Expression of NF-κB and PTEN in osteosarcoma and its clinical significance

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Received May 29, 2017; Accepted September 8, 2017

DOI: 10.3892/ol.2017.6972

Abstract. We investigated the role of nuclear factor-kB $(NF-\kappa B)$ and phosphatase and tensin homolog deleted in chromosome 10 (PTEN) in the pathogenesis of osteosarcoma and its relationship with prognosis. Immunohistochemical method was used to detect the expression of NF- κ B and PTEN in osteosarcoma and adjacent tissues. RT-PCR was used to detect the expression of NF-kB and PTEN mRNA in osteosarcoma and adjacent tissues. Western blotting was used to detect the expression of NF-kB and PTEN in osteosarcoma and adjacent tissues and compare their differences. The expression of NF-kB and PTEN was detected in osteosarcoma and adjacent tissues. The positive rate of NF-KB was 75.3 and 32.9%, respectively; while the positive rate of PTEN was 67.1 and 90.4%, respectively. The positive expression of NF-KB and PTEN was statistically significant. There was a negative correlation between NF-kB and PTEN expression (r=-0.502, p<0.05). The positive and negative expression of NF-KB and PTEN was statistically significant for the five-year survival (p<0.05). At gene and protein level, osteosarcoma tissues had higher expression of NF-kB, and lower expression of PTEN, which was significantly different from the adjacent tissues. In osteosarcoma, NF-KB is highly expressed, but PTEN is expressed at low level, and the two are negatively correlated. This is of great significance for the early diagnosis of osteosarcoma and prognosis.

Introduction

As one of the more common malignant bone tumor, osteosarcoma has high degree of malignancy, and relatively poor prognosis (1). The pathogenesis and development process has not yet been studied fully, and the current clinical diagnosis of osteosarcoma lacks more specific indicators (2). PTEN as a tumor suppressor gene, is associated with the occurrence and development of a variety of malignant tumors (3,4). Overexpression of NF-κB is associated physiologically and pathologically with many tumors, and is a bi-directional regulatory factor (5-7). There are few studies on the expression and correlation of PTEN and NF-κB in osteosarcoma to date. The purpose of this study is to use immunohistochemical method to detect the expression of PTEN and NF-κB in tumor tissues, explore their expression changes at genes and protein levels, and analyze statistically the expression of both and the prognosis of patients, to explore their expression correlation in osteosarcoma and the potential use in clinical diagnosis of osteosarcoma.

Materials and methods

Materials

Experimental materials. Pathologically confirmed osteosarcoma tumor tissues (73 cases) and adjacent tissues (73 cases) were selected. All patients signed an informed consent form. The osteosarcoma tissue and adjacent tissues were treated with paraffin embedding, and then paraffin sectioned with a thickness of ~4 μ m. The patients with osteosarcoma were follow-up for 5-years.

Reagents. Rabbit antihuman nuclear factor-kB (NF-kB) monoclonal antibody (Beijing Dingguo Changsheng Biotechnology Co., Ltd., Beijing, China) rabbit anti-human phosphatase and tensin homolog deleted in chromosome 10 (PTEN) polyclonal antibody, rabbit anti-human glyceraldehyde 3-phosphate dehydrogenase (GAPDH) polyclonal antibody (cat. no. SPC-1303, SPC-1331 and SPC-689; StressMarq Biosciences Inc., Victoria, BC, Canada), DAB coloring reagent (Shanghai Runwell Technology Co., Ltd., Shanghai, China), citrate buffer powder (Shanghai X-Y Biotechnology Co., Ltd., Shanghai, China), reverse transcription kit (GeneCopoeia, Rockville, MD, USA), real-time fluorescence quantitative PCR and Western Blot test kit (Shanghai Biological BestBio Bebo, Shanghai, China), goat anti-rabbit IgG secondary polyclonal antibody (cat. no. ab150077; Abcam, Cambridge, UK), BCA protein quantitative kit (Nanjing SenBeiJia Biotechnology Co., Ltd., Nanjing, China).

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Key words: osteosarcoma, nuclear factor- κ B, phosphatase and tensin homolog deleted in chromosome 10

Experimental methods

Immunohistochemical staining. The paraffin sections were subjected to dewaxing and then washed with phosphate-buffered saline. To reduce the nonspecific background staining caused by endogenous peroxidase, blocking was continued for 20 min in the blocking buffer, then blocked in 10% serum for 10 min at room temperature, and incubated overnight at 4°C with the addition of primary antibody dilution buffer (rabbit anti-human NF-KB monoclonal antibody and PTEN polyclonal antibody, 1:50 dilution). It was washed with phosphate buffer solution, adding the goat anti-rabbit IgG secondary polyclonal antibody dilution buffer (1:50 dilution) and incubated at room temperature for 30 min. It was washed again with phosphate buffer, and then incubated with Streptavidin antibiotic proteinperoxidase solution at room temperature for 30 min, washed with phosphate buffer solution, colored with DAB, washed with distilled water, stained and sealed.

Evaluation of results. One hundred cells were randomly selected to observe the visual field under a light microscope, and the average number of cells in the field of view was obtained as the positive cells expressing the protein in the tissue. PTEN staining was mainly located in the cytoplasm, while the positive expression of NF-κB was mainly located in the nucleus. Depth score: scores 0 to 2, was assigned for no coloring, weak coloring and strong coloring, respectively. Cells stained positive score: 1 to 25% was recorded as 1, 26 to 50% as 2, 51 to 75% as 3, 76 to 100% as 4. The scores of the depth score and the positive rate were multiplied, 1 to 2 were negative, and 3 to 8 were positive.

Real-time PCR method to determine the expression of NF- κB and PTEN mRNA. Total RNA was extracted from both tissues and the concentration and purity were determined. Table I shows the primer sequence used in the synthesis of NF- κB and PTEN gene amplification by Nanjing CoBioer Biotechnology Co., Ltd. (Nanjing, China). Reverse transcription synthesis of cDNA, reaction system 20 μ l.

PCR reaction system: 25 μ l, reaction conditions: 95°C 10 min, 95°C 30 sec, and 59.4°C 30 sec, 40 cycles, 95°C 15 sec, and maintained at 65°C. With GAPDH as the internal control, RT-PCR instrument automatically calculated the NF- κ B and PTEN mRNA relative expression.

Western blot assay for detection of protein. The total protein in the two tissues was extracted according to the instructions in the tissue total protein extraction kit. The concentration of the extracted protein was determined by BCA protein assay, and stored at -70°C. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) was performed on 10% separation gel and 5% concentrated gel. The gel position of the two proteins was selected according to the marker strip. After transmembrane, the PVDF membrane was washed with 1X TBST solution for 5 min, 5% nonfat dry milk blocking at room temperature (1:1,000 dilution) for 1 h, 1X TBST solution for 1 h, 1X TBST solution for 1 h, 1X TBST solution 5 min, total cleaning 3 times. Anti-dark drop ECL luminescent liquid, dark environment for 2 min for development. Finally, the Multi Gauge Ver. 3.0 imaging system was scanned, and ImageJ professional image analysis software (National Institutes of Health, Bethesda, MD, USA) Table I. NF- κ B and PTEN gene amplification primer.

Gene	Primer sequence
NF-ĸB	F: 5'-TACCCTGAGGCTATAACTC-3' R: 5'-GACACTTGATAAGGCTTTG-3'
PTEN	F: 5'-AGTTCCCTCAGCCGTTACCT-3' R: 5'-ATTTGACGGCTCCTCTACTG-3'
GAPDH	F: 5'-TGGGTGTGAACCACGAGAA-3' R: 5'-GGCATGGACTGTGGTCATGA-3'

NF- κ B, nuclear factor- κ B; PTEN, phosphatase and tensin homolog deleted in chromosome 10; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; F, forward; R, reverse.

Table II. PTEN expression comparison in both tissues.

Arm P-value	No. of cases	PT.	EN 	Positive rate %	χ	2
Osteosarcoma Adjacent tissues	73 73	49 66	24 7	67.1 90.4	31.02	<0.05

PTEN, phosphatase and tensin homolog deleted in chromosome 10.

was used for image analysis and the absorbance value was recorded.

Statistical analysis. The statistical software SPSS 17.0 (Beijing Xinmei Jiahong Technology Co., Ltd., Beijing, China) was used in this study for data analysis. The correlation between the expression of NF- κ B and PTEN was tested by Spearman's method, using χ^2 to test the difference of positive rates between arms. The survival analysis was analyzed by GraphPad Prism5 software (GraphPad Software Inc., La Jolla, CA, USA). The difference was considered statistically significant when p<0.05.

Results

The expression of PTEN and NF- κ B in osteosarcoma tissues and adjacent tissues. The immunohistochemical staining showed that PTEN had more positive expression granules in osteosarcoma tissues, while in adjacent tissues, PTEN showed diffuse granule-like expression with expression level significantly higher than that of osteosarcoma tissues. The notable difference of the positive expression rate of PTEN in osteosarcoma tissues and adjacent tissues was statistically significant (p<0.05) (Fig. 1).

Among 73 cases of osteosarcoma tissues, 49 cases were positive in PTEN expression, with positive rate of 67.1%; while in adjacent tissues, 66 cases were positive in PTEN expression, with positive rate of 90.4% (Table II).



Figure 1. Immunohistochemical expression of phosphatase and tensin homolog deleted in chromosome 10 (PTEN) in osteosarcoma tissues and adjacent tissues. Immunohistochemical staining of the two types of tissues. The expression of PTEN in the adjacent tissues was higher than that in the osteosarcoma tissues. (A) Osteosarcoma tissues, (B) adjacent tissues.



Figure 2. Immunohistochemical expression of nuclear factor- κB (NF- κB) in osteosarcoma tissues and adjacent tissues. Immunohistochemical staining of the two tissues. The NF- κB in osteosarcoma tissue had significantly positive expression of granules, while the presence of NF- κB in the adjacent tissue was not obvious. (A) Osteosarcoma tissues, (B) adjacent tissues.

		NF	-кВ			
	No. of			Positive		
Arm	cases	+	-	rate%	χ^2	P-value
Osteosarcoma	73	55	18	75.3	40.57	<0.01
Adjacent tissues	73	24	49	32.9		
NF-κB, nuclear fac	tor-κB.					

Table III. Comparison of NF-κB expression in both tissues.

Immunohistochemical staining showed that NF- κ B had a significant positive expression of granulocyte in osteosarcoma tissues, while NF- κ B expression was not obvious in adjacent tissues. The positive expression of NF- κ B in osteosarcoma tissues was statistically significantly higher than that in adjacent tissues (p<0.05) (Fig. 2).

Out of 73 osteosarcoma tissues, 53 were positive in NF- κ B expression. The positive rate was 75.3%; while 24 adjacent tissues were positive in NF- κ B expression. The positive rate was 32.9% (Table III).

Correlative analysis of PTEN and NF- κ B expression in osteosarcoma. Out of 73 osteosarcoma tissues, 53 were positive in NF- κ B expression. Among them, 16 cases were also positive

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	PT	EN			
NF-κB	+	-	Total	r-value	P-value
+	16	39	55	-0.502	< 0.05
-	14	4	18		
Total	30	43	73		

NF- κ B, nuclear factor- κ B; PTEN, phosphatase and tensin homolog deleted in chromosome 10.

for PTEN, at a rate of 29.1%. The statistical analysis shows a negative correlation between NF- κ B and PTEN (r=-0.502, p<0.05) (Table IV and Fig. 3).

Detecting NF- κ B and PTEN mRNA expression by RT-PCR. The relative expression of NF- κ B mRNA in osteosarcoma tissues (52.9±4.17) was significantly higher than that in adjacent tissues (39.2±5.94), the difference was statistically significant (p<0.05); and the relative expression of PTEN mRNA (59.3±3.19) was significantly lower than that of adjacent tissues (91.4±5.82), and the difference was statistically significant (p<0.05) (Table V and Figs. 4 and 5).

Table V. The relative expression of NF-κB and PTEN mRNA.

Arm	NF-κB	PTEN
Osteosarcoma	52.9±4.17 ^a	59.3±3.19°
Adjacent tissues	39.2±5.94 ^b	91.4±5.82 ^d
P-values	< 0.05	

Comparison of a and b, p<0.05; comparison of c and d, p<0.05. PTEN, phosphatase and tensin homolog deleted in chromosome 10; NF- κ B, nuclear factor- κ B.

Table VI. Expression of NF-κB and PTEN protein.

Arm	NF-ĸB	PTEN	
Osteosarcoma	0.872±0.015ª	1.383±0.047°	
Adjacent tissues	0.403 ± 0.008^{b}	1.902 ± 0.064^{d}	
P-values	< 0.05		

Comparison of a and b, p<0.05; comparison of c and d, p<0.05. PTEN, phosphatase and tensin homolog deleted in chromosome 10; NF- κ B, nuclear factor- κ B.



Figure 4. Expression of nuclear factor- κB (NF- κB) and phosphatase and tensin homolog deleted in chromosome 10 (PTEN) protein. *p<0.05; *p<0.05.



Figure 5. Western blot detection of nuclear factor- κB (NF- κB) and phosphatase and tensin homolog deleted in chromosome 10 (PTEN). In osteosarcoma tissues, NF- κB expression is higher, PTEN expression is lower.



Figure 6. The survival curves of patients with positive and negative expression of nuclear factor- κ B (NF- κ B) and phosphatase and tensin homolog deleted in chromosome 10 (PTEN). The 5-year survival rate was higher for osteosarcoma patients with negative expression of NF- κ B and positive expression of PTEN.

NF-κB positive expression was lower than that of negative expression (χ^2 =8.014, p<0.05) (Fig. 6A). However, the 5-year survival of patients with PTEN positive expression was signifi-



Figure 3. Relative expression of nuclear factor- κB (NF- κB) and phosphatase and tensin homolog deleted in chromosome 10 (PTEN) mRNA. *p<0.05; *p<0.05.

Detecting NF- κ B and PTEN mRNA expression by western blot method. The western blot test indicates the expression of NF- κ B protein in osteosarcoma tissues was higher (0.872±0.015), and the expression of κ B in adjacent tissues was lower (0.403±0.008). The expression of PTEN protein was lower in osteosarcoma tissues (1.383±0.047), and higher in adjacent tissues (1.902±0.064). The difference in expression between the two proteins was statistically significant (p<0.05) (Table VI).

Prognosis of NF-κB and PTEN expression. The survival rate of patients with NF-κB and PTEN positive and negative expression was statistically analyzed. Survival analysis using statistical software showed, the survival rate of those with

cantly higher than that of negative expression. The result was statistically significant (χ^2 =7.625, p<0.05) (Fig. 6B).

Discussion

Osteosarcoma is a type of malignant tumor occurring more frequently in adolescents. Osteosarcoma is developed from the interstitial cell line, with strong infiltration and transfer capacity (8-10) PTEN is a tumor suppressor gene with phosphatase activity (11) and it has been reported that PTEN expression is reduced in a variety of human malignancies, such as breast cancer (12) and prostate cancer (13). PTEN has many important functions in the body, such as the inhibition of cell growth and differentiation (14) and promotion of cell apoptosis (15), making it one of the most noticeable tumor suppressor genes after the p53 gene. NF-kB is a transcription factor with a two-way transcriptional regulation that plays an important role in many pathophysiological processes (16-18). Loftus et al (19) reported a significant increase in NF-KB p65 expression in hepatocellular carcinoma. The study by Lu *et al* (20) showed that the expression of NF- κ B protein in osteosarcoma negative correlates with the apoptotic index of osteosarcoma, and significantly inhibited the apoptosis of osteosarcoma cells. However, reports on the correlation of NF-KB expression and PTEN in osteosarcoma are scarce.

Immunohistochemical method was used in this study. the study results indicated that in osteosarcoma tissues, the positive expression rate of PTEN was significantly lower than that in the adjacent tissues, and the positive expression rate of NF-κB was significantly higher. The difference between the two are statistically significant (r=-0.502, p<0.05). Therefore, we hypothesized that the decrease of PTEN elevated the expression of NF-kB through certain mechanism; or the increased expression of NF-kB leads to decreased expression of PTEN through certain mechanism. In the osteosarcoma tissues and the adjacent tissues, the mRNA expression of NF-κB was 52.9 ± 4.17 and 39.2 ± 5.94 , respectively; the mRNA expression of PTEN was 59.3±3.19 and 91.4±5.82, respectively. The difference in expression of NF-kB and PTEN mRNA was statistically significant (p<0.05). The expression of NF-kB protein in osteosarcoma tissues was higher than that in adjacent tissues, and the expression of PTEN protein in osteosarcoma tissues was lower. The experimental results at both gene and protein levels have proved the NF-KB and PTEN expression was negatively correlated, suggesting these two molecules can act as potential biomarkers for the clinical detection of osteosarcoma.

The patients with osteosarcoma were follow-up for fiveyears. The results showed that the 5-year survival rate was higher in patients with NF- κ B negative expression and those with PTEN positive expression. The difference was statistically significant. The result indicates that combining with detection of NF- κ B and PTEN expression can better assess clinically the prognosis of patients with osteosarcoma, which provides a new direction for the clinical treatment of osteosarcoma.

Acknowledgements

This study was supported by the Natural Science Program of Science and Technology commission of Tianjin (no. 043609011) and the Launching fund project of Youth Doctor from Logistics College of Chinese People's Armed Police Forces (no. WYB201109).

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