

Supplemental Information

8C-like cells capture the human zygotic genome activation program *in vitro*

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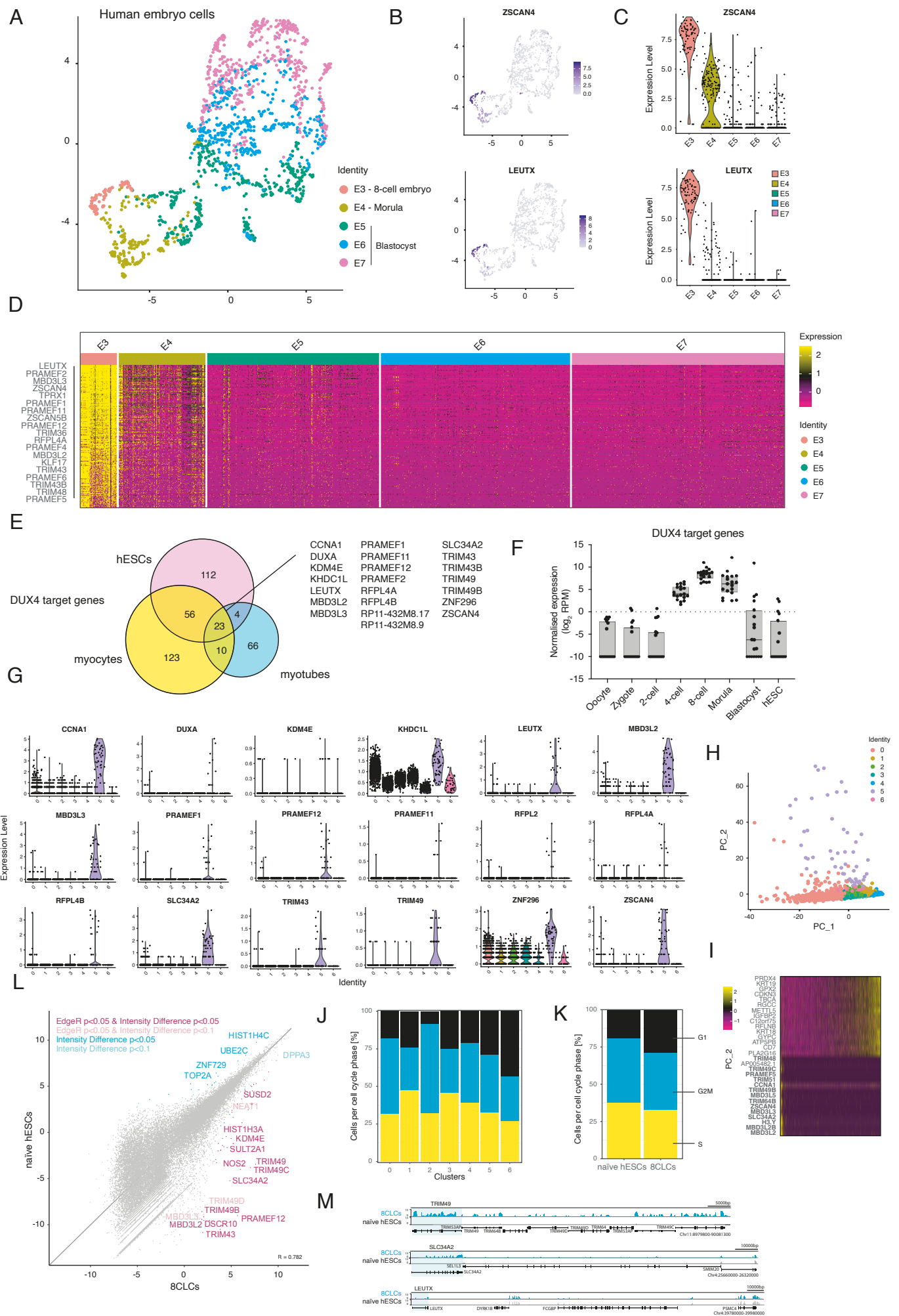


Figure S1

Figure S1, related to Figure 1. Defining human ZGA-like gene expression in 8-cell embryos and stem cells.

(A) UMAP of human 8-cell (E3), morula (E4) and blastocyst (E5 – E7) stage embryo cells based on single cell RNA expression data (Petropoulos et al., 2016). (B) UMAP and (C) Violin plots of normalized, scaled *ZSCAN4* and *LEUTX* expression in human pre-implantation embryo cells. (D) Heatmap of 8-cell embryo signature gene expression (rows) in human pre-implantation embryo cells (E3 – E7, columns). Markers of human ZGA are highlighted. (E) Differentially expressed genes that are upregulated upon *DUX4* overexpression in primed hPSCs (magenta), myocytes (yellow), and endogenous *DUX4* upregulation in human patient myotubes (blue) (Hendrickson et al., 2017; Jiang et al., 2020; Yao et al., 2014). Shared genes across all three datasets are highlighted. (F) Normalized expression of *DUX4* target genes during human pre-implantation development (oocyte to blastocyst) and in hESCs (Yan et al., 2013). (G) Violin plots of selected ZGA markers in clustered human naïve ESCs, based on normalized, scaled single cell RNA expression levels. Clusters were determined as in Figure 1 (B). (H) PC1 and PC2 of clustered naïve human HNES1 ESCs. (I) PC 2 loadings of naïve HNES1 are shown. ZGA marker genes are highlighted in bold. (J) Cell cycle analysis of single cell transcriptome data in naïve hESCs clusters (cluster 0 – 6) or (K) specifically 8CLCs as compared to naïve hESCs. (L) Differentially expressed genes, as defined by EdgeR or Intensity Difference analysis, between 8CLCs (n=5/480, based on the expression of ZGA markers) and naïve H9 PSCs cultured in t2iLGö (Messmer et al., 2019). (M) Genome browser tracks of pseudo-bulked ZGA marker genes (*TRIM49*, *SLC34A2*, *LEUTX*) in 8CLCs (as in G) and naïve H9 hPSCs (Messmer et al., 2019).

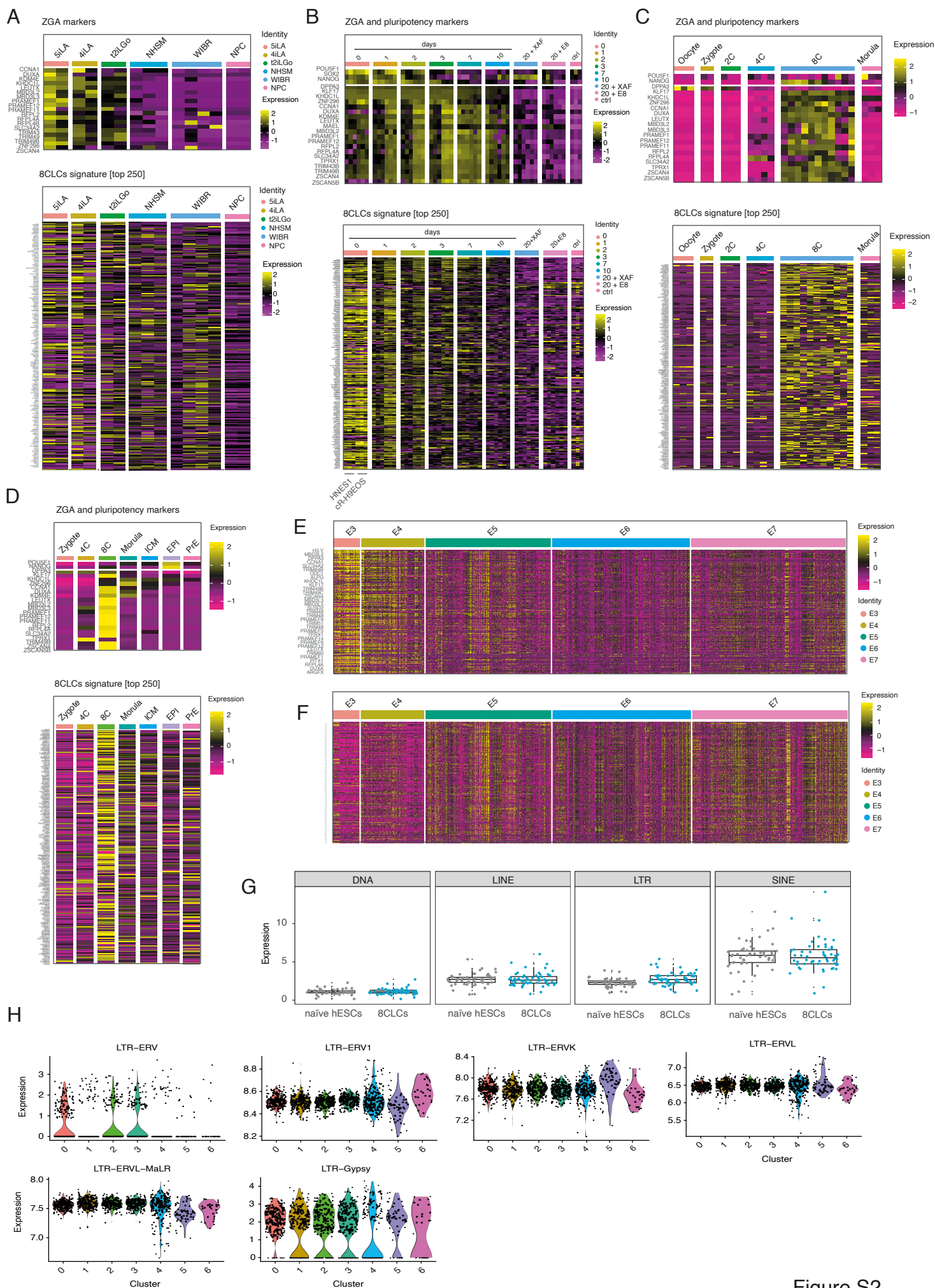


Figure S2

Figure S2, related to Figure 2, 3. 8CLC signature and transposon expression in human embryos and stem cells.

(A) Heatmap of ZGA markers (top panel) and 8CLCs signature genes (bottom panel) of naïve (5iLA, 4iLA, t2iLgö), pseudo-naïve (NHSM), primed (WIBR) and differentiated cells (NPCs, neuronal progenitor cells). The analysis was done on bulk RNA expression data. (B) Heatmap of pluripotency genes (i.e. *POU5F1*, *SOX2*, *NANOG*), ZGA markers (top panel) and 8CLCs signature genes (bottom panel) in two naïve PSC lines (HNES1, cR-H9EOS in triplicate each). Cells were transitioned from the naïve into the primed state, in two different primed media (E8, XAF) (Rostovskaya et al., 2019). The days indicate the days of transition (day 0 – day 10, d20). Gene expression of control primed H9 cells is shown as well (ctrl). (C), (D) Heatmap of pluripotency genes (*POU5F1*, *NANOG*), ZGA markers (top panels) and 8CLCs signature genes (bottom panels) in human pre-implantation embryos from two different studies (C) (Xue et al., 2013) (D) (Stirparo et al., 2018). (E) 8CLCs signature and (F) naïve hPSCs marker gene expression that distinguishes them from 8CLCs, in human embryos (E3 – E7) (Petropoulos et al., 2016). (G) DNA, LTR, LINE, and SINE transposable element expression in naïve hESCs and 8CLCs. Reads are shown as percentage of total reads. (H) Normalized reads of LTR family transposons expressed in 8CLCs and naïve hESCs from single cell RNA sequencing data (for clusters see Figure 1B).

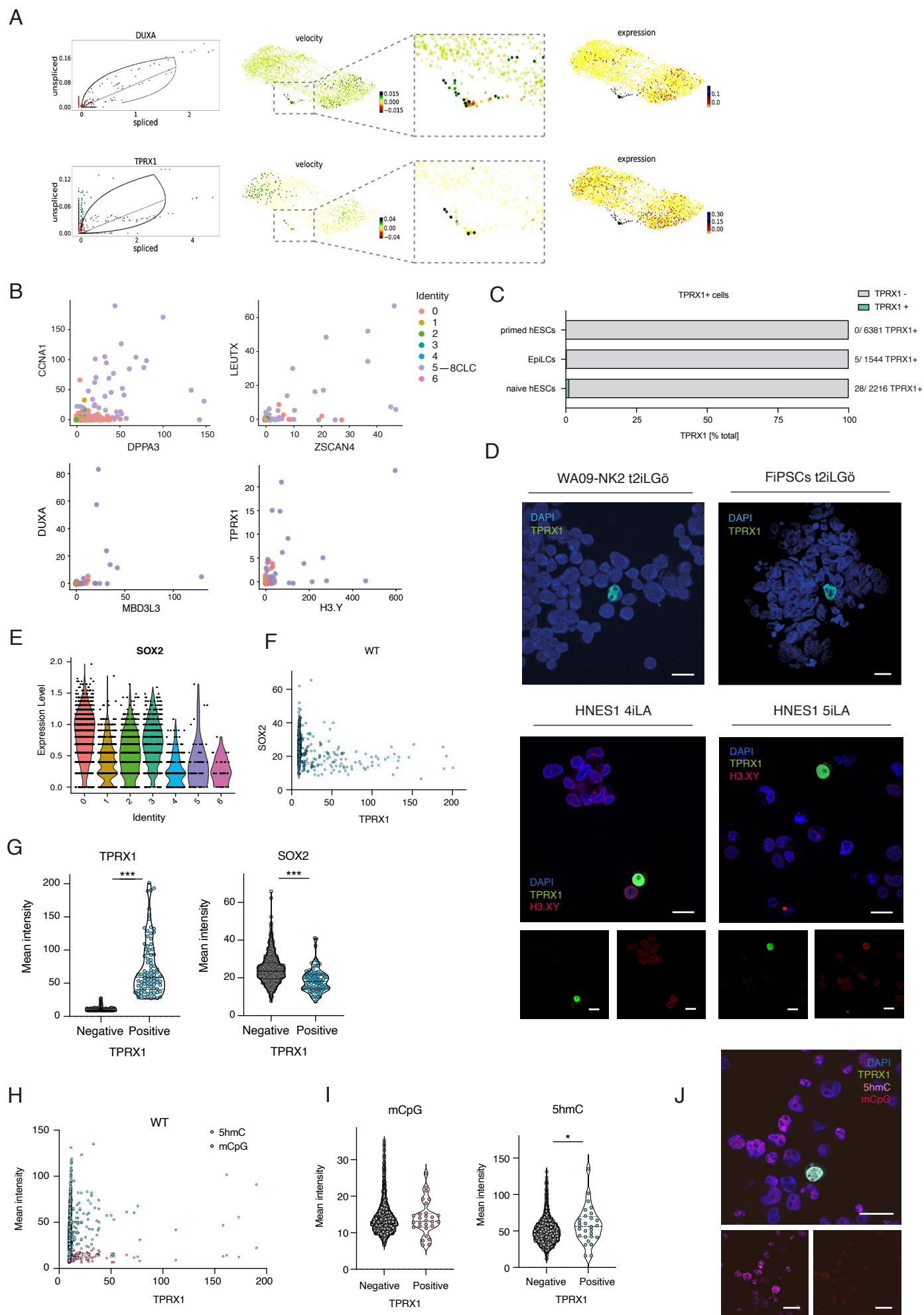


Figure S3

Figure S3, related to Figure 3, 4. Identification of 8CLCs among naïve hESCs by marker protein expression *in vitro*.

(A) Left panel: RNA velocity analysis of *DUXA* and *TPRX1* in individual 8CLCs and naïve hESCs (for clusters see Figure 1B); steady-state ratios (black lines), overall dynamics (black curve) and ratios of unspliced vs. spliced mRNA in single cells; middle panel: RNA velocity of *DUXA* and *TPRX1*; right panel: RNA expression levels of *DUXA* and *TPRX1*. (B) Scatter plots of 8CLC marker gene co-expression in naïve hESCs and 8CLC clusters. (C) Quantification of TPRX1-positive (TPRX1+) naïve hESCs, EpiLCs and primed ESCs as detected by immunofluorescence staining. Percentages are shown, total numbers are indicated on the right-hand side of the graph. (D) Upper panel: IF staining of TPRX1 in reprogrammed naïve H9 PSCs (left) and fibroblast derived naïve iPSCs (FiPSCs, right) cultured in t2iLGö. Bottom panel: IF staining of TPRX1 and H3.X/Y in naïve HNES1 cells cultured in 4iLA (left) or 5iLA (right). Scale bar, 20µm. (E) Violin plot of normalized, scaled *SOX2* expression in clustered naïve human ESCs (for clusters see Figure 1B). (F) TPRX1 and *SOX2* mean fluorescent signal intensity per cell (mid optical slice) in antibody stained, cytopun naïve HNES1 cells cultured in PXGL. TPRX1-positive cells and control cells were selected to assess *SOX2* protein levels. (G) Quantification of mean TPRX1 (left) and *SOX2* (right) intensity levels in TPRX1-positive and TPRX1-negative selected HNES1 cells. Images were taken from four independent experiments and pooled. p-value *P<0.05, **P<0.01, ***P<0.001, absence of stars (non-significant, ns): p-value>0.05; two tailed Mann-Whitney U test. (H) TPRX1 and 5hmC/ mCpG mean fluorescent signal intensity per cell (mid optical slice) in antibody stained, cytopun naïve HNES1 cells cultured in PXGL. (I) Mean mCpG (left) and 5hmC (right) intensity levels in TPRX1-positive and TPRX1-negative HNES1 cells. Images were taken from at least two experiments and pooled. p-value *P<0.05, **P<0.01, ***P<0.001, absence of stars (non-significant, ns): p-value>0.05; unpaired, two tailed Mann-Whitney U test. (J) IF staining of TPRX1, 5hmC and mCpG in naïve HNES1 cells cultured in PXGL. Scale bar, 20µm.

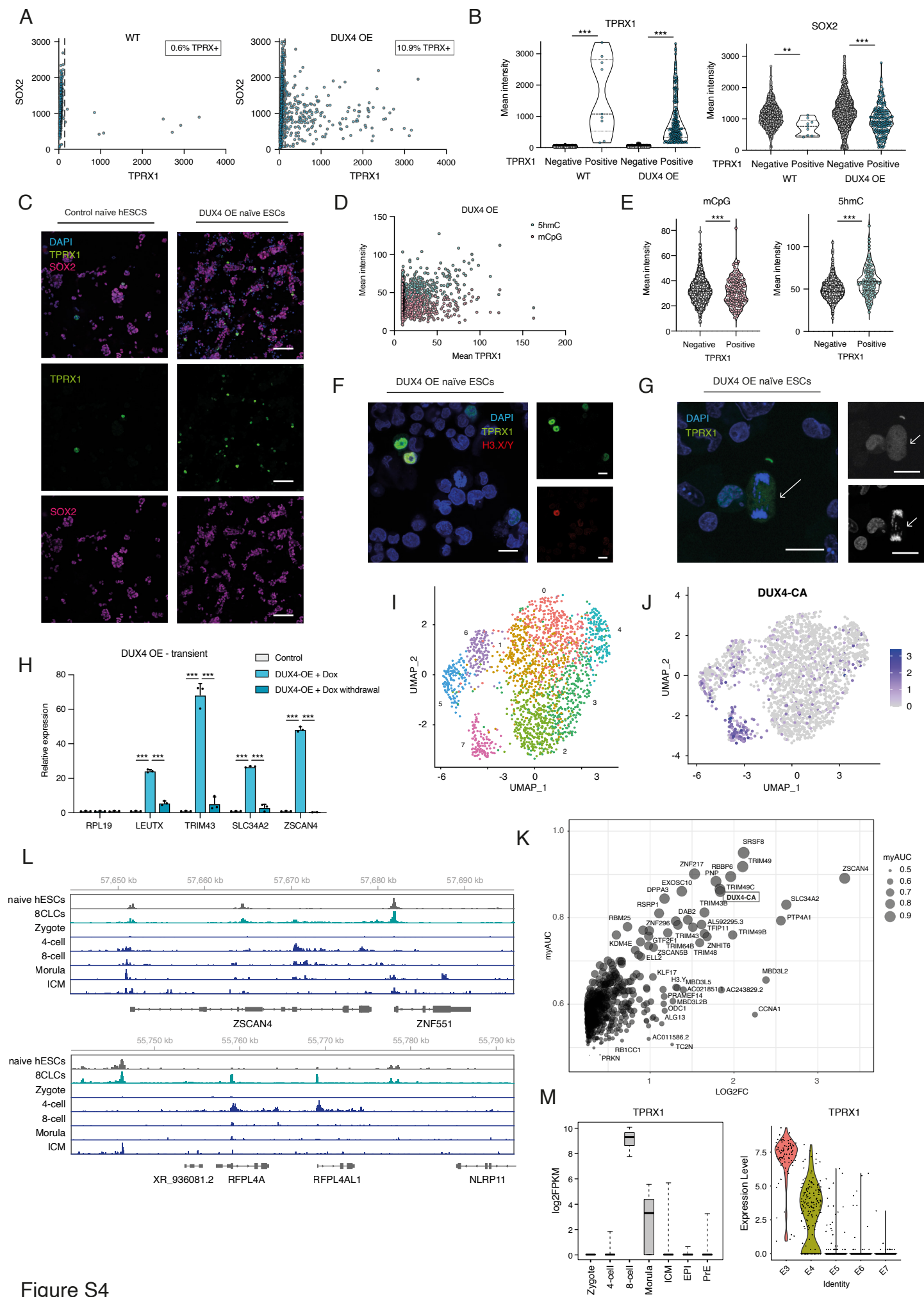


Figure S4

Figure S4, related to Figure 4. Characterization of *DUX4* overexpressing naïve hESCs and 8CLCs.

(A) TPRX1 and SOX2 mean fluorescent signal intensity per cell in antibody-stained control (WT) and *DUX4* overexpressing (*DUX4* OE) naïve HNES1 cells cultured in PXGL and plated on coverslips. The percentage of TPRX1-positive cells is indicated in the graphs, the cut-off is shown as a dashed line ($x=150$). (B) Mean TPRX1 (left) and SOX2 (right) intensity levels in TPRX1-positive and TPRX1-negative control (WT) and *DUX4* overexpressing (*DUX4* OE) HNES1 cells; two independent experiments. Cut-off as in (A). p-value * $P<0.05$, ** $P<0.01$, *** $P<0.001$, absence of stars (non-significant, ns): p-value >0.05 ; two tailed Mann-Whitney U test. (C) IF staining of TPRX1 and SOX2 in control and *DUX4* overexpressing HNES1 cells plated on coverslips. Scale bar, 100 μ m. (D) TPRX1 and 5hmC/ mCpG mean fluorescent signal intensity per cell (mid optical slice) in antibody stained, cytopspun *DUX4* overexpression HNES1 cells cultured in PXGL. (E) Mean mCpG (left) and 5hmC (right) intensity levels in TPRX1-positive and TPRX1-negative *DUX4* overexpressing HNES1 cells; two independent experiments. p-value * $P<0.05$, ** $P<0.01$, *** $P<0.001$, absence of stars (non-significant, ns): p-value >0.05 ; nonparametric Mann-Whitney U test. (F) IF staining of TPRX1 and H3.X/Y in *DUX4* overexpressing HNES1 cells cytopspun onto coverslips. Scale bar, 20 μ m (G) IF staining of TPRX1 in *DUX4* overexpressing cytopspun HNES1 cells. Dividing TPRX1-positive cells (anaphase chromosomes) are indicated (see arrows). Scale bar, 20 μ m. (H) Expression levels of ZGA marker genes measured by RT-qPCR in *DUX4* overexpressing (*DUX4*-OE + Dox, 24h) cells as well as upon Dox withdrawal (*DUX4*-OE + Dox withdrawal, 24h + 72h). Data are shown as mean $\pm$ SD ($n = 3$ technical replicates) of fold-change compared to control HNES1, and are representative of three independent experiments. p-value * $P<0.05$, ** $P<0.01$, *** $P<0.001$, absence of stars (non-significant, ns): p-value >0.05 ; unpaired, two tailed Student's t-test. (I) UMAP of clustered human naïve ESCs and 8CLCs (cluster 7). (J) Normalized scaled *DUX4*-CA expression in clustered hESCs and 8CLCs visualized on a UMAP. (K) Marker gene expression signature of 8CLCs (cluster 7 see S4I). AUC, area under curve; LOG2FC, log2 fold-change. (L) Genome browser views of accessibility at genomic loci of ZGA markers *ZSCAN4* (data range 0.0 – 0.2 stem cells, 0.0 – 1.0 embryo data) and *RFPL4A* (data range 0.0 – 0.1 stem cells, 0.0 – 1.0 embryo data) in naïve hESCs (grey) and 8CLCs (petrol) as compared to human embryos (blue) (Liu et al., 2019). (M) Normalized bulk (left) and single cell (right) *TPRX1* RNA expression levels in preimplantation human embryo cells (Petropoulos et al., 2016; Stirparo et al., 2018). Left: Figure adapted from <https://app.stemcells.cam.ac.uk/human-embryo/>.