

## ORIGINAL RESEARCH

# Acute tetrahydrobiopterin supplementation attenuates sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle of healthy rats

Nicholas G. Jendzjowsky, Timothy P. Just, Kelvin E. Jones &amp; Darren S. DeLorey

Faculty of Physical Education and Recreation, University of Alberta, Edmonton, Alberta, Canada

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**Correspondence**

Darren S. DeLorey, Faculty of Physical Education and Recreation, University of Alberta, E-435 Van Vliet Centre, Edmonton, AB, Canada T6G 2H9.  
Tel: (780)-492-0157  
Fax: (780)-492-2364  
E-mail: darren.delorey@ualberta.ca

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## Introduction

Skeletal muscle blood flow is regulated by a balance between sympathetic nervous system mediated vasoconstriction and local vasodilation (Saltin et al. 1998). Substances released from muscle tissue and/or the

**Abstract**

Tetrahydrobiopterin (BH<sub>4</sub>) is an essential cofactor for the production of nitric oxide (NO) and supplementation with BH<sub>4</sub> improves NO-dependent vasodilation. NO also reduces sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle. Thus, we hypothesized that supplementation with BH<sub>4</sub> would blunt sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle. Sprague-Dawley rats ( $n = 15$ ,  $399 \pm 57$  g) were anesthetized and instrumented with an indwelling brachial artery catheter, femoral artery flow probe, and a stimulating electrode on the lumbar sympathetic chain. Triceps surae muscles were stimulated to contract rhythmically at 30% and 60% of maximal contractile force (MCF). The percentage change of femoral vascular conductance (%FVC) in response to sympathetic stimulations delivered at 2 and 5 Hz was determined at rest and during muscle contraction in control and acute BH<sub>4</sub> supplementation (20 mg·kg<sup>-1</sup> + 10 mg·kg<sup>-1</sup>·h<sup>-1</sup>, IA) conditions. BH<sub>4</sub> reduced ( $P < 0.05$ ) the vasoconstrictor response to sympathetic stimulation (i.e., decrease in FVC) at rest (Control: 2 Hz:  $-28 \pm 5\%$ FVC; 5 Hz:  $-45 \pm 5\%$ ; BH<sub>4</sub>: 2 Hz:  $-17 \pm 4\%$ FVC; 5 Hz:  $-34 \pm 7\%$ FVC) and during muscular contraction at 30% MCF (Control: 2 Hz:  $-14 \pm 6\%$ FVC; 5 Hz:  $-28 \pm 11\%$ ; BH<sub>4</sub>: 2 Hz:  $-6 \pm 6\%$ FVC; 5 Hz:  $-16 \pm 10\%$ ) and 60% MCF (Control: 2 Hz:  $-7 \pm 3\%$ FVC; 5 Hz:  $-16 \pm 6\%$ FVC; BH<sub>4</sub>: 2 Hz:  $-2 \pm 3\%$ FVC; 5 Hz:  $-11 \pm 6\%$ FVC). These data are consistent with our hypothesis that acute BH<sub>4</sub> supplementation decreases sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle.

vascular endothelium can blunt sympathetic vasoconstriction and the inhibition of sympathetic vasoconstriction is an important mechanism for the matching of skeletal muscle blood flow to metabolic demand (DeLorey et al. 2002; VanTeuffelen and Segal 2003; Thomas and Segal 2004; Bagher and Segal 2011).

Indeed, a decline in the ability to blunt sympathetic vasoconstriction may increase vascular resistance and contribute to skeletal muscle hypoperfusion (Saltin and Mortensen 2012). Thus, interventions that improve the blunting of sympathetic vasoconstriction may improve the control of skeletal muscle vascular conductance.

The mechanism(s) responsible for the blunting of sympathetic vasoconstriction has not been definitively established; however, removal of the endothelium or pharmacological blockade of nitric oxide (NO) production enhances the constrictor response to sympathetic stimulation in resting and contracting skeletal muscle, demonstrating that NO inhibits vasoconstriction (Ohyanagi et al. 1992; Habler et al. 1997; Thomas and Victor 1998; Thomas et al. 1998, 2003; Sander et al. 2000; Jendzjowsky and DeLorey 2013b, 2013c). Our laboratory recently demonstrated that exercise training improved the inhibition of sympathetic vasoconstriction in resting and contracting muscle through a NO-dependent mechanism (Jendzjowsky and DeLorey 2013c).

Tetrahydrobiopterin (BH<sub>4</sub>) is an essential cofactor required for NO production by the NO synthase (NOS) group of enzymes (Katusic et al. 2009; Forstermann 2010; Forstermann and Sessa 2012). Decreased BH<sub>4</sub> availability has been associated with reduced NO-dependent vascular function (Beveris et al. 2006; Forstermann 2010; Forstermann and Sessa 2012; Santhanam et al. 2012). In contrast, supplementation with exogenous BH<sub>4</sub> has been shown to improve NO-dependent vasodilation in healthy individuals and to restore ACh and flow-mediated vasodilation in populations with endothelial dysfunction (Higashi et al. 2002; Eskurza et al. 2005; Cosentino et al. 2008).

Exogenous BH<sub>4</sub> treatment may also augment NO-mediated inhibition of sympathetic vasoconstriction in the skeletal muscle vasculature. However, to date, studies of the effects of acute BH<sub>4</sub> supplementation on skeletal muscle vascular control have focused on the effects of BH<sub>4</sub> on endothelium-dependent vasodilation (Gruhn et al. 2001; Higashi et al. 2002; Delp et al. 2008; Sindler et al. 2009). Whether acute BH<sub>4</sub> supplementation can enhance the inhibition of sympathetic vasoconstriction in resting and contracting skeletal muscle has not been investigated.

Therefore, the purpose of this study was to determine the effect of acute BH<sub>4</sub> supplementation on sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle of healthy rats. It was hypothesized that acute BH<sub>4</sub> supplementation would reduce sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle.

## Methods

### Animals and animal care

Male Sprague-Dawley rats ( $n = 15$ ;  $399 \pm 57$  g) were obtained from the institutional breeding colony. Rats were housed in pairs in a 12:12-h light–dark cycle, environmentally controlled (22–24°C, 40–70% humidity) room. Water and rat chow (Lab Diet 5001, PMI Nutrition, Brentwood, MO) were provided ad libitum. All experiments were conducted in accordance with the Canadian Council on Animal Care Guidelines and Policies with approval from the Animal Care and Use Committee: Health Sciences for the University of Alberta.

### Instrumentation

Anesthesia was induced by inhalation of isoflurane (3–3.5%, balance O<sub>2</sub>). During isoflurane anesthesia, the right jugular vein was cannulated and anesthesia was subsequently maintained by infusion of  $\alpha$ -chloralose (8–16 mg·kg<sup>-1</sup>·h<sup>-1</sup>) and urethane (50–100 mg·kg<sup>-1</sup>·h<sup>-1</sup>). The depth of anesthesia was assessed by the stability of arterial blood pressure, heart rate (HR), and the absence of a withdrawal reflex in response to a painful stimulus (i.e., paw-pinch). A tracheotomy was performed to allow spontaneous breathing of room air. We have previously demonstrated that arterial blood gases and acid base status are well maintained at rest and during muscle contraction in this preparation (Jendzjowsky and DeLorey 2013c). Thus, arterial blood gases were checked periodically in these experiments to confirm the maintenance of blood gas and acid base status. Arterial blood pressure was measured by a pressure transducer (Abbott, North Chicago, IL) that was attached to a cannula implanted in the left brachial artery. HR was derived from the blood pressure waveform and mean arterial pressure (MAP) was calculated. The left femoral artery and vein were cannulated for the delivery of solutions and blood sampling. Blood flow was measured using a transit-time flow probe (0.7 V Transonic Systems, Ithaca, NY) placed around the right femoral artery and connected to a flowmeter (T106 Transonic Systems). Core temperature was monitored by rectal probe and maintained at 36–37°C by external heating pad (Physitemp, TCAT-2, Clifton, NJ). Upon completion of all experiments, animals were euthanized by an overdose of the  $\alpha$ -chloralose and urethane anesthetic.

### Muscle contraction

The right sciatic nerve was exposed and instrumented with a nerve cuff electrode. The triceps surae muscle

group was dissected free and attached to a force transducer (Model MLT1030/D, AD Instruments, Colorado Springs, CO) via the calcaneal tendon. Maximal contractile force (MCF) was determined by stimulation of the triceps surae muscle group with 25, 1 ms impulses delivered at  $10 \times$  motor threshold (MT) at a frequency of 100 Hz. The optimal muscle length for tension development was determined by progressively lengthening the muscle and repeating the nerve stimulation until a plateau in tension (peak – baseline) was observed. Rhythmic contractions of the triceps surae muscles were produced at 30% (40 Hz 0.1 ms pulses in 250 ms trains at a rate of 60 trains per min at  $\sim 3 \times$  MT) and 60% MCF (40 Hz 0.1 ms pulses in 250 ms trains at a rate of 60 trains per min at  $\sim 7 \times$  MT).

### Lumbar sympathetic chain stimulation

A laparotomy was performed and the lumbar sympathetic chain distal to the renal branch of the aorta was dissected free with a blunt glass pipette. A bipolar silver-wire-stimulating electrode was attached to the lumbar sympathetic chain between L3 and L4. The electrodes were secured in place and electrically isolated by embedding them in a rapidly curing nontoxic silicone elastomer (Kwiksil, WPI, Sarasota, FL). An isolated constant-current stimulator (Digitimer DS3, Welwyn City, UK) was used to deliver 1 min of stimulation at frequencies of 2 and 5 Hz (1 ms, 1 mAmp) in random order.

### Experimental procedures

The skeletal muscle vasoconstrictor response evoked by stimulation of the lumbar sympathetic chain at 2 and 5 Hz was determined at rest and during muscle contraction at 30% and 60% of MCF. Sympathetic stimulations were delivered in random order in resting skeletal muscle with sufficient time ( $\sim 2$  min) allowed between stimulations to restore baseline hemodynamic values. Bouts of muscle contraction were 8 min in duration, completed in random order, and separated by 30 min of recovery. During each bout of muscle contraction, stimulations of the lumbar sympathetic chain were delivered 3 and 6 min after the onset of contraction.

Following an additional period of recovery ( $\sim 20$  min), a bolus of BH<sub>4</sub> (20 mg·kg<sup>-1</sup>) was injected intra-arterially and followed by continuous intra-arterial infusion (10 mg·kg<sup>-1</sup>·h<sup>-1</sup>) of BH<sub>4</sub> by syringe pump for the duration of the experimental protocol. After 20 min of treatment with BH<sub>4</sub>, lumbar sympathetic stimulations were repeated at rest and during skeletal muscle contraction at 30% and 60% of MCF in random order. We have previously demonstrated that muscle force production and the

cardiovascular response to sympathetic stimulation are not altered over time when bouts of contraction are repeated in this manner (Jendzjowsky and DeLorey 2013c).

### Effectiveness of BH<sub>4</sub> supplementation

To assess the effectiveness of BH<sub>4</sub> supplementation, the vasodilator response to acetylcholine (ACh; 0.1 μg) was measured prior to and following BH<sub>4</sub> treatment. Small volumes (100 μL) of ACh were injected over  $\sim 3$  sec in order to avoid flow-mediated vasodilation. Vehicle injections delivered in this manner did not alter femoral artery blood flow.

The BH<sub>4</sub> treatment regimen and dosing used in this study is consistent with previous investigations in rats (Gruhn *et al.* 2001; Noguchi *et al.* 2010, 2011) and preliminary experiments in our laboratory demonstrated that supplementation performed in this manner improved endothelium-dependent vasodilation (EDD) and did not alter resting MAP, femoral artery blood flow (FBF), and femoral vascular conductance (FVC).

### Plasma NO<sub>x</sub> measurement

Venous blood samples (0.6 mL) were collected in EDTA-containing tubes at rest ( $n = 10$ ) and during the final minute of muscle contraction at 60% MCF ( $n = 10$ ) in Control and BH<sub>4</sub> treatment conditions. Blood samples were immediately centrifuged at 13,000 g for 15 min at 4°C. The plasma was aliquoted, immediately frozen at  $-20^{\circ}\text{C}$ , and subsequently stored at  $-80^{\circ}\text{C}$  until analyzed. Plasma samples were defrosted and the concentration of nitrite+nitrate (NO<sub>x</sub>) was measured using a commercially available enzyme-linked immunoassay kit (Cayman No. 780001, Cayman Chemical Company, Ann Arbor, MI).

### Data analysis

Data were recorded using Chart data acquisition software (AD Instruments). Arterial blood pressure and FBF were sampled at 100 Hz and FVC was calculated as  $\text{FBF} \div \text{MAP}$  (mL·min<sup>-1</sup>·mmHg<sup>-1</sup>). Peak force production and fatigue index ( $([\text{peak force} - \text{end-contraction force}] \div \text{peak force}) \times 100$ ) were calculated for each contractile bout. The magnitude of the vasoconstrictor response to sympathetic stimulation was determined by calculation of the mean of the FVC response to sympathetic stimulation (1 min) and expressing it as a percentage change from FVC prior to the stimulation (1 min) in control and BH<sub>4</sub> conditions. The magnitude of the effect of BH<sub>4</sub> on sympathetic vasoconstriction at rest and during contraction

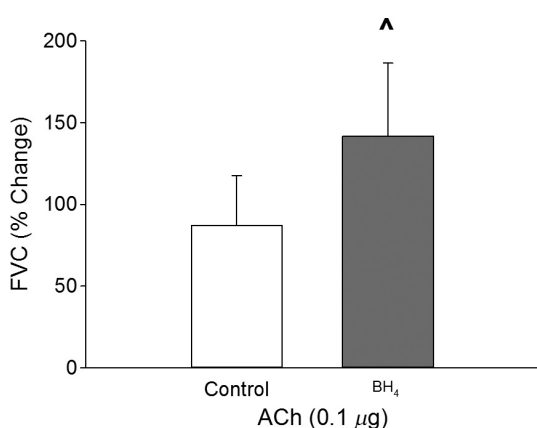
was determined by expressing the constrictor response in the BH<sub>4</sub> condition as a percentage of the constrictor response during control conditions. The response to ACh was calculated as the difference between the peak FVC response (~3 sec average) and the preinfusion baseline (~20 sec average) and expressed as a percentage change from the FVC baseline. All data are expressed as mean ± SD.

## Statistics

The effect of BH<sub>4</sub> supplementation on muscle contractile force, the vasoconstrictor response to sympathetic stimulation, and plasma NO<sub>x</sub> concentration was analyzed by two-way repeated measures ANOVA (drug condition × muscle contractile state). Vasodilator responses to ACh were analyzed by one-way repeated measures ANOVA. When significant F-ratios were detected, Student–Newman–Keuls post-hoc testing was completed. A *P*-value <0.05 was considered statistically significant.

## Results

BH<sub>4</sub> improved (*P* < 0.05) ACh-mediated vasodilation (Fig. 1). Resting HR was reduced (*P* < 0.05) by BH<sub>4</sub> supplementation, whereas MAP, FBF, and FVC were not different (*P* > 0.05) in control and BH<sub>4</sub> conditions (Table 1). Plasma NO<sub>x</sub> concentration was not different (*P* > 0.05) between control and BH<sub>4</sub> conditions at rest or during muscle contraction (Table 2).



**Figure 1.** Percentage change of femoral vascular conductance (FVC) in response to bolus injection of acetylcholine (ACh, 0.1 µg) during control conditions (open bars) and following acute BH<sub>4</sub> supplementation (20 mg·kg<sup>-1</sup> + 10 mg·kg<sup>-1</sup>·h<sup>-1</sup>, IA shaded bars). Values are mean ± SD. ^ indicates a statistically significant difference from the control condition. A *P*-value <0.05 was considered statistically significant.

## Effect of BH<sub>4</sub> on sympathetic vasoconstrictor responses in resting and contracting muscle

The response to sympathetic stimulation at rest and during muscle contraction in a representative rat is shown in Figure 2. BH<sub>4</sub> supplementation reduced (*P* < 0.05) the vasoconstrictor response to sympathetic stimulation delivered at 2 and 5 Hz in resting and contracting skeletal muscle. Compared to the control condition, BH<sub>4</sub> decreased the constrictor response by 40 ± 10% at the 2 Hz stimulation frequency and 26 ± 12% at the 5 Hz stimulation frequency in resting skeletal muscle (Fig. 3).

Muscle contraction reduced (*P* < 0.05) the vasoconstrictor response to sympathetic stimulation at 2 and 5 Hz in a contraction intensity-dependent manner in both control and BH<sub>4</sub> conditions (Table 3 and Fig. 3). During muscle contraction at 30% MCF, BH<sub>4</sub> treatment decreased the constrictor response to sympathetic stimulation at 2 and 5 Hz by 66 ± 48% and 45 ± 24%, respectively. At 60% MCF, BH<sub>4</sub> decreased the constrictor response by 64 ± 37% at the 2 Hz stimulation frequency and by 33 ± 17% at the 5 Hz stimulation frequency. The effect of BH<sub>4</sub> on sympathetic vasoconstriction was greater (*P* < 0.05) in contracting compared to resting muscle, but was not different (*P* > 0.05) between contraction intensities.

## Muscle force production and exercise hyperemia

Muscle force production was not different (*P* > 0.05) between control and BH<sub>4</sub> conditions at 30% MCF (Control: 587 ± 132 g; BH<sub>4</sub>: 561 ± 79 g) and 60% MCF (Control: 983 ± 147 g; BH<sub>4</sub>: 935 ± 157 g). Fatigue index was also not different (*P* > 0.05) between control (30% MCF: 35 ± 15%; 60% MCF: 43 ± 12%) and BH<sub>4</sub> conditions (30% MCF: 33 ± 12%; 60% MCF: 50 ± 13%). The hemodynamic response to muscle contraction at 30% and 60% MCF was not different (*P* > 0.05) between control and BH<sub>4</sub> conditions (Table 4).

## Discussion

The purpose of this investigation was to determine whether acute BH<sub>4</sub> supplementation would blunt sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle. Consistent with our hypothesis, sympathetic vasoconstrictor responsiveness was diminished in resting and contracting skeletal muscle following acute treatment with BH<sub>4</sub> in this study.

**Table 1.** Resting hemodynamics.

Drug condition	HR (beats·min <sup>-1</sup> )	MAP (mmHg)	FBF (mL·min <sup>-1</sup> )	FVC (mL·min <sup>-1</sup> ·mmHg <sup>-1</sup> )
Control	421 ± 35	91 ± 7	3.0 ± 0.7	0.033 ± 0.009
BH <sub>4</sub>	410 ± 11*	90 ± 12	2.6 ± 0.7	0.030 ± 0.007

Heart rate (HR), mean arterial blood pressure (MAP), femoral blood flow (FBF), and femoral vascular conductance (FVC) at rest and during acute tetrahydrobiopterin supplementation (BH<sub>4</sub>, 20 mg·kg<sup>-1</sup> + 10 mg·kg<sup>-1</sup>·h<sup>-1</sup>, IA). Values are mean ± SD.

\*Statistically significant difference from control condition. A *P*-value <0.05 was considered statistically significant.

BH<sub>4</sub> is an essential cofactor for NO production (Forstermann 2010; Forstermann and Sessa 2012) and is required for normal enzymatic function of all NOS isoforms (Katusic and d'Uscio 2004; Katusic et al. 2009). BH<sub>4</sub> acts as an allosteric enzyme that stabilizes the NOS complex and prevents uncoupling of NOS enzyme activity. Increased BH<sub>4</sub> availability may provide additional substrate for NOS enzymes or reduce the uncoupling of eNOS (Vasquez-Vivar et al. 2003; Bevers et al. 2006) and nNOS (Heinzel et al. 1992; Pou et al. 1992; Sun et al. 2008) which limits superoxide (O<sub>2</sub><sup>-</sup>) production, leads to de novo synthesis of NO, and increases NO bioavailability (Bevers et al. 2006; Forstermann 2010; Forstermann and Sessa 2012; Santhanam et al. 2012). The dose of BH<sub>4</sub> utilized in this study has been shown to increase circulating BH<sub>4</sub> concentrations and BH<sub>4</sub> content in skeletal muscle and isolated arteries (Gruhn et al. 2001; Delp et al. 2008; Noguchi et al. 2010, 2011). Functionally, BH<sub>4</sub> supplementation has been shown to increase flow-mediated NO production in isolated arteries (Sindler et al. 2009) and improve ACh- and flow-mediated vasodilation in adults with normal (Higashi et al. 2002) and reduced endothelial function (Higashi et al. 2002; Eskurza et al. 2005; Cosentino et al. 2008). Supplementation with BH<sub>4</sub> has also been shown to increase vascular compliance in older men (Pierce et al. 2012). Collectively, these studies indicate that acute BH<sub>4</sub> supplementation improves NO bioavailability and NO-mediated vascular function. Consistent with previous

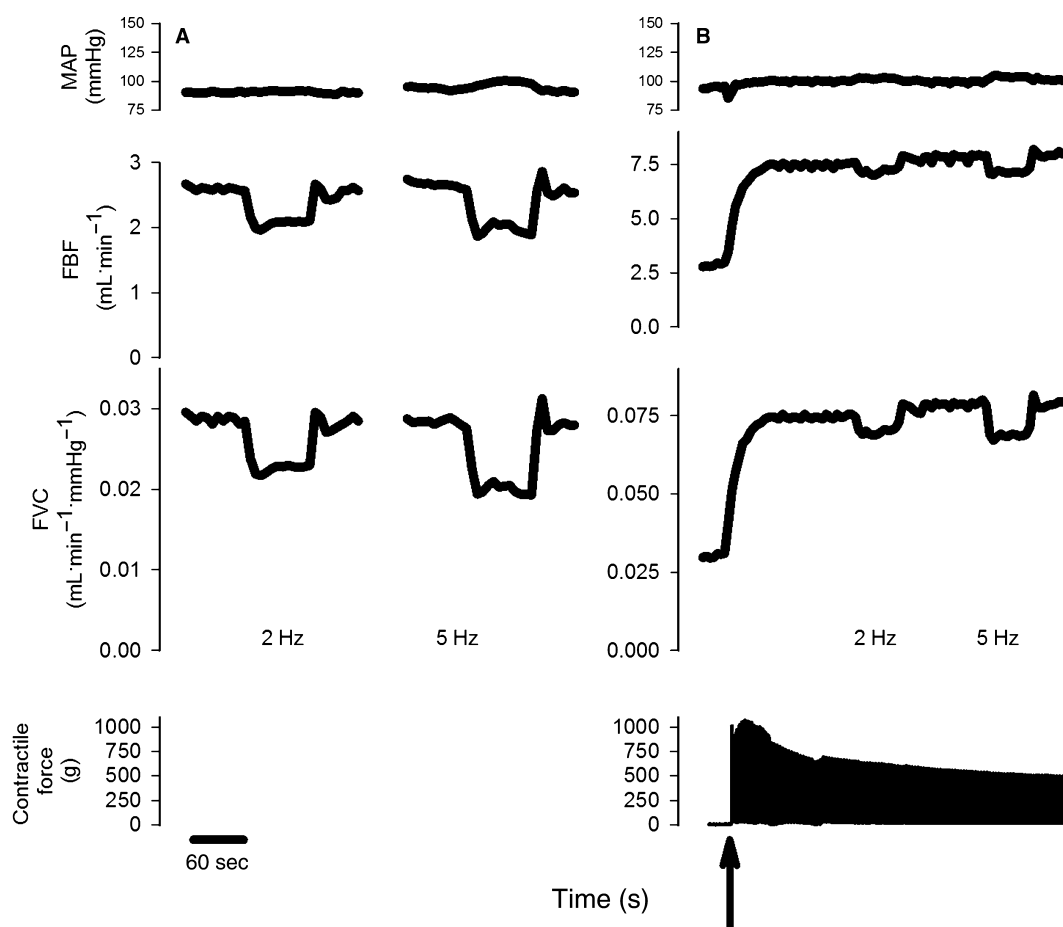
studies, acute treatment with BH<sub>4</sub> improved ACh-mediated vasodilation in this study. Despite improved ACh-mediated vasodilation, BH<sub>4</sub> supplementation did not result in an increase in plasma NO<sub>x</sub> at rest or during muscular contraction. The lack of an increase in plasma NO<sub>x</sub> is difficult to reconcile; however, it is possible that BH<sub>4</sub> supplementation improved tissue NO<sub>x</sub> content and not circulating NO<sub>x</sub> levels. Consistent with the present findings, supplementation with the NO precursor arginine had no effect on plasma NO<sub>x</sub> in humans and sedentary rats (Xiao et al. 2003; Forbes and Bell 2011; Forbes et al. 2013).

Sympathetic vasoconstrictor responsiveness was reduced following treatment with BH<sub>4</sub> in this study, suggesting that supplementation with BH<sub>4</sub> augmented NO-mediated inhibition of sympathetic vasoconstriction. Our laboratory and others have demonstrated that NO inhibits sympathetic vasoconstriction in resting and contracting skeletal muscle (Habler et al. 1997; Thomas and Victor 1998; Chavoshan et al. 2002; Donato et al. 2007; Behnke et al. 2011; Jendzjowsky and DeLorey 2013c). Indeed, NO derived from both eNOS and nNOS has been shown to inhibit sympathetic vasoconstriction in resting and contracting skeletal muscle (Ohyanagi et al. 1992; Habler et al. 1997; Thomas and Victor 1998; Thomas et al. 1998, 2003; Jendzjowsky and DeLorey 2013c). In resting skeletal muscle, NO-mediated inhibition of sympathetic vasoconstriction appears to be predominately mediated by NO derived from eNOS (Grange et al. 2001; Jendzjowsky and DeLorey 2013b). It is well established that BH<sub>4</sub> treatment reduces eNOS uncoupling and improves eNOS-mediated vascular function (Katusic and d'Uscio 2004; Katusic et al. 2009). Thus, the increased blunting of sympathetic vasoconstriction in resting skeletal muscle appears consistent with improved eNOS function following BH<sub>4</sub> supplementation. The augmented inhibition of sympathetic vasoconstriction following BH<sub>4</sub> supplementation suggests that eNOS activity could be a therapeutic target to blunt sympathetic vasoconstriction and reduce vascular resistance. This may be particularly effective in conditions where nNOS expression is reduced, such as muscular dystrophy, heart failure, etc. (Thomas et al. 1998, 2003;

**Table 2.** Plasma NO<sub>x</sub> concentration.

Muscle contraction status	Drug condition	NO <sub>x</sub> (μmol/L)
Rest ( <i>n</i> = 10)	Control	32 ± 16
	BH <sub>4</sub>	28 ± 5
60% MCF ( <i>n</i> = 10)	Control	27 ± 5
	BH <sub>4</sub>	28 ± 8

Plasma NO<sub>x</sub> concentration at rest and during contraction at 60% MCF in control conditions and following acute tetrahydrobiopterin supplementation (BH<sub>4</sub>, 20 mg·kg<sup>-1</sup> + 10 mg·kg<sup>-1</sup>·h<sup>-1</sup>, IA). Values are mean ± SD.

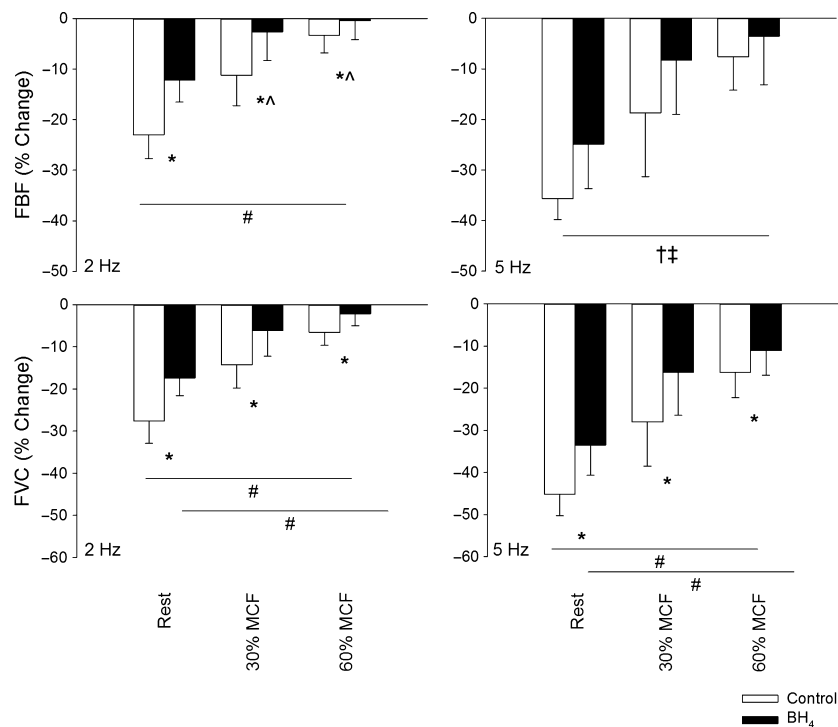


**Figure 2.** Original data from a representative animal illustrating the response of mean arterial blood pressure (MAP), femoral artery blood flow (FBF), femoral vascular conductance (FVC), and muscle contractile force at rest (Panel A) and during muscle contraction at 60% of maximal contractile force (Panel B). The arrow indicates the onset of contraction. Lumbar sympathetic nerve stimulations were delivered at 2 and 5 Hz in random order at rest and during contraction.

Sander *et al.* 2000; Vongpatanasin *et al.* 2011; Notarius *et al.* 2014).

During muscular contraction, increased endothelial shear stress and elevated intracellular [Ca<sup>++</sup>] lead to greater eNOS and nNOS-mediated NO production. In this study, the blunting of sympathetic vasoconstrictor responsiveness during BH<sub>4</sub> treatment was greater in contracting compared to resting skeletal muscle, suggesting that treatment with BH<sub>4</sub> may be particularly effective at increasing “stimulated” NO production. nNOS-derived NO appears to be particularly important for the inhibition of sympathetic vasoconstriction in contracting skeletal muscle (Thomas *et al.* 1998, 2003; Lau *et al.* 2000; Grange *et al.* 2001). The effects of BH<sub>4</sub> on nNOS-mediated vascular function are not well established and have received relatively little attention in the scientific literature. nNOS is primarily localized in the muscle sarcolemma and treatment with BH<sub>4</sub> has been shown to

increase BH<sub>4</sub> content in skeletal muscle cells (Harding *et al.* 2004). Thus, increased BH<sub>4</sub> availability may facilitate increased nNOS-mediated NO production in contracting muscle. However, the increased shear stress will increase eNOS-mediated NO production during exercise and may upregulate eNOS-mediated sympatholysis. Our laboratory recently reported that the contribution of NO derived from eNOS and nNOS to the inhibition of sympathetic vasoconstriction were proportional during skeletal muscle contraction (Jendzjowsky and DeLorey 2013b). This finding indicates that eNOS-derived NO is an important contributor to sympatholysis in healthy rats and suggests that BH<sub>4</sub> supplementation may upregulate eNOS-mediated sympatholysis. Further investigation is required to determine the specific NOS isoform responsible or the relative contribution of each isoform for the improved blunting of sympathetic vasoconstriction following BH<sub>4</sub> treatment. However, regardless of the specific



**Figure 3.** The percentage change of femoral artery blood flow (FBF) and femoral vascular conductance (FVC) in response to sympathetic stimulation at 2 Hz (left) and 5 Hz (right) (left panel) at rest, 30% and 60% maximal contractile force (MCF) during control conditions (open bars) and following BH<sub>4</sub> supplementation (BH<sub>4</sub>, 20 mg·kg<sup>-1</sup> + 10 mg·kg<sup>-1</sup>·h<sup>-1</sup>, IA; filled bars). Values are mean ± SD. \*indicates a significant difference between drug conditions (significant interaction). #indicates a significant difference between all muscle contractile conditions (significant interaction). ^indicates a significant difference from rest during BH<sub>4</sub> supplementation. †indicates a significant main effect of BH<sub>4</sub> supplementation. ‡indicates a significant main effect of contractile force (all contractile conditions different). A *P*-value <0.05 was considered statistically significant.

isoform responsible, the greater blunting of sympathetic vasoconstriction in contracting compared to resting muscle suggests that increased BH<sub>4</sub> availability may be particularly effective for “stimulated” NO production and that BH<sub>4</sub> supplementation may improve skeletal muscle vascular control in conditions where sympatholytic capacity may be diminished (e.g. aging, heart failure, etc.) (Lang et al. 1997; Dinunno et al. 2005; Vongpatanasin et al. 2011; Mortensen et al. 2012; Notarius et al. 2014).

BH<sub>4</sub> may have also reduced sympathetic vasoconstrictor responsiveness through its antioxidant properties. We and others (Zhao et al. 2006; Jendzjowsky and DeLorey 2013a) have shown that treatment with a superoxide scavenger reduces sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle. In vitro, BH<sub>4</sub> has been shown to scavenge O<sub>2</sub><sup>-</sup>, thiol radicals, and peroxynitrite (Heales et al. 1988; Gramsbergen et al. 2002; Kuzkaya et al. 2003; Katusic et al. 2009). However, in vivo, BH<sub>4</sub> does not appear to contribute to the scavenging of O<sub>2</sub><sup>-</sup> and treatment with tetrahydropterin, a BH<sub>4</sub> analogue that has antioxidant properties but does not alter NOS function, did not improve ACh-mediated

vasodilation in the forearm of smokers with reduced endothelial function (Heitzer et al. 2000; Vasquez-Vivar et al. 2002).

### Effect of BH<sub>4</sub> treatment on skeletal muscle blood flow

BH<sub>4</sub> reduced resting HR but did not alter resting MAP, FBF, and FVC, consistent with a previous study that reported no effect of BH<sub>4</sub> on resting limb blood flow (Eskurza et al. 2005). The limb blood flow response (increase in FBF and FVC) to muscle contraction was also not altered by treatment with BH<sub>4</sub>, despite enhanced EDD and greater inhibition of sympathetic vasoconstriction in this study. These findings appear contradictory; however, multiple signaling pathways regulate the hyperemic response to contraction in an integrative and redundant manner and blockade of and/or adaptations in an individual signaling pathway often do not impact the overall blood flow response to exercise (Laughlin and Korzick 2001). Indeed, to our knowledge, it has not been demonstrated that an improvement in EDD consistently

**Table 3.** Hemodynamic responses to sympathetic stimulation at rest and during muscle contraction.

Stimulation frequency	Contractile state	Drug condition	HR (beats·min <sup>-1</sup> )	MAP (mmHg)	FBF (mL·min <sup>-1</sup> )	FVC (mL·min <sup>-1</sup> ·mmHg <sup>-1</sup> )	
2 Hz	Rest	Control	-12 ± 10	6 ± 5	-0.7 ± 0.3	-0.0093 ± 0.0031	
		BH <sub>4</sub>	-4 ± 5*	6 ± 5	-0.3 ± 0.2*	-0.0053 ± 0.0020* <sup>§</sup>	
	30% MCF	Control	-7 ± 7	3 ± 3 <sup>†</sup>	-0.6 ± 0.4	-0.0089 ± 0.0036	
		BH <sub>4</sub>	-4 ± 13*	3 ± 4 <sup>†</sup>	-0.1 ± 0.3* <sup>^</sup>	-0.0037 ± 0.0045* <sup>§</sup>	
	60% MCF	Control	-5 ± 6 <sup>†</sup>	3 ± 3 <sup>†</sup>	-0.3 ± 0.3* <sup>^</sup>	-0.0052 ± 0.00323 <sup>^</sup> <sup>§</sup>	
		BH <sub>4</sub>	-1 ± 7* <sup>†</sup>	2 ± 2 <sup>†</sup>	-0.1 ± 0.3* <sup>^</sup>	-0.0021 ± 0.0022* <sup>§</sup>	
	5 Hz	Rest	Control	-21 ± 11	16 ± 7	-1.1 ± 0.3	-0.0152 ± 0.0049
			BH <sub>4</sub>	-9 ± 13*	12 ± 7*	-0.6 ± 0.2*	-0.0101 ± 0.0032*
30% MCF		Control	-14 ± 10 <sup>†</sup>	12 ± 4 <sup>†</sup>	-1.1 ± 0.7	-0.0182 ± 0.0078 <sup>^</sup>	
		BH <sub>4</sub>	-6 ± 9* <sup>†</sup>	9 ± 5* <sup>†</sup>	-0.5 ± 0.7*	-0.0104 ± 0.0080*	
60% MCF		Control	-14 ± 8 <sup>†</sup>	10 ± 4 <sup>†</sup>	-0.6 ± 0.4	-0.0132 ± 0.0047 <sup>#</sup>	
		BH <sub>4</sub>	-5 ± 10* <sup>†</sup>	7 ± 6* <sup>†</sup>	-0.3 ± 0.6* <sup>†</sup> <sup>‡</sup>	-0.0079 ± 0.0035*	

Absolute change in heart rate (HR), mean arterial blood pressure (MAP), femoral blood flow (FBF), and femoral vascular conductance (FVC) in response to 2 and 5 Hz sympathetic stimulation at rest and during contraction at 30% and 60% of maximal contractile force (MCF) during control conditions and following acute tetrahydrobiopterin supplementation (BH<sub>4</sub>, 20 mg·kg<sup>-1</sup> + 10 mg·kg<sup>-1</sup>·h<sup>-1</sup>, IA). Values are mean ± SD.

\*Statistically significant main effect of BH<sub>4</sub>.

<sup>†</sup>Statistically significant main effect of contractile force (different from rest).

<sup>‡</sup>Statistically significant main effect of contractile force (different from 30% MCF).

<sup>^</sup>Statistically significant difference from rest within specified drug condition (significant interaction).

<sup>#</sup>Statistically significant difference from 30% MCF within specified drug condition (significant interaction).

<sup>§</sup>Statistically significant difference between all contractile states within specified drug condition (significant interaction). A *P*-value <0.05 was considered statistically significant.

**Table 4.** Hemodynamic response to muscle contraction.

Muscle contraction	Drug condition	HR (beats·min <sup>-1</sup> )	MAP (mmHg)	FBF (mL·min <sup>-1</sup> )	FVC (mL·min <sup>-1</sup> ·mmHg <sup>-1</sup> )
30%	Control	10 ± 8	4 ± 6	3.1 ± 0.7	0.033 ± 0.009
	BH <sub>4</sub>	9 ± 13	3 ± 8	3.1 ± 1.1	0.035 ± 0.015
60%	Control	10 ± 6	6 ± 5	4.8 ± 0.9*	0.050 ± 0.010*
	BH <sub>4</sub>	8 ± 14	4 ± 4	4.3 ± 1.4*	0.047 ± 0.015*

Absolute increase of heart rate (HR), mean arterial pressure (MAP), femoral artery blood flow (FBF), and femoral vascular conductance (FVC) in response to muscle contraction at 30% and 60% of maximal contractile force during control conditions and following acute tetrahydrobiopterin supplementation (BH<sub>4</sub>, 20 mg·kg<sup>-1</sup> + 10 mg·kg<sup>-1</sup>·h<sup>-1</sup>, IA). Values are mean ± SD.

\*Significant main effect of contractile force (60% >30% MCF). A *P*-value <0.05 was considered statistically significant.

results in an augmented hyperemic response to exercise. Moreover, the contribution of NO to the regulation of bulk limb blood flow in response to muscle contraction remains controversial (Clifford and Hellsten 2004). Previous studies have reported that NO was not required for a “normal” hyperemic response to exercise (Radegran and Saltin 1999; Frandsenn et al. 2001), whereas others have demonstrated that exercise hyperemia was reduced following NOS inhibition (Hirai et al. 1994; Dietz et al. 1997). Recent evidence suggests that NO derived from nNOS may be involved in the distribution of muscle blood flow during exercise (Copp et al. 2013). It is well recognized that functional sympatholysis is necessary to oppose the increased sympathetic drive during exercise and contributes to the regulation of muscle blood flow and systemic

blood pressure (Buckwalter and Clifford 2001; Delp and O’Leary 2004; Thomas and Segal 2004). However, it has been shown that sympatholysis may facilitate the distribution of blood flow between and within muscles and not alter bulk limb blood flow during contraction (VanTeeffelen and Segal 2003). Consistent with this notion, our laboratory has recently reported that short-term exercise training enhanced NO-mediated inhibition of evoked sympathetic vasoconstriction, while the limb blood flow response to muscle contraction was not altered (Jendzjowsky and DeLorey 2013c). Thus, in this study, it is conceivable that the BH<sub>4</sub>-mediated improvements in EDD and inhibition of sympathetic vasoconstriction may have enhanced the distribution of blood flow between and within muscles at rest and during contraction.



## Experimental considerations and limitations

An additional experimental condition where NO production was inhibited following BH<sub>4</sub> treatment may have provided complementary evidence to the increase in EDD that the effects of BH<sub>4</sub> on sympathetic vasoconstriction were mediated by an NO-dependent mechanism. However, a NOS blockade condition would require a total of six bouts of muscle contraction in each rat. Preliminary experiments in our laboratory involving six bouts of contraction and recovery did not yield reproducible levels of muscle force production and constrictor responses to sympathetic stimulation during the final two bouts of contraction and therefore a NOS inhibition condition was not feasible.

BH<sub>4</sub> is also involved in the synthesis of norepinephrine (NE) in a reaction catalyzed by the enzyme tyrosine hydroxylase (Nagatsu 1983). In the cutaneous vascular bed, BH<sub>4</sub> supplementation has been shown to augment cold induced vasoconstriction without altering postsynaptic receptor function suggesting that the increased vasoconstriction was mediated by increased NE production (Lang *et al.* 2009, 2010). Our finding of reduced sympathetic vasoconstrictor responsiveness suggests that an increase in NE production following treatment with BH<sub>4</sub> was unlikely in this study.

## Conclusions

The current data demonstrate that acute BH<sub>4</sub> supplementation reduced sympathetic vasoconstrictor responsiveness in resting and contracting skeletal muscle. These findings in healthy rats suggest that BH<sub>4</sub> bioavailability may be an important factor in the inhibition of sympathetic vasoconstriction and that BH<sub>4</sub> supplementation may be an appropriate therapy in conditions characterized by elevated sympathetic vascular resistance and diminished sympatholysis. Further investigation is required to identify the cellular mechanism by which BH<sub>4</sub> supplementation reduces sympathetic vasoconstrictor responsiveness.

## Conflict of Interest

The authors have no conflict of interest to disclose.

## References

- Bagher, P., and S. S. Segal. 2011. Regulation of blood flow in the microcirculation: role of conducted vasodilation. *Acta Physiol. (Oxf.)* 202:271–284.
- Behnke, B. J., R. B. Armstrong, and M. D. Delp. 2011. Adrenergic control of vascular resistance varies in muscles composed of different fiber types: influence of the vascular

- endothelium. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 301:R783–R790.
- Beyers, L. M., B. Braam, J. A. Post, A. J. van Zonneveld, T. J. Rabelink, H. A. Koomans, *et al.* 2006. Tetrahydrobiopterin, but not L-arginine, decreases NO synthase uncoupling in cells expressing high levels of endothelial NO synthase. *Hypertension* 47:87–94.
- Buckwalter, J. B., and P. S. Clifford. 2001. The paradox of sympathetic vasoconstriction in exercising skeletal muscle. *Exerc. Sport Sci. Rev.* 29:159–163.
- Chavoshan, B., M. Sander, T. E. Sybert, J. Hansen, R. G. Victor, and G. D. Thomas. 2002. Nitric oxide-dependent modulation of sympathetic neural control of oxygenation in exercising human skeletal muscle. *J. Physiol.* 540:377–386.
- Clifford, P. S., and Y. Hellsten. 2004. Vasodilatory mechanisms in contracting skeletal muscle. *J. Appl. Physiol.* 97:393–403.
- Copp, S. W., C. T. Holdsworth, S. K. Ferguson, D. M. Hirai, D. C. Poole, and T. I. Musch. 2013. Muscle fibre-type dependence of neuronal nitric oxide synthase-mediated vascular control in the rat during high speed treadmill running. *J. Physiol.* 591:2885–2896.
- Cosentino, F., D. Hurlimann, C. Delli Gatti, R. Chenevard, N. Blau, N. J. Alp, *et al.* 2008. Chronic treatment with tetrahydrobiopterin reverses endothelial dysfunction and oxidative stress in hypercholesterolaemia. *Heart* 94:487–492.
- DeLorey, D. S., S. S. Wang, and J. K. Shoemaker. 2002. Evidence for sympatholysis at the onset of forearm exercise. *J. Appl. Physiol.* 93:555–560.
- Delp, M. D., and D. S. O’Leary. 2004. Integrative control of the skeletal muscle microcirculation in the maintenance of arterial pressure during exercise. *J. Appl. Physiol.* 97:1112–1118.
- Delp, M. D., B. J. Behnke, S. A. Spier, G. Wu, and J. M. Muller-Delp. 2008. Ageing diminishes endothelium-dependent vasodilatation and tetrahydrobiopterin content in rat skeletal muscle arterioles. *J. Physiol.* 586:1161–1168.
- Dietz, N. M., K. A. Engelke, T. T. Samuel, R. T. Fix, and M. J. Joyner. 1997. Evidence for nitric oxide-mediated sympathetic forearm vasodilatation in humans. *J. Physiol.* 498(Pt 2):531–540.
- Dinenno, F. A., S. Masuki, and M. J. Joyner. 2005. Impaired modulation of sympathetic alpha-adrenergic vasoconstriction in contracting forearm muscle of ageing men. *J. Physiol.* 567:311–321.
- Donato, A. J., L. A. Lesniewski, and M. D. Delp. 2007. Ageing and exercise training alter adrenergic vasomotor responses of rat skeletal muscle arterioles. *J. Physiol.* 579:115–125.
- Escurza, I., L. A. Myerburgh, Z. D. Kahn, and D. R. Seals. 2005. Tetrahydrobiopterin augments endothelium-dependent dilatation in sedentary but not in habitually exercising older adults. *J. Physiol.* 568:1057–1065.
- Forbes, S. C., and G. J. Bell. 2011. The acute effects of a low and high dose of oral L-arginine supplementation in young active males at rest. *Appl. Physiol. Nutr. Metab.* 36:405–411.

- Forbes, S. C., V. Harber, and G. J. Bell. 2013. The acute effects of L-arginine on hormonal and metabolic responses during submaximal exercise in trained cyclists. *Int. J. Sport Nutr. Exerc. Metab.* 23:369–377.
- Forstermann, U. 2010. Nitric oxide and oxidative stress in vascular disease. *Pflugers Arch.* 459:923–939.
- Forstermann, U., and W. C. Sessa. 2012. Nitric oxide synthases: regulation and function. *Eur. Heart J.* 33: 829–837, 837a–837d.
- Frandsenn, U., J. Bangsbo, M. Sander, L. Hoffner, A. Betak, B. Saltin, et al. 2001. Exercise-induced hyperaemia and leg oxygen uptake are not altered during effective inhibition of nitric oxide synthase with N(G)-nitro-L-arginine methyl ester in humans. *J. Physiol.* 531:257–264.
- Gramsbergen, J. B., M. Sandberg, A. Moller Dall, B. Kornblit, and J. Zimmer. 2002. Glutathione depletion in nigrostriatal slice cultures: GABA loss, dopamine resistance and protection by the tetrahydrobiopterin precursor sepiapterin. *Brain Res.* 935:47–58.
- Grange, R. W., E. Isotani, K. S. Lau, K. E. Kamm, P. L. Huang, and J. T. Stull. 2001. Nitric oxide contributes to vascular smooth muscle relaxation in contracting fast-twitch muscles. *Physiol. Genomics* 5:35–44.
- Gruhn, N., J. Aldershvile, and S. Boesgaard. 2001. Tetrahydrobiopterin improves endothelium-dependent vasodilation in nitroglycerin-tolerant rats. *Eur. J. Pharmacol.* 416:245–249.
- Habler, H. J., G. Wasner, and W. Janig. 1997. Attenuation of neurogenic vasoconstriction by nitric oxide in hindlimb microvascular beds of the rat in vivo. *Hypertension* 30:957–961.
- Harding, C. O., M. Neff, K. Wild, K. Jones, L. Elzaouk, B. Thony, et al. 2004. The fate of intravenously administered tetrahydrobiopterin and its implications for heterologous gene therapy of phenylketonuria. *Mol. Genet. Metab.* 81:52–57.
- Heales, S. J., J. A. Blair, C. Meinschad, and I. Ziegler. 1988. Inhibition of monocyte luminol-dependent chemiluminescence by tetrahydrobiopterin, and the free radical oxidation of tetrahydrobiopterin, dihydrobiopterin and dihydroneopterin. *Cell Biochem. Funct.* 6:191–195.
- Heinzel, B., M. John, P. Klatt, E. Bohme, and B. Mayer. 1992. Ca<sup>2+</sup>/calmodulin-dependent formation of hydrogen peroxide by brain nitric oxide synthase. *Biochem. J.* 281 (Pt 3):627–630.
- Heitzer, T., C. Brockhoff, B. Mayer, A. Warnholtz, H. Mollnau, S. Henne, et al. 2000. Tetrahydrobiopterin improves endothelium-dependent vasodilation in chronic smokers: evidence for a dysfunctional nitric oxide synthase. *Circ. Res.* 86:E36–E41.
- Higashi, Y., S. Sasaki, K. Nakagawa, Y. Fukuda, H. Matsuura, T. Oshima, et al. 2002. Tetrahydrobiopterin enhances forearm vascular response to acetylcholine in both normotensive and hypertensive individuals. *Am. J. Hypertens.* 15:326–332.
- Hirai, T., M. D. Visneski, K. J. Kearns, R. Zelis, and T. I. Musch. 1994. Effects of NO synthase inhibition on the muscular blood flow response to treadmill exercise in rats. *J. Appl. Physiol.* 77:1288–1293.
- Jendzjowsky, N. G., and D. S. DeLorey. 2013a. Acute superoxide scavenging reduces sympathetic vasoconstrictor responsiveness in short-term exercise trained rats. *J. Appl. Physiol.* (1985) 114:1511–1518.
- Jendzjowsky, N. G., and D. S. DeLorey. 2013b. Role of neuronal nitric oxide in the inhibition of sympathetic vasoconstriction in resting and contracting skeletal muscle of healthy rats. *J. Appl. Physiol.* (1985) 115:97–106.
- Jendzjowsky, N. G., and D. S. DeLorey. 2013c. Short-term exercise training enhances functional sympatholysis through a nitric oxide-dependent mechanism. *J. Physiol.* 591:1535–1549.
- Katusic, Z. S., and L. V. d'Uscio. 2004. Tetrahydrobiopterin: mediator of endothelial protection. *Arterioscler. Thromb. Vasc. Biol.* 24:397–398.
- Katusic, Z. S., L. V. d'Uscio, and K. A. Nath. 2009. Vascular protection by tetrahydrobiopterin: progress and therapeutic prospects. *Trends Pharmacol. Sci.* 30:48–54.
- Kuzkaya, N., N. Weissmann, D. G. Harrison, and S. Dikalov. 2003. Interactions of peroxynitrite, tetrahydrobiopterin, ascorbic acid, and thiols: implications for uncoupling endothelial nitric-oxide synthase. *J. Biol. Chem.* 278:22546–22554.
- Lang, C. C., G. H. Rayos, D. B. Chomsky, A. J. Wood, and J. R. Wilson. 1997. Effect of sympathoinhibition on exercise performance in patients with heart failure. *Circulation* 96:238–245.
- Lang, J. A., L. A. Holowatz, and W. L. Kenney. 2009. Local tetrahydrobiopterin administration augments cutaneous vasoconstriction in aged humans. *J. Physiol.* 587:3967–3974.
- Lang, J. A., L. A. Holowatz, and W. L. Kenney. 2010. Tetrahydrobiopterin does not affect end-organ responsiveness to norepinephrine-mediated vasoconstriction in aged skin. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 299:R1651–R1655.
- Lau, K. S., R. W. Grange, E. Isotani, I. H. Sarelius, K. E. Kamm, P. L. Huang, et al. 2000. nNOS and eNOS modulate cGMP formation and vascular response in contracting fast-twitch skeletal muscle. *Physiol. Genomics* 2:21–27.
- Laughlin, M. H., and D. H. Korzick. 2001. Vascular smooth muscle: integrator of vasoactive signals during exercise hyperemia. *Med. Sci. Sports Exerc.* 33:81–91.
- Mortensen, S. P., M. Nyberg, K. Winding, and B. Saltin. 2012. Lifelong physical activity preserves functional sympatholysis and purinergic signalling in the ageing human leg. *J. Physiol.* 590:6227–6236.
- Nagatsu, T. 1983. Biopterin cofactor and monoamine-synthesizing monooxygenases. *Neurochem. Int.* 5:27–38.

- Noguchi, K., N. Hamadate, T. Matsuzaki, M. Sakanashi, J. Nakasone, and M. Tsutsui. 2010. Improvement of impaired endothelial function by tetrahydrobiopterin in stroke-prone spontaneously hypertensive rats. *Eur. J. Pharmacol.* 631:28–35.
- Noguchi, K., N. Hamadate, T. Matsuzaki, M. Sakanashi, J. Nakasone, T. Uchida, et al. 2011. Increasing dihydrobiopterin causes dysfunction of endothelial nitric oxide synthase in rats in vivo. *Am. J. Physiol. Heart Circ. Physiol.* 301:H721–H729.
- Notarius, C. F., P. J. Millar, H. Murai, B. L. Morris, and J. S. Floras. 2014. Inverse relationship between muscle sympathetic activity during exercise and peak oxygen uptake in subjects with and without heart failure. *J. Am. Coll. Cardiol.* 63:605–606.
- Ohyanagi, M., K. Nishigaki, and J. E. Faber. 1992. Interaction between microvascular alpha<sub>1</sub>- and alpha<sub>2</sub>-adrenoceptors and endothelium-derived relaxing factor. *Circ. Res.* 71:188–200.
- Pierce, G. L., K. L. Jablonski, A. E. Walker, S. M. Seibert, A. E. DeVan, S. M. Black, et al. 2012. Tetrahydrobiopterin supplementation enhances carotid artery compliance in healthy older men: a pilot study. *Am. J. Hypertens.* 25:1050–1054.
- Pou, S., W. S. Pou, D. S. Bredt, S. H. Snyder, and G. M. Rosen. 1992. Generation of superoxide by purified brain nitric oxide synthase. *J. Biol. Chem.* 267:24173–24176.
- Radegran, G., and B. Saltin. 1999. Nitric oxide in the regulation of vasomotor tone in human skeletal muscle. *Am. J. Physiol. Heart Circ. Physiol.* 276:H1951–H1960.
- Saltin, B., and S. P. Mortensen. 2012. Inefficient functional sympatholysis is an overlooked cause of malperfusion in contracting skeletal muscle. *J. Physiol.* 590:6269–6275.
- Saltin, B., G. Radegran, M. D. Koskolou, and R. C. Roach. 1998. Skeletal muscle blood flow in humans and its regulation during exercise. *Acta Physiol. Scand.* 162:421–436.
- Sander, M., B. Chavoshan, S. A. Harris, S. T. Iannaccone, J. T. Stull, G. D. Thomas, et al. 2000. Functional muscle ischemia in neuronal nitric oxide synthase-deficient skeletal muscle of children with Duchenne muscular dystrophy. *Proc. Natl Acad. Sci. USA* 97:13818–13823.
- Santhanam, A. V., L. V. d'Uscio, L. A. Smith, and Z. S. Katusic. 2012. Uncoupling of eNOS causes superoxide anion production and impairs NO signaling in the cerebral microvessels of hph-1 mice. *J. Neurochem.* 122:1211–1218.
- Sindler, A. L., M. D. Delp, R. Reyes, G. Wu, and J. M. Muller-Delp. 2009. Effects of ageing and exercise training on eNOS uncoupling in skeletal muscle resistance arterioles. *J. Physiol.* 587:3885–3897.
- Sun, J., L. J. Druhan, and J. L. Zweier. 2008. Dose dependent effects of reactive oxygen and nitrogen species on the function of neuronal nitric oxide synthase. *Arch. Biochem. Biophys.* 471:126–133.
- Thomas, G. D., and S. S. Segal. 2004. Neural control of muscle blood flow during exercise. *J. Appl. Physiol.* 97:731–738.
- Thomas, G. D., and R. G. Victor. 1998. Nitric oxide mediates contraction-induced attenuation of sympathetic vasoconstriction in rat skeletal muscle. *J. Physiol.* 506 (Pt 3):817–826.
- Thomas, G. D., M. Sander, K. S. Lau, P. L. Huang, J. T. Stull, and R. G. Victor. 1998. Impaired metabolic modulation of alpha-adrenergic vasoconstriction in dystrophin-deficient skeletal muscle. *Proc. Natl Acad. Sci. USA* 95:15090–15095.
- Thomas, G. D., P. W. Shaul, I. S. Yuhanna, S. C. Froehner, and M. E. Adams. 2003. Vasomodulation by skeletal muscle-derived nitric oxide requires alpha-syntrophin-mediated sarcolemmal localization of neuronal Nitric oxide synthase. *Circ. Res.* 92:554–560.
- VanTeeffelen, J. W., and S. S. Segal. 2003. Interaction between sympathetic nerve activation and muscle fibre contraction in resistance vessels of hamster retractor muscle. *J. Physiol.* 550:563–574.
- Vasquez-Vivar, J., P. Martasek, J. Whitsett, J. Joseph, and B. Kalyanaraman. 2002. The ratio between tetrahydrobiopterin and oxidized tetrahydrobiopterin analogues controls superoxide release from endothelial nitric oxide synthase: an EPR spin trapping study. *Biochem. J.* 362:733–739.
- Vasquez-Vivar, J., B. Kalyanaraman, and P. Martasek. 2003. The role of tetrahydrobiopterin in superoxide generation from eNOS: enzymology and physiological implications. *Free Radic. Res.* 37:121–127.
- Vongpatanasin, W., Z. Wang, D. Arbique, G. Arbique, B. Adams-Huet, J. H. Mitchell, et al. 2011. Functional sympatholysis is impaired in hypertensive humans. *J. Physiol.* 589:1209–1220.
- Xiao, D. S., L. Jiang, L. L. Che, and L. Lu. 2003. Nitric oxide and iron metabolism in exercised rat with L-arginine supplementation. *Mol. Cell. Biochem.* 252:65–72.
- Zhao, W., S. A. Swanson, J. Ye, X. Li, J. M. Shelton, W. Zhang, et al. 2006. Reactive oxygen species impair sympathetic vasoregulation in skeletal muscle in angiotensin II-dependent hypertension. *Hypertension* 48:637–643.