



GLP-1 Receptor Agonists in Neurodegeneration: Neurovascular Unit in the Spotlight

Giulia Monti ¹, Diana Gomes Moreira ², Mette Richner ¹, Henricus Antonius Maria Mutsaers ³, Nelson Ferreira ¹ and Asad Jan ^{1,*}

- ¹ Department of Biomedicine, Aarhus University, Høegh-Guldbergs Gade 10, DK-8000 Aarhus, Denmark; gm@biomed.au.dk (G.M.); mette.richner@biomed.au.dk (M.R.); ncgferreira@gmail.com (N.F.)
- ² VIB-KU Leuven Center for Brain & Disease Research, Herestraat 49, 3000 Leuven, Belgium; dianaraquel.gomesmoreira@kuleuven.be
- ³ Department of Clinical Medicine, Aarhus University, Palle Juul-Jensens Boulevard 82, DK-8200 Aarhus, Denmark; h.a.m.mutsaers@clin.au.dk
- Correspondence: ajan@aias.au.dk

Abstract: Defects in brain energy metabolism and proteopathic stress are implicated in age-related degenerative neuronopathies, exemplified by Alzheimer's disease (AD) and Parkinson's disease (PD). As the currently available drug regimens largely aim to mitigate cognitive decline and/or motor symptoms, there is a dire need for mechanism-based therapies that can be used to improve neuronal function and potentially slow down the underlying disease processes. In this context, a new class of pharmacological agents that achieve improved glycaemic control via the glucagon-like peptide 1 (GLP-1) receptor has attracted significant attention as putative neuroprotective agents. The experimental evidence supporting their potential therapeutic value, mainly derived from cellular and animal models of AD and PD, has been discussed in several research reports and review opinions recently. In this review article, we discuss the pathological relevance of derangements in the neurovascular unit and the significance of neuron–glia metabolic coupling in AD and PD. With this context, we also discuss some unresolved questions with regard to the potential benefits of GLP-1 agonists on the neurovascular unit (NVU), and provide examples of novel experimental paradigms that could be useful in improving our understanding regarding the neuroprotective mode of action associated with these agents.

Keywords: glucagon-like peptide-1 (GLP-1); Alzheimer's disease; Parkinson's disease; neurodegeneration; neurovascular unit

1. Introduction

The glucagon-like peptide-1 (GLP-1) is a biological product derived from the posttranslational processing of proglucagon. It is secreted by specific cell populations in the gastrointestinal tract (the enteroendocrine L cells), but also by some neuronal populations in the hindbrain (the nucleus of solitary tract-NTS: an important region involved in energy intake) [1–4]. The physiological stimulus for the endogenous GLP-1 secretion is nutrient ingestion and consequent rise in plasma glucose, which leads to a two- to three-fold enhancement in the circulating GLP-1 compared to basal levels [5–7]. This is rapidly followed by an incretin effect (see Glossary) that restores the physiological levels of glucose in circulation [1,3]. The nascent GLP-1 is 37 amino acids in length and has two naturally bioactive forms in circulation, GLP-1 (7–37) and GLP-1 (7–36)-amide [7,8] (Figure 1A). The circulating GLP-1 is rapidly cleared from the circulation due to the proteolytic degradation by the serine protease dipeptidyl peptidase (DPP-4) and neutral endopeptidase 24.11 (NEP 24.11; also known as neprilysin) [1,7]. The direct biological actions of native bioactive forms of GLP-1 are mediated by the activation of the GLP-1 receptor, which belongs to the family B of the G-protein-coupled receptors (GPCRs) [1,4,7,9]. Studies on mRNA



Citation: Monti, G.; Gomes Moreira, D.; Richner, M.; Mutsaers, H.A.M.; Ferreira, N.; Jan, A. GLP-1 Receptor Agonists in Neurodegeneration: Neurovascular Unit in the Spotlight. *Cells* 2022, *11*, 2023. https://doi.org/ 10.3390/cells11132023

Academic Editor: Paola Bagnoli

Received: 5 June 2022 Accepted: 23 June 2022 Published: 25 June 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). and protein expression indicate that GLP-1 receptors are distributed throughout several tissues including the pancreas, kidney, stomach, heart and brain [1,3,10–13]. In the brain, GLP-1 and/or GLP-1 receptor mRNA have been detected in some regions of the thalamus, hypothalamus, hippocampal pyramidal neurons, cortex, cerebellar Purkinje cells and in the brainstem [11,14–17]. The immediate effects of GLP-1 in the cells expressing GLP-1 receptors are attributed to the rise in intracellular cAMP levels and protein kinase A (PKA) activation (Figure 1B), which, in pancreatic β -cells, is directly responsible for calcium influx and insulin secretion [1,2,7,18,19]. Despite the short half-life of the endogenous GLP-1, the interplay of peripheral and central GLP-1 receptor signalling helps achieve glycaemic control and energy intake by slowing gastric emptying and reducing caloric intake (Figure 2) [2,20]. In this context, the gut-derived GLP-1 crosses the blood–brain barrier (BBB) and binds to receptors in the circumventricular organs of the brainstem, transmitting metabolic information to the neurons responsible for feeding behaviour (e.g., in the NTS and hypothalamus) [2,10,20,21]. Furthermore, GLP-1 signalling in the brain is also implicated in cognitive functions, based on the studies showing improved learning and memory performance in rats overexpressing GLP-1 receptors in the hippocampus, or the restoration of learning deficits in GLP-1 receptor-deficient mice following hippocampal *Glp1r* gene transfer [14,22].

The discovery of the insulinotropic glucose-lowering effects of GLP-1 was a major breakthrough that formed the basis for developing the first generation of synthetic GLP-1 analogues and, subsequently, their approval for the treatment of Type 2 diabetes mellitus (T2DM) and obesity in 2005 [2,3]. Since then, a number of DPP-4 degradation-resistant and long-acting synthetic GLP-1 receptor agonists (GLP-1RAs) have been developed that differ in chemical modifications and duration of action (Table 1; see references [2,23] for further details). In brief, the short-acting drugs include lixisenatide and exenatide (Byetta), while the long-acting GLP-1RAs are exemplified by liraglutide, semaglutide, exenatide (Bydureon) and dulaglutide. The main pharmacodynamic difference between the two classes is that the short-acting agonists lower postprandial glucose by delaying gastric emptying, while the long-acting GLP-1RAs have a stronger incretin effect [1,23,24]. Since the short-acting GLP-1RAs are more rapidly cleared from plasma, GLP-1 receptor activation is also short-lived. In contrast, the long-acting GLP-1RAs reach a steady state concentration, with minor fluctuations between the doses, thus causing a continuous stimulation of GLP-1 receptors [3,24]. As a result of this difference, long-acting GLP-1RAs are more effective in lowering fasting plasma glucose and in improving glycaemic status compared to the short-acting compounds, as observed by lower levels of glycated haemoglobin-HbA1c: an index of glycaemic control; see Glossary) [2,25].

The advent of GLP-1RAs led to further discoveries that sustained GLP-1RAs exposure is associated with pro-survival cellular gene expression, purportedly mediated by cAMP responsive element binding (CREB) signalling that involves increased expression and/or activity of receptor tyrosine kinases PI3K/AKT, epidermal growth factor receptor (EGFR) and hypoxia-inducible factor 1-alpha (HIF-1 α) (Figure 1B) [1,4,7,26]. Through these latter mechanisms, GLP-1RAs promote pancreatic β -cell neogenesis, stimulate cell growth and increase insulin synthesis in the β -cells [18,23,27,28]. Furthermore, a growing list of research studies show that GLP-1RAs exert cytoprotective and anti-inflammatory effects in cultured cells and in vivo. For instance, GLP-1RAs modulate autophagy [29–31], reduce endoplasmic reticulum stress (in response to glucotoxicity and lipotoxicity) [32–34], and induce changes in gene expression that lead to a favourable metabolic reprogramming and redox homeostasis [4,26,35–37]. Burgeoning evidence also indicates that GLP-1RAs have neuroprotective properties that can potentially be translated into novel therapies for neurological conditions, in particular neurodegeneration and ischaemic brain injury [14,17,22,38,39]. In a broader context, the long-term enhancement of GLP-1 signalling leads to functional improvements across several tissues including the liver, kidney, heart, skeletal muscle and vascular smooth muscle independent of the incretin effect [3,4,7].





Figure 1. GLP-1 biogenesis and signalling transduction. (A) GLP-1 is liberated from the precursor molecule proglucagon—in the enteroendocrine L cells and in the CNS—by the isoforms of prohormone convertase (PC). GLP-1 (7–36)-amide and GLP-1 (7–37) are the major bioactive forms in circulation and degraded by the serine protease dipeptidyl peptidase (DPP-4) and neutral endopeptidase 24.11 (NEP 24.11). (B) The acute phase of GLP-1 receptor (GLP-1R) stimulation involves the activation of adenylate cyclase (AC) and a rise in the levels of intracellular cyclic (AMP), with subsequent engagement of additional effector mechanisms mediated by protein kinase A (PKA). This culminates in increased calcium influx and the promotion of insulin secretion by the islet β -cells. Long-term (chronic) stimulation of GLP-1R is thought to occur via cAMP-responsive element binding (CREB) signalling that promotes pro-survival gene expression mediated by the receptor tyrosine kinases PI3K/Akt and extracellular signal-regulated kinase (ERK)1/2, among others (see Introduction). In the pancreas, this increases β -cell mass by diminishing apoptosis and increasing β -cell proliferation and differentiation.

Indirect effects of GLP-1R agonists



Figure 2. Indirect effect of GLP-1 receptor agonists (GLP-1RAs). Activation of central GLP-1 receptor signalling in the brain leads to metabolic reprogramming in the peripheral tissues. Shown are the effects on blood glucose homeostasis, lipid metabolism and gastrointestinal motility.

GLP-1RA	Backbone Modification	Frequency of Administration	Half-Life
Exenatide (Byetta/Bydureon)	Exendin-4 (resistant to DPP-4 cleavage, largely due to the substitution of the second amino acid from alanine to glycine)	Twice daily/weekly	3.3–4 h
Lixisenatide (Adlyxin, Lyxumia)	Non-acylated GLP-1 (7–37) analogue based on exendin-4, but is modified by the deletion of one proline residue and with a C-terminal hexa-lysine extension	Daily	2.6 h
Oral Semaglutide (Rybelsus); Semaglutide (Ozempic)	Acylated Human GLP-1 (7–37) analogue	Once daily; weekly	1 week
Liraglutide (Victoza)	Mammalian GLP-1, substitution of lysine for arginine at position 28 with the addition of C-16 fatty acid	Daily	13 h
Dulaglutide (Trulicity)	Mammalian GLP-1; the GLP-1 portion of the molecule is fused to an IgG4 molecule, limiting renal clearance and prolonging activity	Weekly	1 week
Albiglutide (Eperzan and Tanzeum)	Two GLP-1 (7–36) molecules fused in tandem to human serum albumin	Weekly	1 week
Taspoglutide	Modifications designed to delay DPP-4 cleavage and other serine proteases, with greater receptor binding	Weekly	1 week

Table 1. GLP-1RAs: backbone modifications, frequency of administration and half-life.

In this article, we intend to focus on GLP-1RAs in the context of neurodegenerative diseases using examples from Alzheimer's disease (AD) and Parkinson's disease (PD), since there is a growing list of preclinical studies showing GLP-1RAs as potential disease-modifying agents in these disorders. In brief, we discuss the neuroprotective and antiinflammatory effects of GLP-1RAs in the context of neurovascular function and metabolic homeostasis between neurons, astroglia and local elements within the neurovascular unit (NVU). In the sections below, first we provide an overview of the salient findings with regard to the use of GLP1-RAs in experimental models of AD and PD. Then, we introduce the concepts concerning neurovascular coupling and brain energy metabolism, followed by a discussion on relevant aspects in AD and PD. Lastly, we discuss the evidence in support of the hypothesis that the neuroprotective effects of long-term GLP-1RAs may partly be mediated by improved neurovascular coupling, and conclude with outlining some aspects as the outstanding questions.

2. GLP-1RAs in Neurodegenerative Diseases

Late-onset AD (also termed sporadic AD) is the most common neurodegenerative disease and the leading cause of dementia in the elderly population worldwide [40,41], whereas idiopathic PD is the most common neurodegenerative cause of motor disability in the population over 65 years of age [42]. Among the risk factors for sporadic AD and idiopathic PD, advancing age represents the most important risk factor in the vast majority (>85%) of cases, followed by a positive family history and head trauma [40,43]. Clinically, the presenting features of these diseases are remarkably distinct such that AD patients exhibit significant disturbances in recent memory, which eventually progresses to visuospatial disorientation and profound disability in performing routine everyday tasks [41]. In contrast, the presenting features in clinical PD include the slowness of movement initiation and maintenance of posture [44,45]. However, long-standing (>15–20 years) PD is also associated with cognitive decline and dementia [44,46]. The detailed overview of the putative mechanisms underlying neurodegeneration in AD and PD is beyond the scope of this article, and can be found in relevant review opinions elsewhere [42,47,48]. For the present discussion, it suffices to mention that the neuropathology of both AD and PD share certain features, which supports an aetiological role of the proteopathic stress due to protein aggregation and energy dyshomeostasis in the ageing brain. In particular, both diseases are associated with: (i) intracerebral proteinaceous deposits across several cortical and subcortical regions, i.e., amyloid Aß plaques and neurofibrillary tangles (comprised of hyperphosphorylated tau) in AD, and Lewy-related alpha-synuclein pathology in PD; (ii) neuronal loss within specific cell populations and pathological alterations in neuropil that dictate the clinical outcome (i.e., entorhinal cortex and hippocampus in AD and nigrostriatal circuitry in PD); and (iii) pathological alterations within glial cells, including pro-inflammatory phenotypes [40,47,49].

GLP-1RAs have emerged as promising therapeutic candidates for tackling neuronal loss and neuroinflammation since GLP-1 signalling seems to have beneficial effects on several factors implicated in the pathogenesis of neurodegenerative diseases, e.g., ER stress, impaired redox homeostasis, autophagy and chronic inflammation (reviewed in [2,4,17,22,50–52]). In this context, the existing literature (predominantly based on the use of liraglutide or exendin-4) maintains that GLP-1RAs promote neurogenesis, enhance neuronal survival and synaptogenesis, attenuate neuroinflammation and/or reduce the pathological markers of protein aggregation in animal models of neuronal injury [17,53–73]. For instance, in cultures of embryonic cortical neurons, exendin-4 promoted neurite growth and protected against cell death induced by hypoxia and the mitochondrial toxin 6-hydroxydopamine (6-OHDA) [63]. Also in neuronal cultures, liraglutide treatment mitigated the synaptotoxicity of A β oligomers and reduced the tau hyperphosphorylation induced by A β 1-42 [67,74]. Similarly, liraglutide treatment in cultures of neuroblastoma cells exposed to methylglyoxal (a potent glycation stress inducer) enhanced cell viability and reduced cytotoxicity [75]. Besides in vitro evidence, GLP-1RAs have been reported to exert neuroprotective effects in diverse animal models of neuronal damage resulting from acute CNS injury (stroke, traumatic brain injury), sub-acute loss of dopaminergic neurons (chemical parkinsonism; based on injections of mitochondrial toxins 6-OHDA or 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, MPTP) and chronic proteopathic stress (transgenic models of Aβ amyloidosis and tauopathy) [22,51] (Table 2). In transgenic rodent models of AD, liraglutide or exendin-4 treatment has been associated with a reduction in the amyloid plaque burden [62,72,76–78] and tau phosphorylation [56,58,67,72,73,79], which are considered invariable pathogenic factors underlying neurodegeneration in AD. Moreover, liraglutide was shown to prevent the loss

of brain insulin receptors and synapses [74], and reversed cognitive impairments induced by Aβ oligomers in rodents and non-human primates [74,80]. Similarly, in animal models of chemical parkinsonism, liraglutide or exendin-4 treatment have been associated with increased neurogenesis, preservation of dopaminergic neurons and dopamine synthesis in substantia nigra, and the rescue of motor impairments [55,59,60,64]. The knowledge regarding the therapeutic benefits of GLP-1RAs in PD models based on the injections of aggregated alpha-synuclein is currently very limited. One study has reported that a longacting GLP1-RA (NLY01) reduced de novo alpha-synuclein aggregation and dopaminergic neuronal loss in the striatum in a transgenic mouse of synucleinopathy, following the intra-striatal delivery of exogenous aggregated alpha-synuclein [81]. However, another study based on the delivery of aggregated alpha-synuclein in the olfactory bulb showed that a long-acting GLP1 analogue did not significantly reduced alpha-synuclein aggregation in the brain of wild-type mice [82]. In addition, there is a considerable amount of evidence demonstrating that GLP-1RAs reduce brain damage in animal models of ischaemic stroke and traumatic brain injury [38,39,54,61,63], and improve the survival of motor neurons in the mutant superoxide dismutase 1 (SOD1; G93A) model of motor neuron disease [50]. Nevertheless, a single unifying mechanism of action underlying the neuroprotective properties of GLP-1RAs or putative disease modification remains a matter of investigation [4,17,22,38,51,83].

The therapeutic utility of GLP-1RAs on clinical outcomes in neurodegenerative diseases is only recently beginning to emerge (Table 2). In a trial involving patients with impaired glucose tolerance or T2DM, liraglutide administration for 3 weeks improved short-term memory [84]. Although these findings are encouraging, clinical studies in patients with mild cognitive impairment (MCI) or AD have yet to conclude definite beneficial outcomes. For instance, liraglutide administration over 3 months in middle-aged individuals with cognitive complaints and/or a positive family history of AD increased connectivity in the default mode network, as detected by functional magnetic resonance imaging (fMRI) [85]. In a pilot study (NCT01469351, [86]) involving a small cohort of AD patients, 6 months' treatment with liraglutide improved cerebral glucose uptake; however, no significant effects on A β plaque load or cognitive performance were noted [87]. In a phase II study involving patients with mild AD (ELAD, NCT01843075), 12 months' treatment with liraglutide failed to improve cerebral glucose uptake, but there were encouraging indicators with regard to cognitive function and grey matter volume [88]. In addition, in an exploratory study involving AD patients, 18 months' treatment with exenatide produced insignificant outcomes in cognitive measures, cortical thickness and volume, or biomarkers in biological fluids except for a slight reduction in AB42 within neuronal extracellular vesicles (NCT01255163) [89]. In the clinical trials involving PD patients (NCT01971242, NCT01174810), 12 months' treatment with exenatide improved motor and cognitive outcomes [90,91], albeit, these findings have been described as 'low certainty evidence' [92]. In near future, semaglutide is planned to be tested in two large phase III studies involving MCI and AD subjects (EVOKE and EVOKE Plus; NCT04777396 and NCT04777409). Moreover, clinical trials using liraglutide, lixisenatide and semaglutide in PD treatment are also currently on the horizon (NCT02953665, NCT03439943, NCT03659682) [92].

Apart from the common animal models of neurodegeneration specified above, the neuroprotective effects of GLP1-RAs are also reported in some preclinical models of rare neurodegenerative diseases. Huntington's disease is an autosomal dominant neurodegenerative disorder caused by the CAG repeat expansions in the huntingtin (*HTT*) gene, and clinically manifests as hyperkinetic movement disorder and cognitive decline. In a transgenic mouse model overexpressing mutant human huntingtin (N171-82Q), once-daily subcutaneous delivery of exendin-4 reduced huntingtin aggregates in the cortex, along with improvements in motor performance and overall survival [93]. Wolfram syndrome, caused by autosomal recessive inheritance of biallelic variations in the wolframin (*WFS1*) gene, is another rare neurodegenerative disorder that initially presents as diabetes mellitus. In a rat *Wfs1* knockout model, subcutaneous delivery of liraglutide over 6 months attenuated

neuroinflammation and significantly reduced the loss of retinal ganglion cells and optic nerve axons [94]. However, we are not aware of any clinical trials with GLP-1RAs in Huntington's disease or Wolfram syndrome.

Table 2. GLP-1RAs in AD and PD: An overview of the experimental models and clinical trials.

Studies	Experiment	GLP-1RA	Observations	Publications
Preclinical studies	Animal model			
AD features Plaque load	APP/PS1/tau mice 5xFAD mice APP/PS1 mice 3xTg-AD mice	Liraglutide Liraglutide Lixisenatide Exendin-4	Reduction of plaque load Reduction of plaque load Reduction of plaque load Reduction of plaque load	[72,76,77] [78] [76] [62]
Tau phosphorylation	APP/PS1/tau mice hTauP301L mice Aβ injection in mice APP/PS1 x db/db mice Streptozotocin injection in mice	Liraglutide Liraglutide Liraglutide Liraglutide Dulaglutide	Reduction of neurofibrillary tangles Reduced Tau phosphorylation Reduced Tau phosphorylation Reduced Tau phosphorylation Reduced Tau phosphorylation	[56,72] [58] [67] [79] [73]
Cognitive and memory performance	$A\beta$ injection in mice $A\beta$ injection in rats Streptozotocin injection in mice	Liraglutide Lixisenatide Dulaglutide	Împroved cognitive impairment Improved spatial memory Improved memory ability	[74] [80] [73]
Other	Aβ injection in non-human primates	Liraglutide	Reduced synaptic loss	[74]
PD features Dopaminergic neuronal loss	6-OHDA rat model 6-OHDA rat model 6-OHDA rat model	Liraglutide Exendin-4 Exendin-4′	No influence on dopaminergic neuronal loss Neurogenesis Reduced lesions	[59] [55] [60]
Motor performance	MPTP mouse model MPTP mouse model	Liraglutide Lixisenatide	Improved motor control Improved motor control	[64] [64]
α-synuclein aggregation	Preformed fibrils injection in striatum of human A53T α -synuclein mice Preformed fibrils injection in the olfactory bulb of C57BL/6J mice	Exendin-4 (NLY01) Exendin-4	Reduced loss of dopaminergic neurons and improved motor performance No significant reduction of α-synuclein aggregation	[81] [82]
Clinical trials	Trial ID			
AD	NCT02140983	Liraglutide	Increased connectivity in the default mode network	[85]
	NCT01469351 NCT01843075 NCT01255163 NCT04777396 NCT04777409	Liraglutide Liraglutide Exenatide Semaglutide Semaglutide	Improved cerebral glucose uptake Improved cognition No significant changes in cognition Recruiting Recruiting	[86,87] [88] [89]
PD	NCT01971242 NCT01174810 NCT02953665 NCT03439943 NCT04154072 NCT03659682	Exenatide Exenatide Liraglutide Lixisenatide Exenatide Semaglutide	Improved motor and cognitive outcomes Improved motor and cognitive outcomes Active Active Active Not yet recruiting	[90] [91]

3. Neurovascular Unit (NVU) in Ageing and Neurodegeneration

3.1. Neurovascular Coupling and Brain Energy Metabolism

The brain represents 2% of the body mass in adults, yet the energy demands to maintain adequate brain function require 20–25% of total body glucose utilisation and ~20% of the oxygen consumption [95,96]. In the adult brain, glucose is the obligate energy substrate for the production of adenosine triphosphate (ATP), except under particular circumstances where ketone bodies (e.g., during fasting) and lactate (e.g., during intense physical activity) are also utilised [97,98]. Circulating glucose enters the brain parenchyma by facilitated transport via 55-kDa glucose transporter 1 (GLUT1) on the capillary endothelial cells, and is subsequently taken up by neurons (via GLUT3) and glial cells [95,99,100]. The high basal metabolic rate in the brain is thought to be due to the energy demands for maintaining ionic gradients at the plasma membrane of neuronal processes by ATP-consuming pumps (chiefly Na⁺/K⁺-ATPase), as well as for the production and uptake of neurotransmitters [101,102]. Besides being the major substrate for ATP production, glucose is also required for the synthesis of neurotransmitters such as glutamate, γ -aminobutyric acid (GABA), acetylcholine and glycine [101].

The energy supply to the brain is tightly linked with neuronal activity and these phenomena form the basis of neurovascular coupling, and can be directly monitored by acquiring information about the regional brain glucose utilisation through imaging modalities such as fMRI and positron emission tomography (PET) [96,103,104]. Briefly, it is well established that task-dependent neural activity triggers an increase in the local cerebral blood flow (CBF) and glucose uptake in a pattern that is highly restricted to the activated network [105]. This stringent regulation is considered to be due to the lack of locally stored energy reserves that can be mobilised to meet the energy demands (except glycogen in astrocytes, elaborated below), as well as to remove by-products of neural activity (e.g., lactate, CO_2 , A β) and possibly for local temperature regulation [103,106–109]. The physiological basis of functional hyperaemia (see Glossary) is thought to reflect a vasoactive effect on the brain capillaries by the local accumulation of metabolic by-products during synaptic activity (e.g., lactate, CO_2 , H^+) and/or the active generation of vasodilatory mediators (e.g., NO—nitric oxide, adenosine, ATP, K⁺) [106,110,111]. Moreover, glutamate released during synaptic activity raises intracellular calcium in astroglia through metabotropic mGluR5 receptors, with the subsequent stimulation of vasoactive arachidonic acid derivatives generation, which alters the tone of pericytes and vascular smooth muscle cells [99,106,112–114]. Other studies also implicate neuronal ATP release with a subsequent rise in intra-astroglial calcium through the activation of purinergic P2X or P2Y receptors [112,115,116].

The key basis of neurovascular coupling is the anatomical integration between the NVU and the tripartite synapse (i.e., pre- and postsynaptic elements, and astroglial processes) [95,99] (Figure 3A). The intraparenchymal brain capillaries contain a single layer of endothelial cells that are endowed with tight junctions, and are surrounded by a layer of pericytes and astroglial foot processes forming the glia limitans of the BBB. On the synaptic side, astroglial processes ensheath the synapses and are connected to each other via gap junctions. This anatomical arrangement has two implications: (i) the endothelial cells in the intraparenchymal brain capillaries provide a structural barrier for regulating the exchange of substances (e.g., nutrients, peptides, waste products) between blood-to-brain and brain-to-blood via specific efflux and influx transporters [117]; and (ii) the perivascular astrocytes detect the neural activity on the synaptic side (due to the presence of glutamate uptake transporters), while uptaking glucose from capillary blood by facilitated diffusion via 45-kDa GLUT1 transporter in their end feet [100]. Furthermore, the intimate proximity of astroglial end feet to the intraparenchymal brain microvessels is crucial for the adequate expression and proper localisation of transmembrane proteins forming the endothelial tight junctions [118]. With that overview in sight, it is apparent that CBF, glucose transport into brain parenchyma and glucose metabolism are tightly linked via the structural and functional features of the NVU [119]. Accordingly, pathological processes that lead to NVU disruption—either due to structural damage or dysregulation in the levels of local and/or circulating vasoactive factors-can cause brain dysfunction, and in the long-term lead to irreversible brain damage [110].



Figure 3. Overview of the local cerebral blood flow auto-regulation and neuron–glia metabolic coupling. (**A**) Neuronal and astroglia-derived vasoactive mediators act on the intraparenchymal capillaries and arterioles to sustain enhanced delivery of metabolic fuels during functional hyperaemia. Circulating glucose enters into the brain parenchyma by facilitated diffusion via glucose transporters in the capillary endothelial cells (EC), and astroglial foot processes forming the glia limitans of the blood–brain barrier. (**B**) Glutamate uptake at the tripartite synapse by astrocytes is accompanied by sodium (Na⁺) entry, which triggers anaerobic glycolysis and lactate release. The lactate is transferred to the neurons (via a process termed the Astrocyte Neuron Lactate Shuttle) and converted to pyruvate by neuronal lactate dehydrogenase (not shown). The pyruvate enters the citric acid cycle in mitochondria for ATP generation. A significant amount of the ATP generated is used to maintain the function of sodium–potassium pumps (Na⁺/K⁺-ATPase).

3.2. Structural Derangements of NVU in Ageing and Neurodegeneration

Ageing is a significant risk factor for several metabolic, cardiovascular and neurological conditions, including neurodegenerative diseases. Ageing affects cells and tissue both at the molecular and cytoarchitectural levels, such that there is a declining capacity to maintain healthy wear-and-tear, and the capacity to mount an appropriate cellular homeostatic response is also reduced [117,120,121]. The brain is particularly sensitive to the local and systemic effects of the age-related decline in cellular functions that may predispose to cognitive decline [121,122]. In this regard, the concept of 'brain ageing' is quite relevant, since it attempts to correlate the structural and functional alterations in the brain to the cognitive status of an individual [117,123]. In particular, the lack of cognitive and neurological symptoms in the elderly may not necessarily reflect the lack of pathological processes in the brain [122,124]. For instance, a comprehensive post-mortem assessment of 330 brains from cognitively normal elderly donors revealed that more than 50% of the brains exhibited markers of AD pathology, while as many as 40% showed evidence of mixed pathology (AD pathology with microvascular brain damage) or Lewy body pathology. Only about 4% of brains in this study showed lack of pathological markers associated with AD, PD or microvascular damage [121].

Several macroscopic, microscopic and molecular alterations in the brain parenchyma and supporting tissue (e.g., vasculature) hint towards defective energy extraction and/or utilisation during ageing that may adversely affect brain functions [117,125,126]. Neuroimaging studies in the elderly show that there are detectable changes in the volume of both grey and white matter, a reduction in the cortical thickness and/or abnormal functional connectivity in the absence of any severe neurological and/or cognitive disability [127–131]. However, rapid brain ageing in distinct regions (e.g., isocortex and/or hippocampus) is a prognostic indicator of developing cognitive impairment that can progress to overt dementia [132]. Similar observations have been reported with regard to the vulnerability of neuronal populations in substantia nigra to ageing-associated degeneration that may precede clinical PD [133]. In this regard, histological studies reveal that there is a mild to moderate decline in the number of pigmented and tyrosine hydroxylase-positive neurons (10–30% loss; 4–10% per decade) in as many as 1/3 of elderly subjects without clinically defined PD [134,135]. Moreover, it is also suggested that by the time of clinical presentation of PD, the loss of dopaminergic neurons in substantia nigra may exceed 50–70% [133,136]. Conventionally, the selective vulnerability of neuronal populations to disease processes is usually attributed to their inherent properties (i.e., high metabolic rate, ROS generation, and extensive arborisation with increased energy demand) [133,136,137]. However, it can also be argued that in susceptible individuals, distinct neuronal populations experience brain ageing at an accelerated rate, which leads to clinical presentation of a neurodegenerative disorder such as AD or PD.

Thus, during healthy brain ageing, individuals maintain a functional neuronal reserve for a considerably long time before the diagnosis of a neurological and/or cognitive disability is established [136,138]. Conversely, pathological factors such as protein aggregation and proteopathic stress, and possibly the influence of systemic comorbidities, accelerate brain ageing, leading to a decline in the neuronal reserve over time [132,139]. This is supported by epidemiological data suggesting that the dementia risk is greatly aggravated by several factors otherwise commonly associated with cerebrovascular disease, including hypertension, diabetes and obesity [140–142]. Accordingly, a number of MRI-based studies show that intraparenchymal microbleeds (see Glossary) are a feature observed during normal ageing. However, they are more frequent in individuals with MCI, early AD and in as many as 50% of dementia cases [143–148]. Furthermore, MCI and AD are associated with microbleeds in the infratentorial regions, a pattern of vascular damage also seen in individuals suffering from long-standing hypertension [106,143,149]. In addition, several microvascular alterations have long been associated with AD neuropathology, including increased atherosclerotic lesions, capillary tortuosity and rarefaction, thickening of the basement membrane and, in some cases, 'string vessels', which lack endothelial cells [103,150–152]. Similarly, post-mortem studies in long-standing PD cases reveal capillary damage and fragmentation in cortical areas and in brainstem regions, as well as increased endothelial cell nuclei in substantia nigra [153,154]. Moreover, a reduction in

the neocortical CBF has been reported early in the course of PD, which correlates with cognitive impairment in some cohorts of PD patients [155–157].

3.3. Functional Derangements of NVU in Ageing and Neurodegeneration

It is plausible to postulate that accelerated brain ageing could be triggered by aberrations in the neurovascular coupling, thus exacerbating neurometabolic dysfunction in age-related neurodegenerative diseases [106,150]. First, with advancing age, CBF becomes uncoupled from glucose transport and utilisation both spatially and temporally [158–162]. Second, brain imaging studies that measure the metabolic rate of glucose utilisation using fluorodeoxyglucose (FDG-PET) indicate glucose hypometabolism in AD, PD and among the elderly at risk of AD [163–166]. For instance, decreased FDG uptake in the parietotemporal and cingulate cortex has been suggested to be quite specific for AD [167–169] or an indicator of enhanced risk for progression to dementia in individuals with MCI [170,171]. These features are reminiscent of structural and functional changes in the brains of middleaged and elderly individuals with long-standing T2DM, with otherwise normal cognition, who also exhibit variable cortical atrophy and regional decline in the glucose metabolism.

Moreover, a growing body of evidence implicates brain insulin deficiency and/or insulin resistance as a key pathognomic phenomenon in both AD and PD, as indicated by the reduced expression of insulin (mRNA and protein), insulin receptors, decreased receptor affinity and reduction in the downstream signalling pathways (i.e., insulin receptor substrate 1, IRS1; phospho-AKT and phospho-glycogen synthase kinase 3β [172–179]. Insulin is an important growth factor and is involved in dendritic sprouting, neuronal stem cell activation, cell repair and neuroprotection against A β [180,181]. In addition to the defects in central insulin signalling, it has also been suggested that systemic insulin resistance and hyperinsulinemia also impacts brain energy metabolism and memory [178,182]. In particular, T2DM has been described as a risk factor for AD and other neurodegenerative disorders [174,183,184]. For instance, studies show that systemic insulin resistance is associated with lower glucose metabolism in frontal, parietotemporal, and cingulate cortices [185,186]. In AD, decreased brain glucose uptake correlates with lower expression levels of the glucose transporters GLUT1 and neuronal GLUT3 [187–189], as well as with diminished activities of different metabolic enzymes such as glucose-6-phosphate isomerase, lactate dehydrogenase, phosphofruktinase, cytochrome oxidase, and α -ketoglutarate compared to the controls [173,178,190,191].

While the estimates on the prevalence of T2DM in AD are not unequivocally agreed upon, the prevalence of clinically diagnosed T2DM in PD has been reported to be as high as 8–30% and systemic insulin resistance in 50–80% subjects within different cohorts of PD patients [192,193]. Furthermore, T2DM in PD patients is markedly associated with the severity and progression of motor disability, cognitive decline and lower dopamine transporter binding in the striatum [194–196]. Also in PD, the dysregulation of different metabolic factors involved in brain glucose metabolism (e.g., glucose-6-phosphate dehydrogenase and 6-phosphogluconate dehydrogenase) has been reported to occur at early stages in the pathology [197]. Related to this, several loss-of-function mutations in genes known to be associated with early onset PD (e.g., PARK2) are directly linked to proteopathic stress and compromised mitochondrial functions [44]. Hence, it is plausible that systemic insulin resistance contributes to accelerated brain ageing and plays a role in the central energy crisis in the brain, which may be a harbinger of neurodegeneration in individuals at risk [182]. Further evidence of defective energy metabolism in AD and PD is also adduced by studies showing perturbations in the expression and/or activity of mRNA translation machinery required for adequate protein synthesis [198]. Since cellular protein synthesis is a highly energy consuming process, several of the factors involved in translation initiation and elongation phases are sensitive to declining ATP levels. Accordingly, post-mortem studies in AD and PD reveal considerable aberrations within the mammalian target of rapamycin (mTOR) pathway [199], AMP-activated protein kinase (AMPK) [200], elongation

factor-2 kinase (eEF2K) [201,202], and stress-adaptive phosphorylation of the initiation and elongation factors [198,201–203].

4. GLP-1RAs in Neurodegeneration: The Neurovascular Connection

Given the pleiotropic indirect effects of GLP-1 signalling on cardiovascular function, blood glucose homeostasis, fatty acid metabolism and cell growth, the mechanism(s) of GLP-1RAs mediated neuroprotection warrant a careful assessment. As alluded to earlier, the effects of GLP-1RAs have been studied in animal studies of neuronal damage involving acute CNS injury (stroke, traumatic brain injury), sub-acute toxin induced dopaminergic loss and prolonged effects of proteopathic stress (e.g., transgenic models of A β amyloidosis/tau aggregation) [22,38]. A corollary to the neurocentric mode of action (i.e., direct neuroprotection mediated by GLP-1 receptor signalling in neurons or modulation of A β /tau aggregation) would be the ancillary mechanism of improved neurovascular coupling. Accordingly, GLP-1 receptor signalling and/or GLP-1RAs could promote a favourable intercellular milieu at the level of NVU, which in turn is associated with a broad range of beneficial effects in the CNS. For instance, this is supported by observations that exendin-4 treatment in a mouse of atherosclerosis (western-type diet) with human-like lipoprotein metabolism was associated with reduced inflammation, and macrophage influx in the vessel wall and in the liver [204]. Similarly, intraperitoneal delivery of liraglutide in a mouse model of polymicrobial induced sepsis attenuated endothelial dysfunction, vascular inflammation and oxidative stress in the aortic wall [205].

To illustrate this point further, we provide here some examples with the caveat that our understanding of specific cellular effects of long-term GLP-1RAs on the components of NVU is still incomplete. The existing literature supports the view that the predominant direct effect of endogenous GLP-1 receptor signalling and pharmacological GLP-1RAs is to promote improved glycaemic control, reduce systemic insulin resistance and decrease free fatty acids in circulation [1,3,4]. Systemic insulin resistance and consequent dysregulation in homeostatic glycaemic status are the key mediators of the chronic complications associated with uncontrolled T2DM and metabolic syndrome, both in the form of microvascular disease (retinopathy, neuropathy, nephropathy) and macrovascular complications (stroke and coronary artery disease) [206,207]. The underlying metabolic driver of the microvascular complications in prediabetes and T2DM is uncontrolled glucose entry into the cells, which leads to overproduction of mitochondria-derived reactive oxygen species (ROS) due to an increased flux of glucose through the tricarboxylic acid (TCA) cycle. This has directly been implicated in endothelial cell dysfunction and creating a pro-inflammatory milieu, with damage to the vessel wall and normal haemodynamic response [206,207]. For instance, a pathologic factor associated with vascular dysfunction and inflammation is the accumulation of irreversibly glycated proteins (i.e., AGEs-advanced glycosylation end-products). AGEs are known to directly alter endothelial gene expression through transcriptional repression, as well as to activate macrophages by binding to the AGE receptors (RAGEs) and drive downstream inflammatory gene expression involving nuclear factor kappa B (NF- κ B) [208]. In the context of chronic disturbances of glycaemic indices, some studies show that the AGEs accumulation is more frequently encountered in AD brain than in age-matched controls [209,210]. AGEs have been detected in association with Aβ plaques, as well as with neurofibrillary tangles, both in neurons and glial cells [211]. These reports also suggest that AGEs stimulate tau hyperphosphorylation in vivo, and the glycation of A β influences its aggregation [209,211,212]. In addition to the formation of AGEs, intracellular hyperglycaemia leads to the increased formation of diacylglycerol (DAG) from the glycolytic intermediate glyceraldehydes-3-phosphate, which in turn is a potent stimulus for protein kinase C (PKC). The inappropriate activation of PKC affects blood flow and vascular permeability in part via the local synthesis of NO [206,207].

As mentioned above, deciphering the significance of systemic insulin resistance and adequate glycaemic control in neurodegenerative diseases remains an evolving field. For instance, a recent study showed that AD patients exhibit a reduced peripheral insulin sensitivity and hyperinsulinemia, both under fasting state as well as in response to an oral glucose tolerance test [213]. Chronic peripheral hyperinsulinemia is expected to cause the downregulation of insulin receptors at the NVU and reduction in the insulin entry into brain, as reflected in lower brain insulin concentration in AD patients [214,215]. As a consequence, this would lead to aberrations in the glucose transport at the NVU, as reflected by the FDG-PET studies, as well as the reduced expression of GLUT1 in the brain capillaries of AD patients [216]. The significance of glucose transport at the NVU in relation to protein aggregation in the brain has not been extensively studied to date. Nevertheless, studies show that haploinsufficiency of GLUT1 in a transgenic mouse model of A β amyloidosis leads to microvascular degeneration and CBF reduction that precedes progressive neuronal loss and intracerebral A β deposition [217].

5. Unresolved Questions and the Need for Novel Experimental Paradigms

Translating the pleiotropic effects of GLP-1 receptor activation into the neuroprotective properties ascribed to GLP-RAs represents an interesting scientific quest. In addition to the neurocentric context highlighted earlier, it is apparent that novel experimental paradigms are needed to fully establish the therapeutic relevance of GLP-1RAs in neurodegenerative diseases. Taking into consideration the structural and functional perturbations of the NVU in neurodegenerative disorders, some of the approaches could address the direct and indirect effects of GLP-RAs on the physiology of NVU. For instance: (i) probing the effects of GLP-1RAs on the cellular components of the NVU, in particular the metabolic and trophic support activity of astroglia in the context of neurodegenerative proteinopathies and (ii) investigating the relevance of GLP-1RAs activity in the periphery to neuroprotective outcomes in brain pathologies.

5.1. Effects of GLP1-RAs at the NVU

With regard to the first aspect, there is burgeoning evidence that GLP-1 signalling is involved in the modulation of astroglial metabolism and their response to CNS injury. For example, transient cerebral ischaemia in gerbils significantly increased the immunoreactivity of GLP-1 receptors in the pyramidal neurons of the hippocampal CA1 region within the first 2 days after the injury [218]. Intriguingly, this acute phase was followed by a rapid decline in the neuronal GLP-1 receptor expression, and shifted to increasing GLP-1 receptor expression by GFAP-positive astrocytes and some GABAergic interneurons in the CA1 over 4–10 days post-injury [218]. In the same report, the authors showed that exendin-4 administration (2 h prior to surgery and 1 h after reperfusion) prevented neuronal cell loss and infiltration by activated microglia. While the significance of delayed upregulation in astroglial GLP-1R expression remains unsettled, it possibly represents an injury-specific response that may affect the neuron–glia interaction after ischaemia and reperfusion [218]. Furthermore, a long-acting brain penetrant, GLP-1RA (NLY01), prevents the conversion of astrocytes into neurotoxic A1 phenotype in a mouse model of synucleinopathy [81]. Recent evidence also suggests that GLP-1 receptor signalling in astrocytes may also be involved in the neuron-astroglia metabolic coupling, both under physiological and pathological states. Astroglia are integral players in the regulation of brain energy metabolism, and crucial players in the cellular phase of neurodegeneration in AD [48,113]. With their end feet in contact with the capillary endothelial cells, astrocytes are the first cellular barrier encountered by the circulating glucose subsequent to its entry into brain parenchyma [95]. Immunohistochemical analyses show that astroglial processes form closely apposed contacts with GLP-1 receptor-positive axonal processes in the NTS [219]. The same study also demonstrated that exendin-4 increased calcium signalling in astrocytes ex vivo, and localised to NTS astrocytes following in vivo administration [219]. Intriguingly, mice lacking GLP-1 receptors in the glial fibrillary acidic protein (GFAP)-positive astrocytes exhibited increased cerebral glucose uptake, improved memory formation and spontaneous activity in the midbrain dopaminergic neurons [220]. The same report also showed that the genetic deletion of the GLP-1 receptor in astrocytes disrupted mitochondrial integrity, which was

associated with the activation of an integrated stress response involving fibroblast growth factor-21 [220].

It is also noteworthy that neurons and astrocytes differ in the mode of glucose utilisation for ATP generation. In neurons, glucose is processed through ROS-generating oxidative phosphorylation in mitochondria, while astrocytes preferentially utilise glucose via aerobic glycolysis-generating pyruvate and lactate [221,222]. At the glutamatergic synapses (the most common excitatory synapses in the CNS), the neuron-astroglia metabolic coupling during synaptic activation involves neuronal glutamate release and rapid uptake by surrounding astroglia. This triggers aerobic glycolysis and lactate release by the astrocytes, and the released lactate is considered a fuel for ATP generation within neurons—a process commonly referred to as the Astrocyte Neuron Lactate Shuttle (ANLS) [95,223,224] (Figure 3B). In the neurophysiological context, lactate has been implicated in regulating neuronal gene expression for survival, synaptic plasticity and in memory formation [98,109,225,226]. In a recent study, it was shown that liraglutide increased aerobic glycolysis and lactate release, with a concomitant reduction in oxidative phosphorylation by cultured astrocytes. In neuron–astroglia co-cultures, liraglutide also reduced A β neurotoxicity [69]. However, another study showed that liraglutide inhibited glucose uptake in cultured astrocytes, while promoting β -oxidation of fatty acids for ATP generation [220]. Further related to the glucose metabolism, another feature that is peculiar to astroglial cells (although not exclusive) is the glycogen storage, which can be mobilised in the events of increased energy demand and during brain repair [97,221]. The evidence for this is based on the observations that glycogen levels in the brain are increased when neuronal activity is inhibited (e.g., under general anaesthesia) [227]. Moreover, reactive astrocytes in the vicinity of damaged neuropil exhibit the increased accumulation of glycogen [228], which is interesting in view of the upregulation in astroglial GLP-1 receptor expression following ischaemic injury, as mentioned above [218]. While we are not aware of any study to date that has directly investigated the effects of GLP-1RAs on astroglial glycogen storage and/or turnover, studies in human and rodent skeletal muscle and hepatocytes indicate that GLP-1 receptor signalling enhances glycogen synthesis [229,230].

Apart from the metabolic aspects, the glutamate-scavenging function of astroglia is vital to maintain neuronal excitability and to prevent glutamate-induced excitotoxic cell death. Glutamate excitotoxicity is a crucial mediator of neuronal death in ischaemic stroke, and is also widely implicated in some neurodegenerative disorders [48,231]. In this context, the administration of GLP-1RAs (e.g., exendin-4 and liraglutide) prior to or following cerebral ischaemia in animal models of stroke have consistently been reported to reduce infarct size, mitigate oxidative stress and improve endothelial function [38,39,54,61,63]. The neuroinflammatory cascade following the excitotoxic insults within the CNS also affect GABA receptor-mediated inhibitory signalling by eliminating GABAergic synapses, thus exacerbating the imbalance in the neuronal excitation/inhibition and possibly contributing to the NVU dysfunction (i.e., BBB disruption) [232]. Intriguingly, pancreatic β -cells are among the extra-neural sites expressing high levels of glutamate decarboxylase, an enzyme that catalyses the formation of GABA from glutamate [233,234]. Under hyperglycaemic conditions, GLP-1 inhibits glucagon secretion by α -cells through paracrine mechanisms that involve β -cell-derived GABA and insulin [235–237]. While the details of these mechanisms are not fully understood, it is considered that insulin increases the membrane translocation of GABA-A receptors in α -cells, which are activated by the GABA derived from the neighbouring β -cells. This leads to membrane hyperpolarisation and a consequent reduction in the glucagon secretion by the α -cells. Moreover, in cultures of rat hippocampal neurons, insulin stimulation increases the density of postsynaptic GABA-A receptors, presumably mediated by phosphorylated Akt (Thr308) [238]. The interplay of GLP-1 receptor activation and GABA-mediated synaptic effects has largely been studied in the brain centres regulating food intake, energy consumption and glucose disposal [239,240]. Hence, a detailed characterisation of the neuroprotective actions attributed to GLP-1RA in models of

excitotoxic neuronal loss should be carefully evaluated to incorporate excitation/inhibition balance, and possibly in a region-specific manner.

5.2. Effects of GLP1-RAs in Periphery and Inter-Organ Communication

In addition to the direct effects on the cellular elements of the NVU, ancillary mechanisms of neuroprotection by GLP-1RAs may indirectly emanate from the attenuation of pathological processes in the periphery, in particular, favourable outcomes associated with GLP-1RAs on glucose and lipid metabolism, cardiovascular function and systemic inflammation [3,4,241]. As discussed above, the existing literature in animal models of $A\beta$ amyloidosis and tauopathies indicates that the long-term activation of GLP-1 signalling attenuates protein aggregation in the brain (Table 2). Nevertheless, these effects are considered to be the direct result of GLP-1RAs in the CNS and the systemic factors are not usually taken into consideration. Although peripherally administered GLP-1RAs are widely considered to be BBB permeable [3,242], some studies also hint that GLP-1 receptor signalling in the periphery influences central GLP-1 receptors and vice versa; however, the existing literature on the brain-periphery GLP-1 axis is largely limited to the control of food intake and blood glucose disposal [20,37]. It also implies that peripheral actions of GLP-1RAs, in particular reduced systemic inflammation and oxidative stress, may indirectly mitigate pathogenic processes involved in neurodegeneration. For instance, AD is associated with elevated levels of several pro-inflammatory cytokines in circulation (e.g., interleukin 6, IL-6; IL-12, tumour necrosis factor-alpha, TNF- α), that may impact the NVU function [243–245]. This is also supported by epidemiological and neuropathological observations that indicate that there is a considerable overlap between cerebrovascular pathology and the risk of developing dementia [246].

In a broader context with regard to the peripheral effects of GLP1-RAs, aberrations in the physiological inter-organ communication represents an unexplored area in neurological diseases, especially AD and PD. In extreme scenarios, brain function is markedly affected by the health of other organs, as illustrated by the presence of hepatic encephalopathy in patients with cirrhosis [247], and uremic encephalopathy in patients with declining kidney function [248]. In this context, liver or kidney dysfunction has been suggested to increase the risk for AD and PD [249–253]. While the global burden of age-related neurodegenerative disorders is on the rise, this association is especially worrying due to the increasing global prevalence of chronic liver diseases, mainly non-alcoholic fatty liver disease (NAFLD) as well as chronic kidney disease [254,255]. The pathophysiological mechanisms linking these disorders are not well understood; however, it is likely that shared mechanisms such as insulin resistance, inflammation and endothelial dysfunction may play a role. In addition, the pathogenic significance of a reduction in the hepatic/renal clearance and subsequent chronic accumulation of neurotoxic metabolites, either as a result of ageing and/or comorbidity in AD and PD, is largely unknown. For example, polymorphisms in the genes encoding CYP1A1 and CYP2D6, members of the cytochrome P450 family and involved in detoxification reactions, increase the risk of PD [256,257]. Conversely, neuroinflammatory changes in the CNS can also affect peripheral organ function; for instance, damage to the dopaminergic neurons in substantia nigra alters the expression of hepatic P450 enzymes [258]. Thus, it is worthwhile examining whether the putative neuroprotective effects of GLP1-RAs may partly emanate from improving and/or stabilising the functionality of the liver and/or kidney, i.e., excretory and detoxifying functions of the body. Supporting this, Mantovani and colleagues performed a meta-analysis of 11 randomised controlled trials (involving a total of 936 middle-aged individuals) using GLP-1RAs for the treatment of NAFLD or non-alcoholic steatohepatitis (NASH). Their analysis revealed that treatment with GLP-1RAs for a median of 26 weeks was associated with significant reductions in steatosis and serum liver enzyme levels, as well as with greater histological resolution of NASH [259]. However, it remains unclear whether these beneficial effects are due to a direct effect on the liver, since there is still a debate on whether the GLP-1 receptors

are expressed in hepatocytes [260–262]. Nevertheless, there are indications that GLP-1RAs may have therapeutic benefit in the management of patients with NAFLD [263].

Regarding GLP-1RAs and diabetic kidney disease, beneficial effects on composite renal outcomes have been reported, mainly the prevention of new-onset macro-albuminuria, albeit more evidence is needed to establish therapeutic benefits in clinical settings [2,264]. Hence, the ongoing FLOW trial (NCT03819153) is assessing the effects of semaglutide on complex renal outcomes in patients with diabetic kidney disease [265]. Further evidence regarding the therapeutic utility of GLP-1RAs in improving inter-organ communication is hinted by the modulatory effects of GLP-1 receptor activation on the renin-angiotensinaldosterone system (RAAS) [266]. The RAAS is one of the key mechanisms in the longterm regulation of arterial blood pressure through its actions on the vascular smooth muscle tone and renal reabsorption of sodium and water. Perturbations in the RAAS are implicated in the progression of cardiovascular dysfunction and nephropathy associated with poorly controlled T2DM [266]. Biochemical studies also suggest that in addition to the peripheral RAAS axis, the brain harbours a local RAAS system with the expression of several components in neurons and glia [267]. While the detailed discussion of RAAS regulation in health and disease is beyond the scope of this article, existing reports implicate RAAS activity in memory and cognition, as well as in the pathophysiology of AD and PD [267,268]. A direct link between GLP-1RAs and RAAS is adduced by observations that liraglutide increased the expression of angiotensin-converting enzyme 2 (ACE2) and promoted vasodilation in vivo, presumably via increased activation of the Mas receptors and NO synthesis [269,270]. In this regard, it is worth mentioning that a limited number of studies have shown that the activation of Mas receptors with angiotensin (1–7) ameliorates phenotypic defects in the experimental models of ischaemic stroke, 6-OHDA-induced parkinsonism and A β amyloidosis [271–273]. Based on these reports, it remains to be investigated whether GLP-1RAs mitigate cognitive impairment and brain ageing, in part, by maintaining/restoring healthy inter-organ communication.

Another pertinent area for investigation would be the effects of GLP1-RAs on the gut microbiota, since alterations in the composition of gut microbiota or their metabolic products are reported in AD and PD cohorts [274,275]. This is relevant, since systemic metabolic disorders including T2DM affect the composition of gut microbiota [276,277], potentially leading to a milieu that is conducive to pathological processes relevant to neurodegeneration [274,275]. This aspect is also relevant, since there is an emerging notion that some CNS proteinopathies might originate in the periphery. For instance, a number of post-mortem neuropathological and experimental observations suggest that alpha-synuclein aggregation may originate in the periphery (e.g., gut, peripheral nerve afferents) and invade the nervous system in a manner reminiscent of prions [278–280]. However, refined experimental models are needed to fully understand the interaction between peripheral GLP-1 signalling and protein aggregation in the CNS. One potential approach to investigate the peripheral effects of GLP-1RAs could be to employ experimental models of CNS proteinopathies in which protein aggregation is induced in the periphery, for instance by injecting aggregated alpha-synuclein into the gut or propagation involving neuromuscular junction [280–283]. In these models of synucleinopathy, the peripheral induction of alpha-synuclein aggregation is associated with variable degrees of neuronal loss and neuroinflammation in the CNS. Furthermore, the peripheral-to-central propagation of aggregated alpha-synuclein in a transgenic mouse model leads to mechanical allodynia and impaired nociception [280]. Interestingly, enhancing GLP-1 by teneligliptin (DPP-4 inhibitor) in a rat model of partial sciatic nerve transaction ameliorated neuropathic pain and reduced inflammation in the spinal cord [284]. Similar experimental paradigms would be valuable to further elucidate the mechanism(s) that may underlie neuroprotection associated with GLP-1RAs, i.e., investigating whether GLP-1RAs directly affect protein aggregation via peripheral anti-inflammatory effects and/or prevent pathological processes in the CNS predominantly through the central effects.

6. Conclusions

Since the publication of reports showing the involvement of GLP-1 receptor signalling in cognitive function, and encouraging outcomes with GLP-1RAs in experimental models of acute or chronic neuronal toxicity, there is an engaging discussion regarding the mechanism(s) that underlie their neuroprotective mode of action. In addition to their direct pharmacological actions involving GLP-1 signalling and consequent promotion of glycaemic control, a range of indirect effects associated with the long-term use of GLP-1RAs signify tissue-specific mechanisms with translational potential for GLP-1-based therapies (Table 1). We expect that understanding the pharmacological actions of long-term GLP-1RAs use on the structural and functional aspects of the NVU, in particular neuron–astroglia metabolic coupling, will aid in filling critical gaps in the knowledge regarding the therapeutic utility of these agents in neurological conditions. With relevant examples, we have also outlined some experimental paradigms that could be further explored for studying the direct and indirect effects of GLP-1RAs in neurodegenerative diseases that could help elucidate putative disease-modifying effects.

While the outcomes from the clinical trials regarding the therapeutic potential of GLP-1RAs (and DPP-4 inhibitors) in AD and PD are eagerly anticipated in the near future (Table 2), some additional considerations are worth pointing out for the transition into the clinical setting. First, guidelines for patient stratification may be required to identify potential responders and non/weak responders, for example due to inter-individual differences in GLP-1 receptor expression or polymorphisms that may affect response to GLP-1RAs, or clinical evidence indicating advanced stages of neurological dysfunction [2]. Second, it needs to be established whether enhancing GLP-1 receptor signalling would result in clinical improvements via direct central effects, and/or indirectly via promoting healthy brain ageing through a concerted interplay of brain and peripheral organ systems. Hence, a comprehensive framework may be required that incorporates standardised measures of neurological/cognitive functions, as well as markers of metabolic status (e.g., glycaemic control) and co-morbidities (e.g., hypertension, obesity, liver or kidney disease). Third, meticulous post-mortem neuropathological assessments of volunteer donors from these trials would be needed to establish the evidence for disease modification (i.e., reduction in protein aggregation and neuronal loss), as suggested by the animal studies (Table 2). Lastly, refinements in brain imaging modalities may provide additional clues to pinpoint the actions of GLP-1 receptor activation on the NVU. Although this sounds technically challenging, there are examples for assessing NVU function and integrity via fMRI using the blood oxygen level-dependent signal [285], representing potential areas to guide such refinements.

Author Contributions: A.J., N.F. and H.A.M.M. conceptualised the manuscript; G.M., D.G.M., H.A.M.M. and A.J. wrote the manuscript; A.J. and N.F. edited the manuscript; M.R. contributed with figure illustrations in Figures 1–3. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by funding to A.J. in the form of a Marie Skłodowska Curie Fellowship from European Union's Horizon 2020 Research and Innovation Programme (MSCA-IF-2017, grant #786433) and the Lundbeckfonden, Denmark (grant #R250-2017-1131), Lundbeckfonden, Denmark (grant #R368-2021-726) to H.A.M.M. and Lundbeckfonden (grant #R171-2014-591) to N.F.

Institutional Review Board Statement: Not applicable (the article does not contain any experimental data involving human or animal subjects).

Informed Consent Statement: Not applicable (the article does not contain any experimental data involving human subjects).

Data Availability Statement: All of the data cited in the article can be accessed in the original research studies provided in the reference list.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

6-OHDA: 6-hydroxydopamine; AD: Alzheimer's disease; AGEs: advanced glycosylation end-products; AMPK: AMP-activated protein kinase; ATP: adenosine triphosphate; BBB: blood-brain barrier; CBF: cerebral blood flow; CNS: central nervous system; CREB: cAMP responsive element binding; DAG: diacylglycerol; DPP-4: dipeptidyl peptidase; eEF2K: elongation factor-2 kinase; FDG-PET: fluorodeoxyglucose –PET; fMRI: functional magnetic resonance imaging; γ -Aminobutyric acid (GABA); GFAP: glial fibrillary acidic protein; GLP-1: glucagon-like peptide-1; GLP-1Ras: GLP-1 receptor agonists; GLUT1: glucose transporter 1; GPCRs: G-protein-coupled receptors; MCI: mild cognitive impairment; MPTP: 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine; mTOR: mammalian target of rapamycin; NAFLD: non-alcoholic fatty liver disease; NASH: non-alcoholic steatohepatitis; NEP 24.11: neutral endopeptidase 24.11 (Neprilysin); NF-kB: nuclear factor kappa B; NO: nitric oxide; NTS: nucleus of the solitary tract; NVU: neurovascular unit; PD: Parkinson's disease; PET: positron emission tomography; PKA: protein kinase A; PKC: protein kinase C; RAGEs: AGE receptors; ROS: reactive oxygen species; SOD1: superoxide dismutase 1; T2DM: type 2 diabetes mellitus; TCA: tricarboxylic acid; TNF- α : tumour necrosis factor-alpha. Functional hyperaemia: Task-dependent increase in the cerebral blood flow reflecting local neuronal activity in MRI-based brain imaging. Glycated haemoglobin **HbA1c**: Irreversibly glycated haemoglobin resulting from non-enzymatic reactions of sugar molecules with haemoglobin in red blood cells. Used as an index of glycaemic control in diabetic patients. Incretin effect: Potentiation of pancreatic insulin secretion after oral glucose intake (to a much higher degree than parenteral isoglycaemic glucose infusion) mediated by the incretin hormones (GLP-1 and gastric inhibitory polypeptide, GIP). Intraparenchymal microbleeds: Small-vessel disease in the brain causing unregulated extravasation of capillary blood, which can be detected by hemosiderin-laden deposits on T2-weighted MRI. Nucleus of the solitary tract (NTS): Collections of neuronal nuclei situated in the hindbrain region involved in energy balance and cardiovascular functions.

References

- Campbell, J.E.; Drucker, D.J. Pharmacology, physiology, and mechanisms of incretin hormone action. *Cell Metab.* 2013, 17, 819–837. [CrossRef]
- Nauck, M.A.; Quast, D.R.; Wefers, J.; Meier, J.J. GLP-1 receptor agonists in the treatment of type 2 diabetes—State-of-the-art. *Mol. Metab.* 2021, 46, 101102. [CrossRef] [PubMed]
- Nauck, M.A.; Quast, D.R.; Wefers, J.; Pfeiffer, A.F.H. The evolving story of incretins (GIP and GLP-1) in metabolic and cardiovascular disease: A pathophysiological update. *Diabetes Obes. Metab.* 2021, 23 (Suppl. S3), 5–29. [CrossRef] [PubMed]
- 4. Rowlands, J.; Heng, J.; Newsholme, P.; Carlessi, R. Pleiotropic Effects of GLP-1 and Analogs on Cell Signaling, Metabolism, and Function. *Front. Endocrinol.* **2018**, *9*, 672. [CrossRef] [PubMed]
- 5. Kieffer, T.J.; Francis Habener, J. The Glucagon-like Peptides. Endocr. Rev. 1999, 20, 876–913. [CrossRef] [PubMed]
- Elliott, R.M.; Morgan, L.M.; Tredger, J.A.; Deacon, S.; Wright, J.; Marks, V. Glucagon-like peptide-1 (7-36)amide and glucosedependent insulinotropic polypeptide secretion in response to nutrient ingestion in man: Acute post-prandial and 24-h secretion patterns. J. Endocrinol. 1993, 138, 159–166. [CrossRef]
- Pujadas, G.; Drucker, D.J. Vascular Biology of Glucagon Receptor Superfamily Peptides: Mechanistic and Clinical Relevance. Endocr. Rev. 2016, 37, 554–583. [CrossRef]
- 8. Orskov, C.; Rabenhoj, L.; Wettergren, A.; Kofod, H.; Holst, J.J. Tissue and plasma concentrations of amidated and glycine-extended glucagon-like peptide I in humans. *Diabetes* **1994**, *43*, 535–539. [CrossRef]
- Fehmann, H.C.; Habener, J.F. Functional receptors for the insulinotropic hormone glucagon-like peptide-I(7–37) on a somatostatin secreting cell line. FEBS Lett. 1991, 279, 335–340. [CrossRef]
- Alvarez, E.; Martínez, M.D.; Roncero, I.; Chowen, J.A.; García-Cuartero, B.; Gispert, J.D.; Sanz, C.; Vázquez, P.; Maldonado, A.; De Cáceres, J.; et al. The expression of GLP-1 receptor mRNA and protein allows the effect of GLP-1 on glucose metabolism in the human hypothalamus and brainstem. *J. Neurochem.* 2005, 92, 798–806. [CrossRef]
- Farr, O.M.; Sofopoulos, M.; Tsoukas, M.A.; Dincer, F.; Thakkar, B.; Sahin-Efe, A.; Filippaios, A.; Bowers, J.; Srnka, A.; Gavrieli, A.; et al. GLP-1 receptors exist in the parietal cortex, hypothalamus and medulla of human brains and the GLP-1 analogue liraglutide alters brain activity related to highly desirable food cues in individuals with diabetes: A crossover, randomised, placebo-controlled trial. *Diabetologia* 2016, 59, 954–965. [CrossRef] [PubMed]

- 12. Holt, M.K.; Llewellyn-Smith, I.J.; Reimann, F.; Gribble, F.M.; Trapp, S. Serotonergic modulation of the activity of GLP-1 producing neurons in the nucleus of the solitary tract in mouse. *Mol. Metab.* **2017**, *6*, 909–921. [CrossRef] [PubMed]
- 13. Wei, Y.; Mojsov, S. Tissue-specific expression of the human receptor for glucagon-like peptide-I: Brain, heart and pancreatic forms have the same deduced amino acid sequences. *FEBS Lett.* **1995**, *358*, 219–224. [CrossRef]
- During, M.J.; Cao, L.; Zuzga, D.S.; Francis, J.S.; Fitzsimons, H.L.; Jiao, X.; Bland, R.J.; Klugmann, M.; Banks, W.A.; Drucker, D.J.; et al. Glucagon-like peptide-1 receptor is involved in learning and neuroprotection. *Nat. Med.* 2003, *9*, 1173–1179. [CrossRef]
- 15. Hamilton, A.; Holscher, C. Receptors for the insulin-like peptide GLP-1 are expressed on neurons in the CNS. *Neuroreport* 2009, 20, 1161–1166. [CrossRef] [PubMed]
- 16. Hölscher, C. Chapter Thirteen—The Role of GLP-1 in Neuronal Activity and Neurodegeneration. In *Vitamins & Hormones;* Litwack, G., Ed.; Academic Press: Cambridge, MA, USA, 2010; Volume 84, pp. 331–354.
- 17. Perry, T.; Greig, N.H. Enhancing central nervous system endogenous GLP-1 receptor pathways for intervention in Alzheimer's disease. *Curr. Alzheimer Res.* 2005, 2, 377–385. [CrossRef]
- 18. Meloni, A.R.; De Young, M.B.; Lowe, C.; Parkes, D.G. GLP-1 receptor activated insulin secretion from pancreatic beta-cells: Mechanism and glucose dependence. *Diabetes Obes. Metab.* **2013**, *15*, 15–27. [CrossRef]
- 19. Drucker, D.J.; Philippe, J.; Mojsov, S.; Chick, W.L.; Habener, J.F. Glucagon-like peptide I stimulates insulin gene expression and increases cyclic AMP levels in a rat islet cell line. *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 3434–3438. [CrossRef]
- 20. Cendrine, C.; Rémy, B. GLP-1, the Gut-Brain, and Brain-Periphery Axes. Rev. Diabet. Stud. 2011, 8, 418.
- Anderberg, R.H.; Richard, J.E.; Eerola, K.; López-Ferreras, L.; Banke, E.; Hansson, C.; Nissbrandt, H.; Berqquist, F.; Gribble, F.M.; Reimann, F.; et al. Glucagon-like Peptide 1 and Its Analogs Act in the Dorsal Raphe and Modulate Central Serotonin to Reduce Appetite and Body Weight. *Diabetes* 2017, 66, 1062–1073. [CrossRef]
- 22. Holscher, C. Central effects of GLP-1: New opportunities for treatments of neurodegenerative diseases. *J. Endocrinol.* 2014, 221, T31–T41. [CrossRef]
- Drucker, D.J.; Nauck, M.A. The incretin system: Glucagon-like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors in type 2 diabetes. *Lancet* 2006, 368, 1696–1705. [CrossRef]
- 24. Gentilella, R.; Pechtner, V.; Corcos, A.; Consoli, A. Glucagon-like peptide-1 receptor agonists in type 2 diabetes treatment: Are they all the same? *Diabetes Metab. Res. Rev.* 2019, *35*, e3070. [CrossRef] [PubMed]
- 25. Nauck, M.A.; Meier, J.J. Management Of Endocrine Disease: Are all GLP-1 agonists equal in the treatment of type 2 diabetes? *Eur. J. Endocrinol.* **2019**, *181*, R211–R234. [CrossRef] [PubMed]
- Carlessi, R.; Chen, Y.; Rowlands, J.; Cruzat, V.F.; Keane, K.N.; Egan, L.; Mamotte, C.; Stokes, R.; Gunton, J.E.; Bittencourt, P.I.H.; et al. GLP-1 receptor signalling promotes beta-cell glucose metabolism via mTOR-dependent HIF-1alpha activation. *Sci. Rep.* 2017, 7, 2661. [CrossRef]
- Park, M.K. Subchapter 17C—Glucagon-like Peptide-1. In *Handbook of Hormones*; Takei, Y., Ando, H., Tsutsui, K., Eds.; Academic Press: San Diego, CA, USA, 2016; p. 135-e117C-135.
- Cornu, M.; Thorens, B. GLP-1 protects β-cells against apoptosis by enhancing the activity of an IGF-2/IGF1-receptor autocrine loop. *Islets* 2009, 1, 280–282. [CrossRef]
- 29. Arden, C. A role for Glucagon-like Peptide-1 in the regulation of beta-cell autophagy. Peptides 2018, 100, 85–93. [CrossRef]
- Zummo, F.P.; Cullen, K.S.; Honkanen-Scott, M.; Shaw, J.A.M.; Lovat, P.E.; Arden, C. Glucagon-like Peptide 1 Protects Pancreatic Beta-Cells from Death by Increasing Autophagic Flux and Restoring Lysosomal Function. *Diabetes* 2017, 66, 1272–1285. [CrossRef]
- 31. He, Q.; Sha, S.; Sun, L.; Zhang, J.; Dong, M. GLP-1 analogue improves hepatic lipid accumulation by inducing autophagy via AMPK/mTOR pathway. *Biochem. Biophys. Res. Commun.* **2016**, 476, 196–203. [CrossRef]
- Tsunekawa, S.; Yamamoto, N.; Tsukamoto, K.; Itoh, Y.; Kaneko, Y.; Kimura, T.; Ariyoshi, Y.; Miura, Y.; Oiso, Y.; Niki, I. Protection
 of pancreatic beta-cells by exendin-4 may involve the reduction of endoplasmic reticulum stress; in vivo and in vitro studies. J.
 Endocrinol. 2007, 193, 65–74. [CrossRef]
- Yusta, B.; Baggio, L.L.; Estall, J.L.; Koehler, J.A.; Holland, D.P.; Li, H.; Pipeleers, D.; Ling, Z.; Drucker, D.J. GLP-1 receptor activation improves beta cell function and survival following induction of endoplasmic reticulum stress. *Cell Metab.* 2006, 4, 391–406. [CrossRef] [PubMed]
- Cunha, D.A.; Ladriere, L.; Ortis, F.; Igoillo-Esteve, M.; Gurzov, E.N.; Lupi, R.; Marchetti, P.; Eizirik, D.L.; Cnop, M. Glucagon-like peptide-1 agonists protect pancreatic beta-cells from lipotoxic endoplasmic reticulum stress through upregulation of BiP and JunB. *Diabetes* 2009, *58*, 2851–2862. [CrossRef] [PubMed]
- Giacco, F.; Du, X.; Carratu, A.; Gerfen, G.J.; D'Apolito, M.; Giardino, I.; Rasola, A.; Marin, O.; Divakaruni, A.S.; Murphy, A.N.; et al. GLP-1 Cleavage Product Reverses Persistent ROS Generation after Transient Hyperglycemia by Disrupting an ROS-Generating Feedback Loop. *Diabetes* 2015, 64, 3273–3284. [CrossRef] [PubMed]
- Müller, T.D.; Finan, B.; Bloom, S.R.; D'Alessio, D.; Drucker, D.J.; Flatt, P.R.; Fritsche, A.; Gribble, F.; Grill, H.J.; Habener, J.F.; et al. Glucagon-like peptide 1 (GLP-1). *Mol. Metab.* 2019, *30*, 72–130. [CrossRef] [PubMed]
- 37. Burcelin, R.; Serino, M.; Cabou, C. A role for the gut-to-brain GLP-1-dependent axis in the control of metabolism. *Curr. Opin. Pharmacol.* **2009**, *9*, 744–752. [CrossRef]

- Marlet, I.R.; Olmestig, J.N.E.; Vilsboll, T.; Rungby, J.; Kruuse, C. Neuroprotective Mechanisms of Glucagon-like Peptide-1-Based Therapies in Ischaemic Stroke: A Systematic Review Based on Pre-Clinical Studies. *Basic Clin. Pharm. Toxicol.* 2018, 122, 559–569. [CrossRef]
- Maskery, M.P.; Holscher, C.; Jones, S.P.; Price, C.I.; Strain, W.D.; Watkins, C.L.; Werring, D.J.; Emsley, H.C. Glucagon-like peptide-1 receptor agonists as neuroprotective agents for ischemic stroke: A systematic scoping review. J. Cereb. Blood Flow Metab. Off. J. Int. Soc. Cereb. Blood Flow Metab. 2021, 41, 14–30. [CrossRef]
- 40. Gray, F.o.; Duyckaerts, C.; De Girolami, U.; Escourolle, R.; Gray, F.O. *Escourolle & Poirier's Manual of Basic Neuropathology*, 5th ed.; Oxford University Press: Oxford, UK, 2014; p. xiii. 406p.
- 41. Knopman, D.S.; Amieva, H.; Petersen, R.C.; Chetelat, G.; Holtzman, D.M.; Hyman, B.T.; Nixon, R.A.; Jones, D.T. Alzheimer disease. *Nat. Rev. Dis. Primers* 2021, 7, 33. [CrossRef]
- 42. Dickson, D.W. Neuropathology of Parkinson Disease. Parkinsonism Relat. Disord. 2018, 46 (Suppl. S1), S30–S33. [CrossRef]
- Reeve, A.; Simcox, E.; Turnbull, D. Ageing and Parkinson's disease: Why is advancing age the biggest risk factor? *Ageing Res. Rev.* 2014, 14, 19–30. [CrossRef]
- 44. Poewe, W.; Seppi, K.; Tanner, C.M.; Halliday, G.M.; Brundin, P.; Volkmann, J.; Schrag, A.E.; Lang, A.E. Parkinson disease. *Nat. Rev. Dis. Primers* **2017**, *3*, 17013. [CrossRef] [PubMed]
- 45. Schapira, A.H.V. Parkinson's Disease; Oxford University Press: Oxford, UK; New York, NY, USA, 2010; p. xiv. 116p.
- Anang, J.B.; Gagnon, J.F.; Bertrand, J.A.; Romenets, S.R.; Latreille, V.; Panisset, M.; Montplaisir, J.; Postuma, R.B. Predictors of dementia in Parkinson disease: A prospective cohort study. *Neurology* 2014, 83, 1253–1260. [CrossRef] [PubMed]
- 47. Dugger, B.N.; Dickson, D.W. Pathology of Neurodegenerative Diseases. *Cold Spring Harb. Perspect. Biol.* 2017, 9, a028035. [CrossRef]
- 48. De Strooper, B.; Karran, E. The Cellular Phase of Alzheimer's Disease. Cell 2016, 164, 603–615. [CrossRef] [PubMed]
- Braak, H.; Del Tredici, K. Neuroanatomy and pathology of sporadic Parkinson's disease. In Advances in Anatomy Embryology Cell Biology; Springer: Berlin/Heidelberg, Germany, 2009; Volume 201, pp. 1–119.
- Li, Y.; Chigurupati, S.; Holloway, H.W.; Mughal, M.; Tweedie, D.; Bruestle, D.A.; Mattson, M.P.; Wang, Y.; Harvey, B.K.; Ray, B.; et al. Exendin-4 ameliorates motor neuron degeneration in cellular and animal models of amyotrophic lateral sclerosis. *PLoS ONE* 2012, 7, e32008. [CrossRef]
- 51. Holst, J.J.; Burcelin, R.; Nathanson, E. Neuroprotective properties of GLP-1: Theoretical and practical applications. *Curr. Med. Res. Opin.* **2011**, *27*, 547–558. [CrossRef]
- 52. Beal, M.F. Oxidative damage as an early marker of Alzheimer's disease and mild cognitive impairment. *Neurobiol. Aging* 2005, 26, 585–586. [CrossRef]
- Badawi, G.A.; Abd El Fattah, M.A.; Zaki, H.F.; El Sayed, M.I. Sitagliptin and liraglutide reversed nigrostriatal degeneration of rodent brain in rotenone-induced Parkinson's disease. *Inflammopharmacology* 2017, 25, 369–382. [CrossRef]
- 54. Bader, M.; Li, Y.; Tweedie, D.; Shlobin, N.A.; Bernstein, A.; Rubovitch, V.; Tovar, Y.R.L.B.; DiMarchi, R.D.; Hoffer, B.J.; Greig, N.H.; et al. Neuroprotective Effects and Treatment Potential of Incretin Mimetics in a Murine Model of Mild Traumatic Brain Injury. *Front. Cell Dev. Biol.* **2019**, *7*, 356. [CrossRef]
- 55. Bertilsson, G.; Patrone, C.; Zachrisson, O.; Andersson, A.; Dannaeus, K.; Heidrich, J.; Kortesmaa, J.; Mercer, A.; Nielsen, E.; Rönnholm, H.; et al. Peptide hormone exendin-4 stimulates subventricular zone neurogenesis in the adult rodent brain and induces recovery in an animal model of Parkinson's disease. *J. Neurosci. Res.* 2008, *86*, 326–338. [CrossRef]
- Chen, S.; Sun, J.; Zhao, G.; Guo, A.; Chen, Y.; Fu, R.; Deng, Y. Liraglutide Improves Water Maze Learning and Memory Performance While Reduces Hyperphosphorylation of Tau and Neurofilaments in APP/PS1/Tau Triple Transgenic Mice. *Neurochem. Res.* 2017, 42, 2326–2335. [CrossRef] [PubMed]
- 57. Eakin, K.; Li, Y.; Chiang, Y.H.; Hoffer, B.J.; Rosenheim, H.; Greig, N.H.; Miller, J.P. Exendin-4 ameliorates traumatic brain injury-induced cognitive impairment in rats. *PLoS ONE* **2013**, *8*, e82016. [CrossRef] [PubMed]
- Hansen, H.H.; Barkholt, P.; Fabricius, K.; Jelsing, J.; Terwel, D.; Pyke, C.; Knudsen, L.B.; Vrang, N. The GLP-1 receptor agonist liraglutide reduces pathology-specific tau phosphorylation and improves motor function in a transgenic hTauP301L mouse model of tauopathy. *Brain Res.* 2016, 1634, 158–170. [CrossRef]
- Hansen, H.H.; Fabricius, K.; Barkholt, P.; Mikkelsen, J.D.; Jelsing, J.; Pyke, C.; Knudsen, L.B.; Vrang, N. Characterization of liraglutide, a glucagon-like peptide-1 (GLP-1) receptor agonist, in rat partial and full nigral 6-hydroxydopamine lesion models of Parkinson's disease. *Brain Res.* 2016, 1646, 354–365. [CrossRef] [PubMed]
- 60. Harkavyi, A.; Abuirmeileh, A.; Lever, R.; Kingsbury, A.E.; Biggs, C.S.; Whitton, P.S. Glucagon-like peptide 1 receptor stimulation reverses key deficits in distinct rodent models of Parkinson's disease. *J. Neuroinflamm.* **2008**, *5*, 19. [CrossRef]
- 61. He, W.; Wang, H.; Zhao, C.; Tian, X.; Li, L.; Wang, H. Role of liraglutide in brain repair promotion through Sirt1-mediated mitochondrial improvement in stroke. *J. Cell. Physiol.* **2020**, 235, 2986–3001. [CrossRef]
- Li, Y.; Duffy, K.B.; Ottinger, M.A.; Ray, B.; Bailey, J.A.; Holloway, H.W.; Tweedie, D.; Perry, T.; Mattson, M.P.; Kapogiannis, D.; et al. GLP-1 receptor stimulation reduces amyloid-beta peptide accumulation and cytotoxicity in cellular and animal models of Alzheimer's disease. J. Alzheimer's Dis. JAD 2010, 19, 1205–1219. [CrossRef]
- Li, Y.; Perry, T.; Kindy, M.S.; Harvey, B.K.; Tweedie, D.; Holloway, H.W.; Powers, K.; Shen, H.; Egan, J.M.; Sambamurti, K.; et al. GLP-1 receptor stimulation preserves primary cortical and dopaminergic neurons in cellular and rodent models of stroke and Parkinsonism. *Proc. Natl. Acad. Sci. USA* 2009, 106, 1285–1290. [CrossRef]

- 64. Liu, W.; Jalewa, J.; Sharma, M.; Li, G.; Li, L.; Hölscher, C. Neuroprotective effects of lixisenatide and liraglutide in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *Neuroscience* **2015**, *303*, 42–50. [CrossRef]
- 65. Lourenco, M.V.; Clarke, J.R.; Frozza, R.L.; Bomfim, T.R.; Forny-Germano, L.; Batista, A.F.; Sathler, L.B.; Brito-Moreira, J.; Amaral, O.B.; Silva, C.A.; et al. TNF-α mediates PKR-dependent memory impairment and brain IRS-1 inhibition induced by Alzheimer's β-amyloid oligomers in mice and monkeys. *Cell Metab.* 2013, *18*, 831–843. [CrossRef]
- Perry, T.; Holloway, H.W.; Weerasuriya, A.; Mouton, P.R.; Duffy, K.; Mattison, J.A.; Greig, N.H. Evidence of GLP-1-mediated neuroprotection in an animal model of pyridoxine-induced peripheral sensory neuropathy. *Exp. Neurol.* 2007, 203, 293–301. [CrossRef] [PubMed]
- 67. Qi, L.; Ke, L.; Liu, X.; Liao, L.; Ke, S.; Liu, X.; Wang, Y.; Lin, X.; Zhou, Y.; Wu, L.; et al. Subcutaneous administration of liraglutide ameliorates learning and memory impairment by modulating tau hyperphosphorylation via the glycogen synthase kinase-3β pathway in an amyloid β protein induced alzheimer disease mouse model. *Eur. J. Pharm.* 2016, 783, 23–32. [CrossRef] [PubMed]
- Rachmany, L.; Tweedie, D.; Li, Y.; Rubovitch, V.; Holloway, H.W.; Miller, J.; Hoffer, B.J.; Greig, N.H.; Pick, C.G. Exendin-4 induced glucagon-like peptide-1 receptor activation reverses behavioral impairments of mild traumatic brain injury in mice. *Age* 2013, 35, 1621–1636. [CrossRef] [PubMed]
- 69. Zheng, J.; Xie, Y.; Ren, L.; Qi, L.; Wu, L.; Pan, X.; Zhou, J.; Chen, Z.; Liu, L. GLP-1 improves the supportive ability of astrocytes to neurons by promoting aerobic glycolysis in Alzheimer's disease. *Mol. Metab.* **2021**, *47*, 101180. [CrossRef]
- Han, W.N.; Holscher, C.; Yuan, L.; Yang, W.; Wang, X.H.; Wu, M.N.; Qi, J.S. Liraglutide protects against amyloid-beta proteininduced impairment of spatial learning and memory in rats. *Neurobiol. Aging* 2013, 34, 576–588. [CrossRef]
- 71. McClean, P.L.; Parthsarathy, V.; Faivre, E.; Hölscher, C. The diabetes drug liraglutide prevents degenerative processes in a mouse model of Alzheimer's disease. *J. Neurosci. Off. J. Soc. Neurosci.* **2011**, *31*, 6587–6594. [CrossRef] [PubMed]
- Cai, H.-Y.; Yang, J.-T.; Wang, Z.-J.; Zhang, J.; Yang, W.; Wu, M.-N.; Qi, J.-S. Lixisenatide reduces amyloid plaques, neurofibrillary tangles and neuroinflammation in an APP/PS1/tau mouse model of Alzheimer's disease. *Biochem. Biophys. Res. Commun.* 2018, 495, 1034–1040. [CrossRef]
- Zhou, M.; Chen, S.; Peng, P.; Gu, Z.; Yu, J.; Zhao, G.; Deng, Y. Dulaglutide ameliorates STZ induced AD-like impairment of learning and memory ability by modulating hyperphosphorylation of tau and NFs through GSK3β. *Biochem. Biophys. Res. Commun.* 2019, *511*, 154–160. [CrossRef]
- 74. Batista, A.F.; Forny-Germano, L.; Clarke, J.R.; Lyra e Silva, N.M.; Brito-Moreira, J.; Boehnke, S.E.; Winterborn, A.; Coe, B.C.; Lablans, A.; Vital, J.F.; et al. The diabetes drug liraglutide reverses cognitive impairment in mice and attenuates insulin receptor and synaptic pathology in a non-human primate model of Alzheimer's disease. *J. Pathol.* **2018**, 245, 85–100. [CrossRef]
- Sharma, M.K.; Jalewa, J.; Holscher, C. Neuroprotective and anti-apoptotic effects of liraglutide on SH-SY5Y cells exposed to methylglyoxal stress. J. Neurochem. 2014, 128, 459–471. [CrossRef]
- 76. McClean, P.L.; Hölscher, C. Lixisenatide, a drug developed to treat type 2 diabetes, shows neuroprotective effects in a mouse model of Alzheimer's disease. *Neuropharmacology* **2014**, *86*, 241–258. [CrossRef] [PubMed]
- 77. McClean, P.L.; Jalewa, J.; Hölscher, C. Prophylactic liraglutide treatment prevents amyloid plaque deposition, chronic inflammation and memory impairment in APP/PS1 mice. *Behav. Brain Res.* 2015, 293, 96–106. [CrossRef] [PubMed]
- Paladugu, L.; Gharaibeh, A.; Kolli, N.; Learman, C.; Hall, T.C.; Li, L.; Rossignol, J.; Maiti, P.; Dunbar, G.L. Liraglutide Has Anti-Inflammatory and Anti-Amyloid Properties in Streptozotocin-Induced and 5xFAD Mouse Models of Alzheimer's Disease. *Int. J. Mol. Sci.* 2021, 22, 860. [CrossRef] [PubMed]
- 79. Carranza-Naval, M.J.; del Marco, A.; Hierro-Bujalance, C.; Alves-Martinez, P.; Infante-Garcia, C.; Vargas-Soria, M.; Herrera, M.; Barba-Cordoba, B.; Atienza-Navarro, I.; Lubian-Lopez, S.; et al. Liraglutide Reduces Vascular Damage, Neuronal Loss, and Cognitive Impairment in a Mixed Murine Model of Alzheimer's Disease and Type 2 Diabetes. *Front. Aging Neurosci.* 2021, 13, 741923. [CrossRef]
- Cai, H.Y.; Hölscher, C.; Yue, X.H.; Zhang, S.X.; Wang, X.H.; Qiao, F.; Yang, W.; Qi, J.S. Lixisenatide rescues spatial memory and synaptic plasticity from amyloid β protein-induced impairments in rats. *Neuroscience* 2014, 277, 6–13. [CrossRef] [PubMed]
- Yun, S.P.; Kam, T.I.; Panicker, N.; Kim, S.; Oh, Y.; Park, J.S.; Kwon, S.H.; Park, Y.J.; Karuppagounder, S.S.; Park, H.; et al. Block of A1 astrocyte conversion by microglia is neuroprotective in models of Parkinson's disease. *Nat. Med.* 2018, 24, 931–938. [CrossRef]
- Bergkvist, L.; Johnson, M.E.; Mercado, G.; Steiner, J.A.; Meyerdirk, L.; Schulz, E.; Madaj, Z.; Ma, J.; Becker, K.; Li, Y.; et al. An extended release GLP-1 analogue increases alpha-synuclein accumulation in a mouse model of prodromal Parkinson's disease. *Exp. Neurol.* 2021, 341, 113693. [CrossRef]
- 83. Athauda, D.; Foltynie, T. The glucagon-like peptide 1 (GLP) receptor as a therapeutic target in Parkinson's disease: Mechanisms of action. *Drug Discov. Today* **2016**, *21*, 802–818. [CrossRef]
- Vadini, F.; Simeone, P.G.; Boccatonda, A.; Guagnano, M.T.; Liani, R.; Tripaldi, R.; Di Castelnuovo, A.; Cipollone, F.; Consoli, A.; Santilli, F. Liraglutide improves memory in obese patients with prediabetes or early type 2 diabetes: A randomized, controlled study. *Int. J. Obes.* 2020, 44, 1254–1263. [CrossRef]
- Watson, K.T.; Wroolie, T.E.; Tong, G.; Foland-Ross, L.C.; Frangou, S.; Singh, M.; McIntyre, R.S.; Roat-Shumway, S.; Myoraku, A.; Reiss, A.L.; et al. Neural correlates of liraglutide effects in persons at risk for Alzheimer's disease. *Behav. Brain Res.* 2019, 356, 271–278. [CrossRef]

- 86. Egefjord, L.; Gejl, M.; Møller, A.; Brændgaard, H.; Gottrup, H.; Antropova, O.; Møller, N.; Poulsen, H.E.; Gjedde, A.; Brock, B.; et al. Effects of liraglutide on neurodegeneration, blood flow and cognition in Alzheimer's disease—Protocol for a controlled, randomized double-blinded trial. *Dan. Med. J.* 2012, *59*, A4519.
- Gejl, M.; Gjedde, A.; Egefjord, L.; Møller, A.; Hansen, S.B.; Vang, K.; Rodell, A.; Brændgaard, H.; Gottrup, H.; Schacht, A.; et al. In Alzheimer's Disease, 6-Month Treatment with GLP-1 Analog Prevents Decline of Brain Glucose Metabolism: Randomized, Placebo-Controlled, Double-Blind Clinical Trial. *Front. Aging Neurosci.* 2016, *8*, 108. [CrossRef] [PubMed]
- 88. Femminella, G.D.; Frangou, E.; Love, S.B.; Busza, G.; Holmes, C.; Ritchie, C.; Lawrence, R.; McFarlane, B.; Tadros, G.; Ridha, B.H.; et al. Evaluating the effects of the novel GLP-1 analogue liraglutide in Alzheimer's disease: Study protocol for a randomised controlled trial (ELAD study). *Trials* **2019**, *20*, 191. [CrossRef]
- 89. Mullins, J.R.; Mustapic, M.; Chia, W.C.; Carlson, O.; Gulyani, S.; Tran, J.; Li, Y.; Mattson, P.M.; Resnick, S.; Egan, M.J.; et al. A Pilot Study of Exenatide Actions in Alzheimer's Disease. *Curr. Alzheimer Res.* **2019**, *16*, 741–752. [CrossRef] [PubMed]
- Athauda, D.; Maclagan, K.; Skene, S.S.; Bajwa-Joseph, M.; Letchford, D.; Chowdhury, K.; Hibbert, S.; Budnik, N.; Zampedri, L.; Dickson, J.; et al. Exenatide once weekly versus placebo in Parkinson's disease: A randomised, double-blind, placebo-controlled trial. *Lancet* 2017, 390, 1664–1675. [CrossRef]
- Aviles-Olmos, I.; Dickson, J.; Kefalopoulou, Z.; Djamshidian, A.; Ell, P.; Soderlund, T.; Whitton, P.; Wyse, R.; Isaacs, T.; Lees, A.; et al. Exenatide and the treatment of patients with Parkinson's disease. *J. Clin. Investig.* 2013, 123, 2730–2736. [CrossRef] [PubMed]
- 92. Mulvaney, C.A.; Duarte, G.S.; Handley, J.; Evans, D.J.; Menon, S.; Wyse, R.; Emsley, H.C. GLP-1 receptor agonists for Parkinson's disease. *Cochrane Database Syst. Rev.* 2020, 7, CD012990. [CrossRef] [PubMed]
- Martin, B.; Golden, E.; Carlson, O.D.; Pistell, P.; Zhou, J.; Kim, W.; Frank, B.P.; Thomas, S.; Chadwick, W.A.; Greig, N.H.; et al. Exendin-4 improves glycemic control, ameliorates brain and pancreatic pathologies, and extends survival in a mouse model of Huntington's disease. *Diabetes* 2009, *58*, 318–328. [CrossRef]
- Seppa, K.; Toots, M.; Reimets, R.; Jagomae, T.; Koppel, T.; Pallase, M.; Hasselholt, S.; Krogsbaek Mikkelsen, M.; Randel Nyengaard, J.; Vasar, E.; et al. GLP-1 receptor agonist liraglutide has a neuroprotective effect on an aged rat model of Wolfram syndrome. *Sci. Rep.* 2019, *9*, 15742. [CrossRef]
- 95. Bélanger, M.; Allaman, I.; Magistretti, P.J. Brain energy metabolism: Focus on astrocyte-neuron metabolic cooperation. *Cell Metab.* **2011**, *14*, 724–738. [CrossRef]
- 96. Magistretti, P.J.; Pellerin, L. Cellular bases of brain energy metabolism and their relevance to functional brain imaging: Evidence for a prominent role of astrocytes. *Cereb. Cortex* **1996**, *6*, 50–61. [CrossRef]
- Barros, L.F.; San Martín, A.; Ruminot, I.; Sandoval, P.Y.; Baeza-Lehnert, F.; Arce-Molina, R.; Rauseo, D.; Contreras-Baeza, Y.; Galaz, A.; Valdivia, S. Fluid Brain Glycolysis: Limits, Speed, Location, Moonlighting, and the Fates of Glycogen and Lactate. *Neurochem. Res.* 2020, 45, 1328–1334. [CrossRef] [PubMed]
- 98. El Hayek, L.; Khalifeh, M.; Zibara, V.; Abi Assaad, R.; Emmanuel, N.; Karnib, N.; El-Ghandour, R.; Nasrallah, P.; Bilen, M.; Ibrahim, P.; et al. Lactate Mediates the Effects of Exercise on Learning and Memory through SIRT1-Dependent Activation of Hippocampal Brain-Derived Neurotrophic Factor (BDNF). J. Neurosci. Off. J. Soc. Neurosci. 2019, 39, 2369–2382. [CrossRef] [PubMed]
- 99. Beard, E.; Lengacher, S.; Dias, S.; Magistretti, P.J.; Finsterwald, C. Astrocytes as Key Regulators of Brain Energy Metabolism: New Therapeutic Perspectives. *Front. Physiol.* **2021**, *12*, 825816. [CrossRef]
- Patching, S.G. Glucose Transporters at the Blood-Brain Barrier: Function, Regulation and Gateways for Drug Delivery. *Mol. Neurobiol.* 2017, 54, 1046–1077. [CrossRef] [PubMed]
- 101. Mergenthaler, P.; Lindauer, U.; Dienel, G.A.; Meisel, A. Sugar for the brain: The role of glucose in physiological and pathological brain function. *Trends Neurosci.* 2013, *36*, 587–597. [CrossRef] [PubMed]
- 102. Harris, J.J.; Jolivet, R.; Attwell, D. Synaptic energy use and supply. Neuron 2012, 75, 762–777. [CrossRef]
- Iadecola, C. The Neurovascular Unit Coming of Age: A Journey through Neurovascular Coupling in Health and Disease. *Neuron* 2017, 96, 17–42. [CrossRef]
- 104. Figley, C.R.; Stroman, P.W. The role(s) of astrocytes and astrocyte activity in neurometabolism, neurovascular coupling, and the production of functional neuroimaging signals. *Eur. J. Neurosci.* **2011**, *33*, 577–588. [CrossRef]
- 105. Raichle, M.E.; Mintun, M.A. Brain work and brain imaging. Annu. Rev. Neurosci. 2006, 29, 449–476. [CrossRef]
- 106. Sweeney, M.D.; Kisler, K.; Montagne, A.; Toga, A.W.; Zlokovic, B.V. The role of brain vasculature in neurodegenerative disorders. *Nat. Neurosci.* **2018**, *21*, 1318–1331. [CrossRef]
- 107. Tarasoff-Conway, J.M.; Carare, R.O.; Osorio, R.S.; Glodzik, L.; Butler, T.; Fieremans, E.; Axel, L.; Rusinek, H.; Nicholson, C.; Zlokovic, B.V.; et al. Clearance systems in the brain-implications for Alzheimer disease. *Nat. Rev. Neurol.* 2015, 11, 457–470. [CrossRef]
- 108. Zhu, M.; Ackerman, J.J.; Sukstanskii, A.L.; Yablonskiy, D.A. How the body controls brain temperature: The temperature shielding effect of cerebral blood flow. *J. Appl. Physiol.* **2006**, *101*, 1481–1488. [CrossRef]
- Newman, L.A.; Korol, D.L.; Gold, P.E. Lactate produced by glycogenolysis in astrocytes regulates memory processing. *PLoS ONE* 2011, 6, e28427. [CrossRef]
- Zlokovic, B.V. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat. Rev. Neurosci.* 2011, 12, 723–738. [CrossRef]

- 111. Nippert, A.R.; Biesecker, K.R.; Newman, E.A. Mechanisms Mediating Functional Hyperemia in the Brain. *Neurosci. A Rev. J. Bringing Neurobiol. Neurol. Psychiatry* **2018**, 24, 73–83. [CrossRef]
- 112. Mishra, A.; Reynolds, J.P.; Chen, Y.; Gourine, A.V.; Rusakov, D.A.; Attwell, D. Astrocytes mediate neurovascular signaling to capillary pericytes but not to arterioles. *Nat. Neurosci.* 2016, *19*, 1619–1627. [CrossRef]
- Zonta, M.; Angulo, M.C.; Gobbo, S.; Rosengarten, B.; Hossmann, K.A.; Pozzan, T.; Carmignoto, G. Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nat. Neurosci.* 2003, *6*, 43–50. [CrossRef]
- 114. Biesecker, K.R.; Srienc, A.I.; Shimoda, A.M.; Agarwal, A.; Bergles, D.E.; Kofuji, P.; Newman, E.A. Glial Cell Calcium Signaling Mediates Capillary Regulation of Blood Flow in the Retina. J. Neurosci. Off. J. Soc. Neurosci. 2016, 36, 9435–9445. [CrossRef]
- Ko, K.R.; Ngai, A.C.; Winn, H.R. Role of adenosine in regulation of regional cerebral blood flow in sensory cortex. *Am. J. Physiol.* 1990, 259, H1703–H1708. [CrossRef]
- 116. Kur, J.; Newman, E.A. Purinergic control of vascular tone in the retina. J. Physiol. 2014, 592, 491–504. [CrossRef]
- 117. Banks, W.A.; Reed, M.J.; Logsdon, A.F.; Rhea, E.M.; Erickson, M.A. Healthy aging and the blood-brain barrier. *Nat. Aging* **2021**, *1*, 243–254. [CrossRef]
- 118. Watkins, S.; Robel, S.; Kimbrough, I.F.; Robert, S.M.; Ellis-Davies, G.; Sontheimer, H. Disruption of astrocyte-vascular coupling and the blood-brain barrier by invading glioma cells. *Nat. Commun.* **2014**, *5*, 4196. [CrossRef]
- 119. Iadecola, C.; Nedergaard, M. Glial regulation of the cerebral microvasculature. Nat. Neurosci. 2007, 10, 1369–1376. [CrossRef]
- 120. Cunnane, S.; Nugent, S.; Roy, M.; Courchesne-Loyer, A.; Croteau, E.; Tremblay, S.; Castellano, A.; Pifferi, F.; Bocti, C.; Paquet, N.; et al. Brain fuel metabolism, aging, and Alzheimer's disease. *Nutrition* **2011**, *27*, 3–20. [CrossRef]
- 121. Sonnen, J.A.; Santa Cruz, K.; Hemmy, L.S.; Woltjer, R.; Leverenz, J.B.; Montine, K.S.; Jack, C.R.; Kaye, J.; Lim, K.; Larson, E.B.; et al. Ecology of the aging human brain. *Arch. Neurol.* **2011**, *68*, 1049–1056. [CrossRef]
- Mattay, V.S.; Fera, F.; Tessitore, A.; Hariri, A.R.; Berman, K.F.; Das, S.; Meyer-Lindenberg, A.; Goldberg, T.E.; Callicott, J.H.; Weinberger, D.R. Neurophysiological correlates of age-related changes in working memory capacity. *Neurosci. Lett.* 2006, 392, 32–37. [CrossRef]
- 123. Glisky, E.L. Frontiers in Neuroscience. Changes in Cognitive Function in Human Aging. In *Brain Aging: Models, Methods, and Mechanisms*; Riddle, D.R., Ed.; CRC Press: Boca Raton, FL, USA; Taylor & Francis Group, LLC.: Boca Raton, FL, USA, 2007.
- 124. Masliah, E.; Mallory, M.; Hansen, L.; DeTeresa, R.; Terry, R.D. Quantitative synaptic alterations in the human neocortex during normal aging. *Neurology* **1993**, *43*, 192–197. [CrossRef]
- 125. Montagne, A.; Barnes, S.R.; Sweeney, M.D.; Halliday, M.R.; Sagare, A.P.; Zhao, Z.; Toga, A.W.; Jacobs, R.E.; Liu, C.Y.; Amezcua, L.; et al. Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* **2015**, *85*, 296–302. [CrossRef]
- 126. Nelson, A.R.; Sweeney, M.D.; Sagare, A.P.; Zlokovic, B.V. Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim. Biophys. Acta* 2016, 1862, 887–900. [CrossRef]
- 127. Bender, A.R.; Prindle, J.J.; Brandmaier, A.M.; Raz, N. White matter and memory in healthy adults: Coupled changes over two years. *NeuroImage* **2016**, *131*, 193–204. [CrossRef]
- 128. Bi, Q.; Wang, W.; Niu, N.; Li, H.; Wang, Y.; Huang, W.; Chen, K.; Xu, K.; Zhang, J.; Chen, Y.; et al. Relationship between the disrupted topological efficiency of the structural brain connectome and glucose hypometabolism in normal aging. *NeuroImage* 2021, 226, 117591. [CrossRef]
- 129. Fox, N.C.; Schott, J.M. Imaging cerebral atrophy: Normal ageing to Alzheimer's disease. Lancet 2004, 363, 392–394. [CrossRef]
- Resnick, S.M.; Pham, D.L.; Kraut, M.A.; Zonderman, A.B.; Davatzikos, C. Longitudinal magnetic resonance imaging studies of older adults: A shrinking brain. J. Neurosci. Off. J. Soc. Neurosci. 2003, 23, 3295–3301. [CrossRef]
- 131. Salat, D.H.; Buckner, R.L.; Snyder, A.Z.; Greve, D.N.; Desikan, R.S.; Busa, E.; Morris, J.C.; Dale, A.M.; Fischl, B. Thinning of the cerebral cortex in aging. *Cereb. Cortex* 2004, 14, 721–730. [CrossRef]
- 132. Wrigglesworth, J.; Ward, P.; Harding, I.H.; Nilaweera, D.; Wu, Z.; Woods, R.L.; Ryan, J. Factors associated with brain ageing—A systematic review. *BMC Neurol.* 2021, 21, 312. [CrossRef]
- Surmeier, D.J.; Obeso, J.A.; Halliday, G.M. Selective neuronal vulnerability in Parkinson disease. *Nat. Rev. Neurosci.* 2017, 18, 101–113. [CrossRef]
- 134. Rudow, G.; O'Brien, R.; Savonenko, A.V.; Resnick, S.M.; Zonderman, A.B.; Pletnikova, O.; Marsh, L.; Dawson, T.M.; Crain, B.J.; West, M.J.; et al. Morphometry of the human substantia nigra in ageing and Parkinson's disease. *Acta Neuropathol.* 2008, 115, 461–470. [CrossRef]
- 135. Cabello, C.R.; Thune, J.J.; Pakkenberg, H.; Pakkenberg, B. Ageing of substantia nigra in humans: Cell loss may be compensated by hypertrophy. *Neuropathol. Appl. Neurobiol.* **2002**, *28*, 283–291. [CrossRef]
- McCann, H.; Stevens, C.H.; Cartwright, H.; Halliday, G.M. alpha-Synucleinopathy phenotypes. *Parkinsonism Relat. Disord.* 2014, 20 (Suppl. S1), S62–S67. [CrossRef]
- 137. Chan, C.S.; Gertler, T.S.; Surmeier, D.J. A molecular basis for the increased vulnerability of substantia nigra dopamine neurons in aging and Parkinson's disease. *Mov. Disord.* **2010**, *25* (Suppl. S1), S63–S70. [CrossRef]
- Wilson, R.S.; Nag, S.; Boyle, P.A.; Hizel, L.P.; Yu, L.; Buchman, A.S.; Schneider, J.A.; Bennett, D.A. Neural reserve, neuronal density in the locus ceruleus, and cognitive decline. *Neurology* 2013, *80*, 1202–1208. [CrossRef]
- 139. Yan, X.; Hu, Y.; Wang, B.; Wang, S.; Zhang, X. Metabolic Dysregulation Contributes to the Progression of Alzheimer's Disease. *Front. Neurosci.* **2020**, *14*, 530219. [CrossRef]

- 140. Kivipelto, M.; Ngandu, T.; Laatikainen, T.; Winblad, B.; Soininen, H.; Tuomilehto, J. Risk score for the prediction of dementia risk in 20 years among middle aged people: A longitudinal, population-based study. *Lancet. Neurol.* **2006**, *5*, 735–741. [CrossRef]
- 141. Rabin, J.S.; Schultz, A.P.; Hedden, T.; Viswanathan, A.; Marshall, G.A.; Kilpatrick, E.; Klein, H.; Buckley, R.F.; Yang, H.S.; Properzi, M.; et al. Interactive Associations of Vascular Risk and β-Amyloid Burden with Cognitive Decline in Clinically Normal Elderly Individuals: Findings from the Harvard Aging Brain Study. *JAMA Neurol.* 2018, 75, 1124–1131. [CrossRef]
- 142. Saito, S.; Ihara, M. Interaction between cerebrovascular disease and Alzheimer pathology. *Curr. Opin. Psychiatry* **2016**, *29*, 168–173. [CrossRef]
- 143. Brundel, M.; Heringa, S.M.; de Bresser, J.; Koek, H.L.; Zwanenburg, J.J.; Jaap Kappelle, L.; Luijten, P.R.; Biessels, G.J. High prevalence of cerebral microbleeds at 7Tesla MRI in patients with early Alzheimer's disease. J. Alzheimer's Dis. JAD 2012, 31, 259–263. [CrossRef]
- 144. Heringa, S.M.; Reijmer, Y.D.; Leemans, A.; Koek, H.L.; Kappelle, L.J.; Biessels, G.J. Multiple microbleeds are related to cerebral network disruptions in patients with early Alzheimer's disease. J. Alzheimer's Dis. JAD 2014, 38, 211–221. [CrossRef]
- 145. Shams, S.; Wahlund, L.O. Cerebral microbleeds as a biomarker in Alzheimer's disease? A review in the field. *Biomark. Med.* 2016, 10, 9–18. [CrossRef]
- 146. Uetani, H.; Hirai, T.; Hashimoto, M.; Ikeda, M.; Kitajima, M.; Sakamoto, F.; Utsunomiya, D.; Oda, S.; Sugiyama, S.; Matsubara, J.; et al. Prevalence and topography of small hypointense foci suggesting microbleeds on 3T susceptibility-weighted imaging in various types of dementia. *AJNR. Am. J. Neuroradiol.* 2013, 34, 984–989. [CrossRef]
- 147. Poliakova, T.; Levin, O.; Arablinskiy, A.; Vasenina, E.; Zerr, I. Cerebral microbleeds in early Alzheimer's disease. *J. Neurol.* **2016**, 263, 1961–1968. [CrossRef]
- 148. Yates, P.A.; Desmond, P.M.; Phal, P.M.; Steward, C.; Szoeke, C.; Salvado, O.; Ellis, K.A.; Martins, R.N.; Masters, C.L.; Ames, D.; et al. Incidence of cerebral microbleeds in preclinical Alzheimer disease. *Neurology* **2014**, *82*, 1266–1273. [CrossRef]
- 149. Vernooij, M.W.; van der Lugt, A.; Ikram, M.A.; Wielopolski, P.A.; Niessen, W.J.; Hofman, A.; Krestin, G.P.; Breteler, M.M. Prevalence and risk factors of cerebral microbleeds: The Rotterdam Scan Study. *Neurology* **2008**, *70*, 1208–1214. [CrossRef]
- 150. De la Torre, J.C. Pathophysiology of neuronal energy crisis in Alzheimer's disease. Neuro-Degener. Dis. 2008, 5, 126–132. [CrossRef]
- 151. Love, S.; Miners, J.S. Cerebrovascular disease in ageing and Alzheimer's disease. Acta Neuropathol. 2016, 131, 645–658. [CrossRef]
- 152. Roher, A.E.; Esh, C.; Rahman, A.; Kokjohn, T.A.; Beach, T.G. Atherosclerosis of cerebral arteries in Alzheimer disease. *Stroke* 2004, 35, 2623–2627. [CrossRef]
- 153. Faucheux, B.A.; Bonnet, A.M.; Agid, Y.; Hirsch, E.C. Blood vessels change in the mesencephalon of patients with Parkinson's disease. *Lancet* **1999**, 353, 981–982. [CrossRef]
- 154. Guan, J.; Pavlovic, D.; Dalkie, N.; Waldvogel, H.J.; O'Carroll, S.J.; Green, C.R.; Nicholson, L.F. Vascular degeneration in Parkinson's disease. *Brain Pathol.* 2013, 23, 154–164. [CrossRef]
- Al-Bachari, S.; Vidyasagar, R.; Emsley, H.C.; Parkes, L.M. Structural and physiological neurovascular changes in idiopathic Parkinson's disease and its clinical phenotypes. J. Cereb. Blood Flow Metab. Off. J. Int. Soc. Cereb. Blood Flow Metab. 2017, 37, 3409–3421. [CrossRef]
- Borghammer, P.; Chakravarty, M.; Jonsdottir, K.Y.; Sato, N.; Matsuda, H.; Ito, K.; Arahata, Y.; Kato, T.; Gjedde, A. Cortical hypometabolism and hypoperfusion in Parkinson's disease is extensive: Probably even at early disease stages. *Brain Struct. Funct.* 2010, 214, 303–317. [CrossRef]
- 157. Fernandez-Seara, M.A.; Mengual, E.; Vidorreta, M.; Aznarez-Sanado, M.; Loayza, F.R.; Villagra, F.; Irigoyen, J.; Pastor, M.A. Cortical hypoperfusion in Parkinson's disease assessed using arterial spin labeled perfusion MRI. *NeuroImage* 2012, 59, 2743–2750. [CrossRef] [PubMed]
- 158. Alexopoulos, P.; Sorg, C.; Förschler, A.; Grimmer, T.; Skokou, M.; Wohlschläger, A.; Perneczky, R.; Zimmer, C.; Kurz, A.; Preibisch, C. Perfusion abnormalities in mild cognitive impairment and mild dementia in Alzheimer's disease measured by pulsed arterial spin labeling MRI. *Eur. Arch. Psychiatry Clin. Neurosci.* **2012**, 262, 69–77. [CrossRef] [PubMed]
- 159. Dai, W.; Lopez, O.L.; Carmichael, O.T.; Becker, J.T.; Kuller, L.H.; Gach, H.M. Mild cognitive impairment and alzheimer disease: Patterns of altered cerebral blood flow at MR imaging. *Radiology* **2009**, 250, 856–866. [CrossRef]
- den Abeelen, A.S.; Lagro, J.; van Beek, A.H.; Claassen, J.A. Impaired cerebral autoregulation and vasomotor reactivity in sporadic Alzheimer's disease. *Curr. Alzheimer Res.* 2014, 11, 11–17. [CrossRef]
- Hays, C.C.; Zlatar, Z.Z.; Wierenga, C.E. The Utility of Cerebral Blood Flow as a Biomarker of Preclinical Alzheimer's Disease. *Cell. Mol. Neurobiol.* 2016, 36, 167–179. [CrossRef]
- 162. Ruitenberg, A.; den Heijer, T.; Bakker, S.L.; van Swieten, J.C.; Koudstaal, P.J.; Hofman, A.; Breteler, M.M. Cerebral hypoperfusion and clinical onset of dementia: The Rotterdam Study. *Ann. Neurol.* 2005, *57*, 789–794. [CrossRef]
- Costantini, L.C.; Barr, L.J.; Vogel, J.L.; Henderson, S.T. Hypometabolism as a therapeutic target in Alzheimer's disease. BMC Neurosci. 2008, 9 (Suppl. S2), S16. [CrossRef]
- 164. Goyal, M.S.; Vlassenko, A.G.; Blazey, T.M.; Su, Y.; Couture, L.E.; Durbin, T.J.; Bateman, R.J.; Benzinger, T.L.; Morris, J.C.; Raichle, M.E. Loss of Brain Aerobic Glycolysis in Normal Human Aging. *Cell Metab.* 2017, 26, 353–360. [CrossRef]
- 165. Jagust, W.J.; Seab, J.P.; Huesman, R.H.; Valk, P.E.; Mathis, C.A.; Reed, B.R.; Coxson, P.G.; Budinger, T.F. Diminished glucose transport in Alzheimer's disease: Dynamic PET studies. J. Cereb. Blood Flow Metab. Off. J. Int. Soc. Cereb. Blood Flow Metab. 1991, 11, 323–330. [CrossRef]

- 166. Mosconi, L.; Mistur, R.; Switalski, R.; Tsui, W.H.; Glodzik, L.; Li, Y.; Pirraglia, E.; De Santi, S.; Reisberg, B.; Wisniewski, T.; et al. FDG-PET changes in brain glucose metabolism from normal cognition to pathologically verified Alzheimer's disease. *Eur. J. Nucl. Med. Mol. Imaging* 2009, 36, 811–822. [CrossRef]
- 167. Bailly, M.; Destrieux, C.; Hommet, C.; Mondon, K.; Cottier, J.P.; Beaufils, E.; Vierron, E.; Vercouillie, J.; Ibazizene, M.; Voisin, T.; et al. Precuneus and Cingulate Cortex Atrophy and Hypometabolism in Patients with Alzheimer's Disease and Mild Cognitive Impairment: MRI and (18)F-FDG PET Quantitative Analysis Using FreeSurfer. *BioMed Res. Int.* 2015, 2015, 583931. [CrossRef] [PubMed]
- Minoshima, S.; Giordani, B.; Berent, S.; Frey, K.A.; Foster, N.L.; Kuhl, D.E. Metabolic reduction in the posterior cingulate cortex in very early Alzheimer's disease. *Ann. Neurol.* 1997, 42, 85–94. [CrossRef] [PubMed]
- Reiman, E.M.; Chen, K.; Alexander, G.E.; Caselli, R.J.; Bandy, D.; Osborne, D.; Saunders, A.M.; Hardy, J. Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia. *Proc. Natl. Acad. Sci. USA* 2004, 101, 284–289. [CrossRef]
- 170. Iaccarino, L.; Chiotis, K.; Alongi, P.; Almkvist, O.; Wall, A.; Cerami, C.; Bettinardi, V.; Gianolli, L.; Nordberg, A.; Perani, D. A Cross-Validation of FDG- and Amyloid-PET Biomarkers in Mild Cognitive Impairment for the Risk Prediction to Dementia due to Alzheimer's Disease in a Clinical Setting. J. Alzheimer's Dis. JAD 2017, 59, 603–614. [CrossRef]
- 171. Landau, S.M.; Harvey, D.; Madison, C.M.; Koeppe, R.A.; Reiman, E.M.; Foster, N.L.; Weiner, M.W.; Jagust, W.J. Associations between cognitive, functional, and FDG-PET measures of decline in AD and MCI. *Neurobiol. Aging* **2011**, *32*, 1207–1218. [CrossRef]
- 172. Arnold, S.E.; Arvanitakis, Z.; Macauley-Rambach, S.L.; Koenig, A.M.; Wang, H.Y.; Ahima, R.S.; Craft, S.; Gandy, S.; Buettner, C.; Stoeckel, L.E.; et al. Brain insulin resistance in type 2 diabetes and Alzheimer disease: Concepts and conundrums. *Nat. Rev. Neurol.* 2018, 14, 168–181. [CrossRef]
- 173. De la Monte, S.M. Insulin resistance and Alzheimer's disease. BMB Rep. 2009, 42, 475–481. [CrossRef]
- 174. Hoyer, S. Glucose metabolism and insulin receptor signal transduction in Alzheimer disease. *Eur. J. Pharmacol.* **2004**, 490, 115–125. [CrossRef]
- 175. Kellar, D.; Craft, S. Brain insulin resistance in Alzheimer's disease and related disorders: Mechanisms and therapeutic approaches. *Lancet. Neurol.* 2020, 19, 758–766. [CrossRef]
- 176. Liang, W.S.; Reiman, E.M.; Valla, J.; Dunckley, T.; Beach, T.G.; Grover, A.; Niedzielko, T.L.; Schneider, L.E.; Mastroeni, D.; Caselli, R.; et al. Alzheimer's disease is associated with reduced expression of energy metabolism genes in posterior cingulate neurons. *Proc. Natl. Acad. Sci. USA* 2008, 105, 4441–4446. [CrossRef]
- 177. Marcus, D.L.; Freedman, M.L. Decreased brain glucose metabolism in microvessels from patients with Alzheimer's disease. *Ann. N. Y. Acad. Sci.* **1997**, *826*, 248–253. [CrossRef] [PubMed]
- 178. Neth, B.J.; Craft, S. Insulin Resistance and Alzheimer's Disease: Bioenergetic Linkages. *Front. Aging Neurosci.* **2017**, *9*, 345. [CrossRef] [PubMed]
- 179. Rivera, E.J.; Goldin, A.; Fulmer, N.; Tavares, R.; Wands, J.R.; de la Monte, S.M. Insulin and insulin-like growth factor expression and function deteriorate with progression of Alzheimer's disease: Link to brain reductions in acetylcholine. *J. Alzheimer's Dis. JAD* 2005, *8*, 247–268. [CrossRef]
- Chiu, S.L.; Chen, C.M.; Cline, H.T. Insulin receptor signaling regulates synapse number, dendritic plasticity, and circuit function in vivo. *Neuron* 2008, 58, 708–719. [CrossRef] [PubMed]
- De Felice, F.G.; Vieira, M.N.; Bomfim, T.R.; Decker, H.; Velasco, P.T.; Lambert, M.P.; Viola, K.L.; Zhao, W.Q.; Ferreira, S.T.; Klein, W.L. Protection of synapses against Alzheimer's-linked toxins: Insulin signaling prevents the pathogenic binding of Abeta oligomers. *Proc. Natl. Acad. Sci. USA* 2009, 106, 1971–1976. [CrossRef] [PubMed]
- 182. Cholerton, B.; Baker, L.D.; Craft, S. Insulin resistance and pathological brain ageing. Diabet. Med. 2011, 28, 1463–1475. [CrossRef]
- 183. Holscher, C. Diabetes as a risk factor for Alzheimer's disease: Insulin signalling impairment in the brain as an alternative model of Alzheimer's disease. *Biochem. Soc. Trans.* 2011, *39*, 891–897. [CrossRef]
- 184. Arvanitakis, Z.; Wilson, R.S.; Bennett, D.A. Diabetes mellitus, dementia, and cognitive function in older persons. J. Nutr. Health Aging 2006, 10, 287–291.
- 185. Baker, L.D.; Cross, D.J.; Minoshima, S.; Belongia, D.; Watson, G.S.; Craft, S. Insulin resistance and Alzheimer-like reductions in regional cerebral glucose metabolism for cognitively normal adults with prediabetes or early type 2 diabetes. *Arch. Neurol.* 2011, 68, 51–57. [CrossRef]
- Willette, A.A.; Modanlo, N.; Kapogiannis, D.; Alzheimer's Disease Neuroimaging, I. Insulin resistance predicts medial temporal hypermetabolism in mild cognitive impairment conversion to Alzheimer disease. *Diabetes* 2015, 64, 1933–1940. [CrossRef]
- 187. Harr, S.D.; Simonian, N.A.; Hyman, B.T. Functional alterations in Alzheimer's disease: Decreased glucose transporter 3 immunoreactivity in the perforant pathway terminal zone. *J. Neuropathol. Exp. Neurol.* **1995**, *54*, 38–41. [CrossRef] [PubMed]
- 188. Mooradian, A.D.; Chung, H.C.; Shah, G.N. GLUT-1 expression in the cerebra of patients with Alzheimer's disease. *Neurobiol. Aging* **1997**, *18*, 469–474. [CrossRef]
- Simpson, I.A.; Chundu, K.R.; Davies-Hill, T.; Honer, W.G.; Davies, P. Decreased concentrations of GLUT1 and GLUT3 glucose transporters in the brains of patients with Alzheimer's disease. *Ann. Neurol.* 1994, 35, 546–551. [CrossRef] [PubMed]
- 190. Gibson, G.E.; Sheu, K.F.; Blass, J.P.; Baker, A.; Carlson, K.C.; Harding, B.; Perrino, P. Reduced activities of thiamine-dependent enzymes in the brains and peripheral tissues of patients with Alzheimer's disease. *Arch. Neurol.* **1988**, 45, 836–840. [CrossRef]

- 191. Kish, S.J.; Bergeron, C.; Rajput, A.; Dozic, S.; Mastrogiacomo, F.; Chang, L.J.; Wilson, J.M.; DiStefano, L.M.; Nobrega, J.N. Brain cytochrome oxidase in Alzheimer's disease. *J. Neurochem.* **1992**, *59*, 776–779. [CrossRef]
- 192. Bosco, D.; Plastino, M.; Cristiano, D.; Colica, C.; Ermio, C.; De Bartolo, M.; Mungari, P.; Fonte, G.; Consoli, D.; Consoli, A.; et al. Dementia is associated with Insulin Resistance in patients with Parkinson's Disease. *J. Neurol. Sci.* 2012, *315*, 39–43. [CrossRef]
- 193. Santiago, J.A.; Potashkin, J.A. Shared dysregulated pathways lead to Parkinson's disease and diabetes. *Trends Mol. Med.* 2013, 19, 176–186. [CrossRef]
- 194. Pagano, G.; Polychronis, S.; Wilson, H.; Giordano, B.; Ferrara, N.; Niccolini, F.; Politis, M. Diabetes mellitus and Parkinson disease. *Neurology* **2018**, *90*, e1654–e1662. [CrossRef]
- 195. Giuntini, M.; Baldacci, F.; Del Prete, E.; Bonuccelli, U.; Ceravolo, R. Diabetes is associated with postural and cognitive domains in Parkinson's disease. Results from a single-center study. *Parkinsonism Relat. Disord.* **2014**, 20, 671–672. [CrossRef]
- 196. Hu, G.; Jousilahti, P.; Bidel, S.; Antikainen, R.; Tuomilehto, J. Type 2 diabetes and the risk of Parkinson's disease. *Diabetes Care* **2007**, *30*, 842–847. [CrossRef]
- 197. Dunn, L.; Allen, G.F.; Mamais, A.; Ling, H.; Li, A.; Duberley, K.E.; Hargreaves, I.P.; Pope, S.; Holton, J.L.; Lees, A.; et al. Dysregulation of glucose metabolism is an early event in sporadic Parkinson's disease. *Neurobiol. Aging* 2014, 35, 1111–1115. [CrossRef] [PubMed]
- Delaidelli, A.; Jan, A.; Herms, J.; Sorensen, P.H. Translational control in brain pathologies: Biological significance and therapeutic opportunities. *Acta Neuropathol.* 2019, 137, 535–555. [CrossRef] [PubMed]
- 199. Li, X.; Alafuzoff, I.; Soininen, H.; Winblad, B.; Pei, J.J. Levels of mTOR and its downstream targets 4E-BP1, eEF2, and eEF2 kinase in relationships with tau in Alzheimer's disease brain. *FEBS J.* **2005**, *272*, 4211–4220. [CrossRef] [PubMed]
- Vingtdeux, V.; Davies, P.; Dickson, D.W.; Marambaud, P. AMPK is abnormally activated in tangle- and pre-tangle-bearing neurons in Alzheimer's disease and other tauopathies. *Acta Neuropathol.* 2011, 121, 337–349. [CrossRef] [PubMed]
- 201. Jan, A.; Jansonius, B.; Delaidelli, A.; Bhanshali, F.; An, Y.A.; Ferreira, N.; Smits, L.M.; Negri, G.L.; Schwamborn, J.C.; Jensen, P.H.; et al. Activity of translation regulator eukaryotic elongation factor-2 kinase is increased in Parkinson disease brain and its inhibition reduces alpha synuclein toxicity. *Acta Neuropathol. Commun.* 2018, 6, 54. [CrossRef]
- 202. Jan, A.; Jansonius, B.; Delaidelli, A.; Somasekharan, S.P.; Bhanshali, F.; Vandal, M.; Negri, G.L.; Moerman, D.; MacKenzie, I.; Calon, F.; et al. eEF2K inhibition blocks Abeta42 neurotoxicity by promoting an NRF2 antioxidant response. *Acta Neuropathol.* 2017, 133, 101–119. [CrossRef]
- Chang, R.C.; Wong, A.K.; Ng, H.K.; Hugon, J. Phosphorylation of eukaryotic initiation factor-2alpha (eIF2alpha) is associated with neuronal degeneration in Alzheimer's disease. *Neuroreport* 2002, *13*, 2429–2432. [CrossRef]
- 204. Wang, Y.; Parlevliet, E.T.; Geerling, J.J.; van der Tuin, S.J.; Zhang, H.; Bieghs, V.; Jawad, A.H.; Shiri-Sverdlov, R.; Bot, I.; de Jager, S.C.; et al. Exendin-4 decreases liver inflammation and atherosclerosis development simultaneously by reducing macrophage infiltration. *Br. J. Pharmacol.* 2014, 171, 723–734. [CrossRef]
- 205. Helmstadter, J.; Keppeler, K.; Aust, F.; Kuster, L.; Frenis, K.; Filippou, K.; Vujacic-Mirski, K.; Tsohataridis, S.; Kalinovic, S.; Kroller-Schon, S.; et al. GLP-1 Analog Liraglutide Improves Vascular Function in Polymicrobial Sepsis by Reduction of Oxidative Stress and Inflammation. *Antioxidants* 2021, 10, 1175. [CrossRef]
- 206. DeFronzo, R.A.; Ferrannini, E.; Groop, L.; Henry, R.R.; Herman, W.H.; Holst, J.J.; Hu, F.B.; Kahn, C.R.; Raz, I.; Shulman, G.I.; et al. Type 2 diabetes mellitus. *Nat. Rev. Dis. Primers* 2015, 1, 15019. [CrossRef]
- Kitada, M.; Zhang, Z.; Mima, A.; King, G.L. Molecular mechanisms of diabetic vascular complications. J. Diabetes Investig. 2010, 1, 77–89. [CrossRef] [PubMed]
- Kokkola, R.; Andersson, A.; Mullins, G.; Ostberg, T.; Treutiger, C.J.; Arnold, B.; Nawroth, P.; Andersson, U.; Harris, R.A.; Harris, H.E. RAGE is the major receptor for the proinflammatory activity of HMGB1 in rodent macrophages. *Scand. J. Immunol.* 2005, 61, 1–9. [CrossRef] [PubMed]
- 209. Sasaki, N.; Fukatsu, R.; Tsuzuki, K.; Hayashi, Y.; Yoshida, T.; Fujii, N.; Koike, T.; Wakayama, I.; Yanagihara, R.; Garruto, R.; et al. Advanced glycation end products in Alzheimer's disease and other neurodegenerative diseases. *Am. J. Pathol.* **1998**, 153, 1149–1155. [CrossRef]
- Srikanth, V.; Maczurek, A.; Phan, T.; Steele, M.; Westcott, B.; Juskiw, D.; Munch, G. Advanced glycation endproducts and their receptor RAGE in Alzheimer's disease. *Neurobiol. Aging* 2011, 32, 763–777. [CrossRef]
- 211. Luth, H.J.; Ogunlade, V.; Kuhla, B.; Kientsch-Engel, R.; Stahl, P.; Webster, J.; Arendt, T.; Munch, G. Age- and stage-dependent accumulation of advanced glycation end products in intracellular deposits in normal and Alzheimer's disease brains. *Cereb. Cortex* 2005, 15, 211–220. [CrossRef]
- 212. Jana, A.K.; Batkulwar, K.B.; Kulkarni, M.J.; Sengupta, N. Glycation induces conformational changes in the amyloid-beta peptide and enhances its aggregation propensity: Molecular insights. *Phys. Chem. Chem. Phys.* **2016**, *18*, 31446–31458. [CrossRef]
- Craft, S.; Newcomer, J.; Kanne, S.; Dagogo-Jack, S.; Cryer, P.; Sheline, Y.; Luby, J.; Dagogo-Jack, A.; Alderson, A. Memory improvement following induced hyperinsulinemia in Alzheimer's disease. *Neurobiol. Aging* 1996, 17, 123–130. [CrossRef]
- Sartorius, T.; Peter, A.; Heni, M.; Maetzler, W.; Fritsche, A.; Haring, H.U.; Hennige, A.M. The brain response to peripheral insulin declines with age: A contribution of the blood-brain barrier? *PLoS ONE* 2015, *10*, e0126804. [CrossRef]
- 215. Steen, E.; Terry, B.M.; Rivera, E.J.; Cannon, J.L.; Neely, T.R.; Tavares, R.; Xu, X.J.; Wands, J.R.; de la Monte, S.M. Impaired insulin and insulin-like growth factor expression and signaling mechanisms in Alzheimer's disease—Is this type 3 diabetes? *J. Alzheimer's Dis. JAD* 2005, *7*, 63–80. [CrossRef]

- 216. Horwood, N.; Davies, D.C. Immunolabelling of hippocampal microvessel glucose transporter protein is reduced in Alzheimer's disease. *Virchows Arch. Int. J. Pathol.* **1994**, 425, 69–72. [CrossRef]
- 217. Winkler, E.A.; Nishida, Y.; Sagare, A.P.; Rege, S.V.; Bell, R.D.; Perlmutter, D.; Sengillo, J.D.; Hillman, S.; Kong, P.; Nelson, A.R.; et al. GLUT1 reductions exacerbate Alzheimer's disease vasculo-neuronal dysfunction and degeneration. *Nat. Neurosci.* 2015, 18, 521–530. [CrossRef] [PubMed]
- Lee, C.H.; Yan, B.; Yoo, K.Y.; Choi, J.H.; Kwon, S.H.; Her, S.; Sohn, Y.; Hwang, I.K.; Cho, J.H.; Kim, Y.M.; et al. Ischemia-induced changes in glucagon-like peptide-1 receptor and neuroprotective effect of its agonist, exendin-4, in experimental transient cerebral ischemia. *J. Neurosci. Res.* 2011, 89, 1103–1113. [CrossRef] [PubMed]
- Reiner, D.J.; Mietlicki-Baase, E.G.; McGrath, L.E.; Zimmer, D.J.; Bence, K.K.; Sousa, G.L.; Konanur, V.R.; Krawczyk, J.; Burk, D.H.; Kanoski, S.E.; et al. Astrocytes Regulate GLP-1 Receptor-Mediated Effects on Energy Balance. *J. Neurosci. Off. J. Soc. Neurosci.* 2016, *36*, 3531–3540. [CrossRef] [PubMed]
- 220. Timper, K.; Del Rio-Martin, A.; Cremer, A.L.; Bremser, S.; Alber, J.; Giavalisco, P.; Varela, L.; Heilinger, C.; Nolte, H.; Trifunovic, A.; et al. GLP-1 Receptor Signaling in Astrocytes Regulates Fatty Acid Oxidation, Mitochondrial Integrity, and Function. *Cell Metab.* 2020, *31*, 1189–1205.e13. [CrossRef]
- 221. Magistretti, P.J.; Allaman, I. Lactate in the brain: From metabolic end-product to signalling molecule. *Nat. Rev. Neurosci.* 2018, 19, 235–249. [CrossRef]
- 222. Supplie, L.M.; Duking, T.; Campbell, G.; Diaz, F.; Moraes, C.T.; Gotz, M.; Hamprecht, B.; Boretius, S.; Mahad, D.; Nave, K.A. Respiration-Deficient Astrocytes Survive As Glycolytic Cells In Vivo. *J. Neurosci. Off. J. Soc. Neurosci.* 2017, 37, 4231–4242. [CrossRef]
- 223. Karagiannis, A.; Gallopin, T.; Lacroix, A.; Plaisier, F.; Piquet, J.; Geoffroy, H.; Hepp, R.; Naudé, J.; Le Gac, B.; Egger, R.; et al. Lactate is an energy substrate for rodent cortical neurons and enhances their firing activity. *eLife* **2021**, *10*, e71424. [CrossRef]
- 224. Pellerin, L.; Magistretti, P.J. Glutamate uptake into astrocytes stimulates aerobic glycolysis: A mechanism coupling neuronal activity to glucose utilization. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 10625–10629. [CrossRef]
- Berthet, C.; Castillo, X.; Magistretti, P.J.; Hirt, L. New evidence of neuroprotection by lactate after transient focal cerebral ischaemia: Extended benefit after intracerebroventricular injection and efficacy of intravenous administration. *Cerebrovasc. Dis.* 2012, 34, 329–335. [CrossRef]
- 226. Suzuki, A.; Stern, S.A.; Bozdagi, O.; Huntley, G.W.; Walker, R.H.; Magistretti, P.J.; Alberini, C.M. Astrocyte-neuron lactate transport is required for long-term memory formation. *Cell* **2011**, 144, 810–823. [CrossRef]
- 227. Fan, Z.; Zhang, Z.; Zhao, S.; Zhu, Y.; Guo, D.; Yang, B.; Zhuo, L.; Han, J.; Wang, R.; Fang, Z.; et al. Dynamic Variations in Brain Glycogen are Involved in Modulating Isoflurane Anesthesia in Mice. *Neurosci. Bull.* 2020, 36, 1513–1523. [CrossRef] [PubMed]
- Kajihara, H.; Tsutsumi, E.; Kinoshita, A.; Nakano, J.; Takagi, K.; Takeo, S. Activated astrocytes with glycogen accumulation in ischemic penumbra during the early stage of brain infarction: Immunohistochemical and electron microscopic studies. *Brain Res.* 2001, 909, 92–101. [CrossRef]
- Alcantara, A.I.; Morales, M.; Delgado, E.; Lopez-Delgado, M.I.; Clemente, F.; Luque, M.A.; Malaisse, W.J.; Valverde, I.; Villanueva-Penacarrillo, M.L. Exendin-4 agonist and exendin(9-39)amide antagonist of the GLP-1(7-36)amide effects in liver and muscle. *Arch. Biochem. Biophys.* 1997, 341, 1–7. [CrossRef] [PubMed]
- Redondo, A.; Trigo, M.V.; Acitores, A.; Valverde, I.; Villanueva-Penacarrillo, M.L. Cell signalling of the GLP-1 action in rat liver. Mol. Cell Endocrinol. 2003, 204, 43–50. [CrossRef]
- Ambrosi, G.; Cerri, S.; Blandini, F. A further update on the role of excitotoxicity in the pathogenesis of Parkinson's disease. J. Neural. Transm. 2014, 121, 849–859. [CrossRef]
- Garcia, J.D.; Gookin, S.E.; Crosby, K.C.; Schwartz, S.L.; Tiemeier, E.; Kennedy, M.J.; Dell'Acqua, M.L.; Herson, P.S.; Quillinan, N.; Smith, K.R. Stepwise disassembly of GABAergic synapses during pathogenic excitotoxicity. *Cell Rep.* 2021, 37, 110142. [CrossRef]
- Garry, D.J.; Appel, N.M.; Garry, M.G.; Sorenson, R.L. Cellular and subcellular immunolocalization of L-glutamate decarboxylase in rat pancreatic islets. J. Histochem. Cytochem. 1988, 36, 573–580. [CrossRef]
- 234. Sorenson, R.L.; Garry, D.G.; Brelje, T.C. Structural and functional considerations of GABA in islets of Langerhans. Beta-cells and nerves. *Diabetes* **1991**, *40*, 1365–1374. [CrossRef]
- 235. Rorsman, P.; Berggren, P.O.; Bokvist, K.; Ericson, H.; Mohler, H.; Ostenson, C.G.; Smith, P.A. Glucose-inhibition of glucagon secretion involves activation of GABAA-receptor chloride channels. *Nature* **1989**, *341*, 233–236. [CrossRef]
- Wang, C.; Mao, R.; Van de Casteele, M.; Pipeleers, D.; Ling, Z. Glucagon-like peptide-1 stimulates GABA formation by pancreatic beta-cells at the level of glutamate decarboxylase. *Am. J. Physiol. Endocrinol. Metab.* 2007, 292, E1201–E1206. [CrossRef]
- 237. Wendt, A.; Birnir, B.; Buschard, K.; Gromada, J.; Salehi, A.; Sewing, S.; Rorsman, P.; Braun, M. Glucose inhibition of glucagon secretion from rat alpha-cells is mediated by GABA released from neighboring beta-cells. *Diabetes* 2004, 53, 1038–1045. [CrossRef] [PubMed]
- Wang, Q.; Liu, L.; Pei, L.; Ju, W.; Ahmadian, G.; Lu, J.; Wang, Y.; Liu, F.; Wang, Y.T. Control of synaptic strength, a novel function of Akt. *Neuron* 2003, *38*, 915–928. [CrossRef]
- Fortin, S.M.; Lipsky, R.K.; Lhamo, R.; Chen, J.; Kim, E.; Borner, T.; Schmidt, H.D.; Hayes, M.R. GABA neurons in the nucleus tractus solitarius express GLP-1 receptors and mediate anorectic effects of liraglutide in rats. *Sci. Transl. Med.* 2020, 12, eaay8071. [CrossRef] [PubMed]

- Kabahizi, A.; Wallace, B.; Lieu, L.; Chau, D.; Dong, Y.; Hwang, E.S.; Williams, K.W. Glucagon-like peptide-1 (GLP-1) signalling in the brain: From neural circuits and metabolism to therapeutics. *Br. J. Pharmacol.* 2022, 179, 600–624. [CrossRef]
- Laurindo, L.F.; Barbalho, S.M.; Guiguer, E.L.; da Silva Soares de Souza, M.; de Souza, G.A.; Fidalgo, T.M.; Araujo, A.C.; de Souza Gonzaga, H.F.; de Bortoli Teixeira, D.; de Oliveira Silva Ullmann, T.; et al. GLP-1a: Going Beyond Traditional Use. *Int. J. Mol. Sci.* 2022, 23, 739. [CrossRef]
- 242. Göke, R.; Larsen, P.J.; Mikkelsen, J.D.; Sheikh, S.P. Distribution of GLP-1 Binding Sites in the Rat Brain: Evidence that Exendin-4 is a Ligand of Brain GLP-1 Binding Sites. *Eur. J. Neurosci.* **1995**, *7*, 2294–2300. [CrossRef]
- 243. Brosseron, F.; Krauthausen, M.; Kummer, M.; Heneka, M.T. Body fluid cytokine levels in mild cognitive impairment and Alzheimer's disease: A comparative overview. *Mol. Neurobiol.* **2014**, *50*, 534–544. [CrossRef]
- 244. Domingues, C.; da Cruz, E.S.O.A.B.; Henriques, A.G. Impact of Cytokines and Chemokines on Alzheimer's Disease Neuropathological Hallmarks. *Curr. Alzheimer Res.* 2017, 14, 870–882. [CrossRef]
- 245. Galea, I. The blood-brain barrier in systemic infection and inflammation. Cell Mol. Immunol. 2021, 18, 2489–2501. [CrossRef]
- Attems, J.; Jellinger, K.A. The overlap between vascular disease and Alzheimer's disease–lessons from pathology. BMC Med. 2014, 12, 206. [CrossRef]
- 247. Wijdicks, E.F. Hepatic Encephalopathy. N. Engl. J. Med. 2016, 375, 1660–1670. [CrossRef] [PubMed]
- Rosner, M.H.; Husain-Syed, F.; Reis, T.; Ronco, C.; Vanholder, R. Uremic encephalopathy. *Kidney Int.* 2022, 101, 227–241. [CrossRef]
 [PubMed]
- 249. Melendez-Flores, J.D.; Estrada-Bellmann, I. Linking chronic kidney disease and Parkinson's disease: A literature review. *Metab. Brain Dis.* **2021**, *36*, 1–12. [CrossRef] [PubMed]
- Tryc, A.B.; Goldbecker, A.; Berding, G.; Rumke, S.; Afshar, K.; Shahrezaei, G.H.; Pflugrad, H.; Barg-Hock, H.; Strassburg, C.P.; Hecker, H.; et al. Cirrhosis-related Parkinsonism: Prevalence, mechanisms and response to treatments. *J. Hepatol.* 2013, 58, 698–705. [CrossRef]
- 251. Weinstein, G.; Zelber-Sagi, S.; Preis, S.R.; Beiser, A.S.; DeCarli, C.; Speliotes, E.K.; Satizabal, C.L.; Vasan, R.S.; Seshadri, S. Association of Nonalcoholic Fatty Liver Disease with Lower Brain Volume in Healthy Middle-Aged Adults in the Framingham Study. JAMA Neurol. 2018, 75, 97–104. [CrossRef]
- Zhang, C.Y.; He, F.F.; Su, H.; Zhang, C.; Meng, X.F. Association between chronic kidney disease and Alzheimer's disease: An update. *Metab. Brain Dis.* 2020, 35, 883–894. [CrossRef]
- Viggiano, D.; Wagner, C.A.; Martino, G.; Nedergaard, M.; Zoccali, C.; Unwin, R.; Capasso, G. Mechanisms of cognitive dysfunction in CKD. Nat. Rev. Nephrol. 2020, 16, 452–469. [CrossRef]
- 254. Younossi, Z.; Tacke, F.; Arrese, M.; Chander Sharma, B.; Mostafa, I.; Bugianesi, E.; Wai-Sun Wong, V.; Yilmaz, Y.; George, J.; Fan, J.; et al. Global Perspectives on Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis. *Hepatology* 2019, 69, 2672–2682. [CrossRef]
- Collaboration, G.B.D.C.K.D. Global, regional, and national burden of chronic kidney disease, 1990–2017: A systematic analysis for the Global Burden of Disease Study 2017. *Lancet* 2020, 395, 709–733. [CrossRef]
- 256. Cordato, D.J.; Chan, D.K. Genetics and Parkinson's disease. J. Clin. Neurosci. 2004, 11, 119–123. [CrossRef]
- Ur Rasheed, M.S.; Mishra, A.K.; Singh, M.P. Cytochrome P450 2D6 and Parkinson's Disease: Polymorphism, Metabolic Role, Risk and Protection. *Neurochem. Res.* 2017, 42, 3353–3361. [CrossRef] [PubMed]
- 258. Wojcikowski, J.; Daniel, W.A. The brain dopaminergic system as an important center regulating liver cytochrome P450 in the rat. *Expert Opin. Drug Metab. Toxicol.* 2009, *5*, 631–645. [CrossRef] [PubMed]
- Mantovani, A.; Petracca, G.; Beatrice, G.; Csermely, A.; Lonardo, A.; Targher, G. Glucagon-like Peptide-1 Receptor Agonists for Treatment of Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis: An Updated Meta-Analysis of Randomized Controlled Trials. *Metabolites* 2021, 11, 73. [CrossRef] [PubMed]
- Gupta, N.A.; Mells, J.; Dunham, R.M.; Grakoui, A.; Handy, J.; Saxena, N.K.; Anania, F.A. Glucagon-like peptide-1 receptor is present on human hepatocytes and has a direct role in decreasing hepatic steatosis in vitro by modulating elements of the insulin signaling pathway. *Hepatology* 2010, *51*, 1584–1592. [CrossRef]
- Jin, T.; Weng, J. Hepatic functions of GLP-1 and its based drugs: Current disputes and perspectives. Am. J. Physiol. Endocrinol. Metab. 2016, 311, E620–E627. [CrossRef]
- Pyke, C.; Heller, R.S.; Kirk, R.K.; Orskov, C.; Reedtz-Runge, S.; Kaastrup, P.; Hvelplund, A.; Bardram, L.; Calatayud, D.; Knudsen, L.B. GLP-1 receptor localization in monkey and human tissue: Novel distribution revealed with extensively validated monoclonal antibody. *Endocrinology* 2014, 155, 1280–1290. [CrossRef]
- Patel Chavez, C.; Cusi, K.; Kadiyala, S. The Emerging Role of Glucagon-like Peptide-1 Receptor Agonists for the Management of NAFLD. J. Clin. Endocrinol. Metab. 2022, 107, 29–38. [CrossRef]
- Scheen, A.J. Effects of glucose-lowering agents on surrogate endpoints and hard clinical renal outcomes in patients with type 2 diabetes. *Diabetes Metab.* 2019, 45, 110–121. [CrossRef]
- 265. Cohen, S.; Sternlicht, H.; Bakris, G.L. Mineralocorticoid Receptor Antagonists in the Treatment of Diabetic Kidney Disease: Their Application in the Era of SGLT2 Inhibitors and GLP-1 Receptor Agonists. *Curr. Diab. Rep.* 2022, 22, 213–218. [CrossRef]
- 266. Puglisi, S.; Rossini, A.; Poli, R.; Dughera, F.; Pia, A.; Terzolo, M.; Reimondo, G. Effects of SGLT2 Inhibitors and GLP-1 Receptor Agonists on Renin-Angiotensin-Aldosterone System. *Front. Endocrinol.* 2021, 12, 738848. [CrossRef]

- 267. Wright, J.W.; Harding, J.W. The brain renin-angiotensin system: A diversity of functions and implications for CNS diseases. *Pflug. Arch.* **2013**, 465, 133–151. [CrossRef] [PubMed]
- Perez-Lloret, S.; Otero-Losada, M.; Toblli, J.E.; Capani, F. Renin-angiotensin system as a potential target for new therapeutic approaches in Parkinson's disease. *Expert Opin. Investig. Drugs* 2017, 26, 1163–1173. [CrossRef] [PubMed]
- Fandino, J.; Vaz, A.A.; Toba, L.; Romani-Perez, M.; Gonzalez-Matias, L.; Mallo, F.; Diz-Chaves, Y. Liraglutide Enhances the Activity of the ACE-2/Ang(1-7)/Mas Receptor Pathway in Lungs of Male Pups from Food-Restricted Mothers and Prevents the Reduction of SP-A. Int. J. Endocrinol. 2018, 2018, 6920620. [CrossRef] [PubMed]
- Romani-Perez, M.; Outeirino-Iglesias, V.; Moya, C.M.; Santisteban, P.; Gonzalez-Matias, L.C.; Vigo, E.; Mallo, F. Activation of the GLP-1 Receptor by Liraglutide Increases ACE2 Expression, Reversing Right Ventricle Hypertrophy, and Improving the Production of SP-A and SP-B in the Lungs of Type 1 Diabetes Rats. *Endocrinology* 2015, *156*, 3559–3569. [CrossRef] [PubMed]
- 271. Evans, C.E.; Miners, J.S.; Piva, G.; Willis, C.L.; Heard, D.M.; Kidd, E.J.; Good, M.A.; Kehoe, P.G. ACE2 activation protects against cognitive decline and reduces amyloid pathology in the Tg2576 mouse model of Alzheimer's disease. *Acta Neuropathol.* 2020, 139, 485–502. [CrossRef]
- 272. Rabie, M.A.; Abd El Fattah, M.A.; Nassar, N.N.; El-Abhar, H.S.; Abdallah, D.M. Angiotensin 1-7 ameliorates 6-hydroxydopamine lesions in hemiparkinsonian rats through activation of MAS receptor/PI3K/Akt/BDNF pathway and inhibition of angiotensin II type-1 receptor/NF-kappaB axis. *Biochem. Pharm.* 2018, 151, 126–134. [CrossRef]
- Regenhardt, R.W.; Desland, F.; Mecca, A.P.; Pioquinto, D.J.; Afzal, A.; Mocco, J.; Sumners, C. Anti-inflammatory effects of angiotensin-(1-7) in ischemic stroke. *Neuropharmacology* 2013, 71, 154–163. [CrossRef]
- 274. Bairamian, D.; Sha, S.; Rolhion, N.; Sokol, H.; Dorothee, G.; Lemere, C.A.; Krantic, S. Microbiota in neuroinflammation and synaptic dysfunction: A focus on Alzheimer's disease. *Mol. Neurodegener.* **2022**, 17, 19. [CrossRef]
- 275. Romano, S.; Savva, G.M.; Bedarf, J.R.; Charles, I.G.; Hildebrand, F.; Narbad, A. Meta-analysis of the Parkinson's disease gut microbiome suggests alterations linked to intestinal inflammation. NPJ Parkinsons Dis. 2021, 7, 27. [CrossRef]
- Canfora, E.E.; Meex, R.C.R.; Venema, K.; Blaak, E.E. Gut microbial metabolites in obesity, NAFLD and T2DM. Nat. Rev. Endocrinol. 2019, 15, 261–273. [CrossRef]
- Musso, G.; Gambino, R.; Cassader, M. Obesity, diabetes, and gut microbiota: The hygiene hypothesis expanded? *Diabetes Care* 2010, 33, 2277–2284. [CrossRef] [PubMed]
- 278. Del Tredici, K.; Braak, H. Review: Sporadic Parkinson's disease: Development and distribution of alpha-synuclein pathology. *Neuropathol. Appl. Neurobiol.* 2016, 42, 33–50. [CrossRef]
- Jan, A.; Goncalves, N.P.; Vaegter, C.B.; Jensen, P.H.; Ferreira, N. The Prion-like Spreading of Alpha-Synuclein in Parkinson's Disease: Update on Models and Hypotheses. *Int. J. Mol. Sci.* 2021, 22, 8338. [CrossRef]
- Ferreira, N.; Goncalves, N.P.; Jan, A.; Jensen, N.M.; van der Laan, A.; Mohseni, S.; Vaegter, C.B.; Jensen, P.H. Trans-synaptic spreading of alpha-synuclein pathology through sensory afferents leads to sensory nerve degeneration and neuropathic pain. *Acta Neuropathol. Commun.* 2021, 9, 31. [CrossRef]
- 281. Ferreira, N.; Richner, M.; van der Laan, A.; Bergholdt Jul Christiansen, I.; Vaegter, C.B.; Nyengaard, J.R.; Halliday, G.M.; Weiss, J.; Giasson, B.I.; Mackenzie, I.R.; et al. Prodromal neuroinvasion of pathological alpha-synuclein in brainstem reticular nuclei and white matter lesions in a model of alpha-synucleinopathy. *Brain Commun.* 2021, 3, fcab104. [CrossRef]
- 282. Sacino, A.N.; Brooks, M.; Thomas, M.A.; McKinney, A.B.; Lee, S.; Regenhardt, R.W.; McGarvey, N.H.; Ayers, J.I.; Notterpek, L.; Borchelt, D.R.; et al. Intramuscular injection of alpha-synuclein induces CNS alpha-synuclein pathology and a rapid-onset motor phenotype in transgenic mice. *Proc. Natl. Acad. Sci. USA* 2014, 111, 10732–10737. [CrossRef]
- 283. Van Den Berge, N.; Ferreira, N.; Gram, H.; Mikkelsen, T.W.; Alstrup, A.K.O.; Casadei, N.; Tsung-Pin, P.; Riess, O.; Nyengaard, J.R.; Tamguney, G.; et al. Evidence for bidirectional and trans-synaptic parasympathetic and sympathetic propagation of alpha-synuclein in rats. *Acta Neuropathol.* 2019, 138, 535–550. [CrossRef] [PubMed]
- Kuthati, Y.; Rao, V.N.; Busa, P.; Wong, C.S. Teneligliptin Exerts Antinociceptive Effects in Rat Model of Partial Sciatic Nerve Transection Induced Neuropathic Pain. *Antioxidants* 2021, 10, 1438. [CrossRef]
- 285. Hillman, E.M. Coupling mechanism and significance of the BOLD signal: A status report. Annu. Rev. Neurosci. 2014, 37, 161–181. [CrossRef] [PubMed]