Research Article

Mycobacterium bovis in Swine: Spoligotyping of Isolates from Argentina

Soledad Barandiaran,¹ Marcela Martínez Vivot,¹ Eduardo Vicente Moras,¹ Angel Adrián Cataldi,² and Martín José Zumárraga²

¹School of Veterinary of Buenos Aires University, Chorroarín 280, C1427CWO, Buenos Aires, Argentina

² Biotechnology Institute, National Institute of Agricultural Technology (INTA), N. Repetto y De Los Reseros cc25, B1712WAA, Castelar, Buenos Aires, Argentina

Correspondence should be addressed to Martín José Zumárraga, mzumarraga@cnia.inta.gov.ar

Received 11 January 2011; Accepted 16 February 2011

Academic Editor: Mitchell V. Palmer

Copyright © 2011 Soledad Barandiaran et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

A total of 143 *Mycobacterium bovis* isolates of pigs, from the most productive swine area in Argentina, were typed by spoligotyping. Twenty-two different spoligotypes were identified, and 133 (93%) isolates were grouped into 12 clusters. One of them, designed SB0140, was the most frequent because it held 83 (58%) isolates. This spoligotype also grouped 362 (43%) out of 841 isolates from previously typed cattle and, thus, constitutes the most frequent in our country. In addition, 135 (94%) isolates revealed spoligotypes identical to those of cattle, showing an epidemiological link. On the other hand, there were seven novel spoligotypes, six of which were also unique since they had only one isolate each. This study aimed to identify the spoligotypes of *M. bovis* isolated from pigs to contribute to a better understanding of the distribution of bovine tuberculosis in the main productive area of Argentina.

1. Introduction

Bovine tuberculosis (BTB) is a major livestock disease in Latin America. About 70% of the cattle bred are held in areas with high disease prevalence and nearly 17% in areas virtually free from tuberculosis [1].

In Argentina, between 1969 and 2004, an average of 10 million bovine carcasses was annually submitted to official veterinary inspection. During that period, the percentage of animals condemned for tuberculosis decreased from 6.7% to 1.2% [1]. Regarding human tuberculosis, the disease caused by *Mycobacterium bovis* and that caused by *M. tuberculosis* are clinically indistinguishable. A recent work has described an incidence ranging from 0.7% to 6.2% in a main milk region of Argentina where most patients were closely related to rural activities, and a much lower national prevalence [2]. Between 1980 and 2003, the incidence of tuberculosis in Santa Fe, a province with 21% of the national dairy herd and 34% of the nation's milk production, declined

from 48.1 to 20.7 per 100,000 [1]. However, the number of cases due to *M. bovis* remained stable, thus suggesting the relatively constant risk of infection [1]. In the Muñiz Hospital of Buenos Aires city, the percentage of *M. bovis* cases not associated with HIV infection significantly decreased from 0.95% between 1981 and 1991 to 0.22% between 2000–2006, whereas in the HIV/AIDS group the decrease was less pronounced (0.83% to 0.58%) [3]. The first case of person-to-person transmission of *M. bovis* in Argentina has been recently described [4].

In the United States, BTB was the most prevalent infectious disease in bovines and pigs between the XVIII century and the beginning of the XIX century, and the production loss was higher than that caused by all other diseases grouped [5].

Pigs are susceptible to *M. tuberculosis*, *M. bovis*, and the *M. avium* complex. In Argentina, the information about incidence and prevalence of tuberculosis in pigs is scarce. However, *M. bovis* is considered the primary cause of their

infection. The prevalence of lesions found in slaughterhouses from the Pampa Húmeda region during three samplings performed in 1986, 1995, and 1997, were 6.4%, 1.2%, and 0.1%, respectively, and M. bovis was the bacterial agent most frequently isolated in the three cases [6]. In countries where BTB has not been eradicated, such as in Argentina, pigs often become infected from cattle by oral route through ingestion of milk or other dairy products [6, 7]. Therefore, in Argentina, swine tuberculosis underwent a decline parallel to that observed in local cattle [1, 6, 8] because the eradication of BTB from cattle is the main action to avoid the transmission to swine and other hosts. Thus, the percentage of pigs condemned for tuberculosis decreased from 8.4 to 0.7% between 1969 and 2005 [9]. This panorama of the disease is also attributed to the modernization and intensification of breeding systems, which has improved the swine health conditions [9, 10]. Furthermore, in February 2009, pigs were incorporated to the National Control and Eradication Program of Bovine Tuberculosis, implemented for bovines since 1999 [11, 12]. This new resolution considers that those herds with negative skin test results with PPD once a year and the absence of tuberculosis lesions during carcass inspections in slaughterhouses are free of infection [11]. Although the skin test with PPD has a limited usefulness for individual animal diagnosis, it is a good tool to detect infected herds [13].

Approximately 80% of the swine production in Argentina is concentrated in the central region (Buenos Aires, Santa Fe, and Córdoba provinces), and it is closely related to maize production, because pigs are fed mainly on corn (65%) and soy flour (25%).

Molecular epidemiology is a helpful tool that contributes to the understanding of the dynamics of the distribution and spreading of BTB between the different hosts. One of the techniques that has improved and simplified the typing of the *M. tuberculosis* complex is spoligotyping [14]. Spoligotyping is a PCR-based method complemented with reverse line blot hybridization, in which the polymorphism consequence of rearrangements of the direct repeat (DR) region, which is composed of perfect direct 36-bp repeats and variable spacers [14]. The combination of the presence and absence of each 43 representative spacer sequences, shown with spots, represents a spoligotype.

The spoligotyping database of the Biotechnology Institute (BI) of INTA, Argentina, contains the spoligotypes of 1188 *M. bovis* isolates from different hosts from Argentina typed since 1996 and constitutes a valuable source of molecular epidemiology information that may help to contribute to the eradication program of bovine tuberculosis.

The aim of this work was to determine the *M. bovis* spoligotypes circulating among pigs from the main porcine productive region of Argentina.

2. Materials and Methods

A total of 143 isolates of *M. bovis* were obtained from culture of lymph nodes and viscera samples with tuberculosis compatible lesions from pigs from Buenos Aires (n = 66),



FIGURE 1: Map of Argentina. The provinces sampled are denoted in a gray scale according to the number of isolates.

Córdoba (n = 44), Entre Ríos (n = 6), La Pampa (n = 3), Mendoza (n = 1), and Santa Fe (n = 23) provinces (Figure 1), following a convenient sampling. Samples were obtained from the veterinary inspection of 35,000 pigs, between July 2007 and November 2008. The slaughterhouses involved in this sampling were located in Merlo, Moreno, and Tandil cities from Buenos Aires province. The samples were decontaminated using the Petroff's method and cultured in Lowenstein-Jensen and Stonebrink media at 37°C for 60 days [15]. The bacteriological typing of the isolates was performed based on the culture media, incubation temperature, growing time, colony morphology, and Ziehl-Neelsen stain.

Each *M. bovis* strain was isolated from independent animals without apparent epidemiological linkage. A loopful of colonies was transferred into a microcentrifuge tube containing 250 μ L of distilled water and heated at 96°C for 45 min. Colonies were then centrifuged at 12,000 rpm for 10 min, and 5 μ L of the supernatant was used for PCR to amplify the DR region. Spoligotyping [14] was carried out by using the spoligotyping kit (Isogen Biosolutions B.V., Ocimun Biosolutions Company, Ijsselstein, the Netherlands). *M. tuberculosis* H37Rv (ATCC 27294) and *M. bovis* Bacillus Calmette-Guerin (BCG) (ATCC 27289) were included as reference strains in each spoligotyping

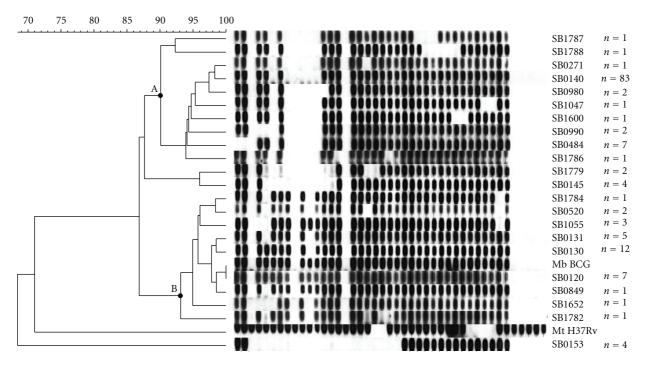


FIGURE 2: Dendrogram showing the relationship between 22 different spoligotypes identified among 143 *M. bovis* isolates of pigs from Argentina. The spoligotypes are detailed on the right of each pattern. The two families of spoligotypes were designated with letters A and B. *n*: number of isolates.

experiment. A cluster analysis of the spoligotype patterns was performed with the BioNumerics software (Windows NT, version 2.5; Applied Maths, Kortrijk, Belgium). The categorical coefficient was used to calculate the similarity of spoligotype patterns, and the UPGMA (unweighted pair-group method with arithmetic averages) method was applied to calculate a dendrogram. Clusters of isolates were defined as two or more M. bovis strains with identical spoligotypes. The different spoligotypes were compared against those included in the BI database, and each was assigned a number (SB code) according to the international database of the Veterinary Laboratories Agency (VLA), the United Kingdom (hosted on University of Sussex servers, http://www.mbovis.org/). When a spoligotype was not found in these databases, it was considered a novel spoligotype. Spoligotypes with only one isolate were regarded as unique spoligotypes. The discriminatory index (D) described by Hunter and Gaston and expressed by the formula of Simpson [16] was calculated to determine the discriminatory power of the spoligotyping in each province (http://insilico.ehu.es/) [17].

3. Results

All the isolates analyzed (n = 143) lacked spacers 3, 9, 16, and 39 to 43, characteristic of *M. bovis* strains. Twentytwo different spoligotypes were found among the 143 *M. bovis* isolates studied (Figure 2 and Table 1). One hundred thirty-three (93%) strains were grouped in 12 clusters; one of them (SB1779) was a novel spoligotype because it had not been previously reported among isolates of other hosts from Argentina or in the VLA database. The main cluster involved 83 (58%) isolates and showed spoligotype SB0140 followed by spoligotype SB0130 with 12 (8.4%) isolates. The remaining spoligotypes grouped seven or fewer isolates. Ten (7%) isolates were unique, and seven of them were also novel (SB0849, SB1600, SB1652, SB1784, SB1786, SB1782, and SB1779) (Figure 2 and Table 1). Additionally, the cluster with the novel spoligotype (SB1779) held two isolates from the cities of Río Cuarto and General Viamonte, both in Córdoba province, which are approximately 150 km from each other. The global discriminatory power of the spoligotyping in this study was 0.65. This index was also individually calculated for each province (Table 1), being 0.62, 0.80, 0.60, and 0.40, for Buenos Aires, Córdoba, Entre Ríos and Santa Fe, respectively. These provinces provided most isolates. These results are related to the size of the clusters and the number of spoligotypes.

The dendrogram grouped the spoligotypes into two main families designed A and B, related to a similarity higher than 90% and 93%, respectively (Figure 2). The inclusion of the two most frequent spoligotypes from Argentina (SB0140 and SB0130) in each family is the main feature of these families. Moreover, 70% and 23% of the isolates were grouped in families A and B, respectively.

4. Discussion

Five of the six provinces studied are located in the most productive swine area in Argentina. When we compared the spoligotypes in the BI database, we found that 135 (94%) isolates revealed spoligotypes identical to bovine isolates,

Numbe	Number of isol	Number of isolates by spoligotype	þe				
0271 0484 0520	0271 0484 0520 0849 0980 0990 1047 1055 1600 1652 1779 1782 1784 1786 1787 1788	0980 0990 1047	7 1055 160	0 1652 177	9 1782 1784 1	786 1787 1788	2
2 1	2 1	1 1	1 1	1	1	1	0.62
1 3	ŝ	2	7	2	1	1 1	0.80
1 1	1 1						0.60
1	1						0.66
							1
	1	1					0.40
1 7 2	7 2 1	2 2 1	3 1	1 2	1 1	1 1 1	0.65

	ă
c.	S
	ō
	-
	9e
	ū
	<u> </u>
	Ē
¢	<i>,</i>
	X.
-	g
	ă
•	1
	Ľ.
	0
	al
	Ĩ
	E
•	Ξ
	5
	IS
	σ
	÷
ĥ	-
	<u> </u>
÷	ali
	Ë
	Ē
•	=
	5
	ar
	S
	ă
	F
	5
	δõ
÷	
	ಹ
	S
-	e
	≥
	g
	5
-	I he
Ē	-
	÷
	Ξ
	1g1
	or
	2
-	6
	ė
	ē,
	otyp
	8
	<u>e</u> p
	0
	ğ
	c C
-	0
	ea
ç	Ĩ.
	0
	cs
	Ĕ
-	Ĕ
	SC
;	Ľ
	õ
	-
	o O
	um
2	ź
	(T)

showing an epidemiological link. Furthermore, seven of twenty two (32%) of the spoligotypes detected in pig isolates were detected in M. bovis human isolates. Conversely, seven (31.8%) spoligotypes had not been previously detected in cattle or other hosts from Argentina. Moreover, these spoligotypes were not reported in the VLA database. This finding could be due to the partial screening of the bovines with BTB in Argentina or to the existence of M. bovis clones circulating exclusively among pigs. Parra et al. [18] also described swine spoligotypes not detected previously in other hosts.

The main cluster detected (SB0140) grouped 43% of the 841 bovine M. bovis isolates from Argentina typed between 1996 and 2009 [19, 20]. This spoligotype is also frequent in Australia and New Zealand, which, like Argentina, introduced several cattle breeds in the 19th century from the United Kingdom, where this spoligotype is also prevalent [21]. Furthermore, spoligotype SB0140 was the most frequent in Buenos Aires (60.6%), Córdoba (43.2%), Entre Ríos (66.7%), and Santa Fe (78.3%) provinces, where most *M. bovis* isolates from pigs were obtained. Additionally, this spoligotype grouped 32.5, 38.9, 62.5, and 52.5% of bovine isolates of these provinces, respectively. These provinces concentrate most of the dairy farms from Argentina. Clustering of isolates has been described as an indication of active transmission of BTB [19]. Spoligotype SB0153, which was 68% related to the other types, was not integrated in families A and B and had been previously detected in only 2.7% of total M. bovis isolates from Argentina. Curiously, 34% of all the M. bovis isolates with spoligotype SB0153 belong to humans.

Taking into account the localization of most of the lesions along the digestive tract of sampled pigs (data not shown), we suggest that the infection via was the digestive route. This could be due to the fact that in Argentina pigs diets are usually supplemented with milk or other dairy derivates without thermal treatment.

Other authors have also found *M. bovis* spoligotypes of cattle in domestic pigs and other hosts (red deer and wild boar), which suggests transmission between species [18, 22, 23].

Spoligotyping is the best option for large-scale screening studies on the distribution of *M. tuberculosis* complex strains [14] and is used worldwide as the first-option typing method for *M. bovis*. Moreover, this technique is useful to identify new types from different host species [24, 25] and to detect preliminary transmission of TB between species. In order to trace transmission chains, it is necessary to perform complementary studies using more discriminatory typing methods such as the recently described VNTRs [26–29].

Future studies must be directed to evaluate the virulence and the fitness of these strains isolated from pigs, since transmission between different host species could be a selective force to increase the virulence of microorganisms. In a previous work carried out in a murine model of tuberculosis, we demonstrated that a particular *M. bovis* strain isolated from a wild boar was the most virulent compared to the *M. bovis* AN5 reference strain and other isolated from cattle and humans [30]. Considering the incidence of *M. bovis* in humans and the prevalence of the disease in cattle, especially in the provinces that hold most dairy herds, pigs could be an additional actor in the transmission chain of bovine tuberculosis to humans and cattle. This potential spillover could be controlled through the recent incorporation of swine to the National Control and Eradication Program of Bovine Tuberculosis of Argentina.

5. Conclusions

Most of the spoligotypes (68%) found in pigs had also been previously detected in cattle. There were seven novel spoligotype detected only in pigs. The most frequent spoligotype among *M. bovis* isolates studied from pigs (SB0140) was also the most prevalent in bovines from Argentina.

This work represents the first large-scale molecular typing study of *M. bovis* isolates from pigs carried out in Argentina and contributes to a better understanding of the features of tuberculosis in pigs in our country.

Acknowledgments

This work was supported by grants from the National Institute of Agricultural Technology of Argentina (INTA) and School of Veterinary of Buenos Aires University. The authors thank Valeria Rocha for her excellent technical help. A. A. Cataldi is a career member of the National Research Council (CONICET) from Argentina.

References

- I. N. De Kantor and V. Ritacco, "An update on bovine tuberculosis programmes in Latin American and Caribbean countries," *Veterinary Microbiology*, vol. 112, no. 2–4, pp. 111– 118, 2006.
- [2] A. Cataldi and M. I. Romano, "Tuberculosis caused by ohter members of the *M. tuberculosis* complex," in *Tuberculosis 2007 From Basic Science to Patient Care*, J. C. Palomino, S. C. Leao, and V. Ritacco, Eds., chapter 8, 1st edition, 2007.
- [3] I. N. de Kantor, M. Ambroggi, S. Poggi et al., "Human Mycobacterium bovis infection in ten Latin American countries," *Tuberculosis*, vol. 88, no. 4, pp. 358–365, 2008.
- [4] I. Etchechoury, G. E. Valencia, N. Morcillo et al., "Molecular typing of *Mycobacterium bovis* isolates in Argentina: first description of a person-to-person transmission case," *Zoonoses and Public Health*, vol. 57, no. 6, pp. 375–381, 2010.
- [5] R. J. Montali, S. K. Mikota, and L. I. Cheng, "Mycobacterium tuberculosis in zoo and wildlife species," OIE Revue Scientifique et Technique, vol. 20, no. 1, pp. 291–303, 2001.
- [6] A. Perez, R. Debenedetti, M. Martínez Vivot, A. Bernardelli, P. Torres, and V. Ritacco, "Tendencia de la tuberculosis porcina y validez de la inspección bromatológica para su detección en áreas de producción intensiva de Argentina," *Revue de Médecine Vétérinaire*, vol. 85, pp. 61–64, 2004.
- [7] M. Martínez Vivot, D. Marticorena, M. Zumárraga et al., "Tuberculosis porcina, caracterización bacteriológica y molecular de micobacterias aisladas de cerdos faenados en La Pampa Húmeda," *InVet*, vol. 7, pp. 246–247, 2005.

- [8] A. Perez and V. Ritacco, "Diagnóstico de la tuberculosis porcina en frigorífico," in *Jornadas de Divulgación Técnico Científicas*, Facultad de Ciencias Veterinarias, U.N.R., Casilda, Santa Fe, Argentina, 2001.
- [9] P. Torres, "Situación de la tuberculosis bovina en la República Argentina," in Programa Nacional de Control y Erradicación de la tuberculosis, SENASA (Servicio Nacional de Sanidad Animal), Secretaría de Agricultura, Buenos Aires, Argentina, 2006.
- [10] L. A. Ottavianoni, A. Bernardelli, and R. J. Dubarry, "Aspectos sobre tuberculosis porcina en la provincia de La Pampa," *Veterinaria Argentina*, vol. 3, pp. 457–462, 1986.
- [11] SENASA, 2009, Resolución 145/2009, http://www.senasa.gov .ar/contenido.php?to=n&in=1403&io=9279.
- [12] Secretaría de Agricultura, Dirección de Sanidad Animal, Argentina, "Plan Nacional de Control y Erradicación de la Tuberculosis Bovina," Resolución no. 115/99 SENASA/ SAGPyA, 1999.
- [13] G. G. Magnano, M. O. Schneider, C. E. Urbani, A. Ambrogi, L. Zapata, and M. C. Jorge, "Comparación de técnicas diagnósticas de tuberculosis porcina en dos establecimientos de cría confinada en Argentina," *InVet*, vol. 12, no. 1, pp. 25– 31, 2010.
- [14] J. Kamerbeek, L. Schouls, A. Kolk et al., "Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* for diagnosis and epidemiology," *Journal of Clinical Microbiol*ogy, vol. 35, no. 4, pp. 907–914, 1997.
- [15] I. N. de Kantor, "Bacteriología de la tuberculosis N°11," CEPANZO OPS/OMS, Buenos Aires, pp. 26–35, 1988.
- [16] P. R. Hunter and M. A. Gaston, "Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity," *Journal of Clinical Microbiology*, vol. 26, no. 11, pp. 2465–2466, 1988.
- [17] S. Rodríguez, B. Romero, J. Bezos et al., "High spoligotype diversity within a *Mycobacterium bovis* population: clues to understanding the demography of the pathogen in Europe," *Veterinary Microbiology*, vol. 141, no. 1-2, pp. 89–95, 2010.
- [18] A. Parra, J. Larrasa, A. García, J. M. Alonso, and J. Hermoso De Mendoza, "Molecular epidemiology of bovine tuberculosis in wild animals in Spain: a first approach to risk factor analysis," *Veterinary Microbiology*, vol. 110, no. 3-4, pp. 293–300, 2005.
- [19] M. J. Zumárraga, C. Martin, S. Samper et al., "Usefulness of spoligotyping in molecular epidemiology of *Mycobacterium bovis*-related infections in South America," *Journal of Clinical Microbiology*, vol. 37, no. 2, pp. 296–303, 1999.
- [20] M. J. Zumárraga, Epidemiología molecular de la tuberculosis bovina, Ph.D. thesis, Facultad de Ciencias Exactas y Naturales, Buenos Aires, Argentina, 2007.
- [21] A. A. Cataldi, A. Gioffré, M. P. Santtangelo et al., "El genotipo de *Mycobacterium bovis* mayoritario en la Argentina lo es también en las Islas Británicas: la tuberculosis bovina provino de Gran Bretaña?" *Revista Argentina de Microbiologia*, vol. 34, no. 1, pp. 1–6, 2002.
- [22] F. Milian-Suazo, B. Harris, C. A. Díaz et al., "Molecular epidemiology of *Mycobacterium bovis*: usefulness in international trade," *Preventive Veterinary Medicine*, vol. 87, no. 3-4, pp. 261–271, 2008.
- [23] E. Costello, D. O'Grady, O. Flynn et al., "Study of restriction fragment length polymorphism analysis and spoligotyping for epidemiological investigation of *Mycobacterium bovis* infection," *Journal of Clinical Microbiology*, vol. 37, no. 10, pp. 3217–3222, 1999.

- [24] M. J. Zumárraga, A. Bernardelli, R. Bastida et al., "Molecular characterization of mycobacteria isolated from seals," *Microbiology*, vol. 145, no. 9, pp. 2519–2526, 1999.
- [25] A. Aranaz, E. Liébana, A. Mateos et al., "Spacer oligonucleotide typing of *Mycobacterium bovis* strains from cattle and other animals: a tool for studying epidemiology of tuberculosis," *Journal of Clinical Microbiology*, vol. 34, no. 11, pp. 2734–2740, 1996.
- [26] S. Roring, A. Scott, D. Brittain et al., "Development of variable-number tandem repeat typing of *Mycobacterium bovis*: comparison of results with those obtained by using existing exact tandem repeats and spoligotyping," *Journal of Clinical Microbiology*, vol. 40, no. 6, pp. 2126–2133, 2002.
- [27] R. A. Skuce, T. P. McCorry, J. F. McCarroll et al., "Discrimination of *Mycobacterium tuberculosis* complex bacteria using novel VNTR-PCR targets," *Microbiology*, vol. 148, no. 2, pp. 519–528, 2002.
- [28] P. Supply, E. Mazars, S. Lesjean, V. Vincent, B. Gicquel, and C. Locht, "Variable human minisatellite-like regions in the *Mycobacterium tuberculosis* genome," *Molecular Microbiology*, vol. 36, no. 3, pp. 762–771, 2000.
- [29] R. Frothingham and W. A. Meeker-O'Connell, "Genetic diversity in the *Mycobacterium tuberculosis* complex based on variable numbers of tandem DNA repeats," *Microbiology*, vol. 144, no. 5, pp. 1189–1196, 1998.
- [30] D. Aguilar León, M. J. Zumárraga, R. Jiménez Oropeza et al., "Mycobacterium bovis with different genotypes and from different hosts induce dissimilar immunopathological lesions in a mouse model of tuberculosis," Clinical and Experimental Immunology, vol. 157, no. 1, pp. 139–147, 2009.