The diabetes-induced functional and distributional changes of the alpha 1-adrenoceptor of the abdominal aorta and distal mesenteric artery from streptozotocin-induced diabetic rats

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Background: The aim of this study was to evaluate the effect of diabetes on the function and distribution of vascular α_1 -adrenoceptors in the abdominal aorta and distal mesenteric artery from streptozotocin (STZ)-induced diabetic rats at the level of the α_1 -adrenoceptor subtypes.

Methods: Diabetes was induced by a single intravenous injection of STZ (60 mg/kg) in 8 week-old male Sprague-Dawley rats (n = 11). Age-matched normal rats (n = 14) were used as a control group. Four weeks after STZ injection, the tilting-induced change of the mean arterial pressure was recorded. The α_1 -adrenoceptor subtypes mediating the contractions of the distal mesenteric artery and abdominal aorta were investigated using the agonist phenylephrine and subtype-selective antagonists that included prazocin, 5-methylurapidil and BMY 7378. The expressions of the α_1 -adrenoceptor subtypes of each artery were examined by immunofluorescence staining using the subtype selective antabodies.

Results: The recovery of the mean arterial pressure was delayed after positional change in the diabetic rats. Compared with that of the normal rats, the contractile response to phenylephrine was increased in the abdominal aortas and it was decreased in the distal mesenteric arteries in the diabetic rats. In addition, compared with the normal rats, the fluorescent intensity of all the α_1 -adrenoceptor subtypes was increased in the abdominal aortas and it was decreased in the mesenteric arteries of the diabetic rats.

Conclusions: Diabetes increased the contractility of the abdominal aorta in response to phenylephrine, yet diabetes decreased that of the mesenteric arteries in the STZ-induced diabetic rats. Those results are mainly based on the overall change of the α_1 -adrenoceptor, and not on the change of the specific α_1 -adrenoceptor subtypes. (Korean J Anesthesiol 2011; 60: 272-281)

Key Words: Abdominal aorta, Alpha-1 adrenergic receptor, Diabetic complications, Experimental diabetes mellitus, Mesenteric artery, Streptozotocin.

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Introduction

Diabetes affects approximately 100 million persons worldwide [1], and it damages the cadiovascular autonomic nerve fibers mainly because of the sympathetic denervation of the heart and the blood vessels (diabetic cardiovascular autonomic neuropathy) [2,3]. This change usually leads to arterial stiffness [4], but it leads to vasodilation in the peripheral arteries [5]. This vascular denervation also changes the blood flow responsiveness to various stimuli. For example, the peripheral vasoconstriction in response to sympathetic stimuli such as standing up or coughing is reduced [2]. Moreover, a previous study has shown that vasopressor support was needed more often during anesthesia in diabetic patients with autonomic dysfunction that in diabetic patients without autonomic dysfunction during anesthesia because the normal autonomic response of vasoconstriction and tachycardia did not completely compensate for the vasodilating effects of anesthesia [6]. Therefore, it has been suggested that preoperative cardiovascular autonomic screening of diabetic patients may be very useful for anesthesiologists [3].

The α_1 -adrenoceptor plays an important role in the vascular sympathetic system to maintain the peripheral vascular resistance and the systemic arterial pressure [7-10]. Moreover, it is generally accepted that there are 3 functional α_1 -adrenoceptor subtypes called α_{1A} , α_{1B} , and α_{1D} and that the distribution of those α_1 -adrenoceptor subtypes is different according to the types of arteries [11,12]. Considering these findings, the diabetes-induced functional and distributional changes of the vascular α_1 -adrenoceptor in different arteries would play a role in the pathogenesis of diabetic vascular autonomic disturbance.

Yet to the best of our knowledge, the previous studies did not show consistent results about adrenergically mediated vasoconstriction in diabetic rats [13-15]. Moreover, there has been no study that has focused on the diabetes-induced distributional change of the α_1 -adrenoceptor or the α_1 -adrenoceptor subtypes according to the different types of arteries.

Therefore, we evaluated the effect of diabetes on the function and distribution of the vascular α_1 -adrenoceptor, at the level of α_1 -adrenoceptor subtypes, using the abdominal aorta and distal mesenteric artery in streptozotocin (STZ)-induced diabetic rats.

Materials and Methods

Animals

Eight-week-old male rats (n = 25) were used for this study. All the rats were housed at $22 \pm 2^{\circ}$ C with 40-60% humidity in a 12/12-h light/dark cycle, and they were maintained on a diet of animal chow and water ad libitum. All the animal studies were carried out in a semi-pathogen-free barrier zone at the Institute for Laboratory Animal Research in Seoul National University Hospital in accordance with the procedure outlined in the Guide for the Care and Use of Laboratory Animals.

Induction of diabetes mellitus

The rats were randomly allocated into either the control group (n = 14) or the diabetic group (n = 11). Diabetes was induced by intravenous administration of a single dose of STZ (60 mg/kg in citrate buffer, pH 4.5) through the tail vein. In the normal group, the same volume of citrate buffer was injected intravenously. Three days after STZ-injection, the blood glucose levels were measured from a drop of blood using a OneTouch Ultra[®] Blood Glucose Meter (LifeScan, Inc) for determining the successful induction of diabetes. Rats with a blood glucose level > 300 mg/dl were considered diabetic. All the rats were maintained for 4 weeks before intervention.

Changes in the arterial blood pressure related to tilting

The rats' blood glucose levels and body weight were measured four weeks after STZ or citrate buffer injection. For each rat, anesthesia was induced with sevoflurane with the rat in a chamber, and the anesthesia was maintained with 2 vol% sevoflurane. Following anesthesia, the rat was placed in a supine position and the four legs and upper front teeth were firmly fixed on a tilt board. The tail artery was cannulated with a 24 G catheter for continuous arterial pressure monitoring. To prevent the effect of postural change on measuring arterial pressure, the transducer was positioned at the level of the heart. After measuring the mean arterial pressure at the supine position (basal MAP), the table was vertically tilted at 45° angle in the head-up direction and the MAPs at 10, 30, 60 and 120 seconds after tilting were recorded. The changes in the MAP related to tilting were compared at each time point between the diabetic and control groups.

Preparation of vascular tissues

After the tilting experiment, the rat was euthanized by cervical dislocation under the deep anesthesia and the third branch of the mesenteric artery and the abdominal aorta were rapidly removed. The arteries were placed on a Petri dish with ice-cold modified Krebs' solution of the following composition (in mM): NaCl: 119, KCl: 4.7, MgSO₄: 1.17, NaHCO₃: 22, CaCl₂: 1.6, HEPES: 8, KH₂PO₄: 1.18, d-glucose: 5, and the arteries were cleaned of fat and connective tissues under a microscope (Zeiss). In addition, the Petri dish was kept cold by surrounding it with crushed ice. Following the cleaning procedure, to exclude the



effect of endothelium-derived relaxing factor, the endothelium of the arteries was removed by gently rubbing the lumen with coarse silk and each artery was cut into 3-mm ring segments for the in vitro contractile studies.

Measurement of isomeric tension

Each ring segment was suspended between two triangleshape tungsten hooks, and this was mounted in a 5 ml waterjacketed temperature-controlled tissue bath (Harvard Apparatus, Holliston, MA, USA) that contained modified Krebs' solution. The lower and upper hooks were attached, respectively, to the bottom of the bath chamber and to a force displacement transducer (NP 72-4493, Harvard Apparatus, Holliston, MA, USA). The tissue bathing solution was gassed with a mixture of 95% O2 and 5% CO2 (pH: 7.38-7.42, pCO2: 34-36 mmHg) and maintained at 37°C. Propranolol $(1 \times 10^{-6} \text{ M})$ and yohimbine $(1 \times 10^{-7} \text{ M})$ were always included in the tissue bathing solution to block the β - and α_2 -adrenoceptors, respectively. The isometric contractions were recorded using an A/D converter (Powerlab[®], ADInstruments, Colorado Springs, CO, USA) and they were analyzed by using Chart for Window Software[®] (ADInstruments, Colorado Springs, CO, USA).

Following applying an initial tension of 2.0 g, which provided the optimum concentration-response relationships to KCl 60 mM, the arterial strips were allowed to equilibrate for 60 min. When the resting tension reached a stable baseline, no attempt was made to adjust the tissue tension. After the equilibrium, tissues were primed by the addition of phenylephrine $(1 \times 10^{-6} \text{ M})$ to the organ bath. After a steady-state contraction was achieved, the bath contents were replaced with drug-free buffer several times. The tissues were then allowed to reach baseline tension, and the priming procedure was repeated once more before constructing the concentration-response curves. This priming procedure reduces tissue variability in response to repeated agonist administration. In addition, the absence of functional endothelium was also confirmed by the loss of the relaxation response to acethylcholine $(1 \times 10^{-6} \text{ M})$ in the pre-contracted rings with a priming dose of phenylephrine $(1 \times 10^{-6} \text{ M})$ during the priming procedure.

Construction of the concentration-response curves

Reproducible cumulative concentration-response curves were constructed by exposing each artery to increasing concentrations (0.5 log unit increments) of phenlyephrine $(1 \times 10^{-9} \text{ to } 1 \times 10^{-4})$ until a sustained maximum response (Emax) was observed. The concentration-response curves to phenylephrine were then obtained (60 min apart each) in the presence of four selective α_1 -adrenoceptor antagonists to each



artery and the procedure was repeated with a high concentration of antagonists in the same preparation. The incubation time for the selective α_1 -adrenoceptor antagonists prazosin (α_1 ; 1, 3 and 10×10^{-9} M), 5-methylurapidil (5-MU; α_{1A} ; 3, 10 and 30×10^{-8} M), and BMY 7378 (α_{1D} ; 3, 10 and 30×10^{-8} M) was 30 min. In all the experiments, one preparation was run in parallel with the experimental tissues, but it received no antagonist, and it was used to determine any time-dependent changes in agonist sensitivity. Propranolol (1×10^{-6} M) and yohimbine (1×10^{-7} M) were always included in the tissue bathing solution to block the β - and α_2 -adrenoceptors, respectively.

Immunofluorescence staining

After removing the vessels and cleaning the extraneous tissues, the arteries were cut into 2-mm rings, embedded in OCT compound (Tissue-Tek[®] OCT[™] Compound, Sakura Finetek) and quickly frozen $(-50^{\circ}C \text{ in isopentane cooled with})$ dry ice). The frozen blocks were cut into 5-µm sections and mounted on the slide. All the sections were stored at -80° C until use. After warming at room temperature for 30 minutes, the tissue sections were fixed in iced cold acetone for 5 minutes. Following rinsing the sections in PBS, a blocking solution with 10% normal donkey serum was placed on the sections and this was incubated for 30 minutes to block the non-specific binding sites of immunoglobulin. After removal of the blocking solution, the tissue section was incubated with the 1:100-diluted primary antibody (α_{1D} -AR cat. no. s.c.-1475, α_{1B} -AR cat. no. s.c.-1476, α_{1A} -AR cat. no. s.c.-1477; Santa Cruz Biotechnologies) for 60 minutes at room temperature. Following washing the primary antibody in PBS, fluoresceinisothiocyanate (FITC)conjugated donkey anti-goat IgG (1:100 dilution, no. s.c.-2024; Santa Cruz Biotechnology, Santa Cruz, CA, USA) secondary antibody was placed on the sections for 60 minutes in the dark at room temperature. After removing the secondary antibody by washing with PBS, mounting media was placed and a glass coverslip was applied. For reducing the technical errors, the staining was performed in one slide for same type of artery from two different groups. Three independent investigators observed the expression of the α_1 -adrenoceptor subtypes of each artery with using a fluorescence microscope (Olympus VENOX-AHBT3, Tokyo, Japan).

Statistical analysis

The data is presented as means \pm SDs, means \pm SEMs, number of rats or number of arterial segments. In each concentration-response curve, the changes in tension related to increasing concentrations of phenylephrine are expressed as the difference of tension (g) between the basal values and the

measured value in a certain concentration of phenylephrine. The curves were fitted to all the data by non-linear regression analysis using GraphPad Prism software (GraphPAD software, San Diego, CA, USA) to calculate the negative logarithm of the concentration of agonist that provokes a half-maximal response (pEC_{50} = agonist potency). The pA₂ values and Schild slopes were calculated by the method of Arunlakshana and Schild [16]. Using three different concentrations of each antagonist, the ratios between the half-maximal concentrations (EC_{50}) of phenylephrine obtained in a specific antagonist concentration and that obtained in its absence (the concentration ratio, R) were respectively calculated when the maximal amplitudes between the concentration-response curves in those two conditions were similar. The data was plotted as negative log-



Fig. 1. Time course for the change in the mean arterial pressure (MAP) after tilting in the streptozotocin-induced diabetic (\blacksquare) and normal (\blacktriangle) rats. Each data point represents the mean ± SD. *P < 0.01 vs. the normal rats.

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concentrations of the antagonist on the x-axis vs. log (R-1) on the y-axis and the data was fitted as a regression line. For calculation purposes, the slope parameter was set to -1.0 when it was statistically not different from unity. The x-intercept of the fitted regression line was the estimated pA₂. pA₂ is the estimated equilibrium dissociation constant for the specific antagonist and also the dose of antagonist that requires a 2-fold increase in the agonist concentration. Student's t-test was used to determine the statistical differences between the diabetic and control groups. A P value < 0.05 was considered statistically significant.

Results

Induction of diabetes mellitus and the changes in arterial blood pressure related to the tilting

The initial body weight was 242.8 ± 18.9 g for the diabetic rats and 244.7 ± 14.8 g for the normal rats, respectively (P = 0.79). Four weeks after diabetic induction, the body weight was 242.3 ± 28.0 g and the blood glucose level was 481.7 ± 110.5 mg/dl in

Table 1. Maximal Response (Emax) and the pEC_{50} Values for Phenyle-phrine-induced Contraction of the Aorta and Distal MesentericArtery from Streptozotocin-induced Diabetic and Normal Rats

| | Diabetic rats | | Normal rats | |
|--------------|--|--|------------------------------------|------------------------------------|
| | Emax (g) | pEC ₅₀ | Emax (g) | pEC ₅₀ |
| Aorta DMA | $2.52 \pm 0.03^{*}$ $2.37 \pm 0.10^{*}$ | $7.44 \pm 0.03^{*}$ $6.06 \pm 0.09^{*}$ | 2.22 ± 0.04 2.79 ± 0.08 | 6.47 ± 0.05 6.74 ± 0.07 |

The data is presented as means \pm SDs. EC₅₀: the concentration of agonist that provokes a half-maximal response on the concentration-response curve, pEC₅₀: negative logarithm of the EC₅₀, DMA: distal mesenteric artery. *P < 0.001 vs. the normal rats.



Fig. 2. Concentration-response curves for phenylephrine-induced contraction of the aorta (A) and the distal mesenteric artery (B) and as obtained from the streptozotocin-induced diabetic (\blacksquare) and normal (\blacktriangle) rats. Tension is expressed as the difference between the measured value and the basal value at each point. Each data point represents the mean ± SEM.



Fig. 3. Effects of prazosin (A), 5-methyurapidil (5-MU,) (B) and BMY 7378 (C) on the concentration-response curve of contraction in response to phenylephrine in the aorta and distal mesenteric arteries from the streptozotocin-induced diabetic and normal rats. Tension is expressed as the difference between the measured value and the basal value at each point. Each data point represents the mean ± SEM.





Fig. 3. Continued.

the diabetic rats. However, the body weight and blood glucose level in the normal rats were 428.5 ± 21.3 g and 133.1 ± 31.5 mg/ dl, respectively. The diabetic rats showed a significantly higher blood glucose level and a lower body weight than that of the normal rats (all P values < 0.001).

After vertical tilting the table at 45° in the head-up direction, the recovery of the mean arterial pressure was significantly slower in the diabetic rats than in that in the normal rats (Fig. 1).

Concentration-response curves

The diabetic rats showed a significantly higher Emax and pEC_{50} than that of the normal rats on the phenylephrineinduced concentration-response curve of the abdominal aorta (all P values < 0.001, Table 1 and Fig. 2A). However, in the distal mesenteric arteries, the diabetic rats showed a lower Emax and pEC_{50} than that of the normal rats (all P values < 0.001, Table 1 and Fig. 2B).

Regardless of the arterial type, prazocin, 5-MU and BMY 7378 shifted the concentration-response curve according to phenylephrine to the right direction for both the diabetic and normal rats (Fig. 3).

In addition, the pA₂ values of prazocin, 5-MU and BMY 7378 for inhibiting the contractile response to phenylephrine



Table 2. The pA_2 Values Against Phenylephrine in the Aorta andDistal Mesenteric Artery from the Streptozotocin-induced Diabeticand Normal Rats

| | Diabetic rats | | Normal rats | |
|----------|-----------------|---------------------|-----------------|-----------------|
| | Aorta | DMA | Aorta | DMA |
| Prazocin | $9.29\pm0.08^*$ | $8.77\pm0.06^*$ | 8.94 ± 0.04 | 9.26 ± 0.05 |
| 5-MU | $8.14\pm0.07^*$ | $7.22 \pm 0.17^{*}$ | 7.79 ± 0.06 | 7.82 ± 0.04 |
| BMY 7378 | $8.49\pm0.01^*$ | $7.59\pm0.12^*$ | 8.02 ± 0.05 | 7.96 ± 0.16 |

The data is expressed as means \pm SDs. pA₂ is the estimated equilibrium dissociation constant for the specific antagonist and also the dose of antagonist that requires a 2-fold increase in the agonist concentration. 5-MU: 5-methylurapidil, DMA: distal mesenteric artery. *P < 0.001 vs. the normal rats.

increased a half order of magnitude in the abdominal aorta from the diabetic rats, but this all decreased by a similar order of magnitude in the distal mesenteric arteries from the diabetic rats, respectively (all P \leq 0.001, Table 2).

Immunofluorescence staining

The diabetic rats showed an increased fluorescent intensity of all the α_1 -adrenoceptor subtypes in the abdominal aorta (Fig. 4A). However, in the distal mesenteric arteries, the fluorescent intensity of all the α_1 -adrenoceptor subtypes was decreased in



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Fig. 4. Immunofluorescence images of the α_{1A^-} (A), α_{1B^-} (B) and α_{1D^-} (C) adrenoceptor subtypes in the abdominal aorta and distal mesenteric arteries from the streptozotocininduced diabetic and normal rats (magnification: ×200). The abdominal aorta showed increased fluorescent intensity in the diabetic rats, but the distal mesenteric arteries showed decreased fluorescent intensity in the diabetic rats.



the diabetic rats (Fig. 4B).

Discussion

Our results showed that diabetes impaired the MAP recovery after positional change in rats, and that the phenylephrinemediated vascular contractility of diabetic rats was increased in the abdominal aorta and decreased in the distal mesenteric arteries compared with that of the normal rats. In addition, the diabetic abdominal aorta showed the increased fluorescent intensity of all the α_1 -adrenoceptor subtypes, but the fluorescent intensity of all the α_1 -adrenoceptor subtypes was decreased in the diabetic distal mesenteric arteries. However, the functional and distributional changes related to diabetes would be based on the increase or decrease of the overall α_1 -adrenoceptors, and it is not based on changes of the specific α_1 -adrenoceptor subtypes.

The vascular α_1 -adrenoceptor plays a primary role in maintaining the systemic arterial pressure and arteriolar resistance [7-10]. Since the first report of strong evidence for pharmacologically distinct α_1 -adrenoceptor subtypes in 1982 [17], numerous studies have assessed the function, distribution and classification of each subtype. It is now well known that there are 3 functional α_1 -adrenoceptor subtypes (α_{1A} , α_{1B} , and α_{1D}) corresponding to the 3-cloned α_1 -adrenoceptor [11]. In

Fig. 4. Continued.

addition, recent studies have shown that the α_{1D} -adrenoceptor plays a major role in the vasoconstriction of the mesenteric arteries [18,19] and that the α_{1A} -adrenoceptor plays a major role in the vascular contraction of the distal mesenteric arteries [19]. Moreover, Asbún-Bojalil et al. [20] suggested that the role of the α_{1D} -adrenoceptor is the most important for the contractile response to phenylephrine in both the abdominal and thoracic aortas. Therefore, in this study, to evaluate the effect of diabetes on the α_1 -adrenoceptor or the α_1 -adrenoceptor subtypes of the different types of artery, the abdominal aorta and the distal mesenteric artery were chosen as the conductive artery and the resistant artery, respectively.

Positional change of the arterial pressure was assessed for testing the appropriateness of our diabetic model; 4 weeks after diabetic induction by STZ intravenous injection, the diabetic rats showed impaired recovery from the tilt-induced decrease in blood pressure. This phenomenon was very comparable with orthostatic hypotension. Considering that orthostatic hypotension is the most serious clinical consequence of diabetes-induced vascular disturbance [2], our diabetic rat model would be appropriate for evaluating the effect of diabetes on the vascular alpha 1-adrenoceptor.

In this study, the responses of the α_1 -adrenoceptor to phenylephrine were different between the abdominal aorta and the mesenteric arteries according to the presence of diabetes.



In the abdominal aorta, diabetes increased the Emax over 10%. In addition, the diabetic abdominal aorta showed an EC_{50} in a 10-times lower concentration of phenylephrine than that of the normal abdominal aorta. This result corresponds with the previous studies where diabetes increased the adrenaline-mediated contractility in the aorta [21-24].

However, the diabetic distal mesenteric arteries showed lower a Emax by over 15% and the EC₅₀ in a 5-times higher phenylephrine concentration than did the normal distal mesenteric arteries. This finding is incompatible with the previous studies where the contractility of diabetic superior mesenteric arteries to α_1 -adrenoceptor agonists was increased [25-27]. However, various contractile responses of the α_1 -adrenoceptor in the superior mesenteric arteries to adrenergic agonist have been reported [28]. It has also been suggested that those various results were caused not only from the different experimental methods, but also from the different vascular responses between the superior and distal mesenteric arteries [28]. In addition, in the study using methoxamine by Makino and Kamato [29], the maximum contractile response was impaired in the diabetic mesenteric arteries. Moreover, the results of our immunofluorescence staining were also in accordance with our different contractile responses to phenylephrine between the abdominal aorta and the distal mesenteric arteries. Therefore, we speculate that the inconsistent results might have been caused by the different vascular characteristics between the superior and distal mesenteric arteries.

There were several limitations in this study. First, in this study, all the results would be based on the overall changes in the α_1 adrenoceptor. Therefore, the authors found no difference of the diabetes-induced functional and distributional changes of the specific α_1 -adrenoceptor subtypes. However, this is still the first study that has evaluated the relationship between diabetes and the α_1 -adrenoceptor subtypes. Second, the distribution of the α_1 -adrenoceptor subtypes was not quantified using Western blotting. In our experiments, we failed to get consistent results for Western blotting. Therefore, we could not quantify our results. However, consider a recent report that used commercial antibodies for the α_1 -adrenoceptor subtypes [30], and that inconsistency might have been caused by the limitations of the commercial antibodies. Moreover, that previous report has also suggested that competition radio-ligand binding is the only reliable approach to quantify the α_1 -adrenoceptor subtype proteins [30]. More studies are needed to overcome the limitations of our study.

In conclusion, progressed diabetes increased the contractile response to phenylephrine in the abdominal aorta, but it decreased the contractile response in the mesenteric arteries in STZ-induced diabetic rats. Those results were caused by the overall change in the α_1 -adrenoceptor, and not from the change

in the specific α_1 -adrenoceptor subtypes. Our results may be helpful to understand one of the complex pathophysiologies of diabetic cardiovascular autonomic neuropathy, and this is a very common and serious complication that clinical anesthesiologists are obligated to deal with.

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