

RESEARCH ARTICLE

Hotspot analysis by confocal microscopy can help to differentiate challenging melanocytic skin lesions

Raquel de Paula Ramos Castro^{1*}, Juliana Casagrande Tavoloni Braga¹, Mariana Petaccia de Macedo², Clóvis Antonio Lopes Pinto², José Humberto Tavares Guerreiro Fregnani³, Gisele Gargantini Rezze⁴

1 Department of Cutaneous Oncology, AC Camargo Cancer Center, São Paulo, Brazil, **2** Department of Pathology, AC Camargo Cancer Center, São Paulo, Brazil, **3** Teaching and Research Institute, AC Camargo Cancer Center, São Paulo, Brazil, **4** Dermaimage Medical Associates, São Paulo, Brazil

* rprcastro@yahoo.com.br



OPEN ACCESS

Citation: Castro RdPR, Casagrande Tavoloni Braga J, Petaccia de Macedo M, Lopes Pinto CA, Tavares Guerreiro Fregnani JH, Gargantini Rezze G (2022) Hotspot analysis by confocal microscopy can help to differentiate challenging melanocytic skin lesions. *PLoS ONE* 17(2): e0263819. <https://doi.org/10.1371/journal.pone.0263819>

Editor: Nikolas K. Haass, University of Queensland Diamantina Institute, AUSTRALIA

Received: May 7, 2021

Accepted: January 27, 2022

Published: February 14, 2022

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: <https://doi.org/10.1371/journal.pone.0263819>

Copyright: © 2022 Castro et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its [Supporting Information](#) files.

Abstract

Some melanocytic lesions do not present enough clinical and dermoscopic features to allow ruling out a possible melanoma diagnosis. These “doubtful melanocytic lesions” pose a very common and challenging scenario in clinical practice and were selected at this study for reflectance confocal microscopy evaluation and subsequent surgical excision for histopathological diagnosis. The study included 110 lesions and three confocal features were statistically able to distinguish benign melanocytic lesions from melanomas: “peripheral hotspot at dermo-epidermal junction”, “nucleated roundish cells at the dermo-epidermal junction” and “sheet of cells”. The finding of a peripheral hotspot (atypical cells in 1mm²) at the DEJ is highlighted because has not been previously reported in the literature as a confocal feature related to melanomas.

Introduction

The diagnosis of melanocytic skin lesions based only on clinical and dermoscopic evaluation can be challenging, even for experienced dermatologists. Some melanocytic lesions do not present enough clinical and dermoscopic features to establish a definitive diagnosis with reliability, imposing the need for biopsy and histopathological evaluation.

The use of additional non-invasive imaging techniques, like *in vivo* reflectance confocal microscopy (RCM) permits a cytoarchitectural evaluation of the epidermis, the dermo-epidermal junction (DEJ) and the upper dermis. Cellular atypia and pleomorphism can also be visualized *in vivo* to aid the diagnosis [1]. As a result, RCM represents a sensitive and specific tool for the early detection of melanomas and other skin tumors [1].

However, only few articles in the literature have described the RCM features present in melanocytic lesions. This study aimed to describe in detail the RCM features of melanocytic lesions with a doubtful diagnosis after clinical and dermoscopic evaluation, and search for new RCM features capable to differentiate them.

Funding: No received specific funding.

Competing interests: The authors have declared that no competing interests exist.

Materials and methods

This retrospective study was approved by the Fundação Antônio Prudente ethical committee (01524/11) and all patients included agreed to participate and signed the informed consent document. A total of 96 patients from the Cutaneous Oncology Department of AC Camargo Cancer Center, São Paulo, Brazil, with 110 doubtful clinical and dermoscopic melanocytic lesions were selected. Only lesions suspicious of superficial spreading melanomas were included because other melanomas subtypes have other dermoscopy and confocal parameters (i.e. lentigo maligna, amelanotic melanoma, nodular melanomas and acral melanomas). Also, lesions located at sites where the VivaScope 1500 RCM device could not be adapted and lesions located at special sites such as the face, scalp and digits were excluded.

The dermoscopic diagnostic method used was the Pattern Analysis, applying the following criteria: eccentric pigmentation, abrupt network loss, poorly defined network, enlarged/atypical pigment network, multiple brown or dark globules with irregular shape and distribution, peppering, multiple colors, negative network, blue-white veil, radial streaks, pseudopods and structureless areas [2]. Melanocytic lesions with few or faint dermoscopy features related to melanoma diagnosis and lacked enough criteria for benign lesion were termed as “doubtful melanocytic lesions”.

These “doubtful melanocytic lesions” were submitted to RCM examination by an experienced dermatologist using the VivaScope® 1500 confocal microscope (Lucid-Tech, Rochester, New York, USA). The examination was carried out in a step-by-step manner, with complete image documentation for subsequent analyses blinded from anatomopathological results. The RCM evaluation was based on features previously described (Table 1) [3–5].

In superficial spreading melanoma, junctional aggregates of melanocytes are commonly found, with high shape and size variability, composed of highly atypical melanocytes and with a tendency to confluence [6]. Due to the non-uniform distribution of these atypical melanocytes aggregates throughout the lesion and in order to quantify the degree and location of higher atypia inside a given lesion, hotspot analyses were included. A hotspot was defined as the 01 x 01 mm area of the lesion where the atypical cells are more aggregated, and was searched at the epidermis and DEJ levels. The selection of the hotspot was defined subjectively by the dermatologist after careful visual inspection of RCM mosaic images from the epidermis and DEJ levels, delimiting the 01 x 01 mm area with higher atypia and classifying its location as central or peripheral. The degree of atypia in a hotspot was based on the number of atypical cells inside its 01 x 01 mm area (absent, ≤ 10 or > 10 atypical cells). Illustrative cases of hotspot location and quantification are shown in Figs 1 and 2.

Subsequently, all lesions were excised for histopathological diagnosis, which was considered the gold standard for final diagnosis, and categorized into benign melanocytic lesions (common melanocytic nevi and atypical melanocytic nevi) or melanomas. Statistical analysis by simple and multiple logistic regressions were conducted using SPSS software (version 21) and p value ≤ 0.05 was considered significant.

Results

All 110 melanocytic lesions were subjected to histopathological examination, and their final diagnoses were: 31 common nevi (28%), 53 atypical (dysplastic) nevi (48%) and 26 melanomas (24%). Intense atypia was present in 3 (6%) dysplastic nevi. Among the 26 melanomas, 18 (69%) were *in situ*, 7 (27%) were thin (Breslow < 1 mm) and 1 (4%) had Breslow of 1.42 mm. Seven (27%) melanomas appeared from a pre-existing nevus.

The features found in the RCM examination were analyzed according to the final histopathological diagnoses (Table 2).

Table 1. Description of RCM features.

RCM features	Description
Honeycomb pattern at suprabasal epidermis: Typical/Atypical	Typical: normal or “preserved” honeycomb pattern; keratinocytes are well-demarcated, “visible”. Atypical: “partial loss” of honeycomb pattern, keratinocytes demarcation are “poorly or not visible”
Atypical cells at epidermis: Presence/Absence	Atypical cells are large, bright and pleomorphic cells
Atypical cells at epidermis: Nucleated roundish/Dendritic	If present, atypical cells were classified: a) nucleated roundish cells—dark nucleus and bright cytoplasm, frequently twice the size of keratinocytes; b) dendritic cells—elongated branching structure extending from the cell body, usually present in melanocytes and Langerhans cells
Hotspot at epidermis: Presence/Absence	Hotspot was defined as the 1 x 1 mm area of the lesion that presents more atypia
Atypical cells at hotspot (epidermis): ≤ 10 or > 10	If present, hotspots were classified according to the number of atypical cells (≤ 10 or > 10 cells)
Location of hotspot at epidermis: Central/Peripheral	If present, hotspots were classified according to their predominantly location at the lesion: central or peripheral
Cobblestone pattern at basal cells: Typical/Atypical	Typical: uniform basal cells distribution; no variation in brightness or cellular outline between individual cells. Atypical: basal cells are not uniformly distributed
Papillae at DEJ: Edged/Non-edged	Edged: demarcated by a rim of bright basal cells (confluent cells). Non-edged: absence of a demarcated rim of bright cells, but separated by a series of large reflecting cells
General atypia at DEJ: Presence/Absence	Present if the normal architecture of the DEJ is partially or completely lost. Described as present if some RCM findings were noted: atypical meshwork pattern, atypical cells (dendritic or roundish cells), sheet of cells and “mitochondria-like structures”
Meshwork pattern at DEJ: Presence/Absence	Characterized by small dark holes surrounded by thickened interpapillary spaces
Meshwork pattern at DEJ: Typical/Atypical	If present, meshwork pattern was classified: a) typical—clearly thickened interpapillary spaces; b) atypical—irregular and enlarged interpapillary spaces by the presence of atypical cells
Atypical cells at DEJ: Presence/Absence	Atypical cells are large, bright and pleomorphic cells
Atypical cells at DEJ: Nucleated roundish/Dendritic	If present, atypical cells were classified: a) nucleated roundish cells—isolated round to oval refractive cells with a dark nucleus, located in the papillary dermis; b) dendritic cells—elongated dendritic cells around dermal papillae
Hotspot at DEJ: Presence/Absence	Hotspot was defined as the 1 x 1 mm area of the lesion that presents more atypia
Atypical cells at hotspot (DEJ): ≤ 10 or > 10	If present, hotspots were classified according to the number of atypical cells (≤ 10 or > 10 cells)
Location of hotspot at DEJ: Central/Peripheral	If present, hotspots were classified according to their predominantly location at the lesion: central or peripheral
Junctional nests: Presence/Absence	Oval compact cellular aggregates, bulging within the dermal papillae connected with the epidermal basal cell layer
Dense and sparse nests: Presence/Absence	Roundish nonreflecting structures with a well-demarcated border, containing isolated round to oval cells with dark nucleus and reflecting cytoplasm; sometimes presenting in a multilobate configuration
Dense (homogeneous) nests: Presence/Absence	Compact aggregates with sharp margin and similar cells in morphology and refractivity
Atypical nests: Presence/Absence	Dense and sparse nests composed by pleomorphic atypical cells
Peripheral nests: Presence/Absence	Enlarging nevus characterized by bulging junctional nests at the lesion periphery
Sheet of cells: Presence/Absence	Atypical pleomorphic melanocytes distributed in sheet-like structures
“Mitochondria-like structures”: Presence/Absence	Elongated dendritic cells crowded around dermal papillae, some of them forming bridges that resembles the mitochondrial aspect

(Continued)

Table 1. (Continued)

RCM features	Description
Short interconnections: Presence/Absence	Junctional thickenings and nests surrounding the papillae
Inflammatory cells: Presence/Absence	Bright particles within the papillae
Melanophages: Presence/Absence	Plump irregularly shaped bright cells with ill-defined borders and usually no visible nucleus in single units or in clusters
Coarse collagen fibers: Presence/Absence	Collagen fibers are packed together forming a coarse web-like architecture

<https://doi.org/10.1371/journal.pone.0263819.t001>

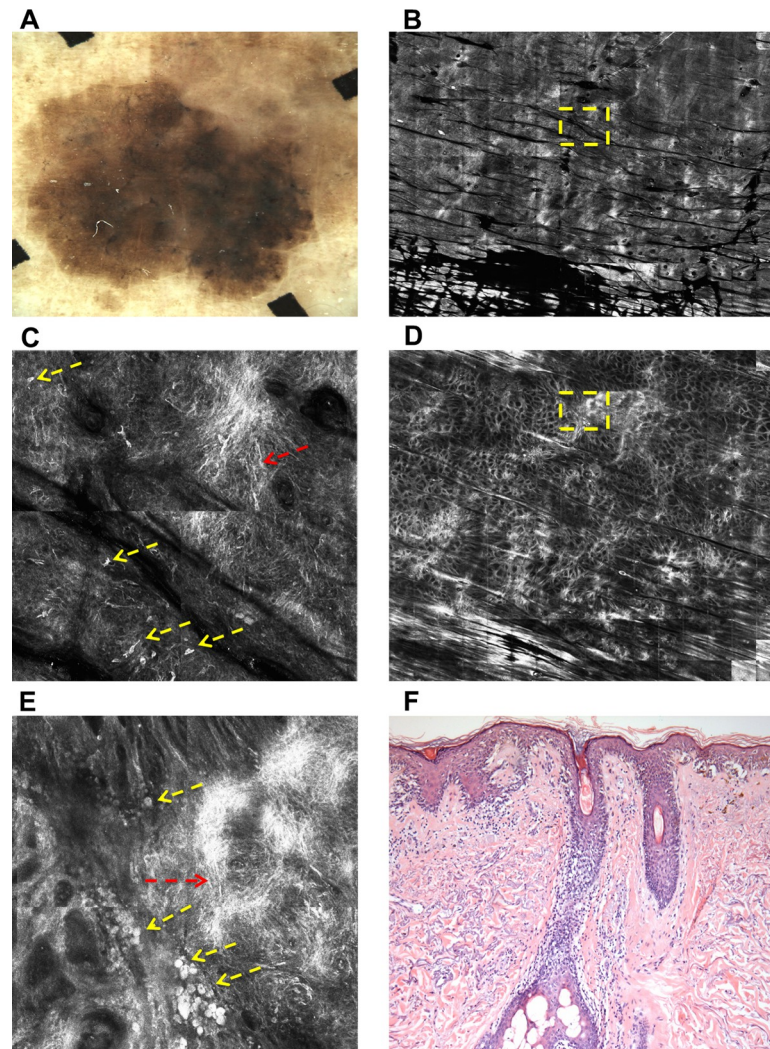


Fig 1. Superficial spreading melanoma *in situ*. (A): Dermoscopy shows atypical network and peppering. (B): RCM mosaic image (8.0 x 8.0 mm) at the spinous layer; visual inspection positioned the hotspot at a central location (1.0 x 1.0 mm square area is marked with yellow dashed outline and better shown in C). (C): Atypical honeycomb pattern and widespread roundish cells (yellow arrows) and dendritic cells (red arrows) in sheet of cells distribution. (D): RCM mosaic image (8.0 X 8.0 mm) at DEJ; visual inspection positioned the hotspot at a peripheral location (1.0 x 1.0 mm square area is marked with yellow dashed outline and better shown in E). (E): Non-edged papillae, roundish cells at DEJ (yellow arrows) and dendritic cells (red arrows) in sheet of cells distribution. (F): Histopathology confirms a superficial spreading melanoma *in situ* (H&E, original magnification x200), with disarrangement of the rete ridge and increased number of atypical melanocytes affecting the adnexae.

<https://doi.org/10.1371/journal.pone.0263819.g001>

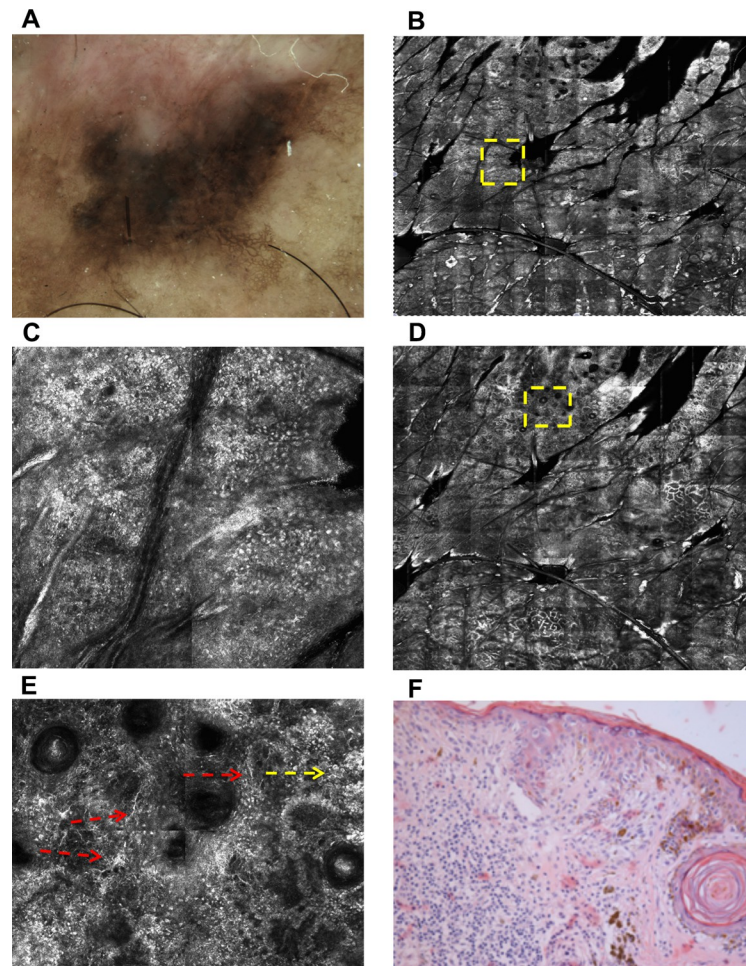


Fig 2. Superficial spreading melanoma. (A): Dermoscopy shows atypical network and peppering. (B): RCM mosaic image (8.0 x 8.0 mm) at epidermis (suprabasal layer); visual inspection positioned the hotspot at a central location (1.0 x 1.0 mm square area is marked with yellow dashed outline and better shown in C). (C): Atypical cobblestone pattern. (D): RCM mosaic image (8.0 X 8.0 mm) at DEJ; visual inspection positioned the hotspot at a peripheral location (1.0 x 1.0 mm square area is marked with yellow dashed outline and better shown in E). (E): Non-edged papillae, roundish cells at DEJ (yellow arrow) and dendritic cells (red arrows). (F): Histopathology confirms a superficial spreading melanoma (H&E, original magnification x200), with Breslow = 0.55 and presence of pagetoid cells.

<https://doi.org/10.1371/journal.pone.0263819.g002>

Statistical analyses of RCM results were performed by simple and multiple logistic regressions. Several RCM findings at the DEJ occurred simultaneously and were labeled as “DEJ general atypia” to better differentiate the groups: atypical meshwork pattern, presence of atypical cells (dendritic or roundish cells), sheet of cells and “mitochondria-like structures”.

After multiple logistic regression, the presence of three of these features remained statistically significant to differentiate benign melanocytic lesions (common and atypical melanocytic nevi) from melanomas: atypical roundish nucleated cells at DEJ ($p = 0.048$, 95% CI = 1.01–47.96), peripheral hotspot at DEJ ($p = 0.032$, 95% CI = 1.18–42.88) and sheet of cells ($p = 0.04$, 95% CI = 1.09–42.35) (Table 3). Only the presence of peripheral hotspot at DEJ was significantly related to melanoma diagnosis, instead predominantly central location of atypical cells at the DEJ hotspot was not.

Based on the final histological diagnosis, each different combination of these statistically significant RCM features was related to the probability of the lesion being a melanoma. The absence of these criteria confers a probability of 5.5%, while lesions that presented only one

Table 2. Frequency of RCM features in common nevi, atypical nevi and melanomas.

RCM features	Common nevus (31)	Atypical nevus (53)	Melanoma (26)
Typical Honeycomb	31 (100%)	48 (91%)	23 (43%)
Atypical Honeycomb	0 (0%)	5 (9%)	3 (12%)
Atypical cells (nucleated roundish or dendritic cells) at epidermis	07 (23%)	32 (60%)	23 (88%)
Nucleated roundish cells at epidermis	0 (0%)	2 (4%)	4 (15%)
Dendritic cells at epidermis	6 (19%)	31 (58%)	20 (77%)
Hotspot \leq 10 atypical cells at epidermis	6 (19%)	33 (62%)	9 (35%)
Hotspot $>$ 10 atypical cells at epidermis	0 (0%)	15 (28%)	14 (54%)
Absent of hotspot at epidermis	25 (81%)	5 (9%)	3 (12%)
Central hotspot at epidermis	5 (16%)	44 (83%)	16 (62%)
Peripheral at epidermis	1 (3%)	05 (6%)	07 (27%)
Typical cobblestone	31 (100%)	42 (79%)	12 (46%)
Atypical cobblestone	0 (0%)	11 (21%)	14 (54%)
Edged papillae	29 (94%)	24 (45%)	6 (23%)
Non-edged papillae	2 (6%)	29 (55%)	20 (77%)
DEJ general atypia	2 (6%)	39 (74%)	26 (100%)
Absent of meshwork pattern	11 (35%)	23 (43%)	13 (50%)
Typical meshwork pattern	20 (65%)	17 (32%)	4 (15%)
Atypical meshwork pattern	0 (0%)	13 (25%)	9 (35%)
Atypical cells (nucleated roundish or dendritic cells) at DEJ	2 (6%)	29 (55%)	20 (77%)
Nucleated roundish cells at DEJ	0 (0%)	6 (11%)	2 (8%)
Dendritic cells at DEJ	2 (6%)	27 (51%)	17 (65%)
Hotspot \leq 10 atypical cells at DEJ	2 (6%)	17 (32%)	3 (12%)
Hotspot $>$ 10 atypical cells at DEJ	0 (0%)	12 (23%)	15 (58%)
Absent of hotspot at DEJ	29 (94%)	24 (45%)	8 (31%)
Central hotspot at DEJ	2 (6%)	20 (38%)	12 (46%)
Peripheral at DEJ	0 (0%)	8 (15%)	6 (23%)
Junctional nests	23 (74%)	47 (89%)	23 (88%)
Dense and sparse nests	11 (39%)	15 (35%)	9 (44%)
Dense (homogeneous) nests	27 (30%)	51 (40%)	25 (60%)
Atypical nests	0 (0%)	3 (10%)	6 (33%)
Peripheral nests	10 (36%)	17 (45%)	5 (28%)
Sheet of cells	0 (0%)	2 (6%)	7 (6%)
Mitochondria-like structures ^a	0 (0%)	6 (11%)	9 (35%)
Short interconnections	4 (13%)	16 (30%)	11 (42%)
Inflammatory cells	14 (45%)	37 (70%)	22 (85%)
Melanophages	13 (42%)	26 (49%)	16 (62%)
Melanophages isolated	13 (42%)	16 (30%)	16 (62%)
Melanophages clusters	4 (13%)	15 (28%)	09 (35%)
Coarse collagen fibers	31 (100%)	52 (98%)	25 (96%)

<https://doi.org/10.1371/journal.pone.0263819.t002>

positive criterion presented a 31.5% to 37.5% probability, the lesions with two positive criteria presented 79.5% to 83.9% probability and the lesions that had three positive features presented 97.5% probability of being melanoma (Table 4).

Discussion

The main advantage of RCM is increasing the sensitivity and specificity of melanoma diagnosis and differentiating them from common and atypical nevi [4, 7, 8]. “Featureless

Table 3. Simple and multiple logistic regressions comparing RCM criteria in common and atypical nevi versus melanomas.

Simple logistic regression				
RCM features	Category	OR	95% CI	P value
Honeycomb	Typical or atypical pattern	2.872	0.71–11.61	0.139
Cobblestone	Typical pattern	1.0	2.86–21.00	<0.0001
Cobblestone	Atypical pattern	7.742	2.86–21.00	<0.0001
Atypical cells at epidermis	Nucleated roundish cells	7.455	1.28–43.39	0.025
Atypical cells at epidermis	Dendritic cells	4.234	1.54–11.61	0.005
Atypical cells at epidermis	Nucleated roundish cells or dendritic cells	8.846	2.47–31.73	0.001
Hotspot at epidermis	Absent	NA	NA	NA
Hotspot at epidermis	≤ 10 cells	5.423	1.35–21.81	0.017
Hotspot at epidermis	> 10 cells	19.939	4.87–81.61	<0.0001
Hotspot location at epidermis	Central	7.667	2.06–2.05	0.002
Hotspot location at epidermis	Peripheral	17.889	3.62–88.41	<0.0001
Papillae at DEJ	Non-edged papillae	5.699	2.07–15.71	0.001
DEJ general atypia	Presence	NA	NA	NA
Meshwork	Absent	NA	NA	NA
Meshwork	Typical pattern	0.283	0.08–0.95	0.041
Meshwork	Atypical pattern	1.811	0.63–5.24	0.27
Atypical cells at DEJ	Nucleated roundish cells	12.3	2.31–65.55	0.003
Atypical cells at DEJ	Dendritic cells	3.582	1.42–9.03	0.007
Atypical cells at DEJ	Nucleated roundish cells or dendritic cells	5.699	2.07–15.71	0.001
Hotspot at DEJ	Absent	NA	NA	NA
Hotspot at DEJ	≤10 cells	1.046	0.25–4.36	0.951
Hotspot at DEJ	> 10 cells	8.281	2.86–23.96	<0.0001
Hotspot location at DEJ	Central	3.682	1.32–10.24	0.012
Hotspot location at DEJ	Peripheral	5.062	1.39–18.45	0.014
Junctional nest	Presence	1.533	0.40–5.82	0.53
Dense and homogeneous nests	Presence	0.446	0.17–1.17	0.102
Dense and sparse nests	Presence	1.181	0.47–2.10	0.726
Atypical nests	Presence	8.1	1.86–35.22	0.005
Location of nests	DEJ /dermis	0.603	0.24–1.52	0.284
Location of nests	Peripheral	0.503	0.17–1.48	0.211
Sheet of cells	Presence	15.105	2.90–78.56	0.001
"Mitochondria-like structures"	Presence	6.882	2.16–21.92	0.001
Short interconnections	Presence	2.347	0.93–5.92	0.071
Inflammatory cells	Presence	3.559	1.13–11.26	0.031
Melanophages	Presence	1.846	0.75–4.54	0.181
Melanophages	Isolated	1.846	0.75–4.54	0.181
Melanophages	Clusters	1.811	0.70–4.71	0.223
Coarse collagen fibers	Presence	3.322	0.20–55.02	0.402
Multiple logistic regression *				
RCM features	Category	OR	95% CI	P value
Atypical cells at DEJ	Nucleated roundish cells	6.973	1.01–47.96	0.048
Hotspot location at DEJ	Peripheral	7.106	1.18–42.88	0.032
Sheet of cells	Presence	6.792	1.09–42.35	0.04

NA = not applicable

* shows only RCM features with $p \leq 0.05$.

<https://doi.org/10.1371/journal.pone.0263819.t003>

Table 4. Probability of melanoma diagnosis, related to the presence of RCM features.

RCM features	Presence/Absence							
	X	✓	X	X	✓	✓	X	✓
Sheet of cells	X	✓	X	X	✓	✓	X	✓
Nucleated roundish cells at DEJ	X	X	✓	X	✓	X	✓	✓
Peripheral hotspot at DEJ	X	X	X	✓	X	✓	✓	✓
Melanoma diagnosis probability	5.5%	31.5%	33.5%	37.5%	79.5%	82.4%	83.9%	97.5%

X = absent, ✓ = present.

<https://doi.org/10.1371/journal.pone.0263819.t004>

melanomas”, a term used in literature to describe melanomas without clinical and dermoscopy signs of malignancy, and melanomas originating from pre-existing nevi, which may have only focal atypia [9], might also benefit from RCM evaluation [10].

With these advantages, RCM can be used with clinical examination and dermoscopy to decrease the number of unnecessary excisions. The ratio of surgically excised benign lesions to reach a single melanoma diagnosis has been related to examination method, with 45:1 by clinical examination [7–10], 17:1 by dermoscopy [10], 4–7:1 by dermoscopy associated to digital monitoring [7–10], and 2:1 by RCM [11].

Some algorithms including RCM evaluation have been proposed in the literature to reach melanoma diagnosis and were related to different sensitivity (86.1% - 93%) and specificity (57–95.3%) [8, 12, 13]. However, the majority of these studies included several subtypes of melanomas, as invasive and amelanotic melanomas.

In our study, we decided to include only melanocytic lesions with few or faint dermoscopy features related to melanoma diagnosis and lacked enough criteria for benign lesion, termed here as “doubtful melanocytic lesions”, because they pose a very common and challenging scenario in clinical practice. Melanocytic lesions that had enough clinical and dermoscopy features to be classified with high probability of being benign or malignant were excluded because their management is already defined (follow-up or excision, respectively). As a result, our approach was highly effective to detect melanomas at their initial stage, with the majority of our melanomas identified as *in situ* (18 out of 26). After simple and multiple logistic regressions, three confocal features were statistically able to distinguish benign melanocytic lesions from melanomas.

The distribution of melanocytes forming aggregates is a frequently pattern found in melanocytic lesions. In benign melanocytic lesions, these aggregates are generally uniform spread throughout the lesion and constituted by non-atypical melanocytes [6]. In superficial spreading melanoma, the melanocyte aggregates are haphazardly distributed, with often highly variable shape and size, constituted by atypical melanocytes, tendency to confluence and located usually at the dermal epidermal junction and / or within the mid-portion and upper levels of the epidermis [14]. Our methods included the hotspot location to study the non-uniform distribution of these atypical melanocytes aggregates at the doubtful melanocytic lesions and also its degree of atypia.

The finding of a peripheral hotspot (atypical cells in 1mm²) at the DEJ was revealed as a specific confocal feature for melanoma diagnosis, however with low sensitivity. On the other hand, the presence of a central hotspot was not statistically more frequent in melanomas. Central hotspots were seen in 12 melanomas (46%) and in 22 (26%) benign melanocytic lesions, while peripheral hotspots were noted in six melanomas (23%) and not in any benign melanocytic lesions. The description of this RCM feature has not been previously reported in the literature.

The detection of atypical nucleated roundish cells at the DEJ have already been reported by other studies and related to 15 times greater risk for a lesion being malignant. The presence of

cells distributed in sheet-like structures disrupting the papillary architecture of the basal layer was also reported as highly specific, but with low sensitivity for melanoma diagnosis [7]. These findings were confirmed in our study: atypical nucleated roundish cells at the DEJ was present in 6 (23.08%) melanomas and in 2 (2.38%) benign lesions ($p = 0.048$, 95% CI = 1.01–47.96); and “sheet of cells” was present in 7 (26.92%) melanomas and in 2 (2.38%) benign lesions ($p = 0.04$, 95% CI = 1.09–42.35).

Conclusion

Our study is focused in RCM evaluation of doubtful melanocytic lesions and identified three confocal features statistically able to distinguish benign melanocytic lesions from melanomas. Among them, the finding of a peripheral hotspot (atypical cells in 1mm^2) at the DEJ is highlighted because has not been previously reported in the literature. Our study included a limited number of cases from a single institution and further research is still necessary to validate to importance and impact of this RCM feature to improve melanoma diagnosis accuracy.

Supporting information

S1 Table. Dermoscopy features of all lesions included at the study.
(XLSX)

S2 Table. Confocal microscopy features of all lesions included at the study.
(XLSX)

Author Contributions

Formal analysis: José Humberto Tavares Guerreiro Fregnani.

Methodology: Juliana Casagrande Tavoloni Braga, Mariana Petaccia de Macedo, Clóvis Antonio Lopes Pinto.

Supervision: Gisele Gargantini Rezze.

Writing – original draft: Raquel de Paula Ramos Castro.

References

1. Rajadhyaksha M., Grossman M., Esterowitz D., Webb R. H., and Anderson R. R. (1995). In vivo confocal scanning laser microscopy of human skin: melanin provides strong contrast. *J. Invest. Dermatol.* 104:946–52. <https://doi.org/10.1111/1523-1747.ep12606215> PMID: 7769264
2. Pehamberger H., Steiner A., and Wolff K. (1987). In vivo epiluminescence microscopy of pigmented skin lesions. Pattern analysis of pigmented skin lesions. *J. Am. Acad. Dermatol.* 17:571–83. [https://doi.org/10.1016/s0190-9622\(87\)70239-4](https://doi.org/10.1016/s0190-9622(87)70239-4) PMID: 3668002
3. Braga J. C., Macedo M. P., Pinto C., Duprat J., Begnami M. D., Pellacani G., et al. (2013). Learning reflectance confocal microscopy of melanocytic skin lesions through histopathologic transversal sections. *PLoS One* 8:e81205. <https://doi.org/10.1371/journal.pone.0081205> PMID: 24339910
4. Pellacani G., Farnetani F., Gonzalez S., Longo C., Cesinaro A. M., Casari A., et al. (2012). In vivo confocal microscopy for detection and grading of dysplastic nevi: a pilot study. *J. Am. Acad. Dermatol.* 66:e109–21. <https://doi.org/10.1016/j.jaad.2011.05.017> PMID: 21742408
5. Scope A. (2007). In vivo reflectance confocal microscopy imaging of melanocytic skin lesions: consensus terminology glossary and illustrative images. *J. Am. Acad. Dermatol.* 57:644–58. <https://doi.org/10.1016/j.jaad.2007.05.044> PMID: 17630045
6. Kutzner H., Metzler G., Argenyi Z., et al. (2012). Histological and genetic evidence for a variant of superficial spreading melanoma composed predominantly of large nests. *Mod Pathol.* 25:838–845. <https://doi.org/10.1038/modpathol.2012.35> PMID: 22388759

7. Pellacani G., Guitera P., Longo C., Avramidis M., Seidenari S., and Menzies S. (2007). The impact of in vivo reflectance confocal microscopy for the diagnostic accuracy of melanoma and equivocal melanocytic lesions. *J. Invest. Dermatol.* 127:2759–65. <https://doi.org/10.1038/sj.jid.5700993> PMID: 17657243
8. Segura S., Puig S., Carrera C., Palou J., and Malvehy J. (2009). Development of a two-step method for the diagnosis of melanoma by reflectance confocal microscopy. *J. Am. Acad. Dermatol.* 61:216–29. <https://doi.org/10.1016/j.jaad.2009.02.014> PMID: 19406506
9. Longo C., Rito C., Beretti F., Cesinaro A. M., Pineiro-Maceira J., Seidenari S., et al. (2011). De novo melanoma and melanoma arising from pre-existing nevus: in vivo morphologic differences as evaluated by confocal microscopy. *J. Am. Acad. Dermatol.* 65:604–14. <https://doi.org/10.1016/j.jaad.2010.10.035> PMID: 21715047
10. Longo C., Pellacani G. (2016). Melanomas. *Dermatologic Clinics.* 34(4):411–419. <https://doi.org/10.1016/j.det.2016.05.004> PMID: 27692447
11. Pellacani G., Pepe P., Casari A., and Longo C. (2014). Reflectance confocal microscopy as a second-level examination in skin oncology improves diagnostic accuracy and saves unnecessary excisions: a longitudinal prospective study. *Br. J. Dermatol.* 171:1044–51. <https://doi.org/10.1111/bjd.13148> PMID: 24891083
12. Pellacani G., Scope A., Gonzalez S., Guitera P., Farnetani F., Malvehy J., et al. (2019). Reflectance confocal microscopy made easy: The 4 must-know key features for the diagnosis of melanoma and nonmelanoma skin cancers. *J. Am. Acad. Dermatol.* 81:520–6. <https://doi.org/10.1016/j.jaad.2019.03.085> PMID: 30954581
13. Borsari S., Pampena R., Benati E., Bombonato C., Kyrgidis A., Moscarella E., et al. (2018). In vivo dermoscopic and confocal microscopy multistep algorithm to detect in situ melanomas. *Br. J. Dermatol.* 179:163–72. <https://doi.org/10.1111/bjd.16364> PMID: 29355898
14. Smoller B. (2006). Histologic criteria for diagnosing primary cutaneous malignant melanoma. *Mod Pathol.* 19(2):S34–40. <https://doi.org/10.1038/modpathol.3800508> PMID: 16446714