Autophagy-assisted antigen cross-presentation

Autophagosome as the argo of shared tumor-specific antigens and DAMPs

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It is generally believed that most tumor antigens are passively released from either health or dying tumor cells as intact soluble antigens, peptide fragments complexed with heat shock proteins (HSPs), or packaged in secretary vesicles in the form of microparticles or exosomes. The passive release of tumor antigens is generally non-inflammatory and non-immunogenic; however, results from others and our laboratories suggest that autophagy is critically involved in immunogenic cell death.

Autophagy is a process of packaging misfolded proteins and damaged organelles in the autophagosomes and degrading them through fusion with lysosomes. During the early development of cancer, autophagy can serve as a tumor suppression mechanism, since it clears abnormal proteins and limits genome damage. Autophagy becomes a mechanism of tumor promotion at later stages, because it helps tumor cells survive during physiological stress of metastases and stress induced by chemotherapy or irradiation.¹

Notably, autophagy in cancer cells also plays a critical role by providing immunogenic tumor antigens and eliciting immune responses required for tumor cell destruction.^{2,3} We have recently identified macroautophagy as one of these critical pathways that regulates antigen delivery, and shown that tumor macroautophagy regulates the efficiency of cross presentation.2 Tumor cells that undergo autophagy release autophagosomes, which are important tumor antigen sources for cross-priming of tumor-specific CD8+ T cells, into the culture media.³ By inducing autophagy and blocking protein degradation through proteasome and lysosome inhibition,

we demonstrated that large amounts of secreted autophagosomes contain abundant ubiquitinated antigens. Using OVA and gp100 as model antigens, we showed that the tumor antigens sequestered in autophagosomes were efficiently cross-presented to naïve transgenic OT-I and pmel-1 CD8+ T cells respectively in vitro and in vivo. Compared with soluble antigens, antigens packaged in the autophagosomes were superior in activating CD8⁺ T cells. We then investigated the mechanisms by which antigens in autophagosomes were cross-presented by DCs. CLEC9A, a novel C-type selectin receptor on a subset of DCs, plays an important role in cross-presentation of dead cell-associated antigens. Interestingly, we detected high expression of CLEC9A ligand on autophagosomes. Blockade of the interaction between the CLEC9A ligand on autophagosomes and CLEC9A on DCs significantly reduced antigen cross-presentation, but did not affect uptake of autophagosomes by DCs. CLEC9A is likely involved in the intracellular process rather than phagocytosis of autophagosomes. Moreover, the size of autophagosomes (200-900 nm) may obligate them to non-acidic intracellular

compartments that favor MHC I presentation on DCs. Utilizing specific inhibitors of distinct types of endocytosis, we showed that cross-presentation of autophagosome derived antigens depended predominantly on the caveolae-mediated endocytosis pathway, which routes antigens to nonacidic compartments. We further demonstrated that tumor antigens present in the autophagosome must exit through the ERAD translocation machinery after which they are degraded by pAPC proteosomes. Thus, the autophagy-assisted antigen cross presentation pathway also requires the phagosome-ER-cytosol route of cross-presentation.4

These unique characteristics of tumor-derived autophagosomes make them an ideal vaccine candidate for cancer immunotherapy. Using the 3LL Lewis lung cancer and B16F10 melanoma models, we showed that tumor-derived autophagosomes loaded onto DCs combined with poly I:C as the adjuvant significantly delayed tumor growth or eradicated tumors in C57BL/6 mice bearing established 3LL lung tumors. We also found potent therapeutic activity in mice with orthotopically transplanted primary

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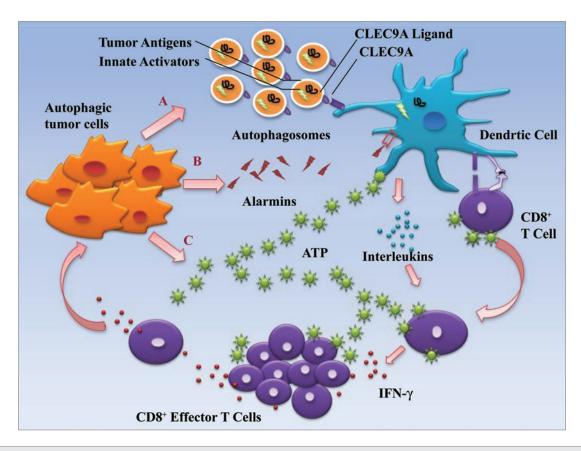


Figure 1. Autophagy-assisted antigen cross presentation and immunogenic cell death. (**A**) Tumor cells dying with autophagy produce autophagosomes that contain a variety of tumor antigens, activators of the innate immune system, and molecules that target autophagosomes to DCs. (**B**) Dying tumor cells release HMGB1 and other alarmins that mobilize and activate DCs. (**C**) Tumor cells dying with autophagy produce ATP and other molecules that directly or indirectly acting on T cells to enhance their activation, proliferation and effector functions.

and metastatic 4T1 mammary carcinomas following immunization with either autologous or allogenic tumor-derived autophagosomes (manuscript is in preparation). The tumor-derived autophagosomes could selectively recruit ubiquitinated proteins via the p62-dependent pathways and we hypothesized many of these ubiquitinated proteins are derived from shared but tumor-specific transcriptome.⁵

Autophagosomes not only sequester tumor antigens for efficient initiation of adaptive immune responses, but also package DAMP molecules that activate innate immune response and enhance DC or T-cell function. In addition to the CLEC9A ligand, other potential DAMP molecules include cellular DNA and RNA, possibly in complexes with bactericidal peptide LL-37 (cathelicidin), HSPs and calreticulin.^{6,7} Oppenheim first coined the term alarmins to describe one

subset of these endogenous innate activators that are rapidly secreted from innate immune cells, particularly neutrophils, or released by tissue damage and necrotic cells.8 They include bactericidal peptides (defensin and cathelicidin), inflammatory cytokines (IL-1α and IL-33), high mobility group box proteins (HMGB1, HMGN1), and cytosolic calcium-binding proteins of the S100 family. Multiple HSPs, High mobility group box 1 protein (HMGBs) and calreticulin are present in autophagosomes. As a bag full of immunological tricks, the tumor-derived autophagosomes are likely to continue to surprise us. Beside these antigens and ability to target CLEC9A cross-presenting dendritic cells, autophagosome vaccines contain many known, and perhaps some unknown, adjuvant components that activate the innate and augment the adaptive immune responses respectively (Fig. 1).

Many chemotherapy drugs and irradiation induce autophagy while killing the tumor cells. Our findings indicate that autophagosomes play an important role as antigen carriers, which suggests that cells dying with autophagy may release autophagosome antigens and inflammation signals into the local environments or body fluids and elicit a strong CD8+ T-cell immune responses. Interestingly, Kroemer's group recently discovered that autophagy provides energy required for priming of anti-tumor T-cell immunity by the release of ATG from dying cells.9 Thus, autophagy in dying tumor cells plays a critical role in induction of anti-tumor immune response through multiple mechanisms.10 Our group is exploiting these mechanisms and evaluating the effectiveness of autophagosomes from cancer cells as vaccines for patients with cancer.

References

- Morselli E, Galluzzi L, Kepp O, Vicencio JM, Criollo A, Maiuri MC, et al. Anti- and pro-tumor functions of autophagy. Biochim Biophys Acta 2009; 1793:1524-32; PMID:19371598; http://dx.doi.org/10.1016/j. bbamcr.2009.01.006.
- Li Y, Wang LX, Yang G, Hao F, Urba WJ, Hu HM. Efficient cross-presentation depends on autophagy in tumor cells. Cancer Res 2008; 68:6889-95; PMID:18757401; http://dx.doi.org/10.1158/0008-5472.CAN-08-0161.
- Li Y, Wang LX, Pang P, Cui Z, Aung S, Haley D, et al. Tumor-derived autophagosome vaccine: mechanism of cross-presentation and therapeutic efficacy. Clin Cancer Res 2011; 17:7047-57; PMID:22068657; http://dx.doi.org/10.1158/1078-0432.CCR-11-0951.
- Kurts C, Robinson BW, Knolle PA. Cross-priming in health and disease. Nat Rev Immunol 2010; 10:403-14; PMID:20498667; http://dx.doi.org/10.1038/ nri2780.

- Twitty CG, Jensen SM, Hu HM, Fox BA. Tumorderived autophagosome vaccine: induction of crossprotective immune responses against short-lived proteins through a p62-dependent mechanism. Clin Cancer Res 2011; 17:6467-81; PMID:21810919; http://dx.doi.org/10.1158/1078-0432.CCR-11-0812.
- Ganguly D, Chamilos G, Lande R, Gregorio J, Meller S, Facchinetti V, et al. Self-RNA-antimicrobial peptide complexes activate human dendritic cells through TLR7 and TLR8. J Exp Med 2009; 206:1983-94; PMID:19703986; http://dx.doi.org/10.1084/ jem.20090480.
- Lande R, Gregorio J, Facchinetti V, Chatterjee B, Wang YH, Homey B, et al. Plasmacytoid dendritic cells sense self-DNA coupled with antimicrobial peptide. Nature 2007; 449:564-9; PMID:17873860; http://dx.doi. org/10.1038/nature06116.
- Oppenheim JJ, Yang D. Alarmins: chemotactic activators of immune responses. Curr Opin Immunol 2005; 17:359-65; PMID:15955682; http://dx.doi.org/10.1016/j.coi.2005.06.002.
- Michaud M, Martins I, Sukkurwala AQ, Adjemian S, Ma Y, Pellegatti P, et al. Autophagy-dependent anticancer immune responses induced by chemotherapeutic agents in mice. Science 2011; 334:1573-7; PMID:22174255; http://dx.doi.org/10.1126/science.1208347.
- Galluzzi L, Senovilla L, Zitvogel L, Kroemer G. The secretally: immunostimulation by anticancer drugs. Nat Rev Drug Discov 2012; 11:215-33; PMID:22301798; http://dx.doi.org/10.1038/nrd3626.