





Review Article

Helicobacter pylori Infection, Virulence Genes' Distribution and Accompanying Clinical Outcomes: The West Africa Situation

Eric Gyamerah Ofori ^{1,2}, Cynthia Ayefoumi Adinortey ¹, Ansumana Sandy Bockarie,³
Foster Kyei ¹, Emmanuel Ayitey Tagoe,^{4,5} and Michael Buenor Adinortey ²

¹Department of Molecular Biology and Biotechnology, School of Biological Sciences, University of Cape Coast, Cape Coast, Ghana

²Department of Biochemistry, School of Biological Sciences, University of Cape Coast, Cape Coast, Ghana

³Department of Internal Medicine, School of Medical Sciences, University of Cape Coast, Cape Coast, Ghana

⁴West Africa Centre for Cell Biology of Infectious Disease and Pathogens, Department of Biochemistry, Cell and Molecular Biology, University of Ghana, Legon, Ghana

⁵Department of Medical Laboratory Sciences, School of Biomedical and Allied Health Sciences, University of Ghana, Ghana

Correspondence should be addressed to Michael Buenor Adinortey; madinortey@ucc.edu.gh

Received 15 May 2019; Revised 18 July 2019; Accepted 20 August 2019; Published 10 December 2019

Academic Editor: Gail B. Mahady

Copyright © 2019 Eric Gyamerah Ofori et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Data on *Helicobacter pylori* (*H. pylori*) infection and virulence factors in countries across West Africa are scattered. This systematic review seeks to present an update on the status of *H. pylori* infection focusing on prevalence rate, distribution of virulent genes, and their link to clinical outcomes across countries in the western part of Africa. This information is expected to broaden the knowledge base of clinicians and researchers regarding *H. pylori* infection and associated virulence factors in West African countries. **Search Method.** A comprehensive search of the scientific literature in PubMed and ScienceDirect was conducted using the search terms including “*Helicobacter pylori* infection in West Africa”. Databases were sourced from January 1988 to December 2018. **Results.** Data on the incidence of *H. pylori* infection and related pathological factors were found for some countries, whereas others had no information on it. Smoking, alcohol, exposure to high levels of carcinogens and diet were reported to be involved in the pathogenesis of gastroduodenal diseases and gastric cancer. Besides the environmental factors and genetic characteristics, there are important characteristics of *H. pylori* such as the ability to infect, replicate, and persist in a host that have been associated with the pathogenesis of various gastroduodenal diseases. **Concluding Remarks.** This systematic search has provided information so far available on *H. pylori* virulence factors and clinical outcomes in West Africa. Accordingly, this piece has identified gaps in the body of knowledge highlighting the need for more studies to clarify the role of *H. pylori* virulence factors and associated clinical outcomes in the burden of this bacterial infection in West Africa, as data from these countries do not give the needed direct relation.

1. Introduction

Helicobacter pylori (*H. pylori*) are common microaerophilic bacteria known to obstinately inhabit the human stomach mucous layer, affecting about half the world's population [1–3]. The pathogen is known to be present in the mucous, on the surface of the stomach lining and its presence causes chronic inflammation, which remains a major cause of prolonged gastritis. Also, *H. pylori* has been identified to increase the risk of developing gastric adenocarcinoma [4]. Apart from the gastrointestinal tract (GIT) related diseases such as gastroesophageal reflux disease, gastric ulcer and duodenal ulcer, infection

from *H. pylori* has also been linked to some other diseases such as iron-deficiency anemia [5, 6], immune thrombocytopenia (ITP), [5, 7] cardiovascular diseases, [8, 9], hepatobiliary diseases [10, 11], diabetes mellitus [12], allergies, and asthma [13] among others. Although Marshall and Warren [14] reported the first isolation of *H. pylori* in 1983, isolation of the organism is still not commonly done in West Africa with only few countries such as Senegal [15] and Ghana [16] having recorded their first successful isolation from gastric biopsy.

There is no certainty in the mode of transmission for *H. pylori* infection; however, various epidemiological studies have made several claims in this regard. The primary means

of spread of the disease has been linked to transmission from one individual to another and usually higher when occurring within a family [17–19]. The spread from person-to-person has been identified to be the most likely and could be by oral-oral, gastro-oral or fecal-oral [20]. In this regard, the practice of good hygiene and improved living conditions becomes an essential factor in reducing the rate of transmission of the infection [21]. Infection occurs in children as well and an infected child maintains a strain, which has a genetic characteristic indistinguishable from that observed in their parents [22–24]. These characteristics remain unchanged upon any alteration in the environment in which they are found.

The prevalence of infection from *H. pylori* varies geographically with the developing world carrying the higher burden [25]. Infection in peptic ulcer diseases (PUDs) patients ranges from almost 25% in countries of the industrialized world, and is anticipated to be around 90% in underdeveloped countries [26, 27]. The prevalence across countries in the West African region is generally high with variations existing from country to country. The reportedly high prevalence observed in Africa (79.1%) and Asia (54.7%) as compared to lower prevalence found in other geographic locations such as Northern America (37.1%) and Oceania (24.4%) [28] have been found not to correlate with the rather low occurrence of gastric cancer [29, 30]. This situation has been described as the “Asian and African enigmas”. These so-called “enigmas” have been explained by several factors including host genetic and immune response, different tumor-inducing potential of explicit strains of *H. pylori* as well as environmental factors [29, 31]. Again, insufficient African population sampling obtained through endoscopy as well as poor access to health care has also been found to contribute to this so-called mystery. In this regard, a stronger and elaborate data on gastric ulcer in Africans and the prevalence of associated cancer have established that the low occurrence is not exactly so [32]. In contrast to the “Asian enigma”, a report by Irino et al. [33] shows that the incidence of gastric cancer (GC) is mainly high in Asian countries, a situation attributed to the high prevalence in infection from *H. pylori*.

It is imperative to mention that an individual that is infected with *H. pylori* faces a strenuous task of getting rid of the bacterium and hence disease eradication, a situation that is largely attributed to the ever-increasing antibiotic resistance [34]. In the near future, the problems associated with *H. pylori* eradication are feared to increase looking at the current increasing infection rates and the gastroduodenal pathological outcomes. It is estimated that about 15% of infected individuals have an increased tendency of developing peptic ulcer [35], while a rate of 1–3% are found to have a bigger propensity of developing gastric malignancy in their lifetime [36, 37]. With the changing epidemiology of infection from this organism, the pattern of other related diseases also keeps changing.

An extensive literature search has revealed that, the prevalence of *H. pylori* in countries across West Africa is generally high and it poses a serious health burden on health care systems. The overall impact varies from country to country.

Several factors have been reported to contribute to this variation such as, host genetic factors, type of *H. pylori* virulence factors and sensitivity of the method of detection employed [38, 39]. There is inadequate information on the role of these factors in *H. pylori* infection rates and associated clinical outcomes across the countries in the Western area of Africa which makes tackling of the growing disease burden increasingly difficult. Assembling data on the type of virulent genes, factors involved in infection and associated clinical outcome is, therefore, important [40, 41]. This systematic review hence looks at available information in the West African zone on the prevalence of *H. pylori* infection, virulence factors and their relation to clinical outcomes.

2. Method of Literature Search

A comprehensive search of the scientific literature in PubMed and ScienceDirect was conducted using the search terms “*Helicobacter pylori* infection in West Africa”. The search was repeated with “West Africa” replaced with each of the following countries; Sierra Leone, Cape Verde, Ghana, Liberia, Benin, Senegal, Sao tome and Principe, Mali, Burkina Faso, Mauritania, Cote D’Ivoire, Guinea Bissau, Niger, Guinea, The Gambia, and Togo. The key words “*Helicobacter pylori*” was replaced with “*H. pylori*” and the search was repeated. Some other keywords employed were “*Helicobacter pylori*”, “epidemiology”, “prevalence”, and “virulent factors”. The search protocol is shown in Figure 1.

The search in PubMed was done with the following activated filters; publication date from 1st January 1988 to 31st December 2018, language filter had English and French activated and the species selected was Human. Filters activated for the search in ScienceDirect database were the selection of the article types; Review articles, Research articles, and mini-reviews as well as the year range of 1988–2018. Duplicate searches were first removed; after which the abstracts of articles retrieved were reviewed for relevance before an attempt was made to retrieve the full paper. Selection of articles was based on the following considerations; (1) Study participants were West Africans. (2) Participants who showed up for endoscopy at a gastroenterology unit and diagnosed with *H. pylori* infection. (3) Studies investigating virulent factors; Vacuolating cytotoxin (*VacA*), Cytotoxin-associated gene A (*cagA*), Outer inflammatory protein (*OipA*), Duodenal ulcer promoting gene (*dupA*), Blood group antigen binding adhesin (*BabA*) and Induced by contact with epithelium (*IceA*) as a contributing factor to disease progression. (4) Obtained clinical features alongside the detection of virulence factors in order to compare how the presence of a factor correlates with disease outcome and manifestation.

Studies excluded were; (1) West Africans participants living outside the study region. (2) Case studies on an individual, and retrospective records review of patients, commentaries, editorials, and letters in response to published articles. (3) Detection of pathogen by stool antigen test. (4) Prevalence among a selected disease group e.g., AIDS and Diabetes patients. (6) Articles that covered prevalence in children alone.

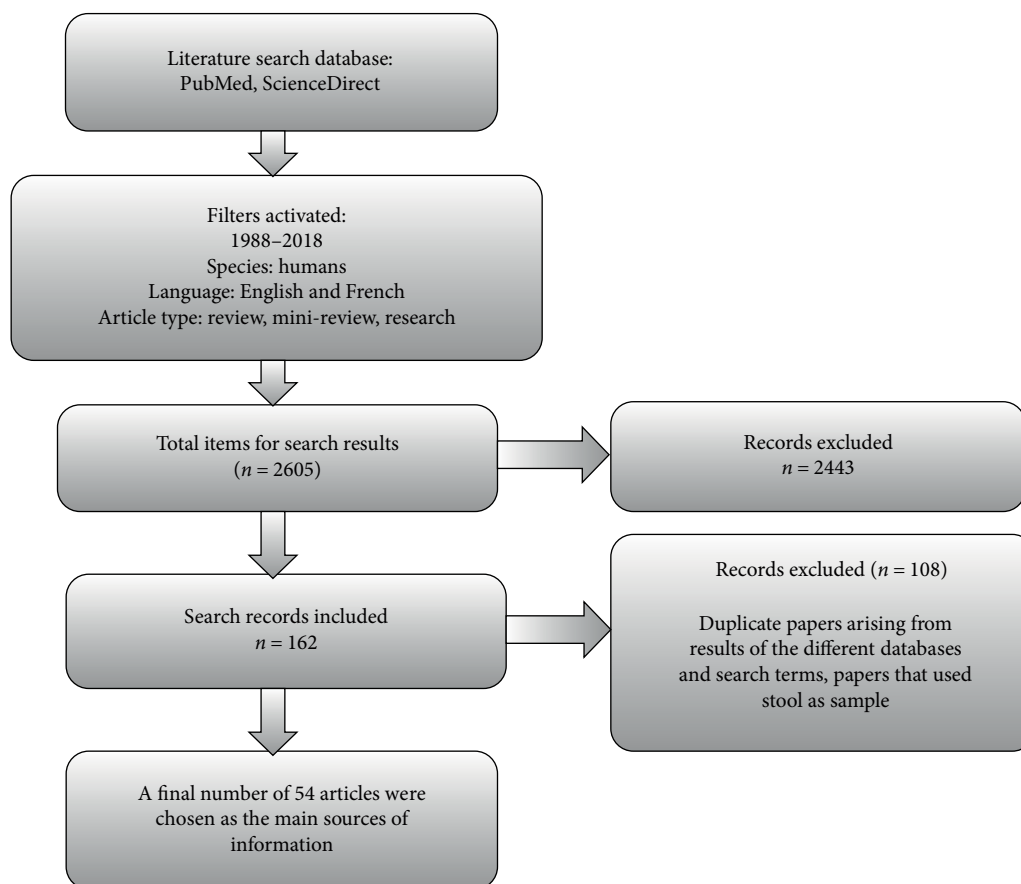


FIGURE 1: Search method employed to identify articles.

3. Results

3.1. Prevalence of *Helicobacter pylori* Infection in West Africa. Infection from *H. pylori* is usually asymptomatic and affects nearly 50–75% of global population [42, 43], its prevalence varies between countries [44]. About 70% of people in developing countries with PUDs are estimated to be affected, though the proportions are slightly lower (25–50%) in the developed countries [42, 45]. The variation across countries is found to be due to an interplay of several factors including host immune response and genetic interaction as well as differences in the potential of specific strains of *H. pylori* to cause cancer. Other studies have attributed the variation in the geographic burden of this infection to age of individual, gender, ethnicity, and factors relating to the environment [29, 31, 46]. These factors are also involved in the variation of the rate of infection that is seen in some countries [3, 47, 48]. Also contributing to this variation in prevalence is socioeconomic status and a dependency on the rate of acquisition in the first 5 years of a person's life [48–51]. In the first 5 years of life, infection with *H. pylori* is reportedly lower in developed countries as compared to developing counterparts. This is possibly due to different degrees in hygiene related practices, which are supposedly better in the developed world at that stage of life. In Burkina Faso for instance, a higher rate of *H. pylori* infection was reported in the primary stages of life [52]. Colonization

in early life by *H. pylori* has been found to predispose infected individuals to the progression of malnutrition and development faltering in The Gambia. These effects however, were found not to persist later in childhood [53]. Studies by Hunt et al. [25] and Kusters et al. [54] however, noted that, generally, infection in the developed countries remains substantially lower in children, but gradually increases with cumulative age. Prevalence in countries in the Middle East are also noted to mostly increase with age and rates are reportedly comparable to each other as well as to those occurring in the United States and Europe. Studies in countries such as Iraq, Iran, Israel, Libya, and Saudi Arabia have shown varied prevalence in various ages with rates in adults reported to be significantly higher than children [55]. Supporting the increasing prevalence in age is a report in The Gambia in which serological indication of *H. pylori* infection in 15% of infants less than 20 months was observed to increase to 46% in those aged 40–60 months [56]. The higher prevalence of *H. pylori* infection reported for this study is attributable to a birth cohort phenomenon.

In the West African region, a disease prevalence of 70–95% has been observed depending on the method used [44]. In Nigeria, a prevalence rate of 93.6% for *H. pylori* was found by serology, while 80.0% was estimated by histology [57]. The prevalence in Nigeria has, however, been found to generally range between 38.0% and 92% [58–62]. Surprisingly, an

unusually low prevalence has been demonstrated in Mali where a rate of 21% was reported among persons with established gastric ulcer, 44% among the casually selected volunteers and 14% in those with gastrointestinal correlated illnesses. This low prevalence in the study population was attributed to the sensitivity of the detection kit [63] due to a possible *H. pylori* strain variability. Generally, the disease prevalence across countries in the West African area varies and there is no clear show of a steady overall increment or decrease with age. In Ghana, the reported prevalence rate is 75% [64–66] while in Senegal 62–97% has been reported [67–72]. A prevalence of 92% was seen in Cote D'Ivoire [73], while in Benin, about 56–72% has been reported [74, 75]. However, a lower prevalence of infection in adults compared to children has been reported in Burkina Faso contrary to earlier reports [52].

Notwithstanding the high rates of *H. pylori* infection globally, there is little evidence of what is being done to investigate or reduce this burden in some West African countries. Currently some countries including Mauritania, Guinea Bissau, Liberia, and Sierra Leone had no published research report that met the inclusion criteria, while others such as Benin registered no updated data in the research area in over a decade. With the high prevalence recorded for studied countries comparable to those of the Middle East and elsewhere in developing countries, overcrowded conditions are major contributing factors. These overcrowded conditions are noted to create closer contacts between mothers and their children as well as the sharing of same bed by siblings which are likely to be a leading reason for the increasing transmission rate and therefore higher infection rates [76].

3.1.1. Influence of Host and *H. pylori* Genetic Factors on Infection. The distribution of *H. pylori* infection and the related pathology are mainly affected by host genetics and *H. pylori* virulence factors. In the progress of gastroduodenal diseases such as gastric cancer (GC), there seems to be an increase in the chances of development resulting from polymorphisms in several virulence genes. *H. pylori* genetic diversity is observed to be widespread and demonstrated in the disease outcome of various strains infection and pathogen interaction with their human host. Although not clearly described, the level of genetic multiplicity is considered as an element for adaptation of *H. pylori* in the stomach of the host and the clinical outcomes of the infection [77]. A high through-put sequencing revealed the diverse nature of *H. pylori* genome, [78] and the bacteria genes' variability was adopted to establish a vibrant phylogeographic distinction, a marker for human migrations [79, 80]. Human genetic polymorphisms also compliment geographical distribution of *H. pylori* and the clinical consequences. For example, persons carrying genetic variation in the proinflammatory interleukin-1-beta (IL-1b) and IL-1 receptor adversary genes are two to three times more probable to develop GC [81]. Similarly, polymorphic tumor necrotic factor (TNF)- α as well as IL-16 genes were found to favor GC development during *H. pylori* infection [82, 83].

3.1.2. Environmental Factors in *Helicobacter pylori* Infection. Several factors have been reported to promote *H. pylori* infection and associated gastroduodenal diseases. Smoking,

excessive alcohol intake, experience with high levels of carcinogens, and diet are proven to be significantly involved in the pathogenesis of gastroduodenal diseases including gastric cancer (GC) [84, 85]. Individuals with a higher intake of refined carbohydrates, pickled, salted, or smoked foods and dried fish and meat are at a higher risk of developing GC as compared to those who consume foods containing higher amounts of fiber, fresh vegetables, and fruits [85]. Antioxidant properties of micronutrients contained in foods with high amounts of fiber and fresh fruits and vegetables are capable of lowering the risk of GC development by exerting a positive effect on the mucosa of gastrointestinal tract. Research on the risk factors of infection in the West African region is very scanty. In Nigeria, while the eating of raw vegetables demonstrated no significance in relation to *H. pylori* infection, drinking of unpasteurized milk recorded a significant association with infection [86]. The study went ahead to demonstrate that the source of drinking water and *H. pylori* infection had no relationship. This is in contrast to another finding in Nigeria that reported a higher prevalence of the infection in individuals who sourced their drinking water from wells, streams, and ponds as compared to tap water [87, 88]. The possibility of fecal contamination was explained to have resulted in the unwholesomeness of water from the other sources besides the tap water.

Meanwhile, isolation of the organism from the intestinal tract of sheep, dogs and cats [89] exists and has been found to survive in a culture of fresh sheep-milk where survival is reported to extend to several days [90]. In a study, conducted in Burkina Faso, healthy individuals visiting a hospital for medical check-up were found to be infected with *H. pylori*, and they were found in shepherds or belonging to families of shepherds [52]. Therefore, study to clarify the role of sheep as zoonotic reservoir of *H. pylori* is highly recommended.

Tobacco usage has also been reported to be a dependent ulcerogenic risk factor for the development of gastroduodenal disorders including peptic ulcers and cancers of the GI tract [91]. Smoking acts indirectly in the promotion of gastroduodenal conditions by adversely affecting the mucosal protective mechanisms of the GI tract and also increasing the risk of *H. pylori* infection [92]. While the relation to the increase in infection may be due to antioxidant reduction or gastroduodenal immune system defense mechanism, the adverse effect on mucosal protective mechanisms can be attributed to an increase in the production of gastric acid coupled with a reduction in the production of bicarbonate [93]. Studies have shown that, smoking inhibits epithelial cell renewal resulting in the alteration of mucosal cell proliferation. This exposes the gastrointestinal tract to various aggressive factors leading to an improved likelihood of the induction of cell apoptosis during ulceration and ulcer healing [94, 95]. A US population-based study obtained within 1997–2003 has revealed that, the prevalence of ulcer disease in present and former smokers (11.43% and 11.52%) is almost twice that of never smokers (6.00%) [96]. Babaei et al. have identified smoking as a habit that is largely associated with PUD patients (85%) than gastritis nonPUD patients (14%). In a study conducted in the USA and Japan to ascertain a possible relationship between patients having functional dyspepsia and smoking, participants did not show any relation of smoking to the condition

[97]. Elsewhere in Nigeria, a study has demonstrated the association where cigarette smoking was found to significantly increase the prevalence of *H. pylori* infection [88].

Alcohol has been recognized to play an active part in the development of gastroduodenal conditions and can do so even at lower concentrations. A lower concentration of alcohol is capable of inducing apoptosis and can increase the expression of alcohol dehydrogenase of the gastric adenocarcinoma cell lines [98]. The carcinogenic and harmful effect of alcohol consumption results from the increase in the expression of aldehyde dehydrogenase, cytochrome P₄₅₀, alcohol dehydrogenase, and by inducing the production of reactive oxygen and nitrogen species [99–101]. In Nigeria, Mnena et al. [86] found no significant association between alcohol consumption and *H. pylori* infection and these findings are similar to those reported in Portugal, China and the United States [102–104]. Contrasting findings obtained from several studies continues to keep inconclusive the consequence of alcohol consumption in *H. pylori* infection. In a study where nondrinkers showed a significantly lower infection rate compared to drinkers, [105] a present study reports of a lower risk of infection in people who drink alcohol, compared to nondrinkers [106]. Effect of alcohol intake on *H. pylori* infection continues to remain a subject open for further investigation.

3.2. Pathogenesis of Infection and Clinical Outcomes. Besides environmental and genetic factors, there are other important characteristics of *H. pylori* such as the ability to enter, replicate, and persist in a host, which have been linked to the pathogenesis of gastroduodenal diseases [107, 108]. These factors come into play after the individual is exposed to the organism. When *H. pylori* first enters the host stomach, it is faced with the task of surviving the hostile acidic condition of gastric acid bath where pH could be as low as 1.5. The organism utilizes its urease activity to break down urea into ammonium ions and carbondioxide. Once in the stomach, *H. pylori* reside on the epithelial surface. Movement into the mucosal lining is flagella-mediated, after which specific interactions between host cell receptors and bacterial adhesins follows. By these interactions, the bacterium latches onto the gastric epithelium and survives a possible displacement by the forces generated from the passing of food down the digestive tract.

H. pylori possess several strain specific virulent genes aside the typical virulence factors such as urease (*UreA*) and outer membrane protein (OMP) [109]. These strain specific genes include Vacuolating cytotoxin gene A (*vacA*), Cytotoxin-associated gene A (*cagA*), Outer inflammatory protein (*OipA*), (Duodenal ulcer promoting gene (*dupA*), Blood group antigen binding adhesin (*BabA*) and Induced by contact with epithelium (*IceA*). Among these, *cagA* and *vacA* are considered most frequently reported entities associated with clinical outcomes of infection with *CagA* being the most studied. An added advantage of the close proximity of the bacteria to gastric epithelium besides the promotion of survival in the harsh pH environment allows for ease in scavenging for nutrients from host. The nutrients are made available when toxins such as cytotoxin-associated gene A (*cagA*) and the vacuolating cytotoxin gene A (*vacA*) from the bacteria, effectively harm the host tissues. The complement of strain-specific *H. pylori* toxin

gene is a mark of its virulence and the damage triggered by these toxins may eventually result in the onset of clinical symptoms. For example, some studies have shown that *cagA* which is a powerful bacterial toxin, is particularly linked to acute gastritis, and gastric cancer development [110–114] while *vacA* is related to gastric adenocarcinoma [115, 116]. As such, the measure of virulence and explanation to the manifestation of clinical outcome of several cases of infection has primarily been linked to the capacity of the organism to produce any of these virulence factors.

The cytotoxic activities of *vacA* and *cagA* are reported to have a high correlation although their genes occupy different genomic regions [117]. An important relationship therefore exist between *vacA* and *cagA* [81] and *H. pylori* strains that express a combination of the alleles of *vacA* s1m1 and *cagA* represent the highest in virulence [118, 119]. Infections involving the expression of a combination of these alleles may lead to a serious epithelial damage [120, 121] which can lead to the development of severe gastric diseases as depicted in Table 1. Among the strains of *vacA*, studies have shown that infections involving subtype *vacA* s1m1 account for higher levels of inflammation in the gastric mucosa and increases the risk for carcinoma and gastric atrophy, as compared to the less virulent *vacA* s2m2 strains [83]. Nonetheless, the association of *vacA* subtypes with disease outcome is not always consistent as may be seen in reports from many countries [122, 123].

In the West African region, there is paucity of studies existing on the genotyping of although a few studies from countries such as Ghana, Senegal, Nigeria, and The Gambia show that majority of *H. pylori* strains have virulence factors [44]. Reports show inconsistent observations concerning the link of *vacA* and *cagA* with the sternness of disease as they occur at different geographic regions [124]. This means, there is no definite property of the bacterium in terms of virulence of the genotypes and that the selective inactivation of certain genes of virulence is an adaptation for host specificity [125]. Nonetheless, there is the likelihood of a global similarity in the mechanism responsible for differential antibiotic resistance.

3.2.1. Cytotoxin-Associated Gene A. *CagA* is encoded on the *cag* pathogenicity island (*CagPAI*). *CagPAI* is a 40kb region of chromosomal DNA that encodes nearly 31 genes forming a type IV secretion system. This system is noted for the injection of the oncoprotein, *cagA*, into mammalian cells [126] where it triggers cytokine production. Exploration of *cagA* gene is very popular among the *cagPAI* and constitutes the most documented virulence factor.

In line with a novel insertion sequence, *cagA* can be separated into two parts (regions) namely *cag* I and *cag* II [127] and depending on the repeat sequences of the 3' region containing the Glu-Pro-Ile-Tyr-Ala (EPIYA) motifs, this gene can also be grouped as East-Asian-type or Western-type [127]. Although some specific EPIYA-like motifs (ESIYA and ESIYT) have been reported [128] in the C-terminal region of some isolated *CagA* strains elsewhere, the carcinogenic potential of *CagA* is greatly linked to its main polymorphic EPIYA motif variants, C and D [129]. *In vitro* studies have shown that the East-Asian-type *CagA* which contains EPIYA-D segments has

TABLE 1: *H. pylori* virulence factors and related clinical outcome in some West Africa countries.

Country	<i>H. pylori</i> genotypes	Clinical outcomes	Reference
Ghana	<i>vacAs1m1</i>	Increased risk of duodenal ulcer disease Duodenal ulcer (reduced risk as compared to <i>vacAs1m1</i>)	[66]
	<i>vacAs1m2</i>		
	<i>cagA13</i> -(hydrophilic region)	Duodenal ulcer	
	<i>cagA24</i> -(region of internal duplication)	Erosive gastritis	
	<i>iceA1</i>	Normal pathology	
	<i>iceA2</i>	Normal pathology	
	<i>vacAs1m1</i>	Normal pathology	
Nigeria	<i>vacAs1m2</i>	Normal pathology	[61]
	<i>vacAs2m1</i>	Normal pathology	
	<i>vacAs2m2</i>	Normal pathology	
	<i>cagA</i>	Normal pathology	
	<i>cagA</i>	Gastric cancer	
Senegal	<i>vacAs1</i>	Gastric cancer	[70]
	<i>vacAm1</i>	Not associated with an enhanced risk of Gastric cancer	
	<i>vacAs1</i> <i>vacAs2</i>		
Gambia	<i>vacAm1</i> <i>vacAm2</i>	Not correlated with disease outcomes	[146]
	<i>cagA</i>		
	<i>iceA1</i>		
	<i>iceA2</i>		

an increased ability to induce morphological changes in epithelial cells and promote gastric cancer or peptic ulcer development than their Western-type counterpart that contains EPIYA-C segments [130]. In a different study however, EPIYA-C was identified to be an important factor in identifying patients with an increased risk of developing gastric cancer [131, 132]. The study of EPIYA motifs is poorly investigated in West Africa and, therefore, the link of the type that may be present to cause the disease cannot be clearly stated. Harrison et al. [61] identified that majority of the 111 study participants had a KDKGPE motif that was upstream an EPIYA-A motif. The study, however, did not investigate the function of this motif, and hence the link to pathology among Nigerians could not be clarified. Once *cagA* is introduced into the cells, it goes through phosphorylation by kinases of the host cell affecting cytoskeletal and tissue structure along with cell proliferation [83, 133]. *H. pylori* strains identified with this gene are capable of inducing apoptosis of the epithelial cells through the mitochondrial pathway and this compromises the barrier responsible for the protection of the epithelium against luminal acid and pepsin [134].

The *cagA* gene is associated with greater outcomes of inflammation and is involved in some severe forms of gastrointestinal diseases such as peptic ulcer and GC [111–113]. Elsewhere in the more developed world, reports have shown that persons infected with *H. pylori* that are positive for *cagA* strains are at a greater risk of developing peptic ulcer or GC than those that are *cagA*-negative [135]. Meanwhile, in East Asia, most strains of *H. pylori* have the *cagA* gene regardless of the disease [136].

Among the few countries in the West African region to sequence the *H. pylori* gene, there have been reported associations between *cagA* and the prevalence of the various diseases. For instance, in a cross-sectional study of 113 *H. pylori* positive Ghanaian population with dyspepsia, the prevalence of the organism harboring the *cagA* virulence factor was found to be 74.8% and a persistent association existed between *cagA*-(hydrophilic region) and duodenal ulcer (Table 1) [66]. A high prevalence 90–97% has also been demonstrated in Nigeria [61, 137, 138]. A study by Smith et al. found the presence of *cagA* infection in 91% of patients in Nigeria with nonulcer dyspepsia and 95% of them had duodenal ulcer. The study, however, concluded that no association existed between the studied genotypes and duodenal ulcer disease in that particular population [139]. These reported observations on prevalence are consistent with studies in India (96% among duodenal ulcer patients), [140] Gauteng (87% among asymptomatic children age between 6 and 15 years), [141] Alaskans-US (85%), [142] China (89.3% in patients with upper gastrointestinal diseases), and Taiwan (83% in isolates from patients with chronic gastritis and peptic ulcer) [143]. Another report in Nigeria by Mnena et al. [86], however, identified only 29% out of 22 *H. pylori* positive patients to have the *cagA* gene. The rather low prevalence of *cagA* in that population explains the low rate of recurrence of severe gastrointestinal disorders among the studied patients. Elsewhere in Senegal, a total of 117 *H. pylori* culture-positive patients yielded 73.3% of the *cagA* gene and this was observed to be strongly related to GC. Breurec et al. [70] also found 73.3% of isolates from Senegalese patients to be positive for *cagA* gene which was also associated with GC. These findings are synonymous to findings reported elsewhere on the African continent [78, 144]. Similar results also exist for bacterial strains with *cagA* in countries such as Iraq, Iran, and Turkey with a reported prevalence of 71%, 76%, and 78%, respectively. The presence of *cagA* identified was found to have a significant association with the incidence of peptic ulcer disease in Turkey and Iraq but not in Iran [55, 145]. Also, in The Gambia, similar results were obtained for the proportion of samples that were *cagA* positive using DNA from biopsies and culture. A prevalence of 58.3% and 61.7% were recorded for DNA from biopsies and culture, respectively [61, 146, 147]. Generally, 61.2% of these Gambian patients were found to have the *cagA* gene only, while 17.4% were positive for the *cag* empty site only. A rate of 19% was also found to be positive for both. The study, however, did not state clearly the relation of the detected gene to the pathology of the disease.

3.2.2. *Vacuolating Cytotoxin Gene A (vacA)*. The *vacA* is a 140-kDa polypeptide that is secreted from the bacteria and

delivered in an active form to host cells, where it exerts its activity [148]. All *H. pylori* strains contain the gene that encodes *vacA* although not all are fully cytotoxic [135, 149]. The *vacA* gene has three regions of genetic allelic diversity namely; intermediate (i1 and i2), the signal (s1 and s2), and the middle regions (m1 and m2) which determines the difference in vacuolating abilities [115, 121]. Damage to epithelial cells by the *vacA* gene is achieved by inducing the formation of vacuoles. The degree of cytotoxic activity of the toxin varies from strain to strain [83, 150] with the highest vacuolating activity occurring in s1/m1 genotypes. Activity is intermediate in s1/m2 genotypes and absent in s2/m2 genotypes [135]. Among the types, the s1/i1/m1 *vacA* is repeatedly associated with genopositive *cagA* [151] and neither of the virulence indicators is considered a self-regulating influence for the outcome of the disease [54]. The risk of severe clinical outcome is therefore greater when several virulence markers exist. Apart from the impact of the differences in *vacA* toxicity among strains, the expression of *vacA* during *H. pylori* infection varies widely and is associated with the degree of inflammation and presence of atrophy [152]. This implies that the risk of development of disease by an *H. pylori* infected person is not only reliance on the type of *vacA* of the infecting strain but also on the level of expression of the gene [152]. The study by Sinnet et al. and Amilon et al. [153] has shown that *vacA* expression level and gastric inflammation is associated with polymorphism at nucleotide +28 with the *vacA* 5' untranslated region of the transcript. Such effects on *vacA* transcript levels are important in their provision of possible additional risk markers for determining patients at a higher risk of developing severe duodenal or gastric diseases [152].

The presence of *vacA* is therefore very vital in the disease outcome of *H. pylori* infection and hence its identification in infected individuals could be very helpful in prediction, diagnosis, and subsequent treatment approach of infection. In several of the research performed on *H. pylori* in the studied countries, only a few delve into the virulence factors. Among Ghanaian patients presenting with dyspepsia, the prevalence of *vacAs1m1* was found to be 25.2% and that of *vacAs1m2* was 8.2%. Majority of patients who recorded a positive result for the presence of *vacAs1* were also positive for *cagA* [66]. Infection with *vacA s1m1 H. pylori* genotypes have been found to be associated with an increased risk of duodenal ulcer disease and this was evident in the study by Archampong et al. [142]. The association identified in this study was consistent with other studies [46, 154].

In Nigeria, Smith et al. detected 98% of *vacAs1* in 40 *H. pylori* strains [139] while Harrison et al. found 92.8% detectable *vacA* levels [61] similar to the 90.6% obtained in South Africa [141]. In the study by Harrison et al., a bulk of the isolates harbored the *vacA* s1, m2 genotype, followed by the s1, m1 genotype. There was no detection of the *vacA* s2 genotype, consistent with the outcome of a study by Wei et al. [155]. Harrison et al. also detected no *vacA* m1 and results obtained for the two patient groups being those presenting with duodenal ulcer disease and nonulcer dyspepsia showed no significant difference. Mnena et al. [86] found the following among Nigerian dyspeptic patients; For the *vacA* genotypes, the s1c/m2 genotype formed 79% of the *H. pylori* infections

while 8% was found for s1b/m2 genotype similar to findings in China in which 69.5% and 2.5% were detected for s1cm2 and s1bm2, respectively [155]. Occurring at 4% each were three different genotypes; s1c/m1, s1c/m1/m2, and s1c/s2/m2. The most prevalent (83%) among the various genotypes was the moderate virulence type of s1m2. The most virulent (s1m1) and least virulent (s2m2) genotypes were found to be 8% and 4% respectively. In the Gambia, the more toxigenic *VacA* s1 and m1 gene were demonstrated in 76.9% and 45.5% of subjects, respectively [146, 147], although the relation to clinical outcome was not clearly stated. *vacAs2* and *vacAm2* were, however, found to be at a rate of 19% and 29.8% respectively. Among Senegalese patients, there was a detection of high-vacuolization isotypes in which 57.1% were s1im1 subtype, while 21.9% had the s1im2 subtype. The s1 *vacA* allele were found to be associated with GC [70].

3.2.3. Induced by Contact with Epithelium Gene A (*IceA*). *IceA* has two central allelic variants namely, *iceA1* and *iceA2* [156, 157] and the relationship between them and clinical outcome is quite controversial. While some studies have proven that *iceA1/iceA2* may be directly involved in diseases of the gastrointestinal system, [158, 159] others have demonstrated contrary findings [115, 116]. *IceA1* is, however, upregulated upon the contact of *H. pylori* with the gastric epithelium and has been found to be a probable marker for peptic ulcer disease while *IceA2* is not considered a molecular marker of more virulent *H. pylori* strains [160, 161]. In a study by Smith et al. [139] conducted in Nigeria, all *H. pylori* isolates contained the *iceA* gene. In total, a considerably higher rate of 90.2% (37 isolates) were positive for *iceA1* out of which 94.7% (18 isolates) were obtained from duodenal ulcer patients while 86.4% (19 isolates) were from nonulcer dyspepsia patients. Only one isolate from a nonulcer dyspepsia patient yielded both *iceA1* and *iceA2* in the PCR product. Babaei et al. [91] have confirmed that there is a strong relationship between duodenal ulcer and the genotypes of *iceA1(+)/iceA2(-)* [91]. The study identified a 61% *iceA1(+)/iceA2(-)* prevalence in peptic ulcer patients and 17.6% prevalence of this genotype for *H. pylori* infected nonpeptic ulcer disease patients. Wei et al. [155] have also recorded a gastric cancer associated *iceA1* prevalence of 70.1% which were stated to be consistent with findings from Korea, China, Tunisia, and Thailand.

4. Concluding Remarks and Further Studies

The understanding of *H. pylori* infection and disease progression has improved over the past few years in countries of West Africa although prevalence of infection is generally high. Available data indicate that studies on virulence factors are inadequate although few countries with research data on it studied factors such as *cagA* and *vacA* and observed a high prevalence. While information in literature has demonstrated the role of other factors such as *BabA2*, *OipA*, *dupA* *IceA*, the genes that encode for glycosyl transferases and the allelic variants of some of the studied factors on the pathology of *H. pylori* infection, these areas are insufficiently dealt with in studies across countries of the Western part of the African

continent. Expounding the roles of *H. pylori* virulence factors in pathogenesis and clinical outcomes would greatly benefit vaccine and alternative drug therapy development.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] A. Tonkic, M. Tonkic, P. Lehours, and F. Mégraud, "Epidemiology and diagnosis of *Helicobacter pylori* infection," *Helicobacter*, vol. 17, pp. 1–8, 2012.
- [2] P. Malfertheiner, F. Megraud, C. A. O'Morain et al., "Management of *Helicobacter pylori* infection-the Maastricht V/Florence consensus report," *Gut*, vol. 66, no. 1, pp. 6–30, 2017.
- [3] H. M. Mitchell, *Epidemiology of Infection*, chapter 2, ASM Press, Washington (DC), 2001.
- [4] S. Suerbaum and P. Michetti, "*Helicobacter pylori* infection," *New England Journal of Medicine*, vol. 347, no. 15, pp. 1175–1186, 2002.
- [5] M. Banić, F. Franceschi, Z. Babić, and A. Gasbarrini, "Extragastric Manifestations of *Helicobacter pylori* Infection," *Helicobacter*, vol. 17, pp. 49–55, 2012.
- [6] W. Xia, X. Zhang, J. Wang, C. Sun, and L. Wu, "Survey of anaemia and *Helicobacter pylori* infection in adolescent girls in Suihua, China and enhancement of iron intervention effects by *H. pylori* eradication," *British Journal of Nutrition*, vol. 108, no. 2, pp. 357–362, 2012.
- [7] K. Kohda, T. Kuga, K. Kogawa et al., "Effect of *Helicobacter pylori* eradication on platelet recovery in Japanese patients with chronic idiopathic thrombocytopenic purpura and secondary autoimmune thrombocytopenic purpura," *British Journal of Haematology*, vol. 118, no. 2, pp. 584–588, 2002.
- [8] M. Rogha, M. Nikvarz, Z. Pourmoghaddas, K. Shirneshan, D. Dadkhah, and M. Pourmoghaddas, "Is *Helicobacter pylori* infection a risk factor for coronary heart disease?," *ARYA Atherosclerosis*, vol. 8, no. 1, p. 5, 2012.
- [9] F. Franceschi, G. Niccoli, G. Ferrante et al., "CagA antigen of *Helicobacter pylori* and coronary instability: insight from a clinico-pathological study and a meta-analysis of 4241 cases," *Atherosclerosis*, vol. 202, no. 2, pp. 535–542, 2009.
- [10] G. Isaeva, E. Abuzarova, I. Valeeva, O. Pozdeev, and E. Murav'eva, "*Helicobacter pylori* in patients with disorders of hepatobiliary system," *Zhurnal Mikrobiologii, Epidemiologii, I Immunobiologii*, vol. 2, pp. 96–101, 2009.
- [11] T. Pirouz, L. Zounubi, H. Keivani, N. Rakhshani, and M. Hormazdi, "Detection of *Helicobacter pylori* in paraffin-embedded specimens from patients with chronic liver diseases, using the amplification method," *Digestive Diseases and Sciences*, vol. 54, no. 7, pp. 1456–1459, 2009.
- [12] D. Ortiz-Princz, G. Daoud, A. Salgado-Sabel, and M. Cavazza, "*Helicobacter pylori* infection in children: should it be carefully assessed?," *European Review for Medical Pharmacological Sciences*, vol. 20, no. 9, pp. 1798–1813, 2016.
- [13] D. Jaspersen, M. Kulig, J. Labenz et al., "Prevalence of extra-oesophageal manifestations in gastro-oesophageal reflux disease: an analysis based on the ProGERD study," *Alimentary Pharmacology and Therapeutics*, vol. 17, no. 12, pp. 1515–1520, 2003.
- [14] B. Marshall and J. R. Warren, "Unidentified curved bacilli in the stomach of patients with gastritis and peptic ulceration," *The Lancet*, vol. 323, no. 8390, pp. 1311–1315, 1984.
- [15] A. Seck, M. Mbengue, A. Gassama-Sow, L. Diouf, M. M. Ka, and C. S.-B. Boye, "Antibiotic susceptibility of *Helicobacter pylori* isolates in Dakar, Senegal," *The Journal of Infection in Developing Countries*, vol. 3, no. 2, pp. 137–140, 2009.
- [16] M. B. Adinortey, C. Ansah, C. A. Adinortey, A. S. Bockarie, M. T. Morna, and D. H. Amewowor, "Isolation of *Helicobacter pylori* from gastric biopsy of dyspeptic patients in Ghana and in vitro preliminary assessment of the effect of dissotis rotundifolia extract on its growth," *Journal of Tropical Medicine*, vol. 2018, pp. 1–6, 2018.
- [17] Y. Urita, T. Watanabe, N. Kawagoe et al., "Role of infected grandmothers in transmission of *Helicobacter pylori* to children in a Japanese rural town," *Journal of Paediatrics and Child Health*, vol. 49, no. 5, pp. 394–398, 2013.
- [18] A. Mentis, P. Lehours, and F. Mégraud, "Epidemiology and diagnosis of *Helicobacter pylori* infection," *Helicobacter*, vol. 20, pp. 1–7, 2015.
- [19] L. M. Brown, "*Helicobacter pylori*: epidemiology and routes of transmission," *Epidemiologic Reviews*, vol. 22, no. 2, pp. 283–297, 2000.
- [20] K. L. Goh, W. K. Chan, S. Shiota, and Y. Yamaoka, "Epidemiology of *Helicobacter pylori* infection and public health implications," *Helicobacter*, vol. 16, pp. 1–9, 2011.
- [21] F. Vale and J. Vitor, "Transmission pathway of *Helicobacter pylori*: does food play a role in rural and urban areas?," *International Journal of Food Microbiology*, vol. 138, no. 1–2, pp. 1–12, 2010.
- [22] A. Covacci, J. L. Telford, G. Del Giudice, J. Parsonnet, and R. Rappuoli, "*Helicobacter pylori* virulence and genetic geography," *Science*, vol. 284, no. 5418, pp. 1328–1333, 1999.
- [23] M. Go, "Natural history and epidemiology of *Helicobacter pylori* infection," *Alimentary Pharmacology & Therapeutics*, vol. 16, pp. 3–15, 2002.
- [24] M. M. Khalifa, R. R. Sharaf, and R. K. Aziz, "*Helicobacter pylori*: a poor man's gut pathogen?," *Gut pathogens*, vol. 2, no. 1, p. 2, 2010.
- [25] R. Hunt, S. Xiao, F. Megraud et al., "*Helicobacter pylori* in developing countries. World Gastroenterology Organisation Global Guideline," *Journal of Gastrointestinal and Liver Diseases*, vol. 20, no. 3, pp. 299–304, 2011.
- [26] P. K. Bardhan, "Epidemiological features of *Helicobacter pylori* infection in developing countries," *Clinical Infectious Diseases*, vol. 25, no. 5, pp. 973–978, 1997.
- [27] M. Zamani, F. Ebrahimitabar, V. Zamani et al., "Systematic review with meta-analysis: the worldwide prevalence of *Helicobacter pylori* infection," *Alimentary Pharmacology & Therapeutics*, vol. 47, no. 7, pp. 868–876, 2018.
- [28] J. K. Hooi, W. Y. Lai, W. K. Ng et al., "Global prevalence of *Helicobacter pylori* infection: systematic review and meta-analysis," *Gastroenterology*, vol. 153, no. 2, pp. 420–429, 2017.
- [29] C. Holcombe, "*Helicobacter pylori*: the African enigma," *Gut*, vol. 33, no. 4, pp. 429–431, 1992.
- [30] K. Singh and U. C. Ghoshal, "Causal role of *Helicobacter pylori* infection in gastric cancer: an Asian enigma," *World Journal of Gastroenterology*, vol. 12, no. 9, p. 1346, 2006.

- [31] U. C. Ghoshal, R. Chaturvedi, and P. Correa, "The enigma of *Helicobacter pylori* infection and gastric cancer," *Indian Journal of Gastroenterology*, vol. 29, no. 3, pp. 95–100, 2010.
- [32] D. Y. Graham, H. Lu, and Y. Yamaoka, "African, Asian or Indian enigma, the East Asian *Helicobacter pylori*: facts or medical myths," *Journal of Digestive Diseases*, vol. 10, no. 2, pp. 77–84, 2009.
- [33] T. Irino, H. Takeuchi, M. Terashima, T. Wakai, and Y. Kitagawa, "Gastric cancer in Asia: unique features and management," *American Society of Clinical Oncology Educational Book*, vol. 37, pp. 279–291, 2017.
- [34] Y. Hu, Y. Zhu, and N.-H. Lu, "Novel and effective therapeutic regimens for *Helicobacter pylori* in an era of increasing antibiotic resistance," *Frontiers in Cellular and Infection Microbiology*, vol. 7, 2017.
- [35] E. Kuipers, J. Thijs, and H. Festen, "The prevalence of *Helicobacter pylori* in peptic ulcer disease," *Alimentary Pharmacology & Therapeutics*, vol. 9, pp. 59–69, 1995.
- [36] M. Gasparetto, M. Pescarin, and G. Guariso, "*Helicobacter pylori* eradication therapy: current availabilities," *ISRN Gastroenterology*, vol. 2012, pp. 1–8, 2012.
- [37] L. Boyanova, "*Helicobacter pylori* resistance to antibiotics *Helicobacter pylori*," vol. 201, 2011.
- [38] F. Aziz, X. Chen, X. Yang, and Q. Yan, "Prevalence and correlation with clinical diseases of *Helicobacter pylori* cagA and vacA genotype among gastric patients from Northeast China," *BioMed Research International*, vol. 2014, 2014.
- [39] M. Akeel, E. Elmakki, A. Shehata et al., "Prevalence and factors associated with *H. pylori* infection in Saudi patients with dyspepsia," *Electronic Physician*, vol. 10, no. 9, pp. 7279–7286, 2018.
- [40] J. G. Fox and T. C. Wang, "Inflammation, atrophy, and gastric cancer," *The Journal of Clinical Investigation*, vol. 117, no. 1, pp. 60–69, 2007.
- [41] J. Alam, S. Maiti, P. Ghosh et al., "Significant association of the dupA gene of *Helicobacter pylori* with duodenal ulcer development in a South-east Indian population," *Journal of Medical Microbiology*, vol. 61, no. Pt_9, pp. 1295–1302, 2012.
- [42] M. Safavi, R. Sabourian, and A. Foroumadi, "Treatment of *Helicobacter pylori* infection: current and future insights," *World Journal of Clinical Cases*, vol. 4, no. 1, p. 5, 2016.
- [43] V. Conteduca, D. Sansonno, G. Lauletta, S. Russi, G. Ingravallo, and F. H. Dammacco, "Pylori infection and gastric cancer: state of the art," *International Journal of Oncology*, vol. 42, no. 1, pp. 5–18, 2013.
- [44] F. Abdulkareem, K. Badmos, and N. Awolola, "*Helicobacter pylori* infections and gastric cancer: the West African experience Niger," *Journal Gastroenterology Hepatology*, vol. 7, no. 1, pp. 25–30, 2015.
- [45] B. Zhao, J. Zhao, W.-F. Cheng et al., "Efficacy of *Helicobacter pylori* eradication therapy on functional dyspepsia," *Journal of Clinical Gastroenterology*, vol. 48, no. 3, pp. 241–247, 2014.
- [46] S. Kabir, "Effect of *Helicobacter pylori* eradication on incidence of gastric cancer in human and animal models: underlying biochemical and molecular events," *Helicobacter*, vol. 14, no. 3, pp. 159–171, 2009.
- [47] H. M. Malaty, D. G. Evans, D. J. Evans, and D. Y. Graham, "*Helicobacter pylori* in Hispanics: comparison with blacks and whites of similar age and socioeconomic class," *Gastroenterology*, vol. 103, no. 3, pp. 813–816, 1992.
- [48] H. M. Malaty, D. Y. Graham, W. A. Wattigney, S. R. Srinivasan, M. Osato, and G. S. Berenson, "Natural history of *Helicobacter pylori* infection in childhood: 12-year follow-up cohort study in a biracial community," *Clinical Infectious Diseases*, vol. 28, no. 2, pp. 279–282, 1999.
- [49] B. A. Salih, "*Helicobacter pylori* infection in developing countries: the burden for how long?," *Saudi Journal of Gastroenterology: Official Journal of the Saudi Gastroenterology Association*, vol. 15, no. 3, p. 201, 2009.
- [50] M. A. Tkachenko, N. Z. Zhannat, L. V. Erman et al., "Dramatic changes in the prevalence of *Helicobacter pylori* infection during childhood: a 10-year follow-up study in Russia," *Journal of Pediatric Gastroenterology and Nutrition*, vol. 45, no. 4, pp. 428–432, 2007.
- [51] C. Malcolm, W. MacKay, A. Shepherd, and L. Weaver, "*Helicobacter pylori* in children is strongly associated with poverty," *Scottish Medical Journal*, vol. 49, no. 4, pp. 136–138, 2004.
- [52] F. Cataldo, J. Simpoire, P. Greco, D. Ilboudo, and S. Musumeci, "*Helicobacter pylori* infection in Burkina Faso: an enigma within an enigma," *Digestive and Liver Disease*, vol. 36, no. 9, pp. 589–593, 2004.
- [53] J. Thomas, A. Dale, and J. Bunn, "Early *Helicobacter pylori* colonisation: the association with growth faltering in The Gambia," *Archives of Disease in Childhood*, vol. 89, no. 12, pp. 1149–1154, 2004.
- [54] J. G. Kusters, A. H. van Vliet, and E. J. Kuipers, "Pathogenesis of *Helicobacter pylori* infection," *Clinical Microbiology Reviews*, vol. 19, no. 3, pp. 449–490, 2006.
- [55] N. R. Hussein, M. Mohammadi, Y. Talebkhan et al., "Differences in virulence markers between *Helicobacter pylori* strains from Iraq and those from Iran: potential importance of regional differences in *H. pylori*-associated disease," *Journal of Clinical Microbiology*, vol. 46, no. 5, pp. 1774–1779, 2008.
- [56] P. Sullivan, J. Thomas, D. Wight et al., "*Helicobacter pylori* in Gambian children with chronic diarrhoea and malnutrition," *Archives of Disease in Childhood*, vol. 65, no. 2, pp. 189–191, 1990.
- [57] A. Olokoba, W. Gashau, S. Bwala, A. Adamu, and F. Salawu, "*Helicobacter pylori* infection in Nigerians with dyspepsia," *Ghana Medical Journal*, vol. 47, no. 2, pp. 79–81, 2013.
- [58] E. Ray-Offor, C. Obiorah, "*Helicobacter pylori* and precancerous lesions of the stomach in a Nigerian metropolis: a cohort study," *Nigerian Journal of Clinical Practice*, vol. 21, no. 3, pp. 375–379, 2018.
- [59] A. C. Jemilohun, J. A. Otegbayo, S. O. Ola, O. A. Oluwasola, and A. Akere, "Prevalence of *Helicobacter pylori* among Nigerian patients with dyspepsia in Ibadan," *Pan African Medical Journal*, vol. 6, no. 1, 2010.
- [60] M. Tanko, A. Manasseh, and G. Echejoh, "Relation between *Helicobacter pylori*, inflammatory (neutrophil) activity, chronic gastritis, gastric atrophy and intestinal metaplasia," *Nigerian Journal of Clinical Practice*, vol. 11, no. 3, pp. 270–274, 2008.
- [61] U. Harrison, M. A. Fowora, A. T. Seriki et al., "*Helicobacter pylori* strains from a Nigerian cohort show divergent antibiotic resistance rates and a uniform pathogenicity profile," *PLoS One*, vol. 12, no. 5, p. e0176454, 2017.
- [62] O. O. Lawal, O. Rotimi, and I. Okeke, "*Helicobacter pylori* in gastroduodenal diseases," *Journal of the National Medical Association*, vol. 99, no. 1, p. 31, 2007.

- [63] I. Austarheim, K. T. Inngjerdingen, B. S. Paulsen et al., "Chromatographic immunoassays for *Helicobacter pylori* detection - are they reliable in Mali, West Africa?," *Pan African Medical Journal*, vol. 2, 2009.
- [64] T. Archampong, R. H. Asmah, E. K. Wiredu, R. K. Gyasi, K. N. Nkrumah, and K. Rajakumar, "Epidemiology of *Helicobacter pylori* infection in dyspeptic Ghanaian patients," *Pan African Medical Journal*, vol. 20, no. 1, 2015.
- [65] B. Baako and R. Darko, "Incidence of *Helicobacter pylori* infection in Ghanaian patients with dyspeptic symptoms referred for upper gastrointestinal endoscopy," *West African Journal of Medicine*, vol. 15, no. 4, pp. 223–227, 1996.
- [66] T. N. Archampong, R. H. Asmah, E. K. Aidoo et al., "*Helicobacter pylori* cagA and vacA genes in dyspeptic Ghanaian patients," *BMC Research Notes*, vol. 10, no. 1, 2017.
- [67] P. Mbaye, A. Diallo, F. Klotz, and G. Michel, "*Helicobacter pylori* and upper digestive disease at the Main Hospital of Dakar. Study apropos of 105 consecutive endoscopies," *Dakar Medical*, vol. 40, no. 2, pp. 187–191, 1995.
- [68] M. Mbengue, A. Seck, D. Dia et al., "Gastroduodenal peptic ulcer: descriptive study," *Dakar Medical*, vol. 48, no. 3, pp. 176–180, 2003.
- [69] K. Doh, I. Thiam, A. Halim, R. Takin, and G. Woto-Gaye, "Pathological overview of chronic gastritis in Senegal: results of upper gastrointestinal tract endoscopies," *Médecine et Sante Tropicales*, vol. 27, no. 4, pp. 439–442, 2017.
- [70] S. Breurec, R. Michel, A. Seck et al., "Clinical relevance of cagA and vacA gene polymorphisms in *Helicobacter pylori* isolates from Senegalese patients," *Clinical Microbiology and Infection*, vol. 18, no. 2, pp. 153–159, 2012.
- [71] D. Dia, A. Seck, and M. Mbengue, "*Helicobacter pylori* and gastroduodenal lesions in Dakar, Senegal," *Medecine tropicale: revue du Corps de sante colonial*, vol. 70, no. 4, pp. 367–370, 2010.
- [72] M. Mbengue, M. Diouf, and J. Dangou, "Frequency of *Helicobacter pylori* infection in symptomatic patients in Senegal," *Medecine tropicale: revue du Corps de sante colonial*, vol. 57, no. 3, pp. 256–258, 1997.
- [73] M. Diomande, J. Flejou, and F. Potet, "Chronic gastritis and *Helicobacter pylori* infection on the Ivory Coast. A series of 277 symptomatic patients," *Gastroenterologie clinique et biologique*, vol. 15, no. 10, pp. 711–716, 1991.
- [74] N. Kodjoh, A. Hountondji, and B. Addra, "The contribution of endoscopy in the diagnosis of esophago-gastro-duodenal disorders in a tropical milieu. Experience in Benin with 930 examinations," *Annales de gastroenterologie et d'hepatologie*, vol. 27, no. 6, pp. 261–267, 1991.
- [75] B. Aguemou, M. Struelens, A. Massougbdji, and E. M. Ouendo, "Prevalence and risk-factors for *Helicobacter pylori* infection in urban and rural Beninese populations," *Clinical Microbiology and Infection*, vol. 11, no. 8, pp. 611–617, 2005.
- [76] B. Salih, "*Helicobacter pylori* infection in developing countries: the burden for how long?," *Saudi Journal of Gastroenterology*, vol. 15, no. 3, pp. 201–207, 2009.
- [77] B. Roesler, E. M. Rabelo-Gonçalves, and J. M. Zeitune, "Virulence factors of *Helicobacter pylori*: a review," *Clinical Medicine Insights Gastroenterology*, vol. 7, CGast S13760 pages, 2014.
- [78] F.-C. Han, H.-C. Ng, and B. Ho, "Stability of randomly amplified polymorphic DNA fingerprinting in genotyping clinical isolates of *Helicobacter pylori*," *World Journal of Gastroenterology*, vol. 9, no. 9, p. 2021, 2003.
- [79] D. A. Israel, N. Salama, U. Krishna et al., "*Helicobacter pylori* genetic diversity within the gastric niche of a single human host," *Proceedings of the National Academy of Sciences*, vol. 98, no. 25, pp. 14625–14630, 2001.
- [80] D. Falush, T. Wirth, and B. Linz, "Traces of human migrations in *Helicobacter pylori* populations," *Science*, vol. 299, no. 5612, pp. 1582–1585, 2003.
- [81] A. Covacci, S. Censini, M. Bugnoli et al., "Molecular characterization of the 128-kDa immunodominant antigen of *Helicobacter pylori* associated with cytotoxicity and duodenal ulcer," *Proceedings of the National Academy of Sciences*, vol. 90, no. 12, pp. 5791–5795, 1993.
- [82] A. Nomura, G. N. Stemmermann, P.-H. Chyou, I. Kato, G. I. Perez-Perez, and M. J. Blaser, "*Helicobacter pylori* infection and gastric carcinoma among Japanese Americans in Hawaii," *New England Journal of Medicine*, vol. 325, no. 16, pp. 1132–1136, 1991.
- [83] L. E. Wroblewski, R. M. Peek, and K. T. Wilson, "*Helicobacter pylori* and gastric cancer: factors that modulate disease risk," *Clinical Microbiology Reviews*, vol. 23, no. 4, pp. 713–739, 2010.
- [84] G. La Torre, G. Chiaradia, F. Gianfagna et al., "Smoking status and gastric cancer risk: an updated meta-analysis of case-control studies published in the past ten years," *Tumori Journal*, vol. 95, no. 1, pp. 13–22, 2009.
- [85] C. A. Gonzalez, L. Lujan-Barroso, M. Jenab et al., "Fruit and vegetable intake and the risk of gastric adenocarcinoma: a reanalysis of the European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST) study after a longer follow-up," *International Journal of Cancer*, vol. 131, no. 12, pp. 2910–2919, 2012.
- [86] E. Mnena, U. Ebele, and Emmanuel N. Risk, "Factors associated with *Helicobacter pylori* infections in Makurdi Northcentral Nigeria," *Journal of Infectious Diseases Therapy*, vol. 5, no. 325, Article ID 1000325, pp. 2332–0877, 2017.
- [87] D. Jemikalajah and G. Okogun, "Health point prevalence of *Helicobacter pylori* in central hospital Warri, Nigeria," *African Journal of Cellular Pathology*, vol. 3, no. 12, pp. 57–60, 2014.
- [88] A. K. Bello, A. B. Umar, and M. M. Borodo, "Prevalence and risk factors for *Helicobacter pylori* infection in gastroduodenal diseases in Kano, Nigeria," *African Journal of Medical and Health Sciences*, vol. 17, no. 1, p. 41, 2018.
- [89] M. P. Dore, A. R. Sepulveda, H. El-Zimaity et al., "Isolation of *Helicobacter pylori* from sheep-implications for transmission to humans," *The American Journal of Gastroenterology*, vol. 96, no. 5, pp. 1396–1401, 2001.
- [90] M. P. Dore, A. R. Sepulveda, M. S. Osato, G. Realdi, and D. Y. Graham, "*Helicobacter pylori* in sheep milk," *The Lancet*, vol. 354, no. 9173, p. 132, 1999.
- [91] V. Babaei, Y. Saghaei, H. Z. Gohardani, F. Vali, and S. Teimourian, "Effects of different environmental factors and virulence factors, dupA and iceA genes, of *Helicobacter pylori* on peptic Ulcer," *Jundishapur Journal of Microbiology*, vol. 10, no. 5, 2017.
- [92] G. Parasher and G. L. Eastwood, "Smoking and peptic ulcer in the *Helicobacter pylori* era," *European Journal of Gastroenterology & Hepatology*, vol. 12, no. 8, pp. 843–853, 2000.
- [93] P. Maity, K. Biswas, S. Roy, R. K. Banerjee, and U. Bandyopadhyay, "Smoking and the pathogenesis of gastroduodenal ulcer—recent mechanistic update," *Molecular and Cellular Biochemistry*, vol. 253, no. 1–2, pp. 329–338, 2003.

- [94] W. K. Wu and C. H. Cho, "The pharmacological actions of nicotine on the gastrointestinal tract," *Journal of Pharmacological Sciences*, vol. 94, no. 4, pp. 348–358, 2004.
- [95] L. Zhang, J. W. Ren, C. C. M. Wong et al., "Effects of cigarette smoke and its active components on ulcer formation and healing in the gastrointestinal mucosa," *Current Medicinal Chemistry*, vol. 19, no. 1, pp. 63–69, 2012.
- [96] D. Garrow and M. H. Deleage, "Risk factors for gastrointestinal ulcer disease in the US population," *Digestive Diseases and Sciences*, vol. 55, no. 1, pp. 66–72, 2010.
- [97] T. Shimamoto, N. Yamamichi, S. Kodashima et al., "No association of coffee consumption with gastric ulcer, duodenal ulcer, reflux esophagitis, and non-erosive reflux disease: a cross-sectional study of 8,013 healthy subjects in Japan," *PLoS One*, vol. 8, no. 6, p. e65996, 2013.
- [98] L. Wu, S. Chen, Y. Zhang, and H. Pan, "The effect of low concentrations of ethanol on gastric adenocarcinoma cell lines," *Archives of Biological Sciences*, vol. 66, no. 1, pp. 317–321, 2014.
- [99] M. Tamura, H. Matsui, T. Kaneko, and I. Hyodo, "Alcohol is an oxidative stressor for gastric epithelial cells: detection of superoxide in living cells," *Journal of Clinical Biochemistry and Nutrition*, vol. 53, no. 2, pp. 75–80, 2013.
- [100] H. Ahmad, A. Wadud, N. Jahan, M. Khazir, and G. Sofi, "Evaluation of anti-ulcer activity of hydro alcoholic extract of Post Sumaq (*Rhus coriaria* Linn.) in Ethanol induced Gastric ulcer in experimental rats," *International Research Journal of Medical Sciences*, vol. 1, no. 10, pp. 7–12, 2013.
- [101] T.-L. M. Nguyen, S. S. Khurana, C. J. Bellone et al., "Autoimmune gastritis mediated by CD4+ T cells promotes the development of gastric cancer," *Cancer Research*, vol. 73, no. 7, pp. 2117–2126, 2013.
- [102] Group ES, "Epidemiology of, and risk factors for, *Helicobacter pylori* infection among 3194 asymptomatic subjects in 17 populations. The EUROGAST study group," *Gut*, vol. 34, pp. 1672–1676, 1993.
- [103] O. Amaral, I. Fernandes, N. Veiga et al., "Living conditions and *Helicobacter pylori* in adults," *BioMed Research International*, vol. 2017, pp. 1–5, 2017.
- [104] B. Qu, J. Su, Z. Wang et al., "Effect of *H. pylori* infection on cytokine profiles and oxidative balance in subjects with chronic alcohol ingestion," *PLoS One*, vol. 10, no. 6, p. e0129352, 2015.
- [105] L. Zhang, G. D. Eslick, H. H.-X. Xia, C. Wu, N. Phung, and N. J. Talley, "Relationship between alcohol consumption and active *Helicobacter pylori* infection," *Alcohol and Alcoholism*, vol. 45, no. 1, pp. 89–94, 2009.
- [106] S.-Y. Liu, X.-C. Han, J. Sun, G.-X. Chen, X.-Y. Zhou, and G.-X. Zhang, "Alcohol intake and *Helicobacter pylori* infection: a dose-response meta-analysis of observational studies," *Infectious Diseases*, vol. 48, no. 4, pp. 303–309, 2016.
- [107] A. T. B. Abadi, "Strategies used by *Helicobacter pylori* to establish persistent infection," *World Journal of Gastroenterology*, vol. 23, no. 16, p. 2870, 2017.
- [108] Y. Huang, Q.-L. Wang, D.-D. Cheng, W.-T. Xu, and N.-H. Lu, "Adhesion and invasion of gastric mucosa epithelial cells by *Helicobacter pylori*," *Frontiers in Cellular and Infection Microbiology*, vol. 6, 2016.
- [109] A. Alvi, S. A. Ansari, N. Z. Ehtesham et al., "Concurrent proinflammatory and apoptotic activity of a *Helicobacter pylori* protein (HP986) points to its role in chronic persistence," *PLoS One*, vol. 6, no. 7, p. e22530, 2011.
- [110] J. I. Matos, H. A. de Sousa, R. Marcos-Pinto, and M. Dinis-Ribeiro, "*Helicobacter pylori* CagA and VacA genotypes and gastric phenotype: a meta-analysis," *European Journal of Gastroenterology & Hepatology*, vol. 25, no. 12, pp. 1431–1441, 2013.
- [111] J. Parsonnet, G. Friedman, N. Orentreich, and H. Vogelman, "Risk for gastric cancer in people with CagA positive or CagA negative *Helicobacter pylori* infection," *Gut*, vol. 40, no. 3, pp. 297–301, 1997.
- [112] S.-K. Wang, H.-F. Zhu, and B.-S. He, "CagA+*H. pylori* infection is associated with polarization of T helper cell immune responses in gastric carcinogenesis," *World Journal of Gastroenterology*, vol. 13, no. 21, p. 2923, 2007.
- [113] B. Roesler, S. Costa, and J. Zeitune, "Virulence factors of *Helicobacter pylori* and their relationship with the development of early and advanced distal intestinal type gastric adenocarcinoma," *Gastritis and Gastric Cancer-New Insights in Gastroprotection, Diagnosis and Treatments: InTech*, 2011.
- [114] Y. Yamaoka, T. Kodama, O. Gutierrez, J. G. Kim, K. Kashima, and D. Y. Graham, "Relationship between *Helicobacter pylori* iceA, cagA, and vacA status and clinical outcome: studies in four different countries," *Journal of Clinical Microbiology*, vol. 37, no. 7, pp. 2274–2279, 1999.
- [115] J. L. Rhead, D. P. Letley, M. Mohammadi et al., "A new *Helicobacter pylori* vacuolating cytotoxin determinant, the intermediate region, is associated with gastric cancer," *Gastroenterology*, vol. 133, no. 3, pp. 926–936, 2007.
- [116] C. Chung, A. Olivares, E. Torres, O. Yilmaz, H. Cohen, and G. Perez-Perez, "Diversity of VacA intermediate region among *Helicobacter pylori* strains from several regions of the world," *Journal of Clinical Microbiology*, vol. 48, no. 3, pp. 690–696, 2010.
- [117] M. Höcker and P. Hohenberger, "*Helicobacter pylori* virulence factors—one part of a big picture," *The Lancet*, vol. 362, no. 9391, pp. 1231–1233, 2003.
- [118] S. Miehke, C. Kirsch, K. Agha-Amiri et al., "The *Helicobacter pylori* vacA s1, m1 genotype and cagA is associated with gastric carcinoma in Germany," *International Journal of Cancer*, vol. 87, no. 3, pp. 322–327, 2000.
- [119] A. Leanza, M. Matteo, O. Crespo, P. Antelo, J. Olmos, and M. Catalano, "Genetic characterisation of *Helicobacter pylori* isolates from an Argentinean adult population based on cag pathogenicity island right-end motifs, lspA-glmM polymorphism and iceA and vacA genotypes," *Clinical Microbiology and Infection*, vol. 10, no. 9, pp. 811–819, 2004.
- [120] P. Ghiara, M. Marchetti, M. J. Blaser et al., "Role of the *Helicobacter pylori* virulence factors vacuolating cytotoxin, CagA, and urease in a mouse model of disease," *Infection and Immunity*, vol. 63, no. 10, pp. 4154–4160, 1995.
- [121] J. Atherton, R. Peek, K. Tham, T. Cover, and M. Blaser, "Clinical and pathological importance of heterogeneity in vacA, the vacuolating cytotoxin gene of *Helicobacter pylori*," *Gastroenterology*, vol. 112, no. 1, pp. 92–99, 1997.
- [122] S. Shiota, K. Murakawi, R. Suzuki, T. Fujioka, and Y. Yamaoka, "*Helicobacter pylori* infection in Japan," *Expert Review of Gastroenterology & Hepatology*, vol. 7, no. 1, pp. 35–40, 2013.
- [123] S. Shiota, R. Suzuki, and Y. Yamaoka, "The significance of virulence factors in *Helicobacter pylori*," *Journal of Digestive Diseases*, vol. 14, no. 7, pp. 341–349, 2013.
- [124] P. Jenks, F. Megraud, and A. Labigne, "Clinical outcome after infection with *Helicobacter pylori* does not appear to be

- reliably predicted by the presence of any of the genes of the cag pathogenicity island," *Gut*, vol. 43, no. 6, pp. 752–758, 1998.
- [125] A. van der Ende, Z.-J. Pan, A. Bart et al., "cagA-positive *Helicobacter pylori* populations in China and The Netherlands are distinct," *Infection and Immunity*, vol. 66, no. 5, pp. 1822–1826, 1998.
- [126] M. Rohde, J. Püls, R. Buhrdorf, W. Fischer, and R. Haas, "A novel sheathed surface organelle of the *Helicobacter pylori* cag type IV secretion system," *Molecular Microbiology*, vol. 49, no. 1, pp. 219–234, 2003.
- [127] S. Censini, C. Lange, Z. Xiang et al., "Cag, a pathogenicity island of *Helicobacter pylori*, encodes type I-specific and disease-associated virulence factors," *Proceedings of the National Academy of Sciences*, vol. 93, no. 25, pp. 14648–14653, 1996.
- [128] Y. Xia, Y. Yamaoka, Q. Zhu, I. Matha, and X. Gao, "A comprehensive sequence and disease correlation analyses for the C-terminal region of CagA protein of *Helicobacter pylori*," *PLoS One*, vol. 4, no. 11, p. e7736, 2009.
- [129] A. Vilar e Silva, M. R. S. da Junior, R. M. D. F. Vinagre et al., "Evaluation of the pattern of EPIYA motifs in the *Helicobacter pylori* cagA gene of patients with gastritis and gastric adenocarcinoma from the Brazilian Amazon region," *International Journal of Bacteriology*, vol. 2014, pp. 1–6, 2014.
- [130] M. Miura, N. Ohnishi, S. Tanaka, K. Yanagiya, and M. Hatakeyama, "Differential oncogenic potential of geographically distinct *Helicobacter pylori* CagA isoforms in mice," *International Journal of Cancer*, vol. 125, no. 11, pp. 2497–2504, 2009.
- [131] M. Naito, T. Yamazaki, R. Tsutsumi et al., "Influence of EPIYA-repeat polymorphism on the phosphorylation-dependent biological activity of *Helicobacter pylori* cagA," *Gastroenterology*, vol. 130, no. 4, pp. 1181–1190, 2006.
- [132] M. El Khadir, S. A. Boukhris, D.-A. Benajah et al., "*Helicobacter pylori* CagA EPIYA-C motifs and gastric diseases in Moroccan patients," *Infection, Genetics and Evolution*, vol. 66, pp. 120–129, 2018.
- [133] P. Correa and M. B. Piazuelo, "*Helicobacter pylori* infection and gastric adenocarcinoma," *US Gastroenterology & Hepatology Review*, vol. 7, no. 1, p. 59, 2011.
- [134] S. Maeda, H. Yoshida, and Y. Mitsuno, "Analysis of apoptotic and antiapoptotic signalling pathways induced by *Helicobacter pylori*," *Gut*, vol. 50, no. 6, pp. 771–778, 2002.
- [135] J. Atherton, P. Cao, R. M. Peek, M. K. Tummuru, M. J. Blaser, and T. L. Cover, "Mosaicism in vacuolating cytotoxin alleles of *Helicobacter pylori* association of specific vacA types with cytotoxin production and peptic ulceration," *Journal of Biological Chemistry*, vol. 270, no. 30, pp. 17771–17777, 1995.
- [136] D. Letley, A. Lastovica, J. Louw, C. Hawkey, and J. Atherton, "Allelic diversity of the *Helicobacter pylori* vacuolating cytotoxin gene in South Africa: rarity of the vacA s1a genotype and natural occurrence of an s2/m1 allele," *Journal of Clinical Microbiology*, vol. 37, no. 4, pp. 1203–1205, 1999.
- [137] M. Kidd, A. Lastovica, J. Atherton, and J. Louw, "Heterogeneity in the *Helicobacter pylori* vacA and cagA genes: association with gastroduodenal disease in South Africa?," *Gut*, vol. 45, no. 4, pp. 499–502, 1999.
- [138] M. A. Fowora, U. Breithaupt, and J. A. Otegbayo, "Molecular characterization of *Helicobacter pylori* infections in Nigeria," *Helicobacter*, vol. 1, no. 17, 2012.
- [139] S. I. Smith, C. Kirsch, K. S. Oyedemi et al., "Prevalence of *Helicobacter pylori* vacA, cagA and iceA genotypes in Nigerian patients with duodenal ulcer disease," *Journal of Medical Microbiology*, vol. 51, no. 10, pp. 851–854, 2002.
- [140] H. J. Arachchi, V. Kalra, B. Lal et al., "Prevalence of duodenal ulcer-promoting gene (dupA) of *Helicobacter pylori* in patients with duodenal ulcer in North Indian population," *Helicobacter*, vol. 12, no. 6, pp. 591–597, 2007.
- [141] A. Idowu, A. Mzukwa, U. Harrison et al., "Detection of *Helicobacter pylori* and its virulence genes (cagA, dupA, and vacA) among patients with gastroduodenal diseases in Chris Hani Baragwanath Academic Hospital, South Africa," *BMC Gastroenterology*, vol. 19, no. 1, 2019.
- [142] K. Miernyk, J. Morris, D. Bruden et al., "Characterization of *Helicobacter pylori* cagA and vacA genotypes among Alaskans and their correlation with clinical disease," *Journal of Clinical Microbiology*, vol. 49, no. 9, Article ID JCM-00469, pp. 3114–3121, 2011.
- [143] H.-J. Lin, C.-L. Perng, and W.-C. Lo, "*Helicobacter pylori* cagA, iceA and vacA genotypes in patients with gastric cancer in Taiwan," *World Journal of Gastroenterology: WJG*, vol. 10, no. 17, p. 2493, 2004.
- [144] O. Sanz-Pelaez, E. Santana-Rodriguez, A. Maroto et al., "*Helicobacter pylori* and cagA seroprevalence in sub-Saharan immigrants recently arrived to Gran Canaria (Spain)," *Scandinavian Journal of Infectious Diseases*, vol. 40, no. 9, pp. 756–758, 2008.
- [145] H. Saribasak, B. A. Salih, Y. Yamaoka, and E. Sander, "Analysis of *Helicobacter pylori* genotypes and correlation with clinical outcome in Turkey," *Journal of Clinical Microbiology*, vol. 42, no. 4, pp. 1648–1651, 2004.
- [146] O. Secka, M. Antonio, M. Tapgun et al., "PCR-based genotyping of *Helicobacter pylori* of Gambian children and adults directly from biopsy specimens and bacterial cultures," *Gut Pathogens*, vol. 3, no. 1, p. 5, 2011.
- [147] O. Secka, M. Antonio, D. E. Berg et al., "Mixed infection with cagA positive and cagA negative strains of *Helicobacter pylori* lowers disease burden in the Gambia," *PLoS One*, vol. 6, no. 11, p. e27954, 2011.
- [148] R. Leunk, P. Johnson, B. David, W. Kraft, and D. Morgan, "Cytotoxic activity in broth-culture filtrates of *Campylobacter pylori*," *Journal of Medical Microbiology*, vol. 26, no. 2, pp. 93–99, 1988.
- [149] T. L. Cover, M. K. Tummuru, P. Cao, S. A. Thompson, and M. J. Blaser, "Divergence of genetic sequences for the vacuolating cytotoxin among *Helicobacter pylori* strains," *Journal of Biological Chemistry*, vol. 269, no. 14, 10573 pages, 1994.
- [150] M. R. Amieva and E. M. El-Omar, "Host-bacterial interactions in *Helicobacter pylori* infection," *Gastroenterology*, vol. 134, no. 1, pp. 306–323, 2008.
- [151] M. Sugimoto, M. R. Zali, and Y. Yamaoka, "The association of vacA genotypes and *Helicobacter pylori*-related gastroduodenal diseases in the middle East," *European Journal of Clinical Microbiology & Infectious Diseases*, vol. 28, no. 10, pp. 1227–1236, 2009.
- [152] C. G. Sinnett, D. P. Letley, G. L. Narayanan et al., "*Helicobacter pylori* vacA transcription is genetically-determined and stratifies the level of human gastric inflammation and atrophy," *Journal of Clinical Pathology*, vol. 69, no. 11, pp. 968–973, 2016.

- [153] K. R. Amilon, D. P. Letley, J. A. Winter, K. Robinson, and J. C. Atherton, "Expression of the *Helicobacter pylori* virulence factor vacuolating cytotoxin A (vac A) is influenced by a potential stem-loop structure in the 5' untranslated region of the transcript," *Molecular Microbiology*, vol. 98, no. 5, pp. 831–846, 2015.
- [154] S. Maeda, K. Ogura, H. Yoshida et al., "Major virulence factors, VacA and CagA, are commonly positive in *Helicobacter pylori* isolates in Japan," *Gut*, vol. 42, no. 3, pp. 338–343, 1998.
- [155] G.-C. Wei, J. Chen, A.-Y. Liu et al., "Prevalence of *Helicobacter pylori* vacA, cagA and iceA genotypes and correlation with clinical outcome," *Experimental and Therapeutic Medicine*, vol. 4, no. 6, pp. 1039–1044, 2012.
- [156] L. J. van Doorn, C. Figueiredo, R. Sanna et al., "Clinical relevance of the cagA, vacA, and iceA status of *Helicobacter pylori*," *Gastroenterology*, vol. 115, no. 1, pp. 58–66, 1998.
- [157] J. R. Peek, S. A. Thompson, J. P. Donahue et al., "Adherence to gastric epithelial cells induces expression of a *Helicobacter pylori* gene, iceA, that is associated with clinical outcome," *Proceedings of the Association of American Physicians*, vol. 110, no. 6, pp. 531–544, 1998.
- [158] M. Rizwan, A. Alvi, and N. Ahmed, "Novel protein antigen (JHP940) from the genomic plasticity region of *Helicobacter pylori* induces tumor necrosis factor alpha and interleukin-8 secretion by human macrophages," *Journal of Bacteriology*, vol. 190, no. 3, pp. 1146–1151, 2008.
- [159] A. Axon, "Are all helicobacters equal? Mechanisms of gastroduodenal pathology and their clinical implications," *Gut*, vol. 45, no. supplement 1, pp. i1–i4, 1999.
- [160] A. A. R. Ashour, G. B. Collares, E. Nogueira Mendes et al., "iceA genotypes of *Helicobacter pylori* Strains Isolated from Brazilian children and adults," *Journal of Clinical Microbiology*, vol. 39, no. 5, pp. 1746–1750, 2001.
- [161] K. B. Mansour, C. Fendri, M. Zribi et al., "Prevalence of *Helicobacter pylori* vacA, cagA, iceA and oipA genotypes in Tunisian patients," *Annals of Clinical Microbiology and Antimicrobials*, vol. 9, no. 1, p. 10, 2010.