

Prion protein in Alzheimer's pathogenesis: a hot and controversial issue

Iryna Benilouva^{1,2}, Bart De Strooper^{1,2*}

Keywords: Amyloid-beta (A β); oligomers; A β receptor; prion protein (PrP); neurotoxicity

See related article in EMBO Mol Med (Calella AM et al (2010) EMBO Mol Med 2: 306–314)

The role for cellular prion protein PrP^c in β -amyloid (A β) oligomer-induced synaptic impairment is a topic of great interest and some controversy. In this issue of EMBO Molecular Medicine Aguzzi and co-workers explore the contribution of PrP^c to deficient long term potentiation (LTP) and soluble A β levels in an Alzheimer's disease mouse model and show that the role of prions in A β related toxicity is far from 'black and white' suggesting complex interpretations of the data available thus far.

Pathogenic amyloid formation is characteristic of several neurodegenerative disorders including Alzheimer's and Parkinson's disease, transmissible spongiform encephalopathies and others (Aguzzi & O'Connor, 2010). The prion diseases are propagated via conversion of the cellular prion protein PrP^c into an abnormal β -sheet enriched isoform PrP^{Sc} (Aguzzi & O'Connor, 2010). In Alzheimer's disease (AD), β - and γ -secretases cleave the amyloid precursor protein (APP), resulting in the generation of A β peptides that aggregate in β -sheet enriched A β fibrils (De Strooper, 2010)

and form the characteristic amyloid plaques in the brain of AD patients. Recent insights suggest that small oligomeric assemblies of A β , in contrast to monomeric and fibrillar species, are toxic for neuronal synapses, but the molecular targets of these assemblies and the mechanism of toxicity remain very controversial topics (Ashe & Zahs, 2010, see also supplemental data there). The main problem is that oligomeric A β assemblies are in a dynamic equilibrium with monomeric and fibrillar A β assemblies, implying that various biophysical parameters determine the relative abundance of different aggregation states. The dynamic nature of this process makes the definition of such toxic assemblies elusive and probably also explains why so many various direct and indirect interactions of A β peptides with membrane bound and intracellular proteins have been described (Ashe & Zahs, 2010).

One of the most spectacular candidates in the series of candidate receptors for these toxic assemblies is, without doubt, the prion protein (Lauren et al, 2009). Indeed, an interaction between A β and the prion protein suggests a potential common molecular substratum for the neurotoxicity seen in both diseases. Prion protein (PrP) was identified in an unbiased screening for receptors that could bind A β ₄₂ oligomers prepared according to a particular protocol to yield A β -derived diffusible ligands (ADDL) (Lambert et al, 1998). Such ADDLs are neurotoxic, interfere with LTP and are considered a more or less stable form

»» Calella et al have (...) investigated the potential role of PrP in A β neurotoxicity in a series of elegant genetic experiments... ««

among several toxic species along the A β aggregation pathway. The interactions between ADDLs and cellular PrP along with other A β binding molecules that might mediate AD pathogenesis are depicted in Figure 1. These oligomers failed to impair LTP in mouse hippocampal slices lacking PrP (Lauren et al, 2009) and the same authors have recently demonstrated that in an AD transgenic mouse model (APP^{swe}/Psen1 Δ E9) characterized by amyloid plaques formation and learning and memory deficits, deletion of the endogenous PrP gene prevented the development of the functional deficits despite unchanged levels of A β generation and A β deposition in their brains (Gimbel et al, 2010). The temptation to extrapolate these interesting findings towards real AD is obvious but it requires some caution as other researchers (Balducci et al, 2010) did not observe any protection in prion deficient animals with regard to acute memory impairments when injecting different A β oligomer preparations. Calella et al have now revisited this issue and investigated the potential role of PrP in A β neurotoxicity in an extensive series of elegant genetic experiments, crossing loss- and gain-of function PrP mouse strains with a transgenic AD mouse model

(1) Department for Molecular and Developmental Genetics, Flanders Institute for Biotechnology (VIB), Leuven, Belgium.

(2) Center for Human Genetics, KULeuven, Leuven, Belgium.

*Corresponding author: Tel: +3216346227;

Fax: +3216347181;

E-mail: bart.destrooper@med.kuleuven.be

DOI 10.1002/emmm.201000088

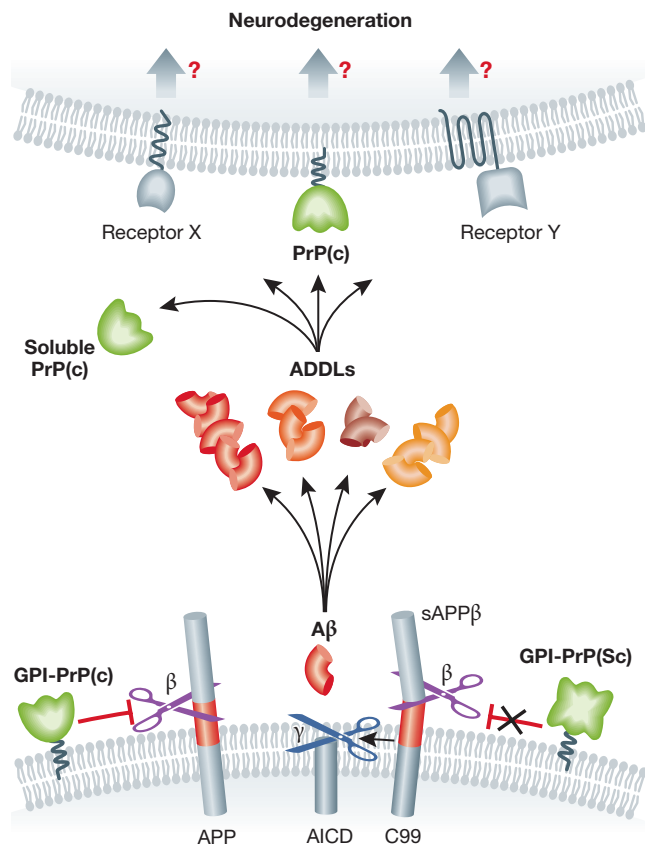


Figure 1. Cellular PrP in amyloid-beta induced neurodegeneration.

(APPKM670/671NL/Psen1L166P). These authors do not find any significant modulation of LTP formation by the presence or absence of PrP^c. The study intelligently explores any genetic confounders that could be blurring the effects and rules out the possibility that PrP^c is the direct mediator of the synaptotoxicity caused by Aβ in this model. They conclude, appealingly 'The hypothesis of PrP^c being a crucial mediator of Aβ synaptotoxicity might be not universal'.

» ...PrP has a remarkable good affinity for Aβ peptides... «

The story is however far from finished. All groups involved agree that PrP has a remarkable good affinity for Aβ peptides tested in various conformations (Balducci et al, 2010; Calella et al, 2010; Lauren et al, 2009). Calella et al investigated the effects

of expressing a soluble form of PrP (without its GPI-anchor) in their AD mouse model. In this case, whereas the levels of soluble and insoluble Aβ remain unchanged, LTP is less affected. Although seemingly contradictory with Lauren et al, the finding suggests that secreted PrP might interfere with Aβ mediated toxic pathways by directly binding to the peptide, not unlike the effect of Aβ antibodies in similar experiments. Whether such a protective effect is also observed with endogenously expressed (soluble) PrP remains obviously unaddressed.

» Other aspects of PrP and APP biology also suggest that the situation might be more complicated. «

Other aspects of PrP and APP biology also suggest that the situation might be more complicated. Parkin et al, 2007

showed for instance that cellular PrP can inhibit β-secretase-mediated cleavage of APP. The prediction that the lack of functional PrP^c would lead to a rise in Aβ levels was confirmed by analysing the brain of PrP knock out mice (Parkin et al, 2007). However, the PrP gene is located close to a quantitative trait locus (QTL) for Aβ levels (Ryman et al, 2008) and comparing Aβ levels between inbred wild type and knock out strains might still be confounded by such genes closely linked to the targeted locus. Calella et al (2010) demonstrate in their paper how such QTL can dramatically alter Aβ levels over various generations.

Finally, while highlighting the PrP^c interaction, Lauren et al (2009) have clearly shown that PrP^c is not the only cell-surface molecule binding Aβ oligomers, as a high level of Aβ binding signals was still observed in *Prnp*^{-/-} hippocampal neurons (50% compared to wild type). Furthermore, as discussed above, the in vivo generated Aβ oligomer pool is likely more complex than any in vitro generated Aβ oligomer mixture, and may therefore contain several 'strains' of toxic and less toxic conformers, somewhat resembling PrP^{Sc} (Aguzzi, 2008). Each of these conformations might act via different pathways. Therefore, and in conclusion, one cannot exclude that a remarkably high affinity of PrP to Aβ could be ascribed to a sub-pool of amyloid species, which is not necessarily the (most) toxic one.

The authors declare that they have no conflict of interest.

References

- Aguzzi A (2008) *Proc Natl Acad Sci USA* 105: 11-12
- Aguzzi A et al (2010) *Nat Rev Drug Discov* 9: 237-248
- Ashe KH et al (2010) *Neuron* 66: 631-645
- Balducci C et al (2010) *Proc Natl Acad Sci USA* 107: 2295-2300
- Calella AM et al (2010) *EMBO Mol Med* 2: 306-314
- De Strooper B (2010) *Physiol Rev* 90: 465-494
- Gimbel DA, et al (2010) *J Neurosci* 30: 6367-6374
- Lambert MP et al (1998) *Proc Natl Acad Sci USA* 95: 6448-6453
- Lauren J et al (2009) *Nature* 457: 1128-1132.
- Parkin ET et al (2007) *Proc Natl Acad Sci USA* 104: 11062-11067
- Ryman D et al (2008) *Neurobiol Aging* 29: 1190-1198