTA was remarkably associated with high infiltration of CD8 (+) T cell and M1 macrophages. The results revealed that VISTA could be served as a potential biomarker for prognostic prediction and immune infiltration in TNBC.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories or accession number(s) can be found below: https://portal.gdc.cancer.gov, The Cancer Genomic Atlas; https://www.ncbi.nlm.nih.gov/geo/, Gene Expression Omnibus, accession number GSE83937; http://kmplot. com/analysis, Kaplan-Meier plotter; http://www.omicsnet.org/ bcancer/, Breast cancer integrative platform; http://biocc.hrbmu. edu.cn/CancerSEA/home.jsp, Cancer single-cell state atlas; http:// timer.cistrome.org/, TIMER 2.0.

Ethics statement

Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Author contributions

Research design: MZ, JZ and JY. Data collection and drafting: MZ, NL and BW. Statistical analysis: MZ, JZ and YZ. Manuscript polishing: MZ and JZ. Building figures: MZ and JZ. Manuscript editing: MZ and JY. Manuscript revision: MZ, JZ and JY. All authors have read and approved the manuscript. All authors contributed to the article and approved the submitted version.

Acknowledgments

We would like to thank TCGA, GEO, CancerSEA, TIMER, METABRIC databases, and FUSCC for providing data.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the

References

1. Siegel RL, Miller KD. Cancer statistics, 2021. CA Cancer J Clin (2021) 71:7-33.

2. Rastelli F, Biancanelli S, Falzetta A, Martignetti A, Casi C, Bascioni R, et al. Triple-negative breast cancer: current state of the art. *Tumori* (2010) 96:875–88.

3. Schmid P, Rugo HS, Adams S, Schneeweiss A, Barrios CH, Iwata H, et al. Atezolizumab plus nab-paclitaxel as first-line treatment for unresectable, locally advanced or metastatic triple-negative breast cancer (IMpassion130): updated efficacy results from a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol* (2020) 21:44–59.

4. Haslam A, Prasad V. Estimation of the percentage of US patients with cancer who are eligible for and respond to checkpoint inhibitor immunotherapy drugs. *JAMA Netw Open* (2019) 2:e192535.

5. Kwa MJ, Adams S. Checkpoint inhibitors in triple-negative breast cancer (TNBC): Where to go from here. *Cancer* (2018) 124:2086–103.

6. Saleh R, Elkord E. Treg-mediated acquired resistance to immune checkpoint inhibitors. *Cancer Lett* (2019) 457:168–79.

7. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell* (2017) 168:707–23.

8. Hou Z, Pan Y, Fei Q, Lin Y, Zhou Y, Liu Y, et al. Prognostic significance and therapeutic potential of the immune checkpoint VISTA in pancreatic cancer. J Cancer Res Clin Oncol (2021) 147:517–31.

9. Wang L, Rubinstein R, Lines JL, Wasiuk A, Ahonen C, Guo Y, et al. VISTA, a novel mouse ig superfamily ligand that negatively regulates T cell responses. *J Exp Med* (2011) 208:577–92.

10. Lines JL, Pantazi E, Mak J, Sempere LF, Wang L, O'Connell S, et al. VISTA is an immune checkpoint molecule for human T cells. *Cancer Res* (2014) 74:1924–32.

11. Lines JL, Sempere LF, Broughton T, Wang L, Noelle R. VISTA is a novel broad-spectrum negative checkpoint regulator for cancer immunotherapy. *Cancer Immunol Res* (2014) 2:510–7.

12. Xu W, Hiếu T, Malarkannan S, Wang L. The structure, expression, and multifaceted role of immune-checkpoint protein VISTA as a critical regulator of anti-tumor immunity, autoimmunity, and inflammation. *Cell Mol Immunol* (2018) 15:438–46.

13. Liu J, Yuan Y, Chen W, Putra J, Suriawinata AA, Schenk AD, et al. Immunecheckpoint proteins VISTA and PD-1 nonredundantly regulate murine T-cell responses. *Proc Natl Acad Sci U.S.A.* (2015) 112:6682–7.

14. Kondo Y, Ohno T, Nishii N, Harada K, Yagita H, Azuma M. Differential contribution of three immune checkpoint (VISTA, CTLA-4, PD-1) pathways to antitumor responses against squamous cell carcinoma. *Oral Oncol* (2016) 57:54–60.

reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fonc.2022.961374/full#supplementary-material

SUPPLEMENTARY FIGURE 1

Functional relevance of VISTA in BC. (A) Correlations between VISTA expression and 14 functional states in BC single-cell datasets. No.cells refers to the number of cells. (B) VISTA expression exhibited a positive correlation with inflammation, differentiation, angiogenesis, quiescence and metastasis in BC (***P < 0.001, **P < 0.01, *P < 0.05).

SUPPLEMENTARY FIGURE 2

Correlation between VISTA expression and PD-1 (A) and PD-L1 (B) expression in basal-like BC.

15. Villarroel-Espindola F, Yu X, Datar I, Mani N, Sanmamed M, Velcheti V, et al. Spatially resolved and quantitative analysis of VISTA/PD-1H as a novel immunotherapy target in human non-small cell lung cancer. *Clin Cancer Res* (2018) 24:1562–73.

16. Loeser H, Kraemer M, Gebauer F, Bruns C, Schröder W, Zander T, et al. The expression of the immune checkpoint regulator VISTA correlates with improved overall survival in pT1/2 tumor stages in esophageal adenocarcinoma. *Oncoimmunology* (2019) 8:e1581546.

17. Kuklinski LF, Yan S, Li Z, Fisher JL, Cheng C, Noelle RJ, et al. VISTA expression on tumor-infiltrating inflammatory cells in primary cutaneous melanoma correlates with poor disease-specific survival. *Cancer Immunol Immunother* (2018) 67:1113–21.

18. Jiang YZ, Ma D, Suo C, Shi J, Xue M, Hu X, et al. Genomic and transcriptomic landscape of triple-negative breast cancers: Subtypes and treatment strategies. *Cancer Cell* (2019) 35:428–40.e5.

19. Yuan H, Yan M, Zhang G, Liu W, Deng C, Liao G, et al. CancerSEA: A cancer single-cell state atlas. *Nucleic Acids Res* (2019) 47:D900–d908.

20. Aran D, Hu Z, Butte AJ. xCell: digitally portraying the tissue cellular heterogeneity landscape. *Genome Biol* (2017) 18:220.

21. Li T, Fu J, Zeng Z, Cohen D, Li J, Chen Q, et al. TIMER2.0 for analysis of tumor-infiltrating immune cells. *Nucleic Acids Res* (2020) 48:W509–514.

22. Zong Z, Zou J, Mao R, Ma C, Li N, Wang J, et al. M1 macrophages induce PD-L1 expression in hepatocellular carcinoma cells through IL-1 β signaling. Front Immunol (2019) 10:1643.

23. Marques P, Barry S, Carlsen E, Collier D, Ronaldson A, Dorward N, et al. The role of the tumour microenvironment in the angiogenesis of pituitary tumours. *Endocrine* (2020) 70:593–606.

24. Rakaee M, Busund LR, Jamaly S, Paulsen EE, Richardsen E, Andersen S, et al. Prognostic value of macrophage phenotypes in resectable non-small cell lung cancer assessed by multiplex immunohistochemistry. *Neoplasia* (2019) 21:282–93.

25. Marques P, Barry S, Carlsen E, Collier D, Ronaldson A, Grieve J, et al. The expression of neural cell adhesion molecule and the microenvironment of pituitary neuroendocrine tumours. *Neuroendocrinol* (2021) 33:e13052.

26. Zou Y, Zou X, Zheng S, Tang H, Zhang L, Liu P, et al. Efficacy and predictive factors of immune checkpoint inhibitors in metastatic breast cancer: a systematic review and meta-analysis. *Ther Adv Med Oncol* (2020) 12:1758835920940928.

27. Schlichtner S, Yasinska IM, Ruggiero S, Berger SM, Aliu N, Prunk M, et al. Expression of the immune checkpoint protein VISTA is differentially regulated by the TGF- β 1 - Smad3 signaling pathway in rapidly proliferating human cells and T lymphocytes. *Front Med (Lausanne)* (2022) 9:790995.

28. Deng J, Li J, Sarde A, Lines JL, Lee YC, Qian DC, et al. Hypoxia-induced VISTA promotes the suppressive function of myeloid-derived suppressor cells in the tumor microenvironment. *Cancer Immunol Res* (2019) 7:1079–90.

29. Yasinska IM, Meyer NH, Schlichtner S, Hussain R, Siligardi G, Casely-Hayford M, et al. Ligand-receptor interactions of galectin-9 and VISTA suppress human T lymphocyte cytotoxic activity. *Front Immunol* (2020) 11:580557.

30. Wang J, Wu G, Manick B, Hernandez V, Renelt M, Erickson C, et al. VSIG-3 as a ligand of VISTA inhibits human T-cell function. *Immunology* (2019) 156:74–85.

31. Tagliamento M, Agostinetto E, Borea R. VISTA: A promising target for cancer immunotherapy? *Immunotargets Ther* (2021) 10:185–200.

32. Gao J, Ward JF, Pettaway CA, Shi LZ, Subudhi SK, Vence LM, et al. VISTA is an inhibitory immune checkpoint that is increased after ipilimumab therapy in patients with prostate cancer. *Nat Med* (2017) 23:551–5.

33. Kakavand H, Jackett LA, Menzies AM, Gide TN, Carlino MS, Saw RPM, et al. Negative immune checkpoint regulation by VISTA: a mechanism of acquired resistance to anti-PD-1 therapy in metastatic melanoma patients. *Mod Pathol* (2017) 30:1666–76.

34. Zong L, Mo S, Yu S. Expression of the immune checkpoint VISTA in breast cancer. *Immunol Immunother* (2020) 69:1437–46.

35. Cao X, Ren X, Zhou Y, Mao F, Lin Y, Wu H, et al. VISTA expression on immune cells correlates with favorable prognosis in patients with triple-negative breast cancer. *Front Oncol* (2020) 10:583966.

36. He XL, Zhou Y, Lu HZ, Li QX, Wang Z. Prognostic value of VISTA in solid tumours: a systematic review and meta-analysis. *Sci Rep* (2020) 10:2662.

37. Morein D, Erlichman N, Ben-Baruch A. Beyond cell motility: The expanding roles of chemokines and their receptors in malignancy. *Front Immunol* (2020) 11:952.

38. Mollica Poeta V, Massara M, Capucetti A, Bonecchi R. Chemokines and chemokine receptors: New targets for cancer immunotherapy. *Front Immunol* (2019) 10:379.

39. Tanaka S. Baba y. B Cell Receptor Signaling Adv Exp Med Biol (2020) 1254:23-36.

40. Imai K, Matsuyama S, Miyake S, Suga K, Nakachi K. Natural cytotoxic activity of peripheral-blood lymphocytes and cancer incidence: an 11-year followup study of a general population. *Lancet* (2000) 356:1795–9.

41. Abel AM, Yang C, Thakar MS, Malarkannan S. Natural killer cells: Development, maturation, and clinical utilization. *Front Immunol* (2018) 9:1869.

42. Zhao X, Kolawole EM. Tuning T cell receptor sensitivity through catch bond engineering. *Science* (2022) 376:eabl5282.

43. Pang J, Zhou H, Dong X, Wang S, Xiao Z. Relationship between the neutrophil to lymphocyte ratio, stromal tumor-infiltrating lymphocytes, and the prognosis and response to neoadjuvant chemotherapy in triple-negative breast cancer. *Clin Breast Cancer* (2021) 21:e681–7.

44. Ali HR, Provenzano E, Dawson SJ, Blows FM, Liu B, Shah M, et al. Association between CD8+ T-cell infiltration and breast cancer survival in 12,439 patients. *Ann Oncol* (2014) 25:1536–43.

45. Zhang M, Pang HJ, Zhao W, Li YF, Yan LX, Dong ZY, et al. VISTA expression associated with CD8 confers a favorable immune microenvironment and better overall survival in hepatocellular carcinoma. *BMC Cancer* (2018) 18:511.

46. Qiu SQ, Waaijer SJH, Zwager MC, de Vries EGE, van der Vegt B, Schröder CP. Tumor-associated macrophages in breast cancer: Innocent bystander or important player? *Cancer Treat Rev* (2018) 70:178–89.

47. Yang Q, Guo N, Zhou Y, Chen J, Wei Q, Han M. The role of tumorassociated macrophages (TAMs) in tumor progression and relevant advance in targeted therapy. *Acta Pharm Sin B* (2020) 10:2156–70.

48. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* (2010) 141:39–51.

49. Fridman WH, Zitvogel L, Sautès-Fridman C, Kroemer G. The immune contexture in cancer prognosis and treatment. *Nat Rev Clin Oncol* (2017) 14:717–34.

50. Väyrynen JP, Haruki K. The prognostic role of macrophage polarization in the colorectal cancer microenvironment. *Cancer Immunol Res* (2021) 9:8–19.