



## Molecular detection of *Bartonella rochalimae* and *Hepatozoon canis* in red foxes (*Vulpes vulpes*) from China

Shiyi Wang<sup>a</sup>, Nannan Cui<sup>a</sup>, Ziman Lv<sup>a</sup>, Nan Wang<sup>a</sup>, Gang Liu<sup>a</sup>, Shanshan Zhao<sup>a</sup>, Changqing Liu<sup>b</sup>, Yuanzhi Wang<sup>a,\*</sup>

<sup>a</sup> Key Laboratory for Prevention and Control of Emerging Infectious Diseases and Public Health Security, the XPCC, School of Medicine, Shihezi University, Shihezi, Xinjiang, Uygur Autonomous Region, China

<sup>b</sup> Forestry and grassland Resources Monitoring Center of Xinjiang Production and Construction Corps, Shihezi University, Shihezi City, Xinjiang, Uygur Autonomous Region, 832002, China

### ARTICLE INFO

#### Keywords:

*Bartonella rochalimae*  
*Hepatozoon canis*  
Red fox  
China

### ABSTRACT

Red foxes (*Vulpes vulpes*) have been recognized as natural reservoirs for multiple pathogens and a source of infection for domestic animals, wildlife and humans. To date, no reports are available on the *Bartonella rochalimae* and *Hepatozoon canis* infection in red foxes from China. In 2018–2022, a total of 16 red foxes were sampled in two counties and a city in Xinjiang Uygur Autonomous Region (XUAR) in northwest China. Subsequently analyzed by DNA extraction amplified by polymerase chain reaction (PCR). In the present study, based on nucleotide sequence and phylogenetic tree analyses, *B. rochalimae* and *H. canis* were molecularly identified in red foxes. Our findings provide the first molecular evidence of *B. rochalimae* and *H. canis* in red foxes from China.

### 1. Introduction

Canine vector-borne diseases (CVBD) are caused by numerous viral, bacterial and protozoal pathogens, which are transmitted by blood-sucking arthropods including ticks, fleas, mosquitos, and flies and pose considerable public health challenges in animals and humans globally (Otranto et al., 2009). As is well known, the majority of CVBD are closely associated with human population growth, global warming, illicit trade exchange, and habitat destruction, and these factors may have an impact on their prevalence and distribution (Kilpatrick et al., 2012; Wilke et al., 2021). However, there has been epidemiological research on CVBD in companion animals mainly focused on dogs (Hamel et al., 2012; Otranto and Dantas-Torres, 2010; Xu et al., 2015), data on the occurrence and prevalence of CVBD in red foxes in China remain scarce.

Recently, multiple fox species were reported to harbor vector-borne pathogens of public health concern, including spotted fever group rickettsiae (Dall'Agnol et al., 2018), *Borrelia burgdorferi* (Lledó et al., 2016), *Ehrlichia canis* (Fishman et al., 2004), *Leishmania infantum* (Davoust et al., 2014), and *Babesia vulpes* (Checa et al., 2018). In China, the red fox (*Vulpes vulpes*) can transmit classical zoonotic diseases, such as rabies (Yakovchits et al., 2021), echinococcosis (Jiang et al., 2012),

and rickettsioses (Liu et al., 2021). In this study, we aimed to investigate the molecular characterization of *Bartonella* and *Hepatozoon* in red foxes from China.

### 2. Materials and methods

In 2018–2022, a total of 16 red foxes (five adults and eleven pups) were found dead as a result of road kills and poaching, and subsequently sampled in two counties and a city (Manas County, Nilka County, and Alataw City) in Xinjiang Uygur Autonomous Region (XUAR) in northwest China. The red foxes were morphologically identified by an experienced zoologist. The study had formal ethical approval from Medical College of Shihezi University.

All fox organs, including the hearts, lungs, livers, spleens, and kidneys, were removed. The spleen DNA were extracted using the TIANamp Genomic DNA Kit (TIANGEN, Beijing, China). After the DNA concentration was qualified, and subsequent polymerase chain reaction (PCR) experiments were carried out. Two genes, *gltA* and *rpoB*, were used to detect *Bartonella*, whereas one gene, *18S rRNA*, was used to detect *Hepatozoon*. The primer sequences were as follows: *gltA*-F: 5'-GCTTTA-CAAAATTCTAAAAACCATATA-3', and *gltA*-R: 5'-TGTCTATCAATTCACAACCTGCGGT-3'. The PCR cycling conditions for *Bartonella* detection

\* Corresponding author.

E-mail address: [wangyuanzhi621@126.com](mailto:wangyuanzhi621@126.com) (Y. Wang).

<https://doi.org/10.1016/j.ijppaw.2024.100925>

Received 21 December 2023; Received in revised form 29 February 2024; Accepted 15 March 2024

Available online 16 March 2024

2213-2244/© 2024 The Authors. Published by Elsevier Ltd on behalf of Australian Society for Parasitology. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

consisted of an initial 5-min denaturation at 95 °C, followed by 35 cycles at 95 °C for 30 s, 52 °C for 30s, and 72 °C for 30s, with a final extension at 72 °C for 8 min. To confirm the positivity of this PCR, it was attempted to amplify an additional genetic marker, an approximately 330-bp-long fragment of the *rpoB* gene primers *rpoB-F*: 5'-GCTCTTGCAACTTC-TATGTT-3', and *rpoB-R*: 5'-CATTGTTCGTCAGGTTGGCG-3'. The PCR conditions were the same as in the *gltA*, except that the annealing temperature was 53 °C. The primer sequences of 18S rRNA: 18S rRNA-F5'-ATGGCTCATTACAACAGT-3', and 18S rRNA-R: 5'-AAGTTT-CAGCCTTGCGACCATACTC-3'. The PCR cycling conditions for *Hepatozoon* detection consisted of an initial 5-min denaturation at 95 °C, followed by 37 cycles at 95 °C for 40 s, 50 °C for 40s, and 72 °C for 50s, with a final extension at 72 °C for 8 min. The negative control was double distilled water, and the positive controls were *Bartonella elizabethae* DNA from fleas and *Hepatozoon* sp. DNA from great gerbils obtained in our previous works (Ji et al., 2021; Yin et al., 2019). Purification and sequencing of the PCR products were performed as described previously (Farkas et al., 2014).

Sequences were manually edited, aligned and compared with GenBank sequences (<https://blast.ncbi.nlm.nih.gov>). Two phylogenetic trees were constructed using the neighbor-joining method, bootstrap analyses 1000 replicates were conducted to determine the relative support for clades in the consensus trees in MEGA 7.0 software.

### 3. Results

Out of the 16 fox specimens collected from the XUAR, after molecular detection, *Bartonella* was found in two juvenile red foxes, whereas *Hepatozoon* was found in one adult red fox. Based on sequencing and phylogenetic analysis, the *Bartonella* that was identified in the red foxes was most closely related to *B. rochalimae*, which was from grey foxes (*Urocyon cinereoargenteus*) (GenBank accession No. FN645459), with a 99.40% (352/354) shared sequence identity in the *gltA* gene. Meanwhile, its *rpoB* gene was most closely related to *B. rochalimae* (DQ676489) from a dog sampled in USA, with 99.71% (345/346) shared sequence identity. The phylogenetic tree of the *Bartonella gltA-rpoB* concatenated sequences provide additional support for this conclusion (shown in Fig. 1). In addition, the *Hepatozoon* sp. identified in the red foxes was most closely related to *H. canis* from a golden jackal (*Canis aureus*) sampled in the Czech Republic (KX712124), with 98.55% (1085/1101) shared sequence identity in the 18S rRNA gene (shown in Fig. 2). All obtained sequences from this study were deposited in the GenBank (*B. rochalimae gltA*: OQ834668, *B. rochalimae rpoB*: OQ868002 and *H. canis 18S rRNA*: OQ859899).

### 4. Discussion

Wild animals are important indicators of vector-borne diseases. Red foxes live near ruminants (cattle, sheep, and goats), companion animals

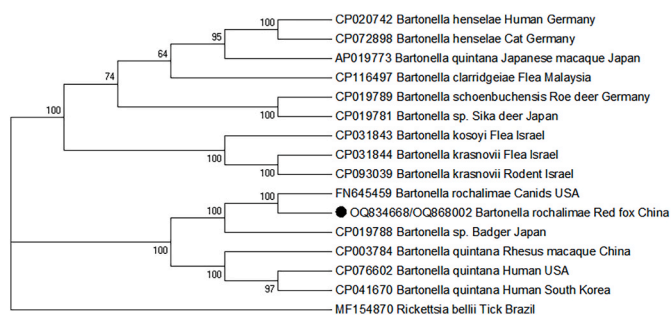


Fig. 1. The NJ phylogenetic tree of the *gltA-rpoB* concatenated sequences of *Bartonella* spp. (NJ; bootstrap replicates: 1000). Branch lengths correlate to the number of substitutions inferred according to the scale shown. Sequences of obtained in this study are indicated by solid cycle (●).

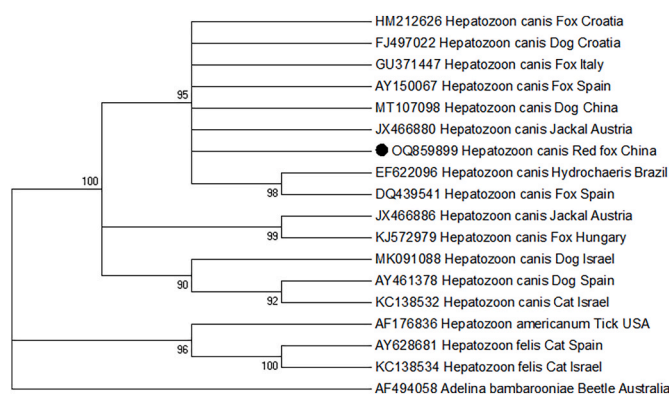


Fig. 2. The NJ phylogenetic tree of the 18S rRNA of *Hepatozoon* spp. (NJ; bootstrap replicates: 1000). Branch lengths correlate to the number of substitutions inferred according to the scale shown. Sequences of obtained in this study are indicated by solid cycle (●).

(dogs and cats), and wildlife (rodents), and in close proximity to humans. Red foxes usually display strong migratory behavior associated with habitat destruction, food loss, and climate change. Previously, red foxes were reported to harbor several vector-borne pathogens, including tick-borne encephalitis virus, *H. canis*, *Rickettsia raoultii*, *Rickettsia sibirica*, *Candidatus Rickettsia barbariae*, *Babesia vulpes*, *Babesia vogeli* and et al. (Wurm et al., 2000; Mierzejewska et al., 2021; Sang et al., 2021; Liu et al., 2021). In China and adjacent countries, some pathogens such as rabies virus, novel norovirus, *Echinococcus* spp., *Babesia* spp, and *H. canis*, were found in red foxes, corsac foxes, wolves and dogs (Wang et al., 2022; Altay et al., 2023; Xu et al., 2015). In this study, *B. rochalimae* and *H. canis* were detected for the first time in red foxes from China.

*Bartonella rochalimae*, a zoonotic agent, was first isolated from a patient who had a history of travel to Peru, had sustained multiple insect bites, and exhibited fever, rash, myalgia, anemia, and splenomegaly (Eremeeva et al., 2007). Subsequently, *B. rochalimae* was detected in many wild animals. In the canid red fox, *B. rochalimae* has been reported in France, Spain, and the USA (Greco et al., 2021). In this study, *B. rochalimae* were found in approximately 4-week-old juvenile red foxes in Manas County in XUAR, China. Juvenile mammals may be more susceptible to *Bartonella* transmitted through flea-biting than adult mammals, and this finding is consistent with a previous study (Chomel et al., 2006). In previous reports, *B. rochalimae* was vectored by *Pulex simulans*, *Paraceras melis*, *Xenopsylla gerbilli minax*, and *Xenopsylla conformis conformis* (Yin et al., 2019), implying that this pathogen may be vectored by flea species. Here, *B. rochalimae* was detected in red foxes in China for the first time.

*Hepatozoon canis*, a known veterinary parasite, has been mainly reported in canid animals, including the African wild dog (*Lycya pictus*), red fox (*V. vulpes*) crab-eating fox (*Cerdocoyon thous*), grey wolf (*Canis lupus*), golden jackal (*Canis aureus*), raccoon (*Procyon lotor*), and spotted hyena (*Crocota crocuta*) (Gabriel et al., 2009; Allen et al., 2011; Baneth, 2011; Gerrikagoitia et al., 2012). In China, *H. canis* was previously only reported in domestic dogs (Guo et al., 2020). In the present study, *H. canis* was detected in the red fox in China for the first time. In addition, as a tick-borne pathogen, *H. canis* has also been documented in several tick species, such as *Rhipicephalus sanguineus*, *Haemaphysalis* sp., and *Dermacentor reticulatus* (Giannelli et al., 2017; Mierzejewska et al., 2021; Najm et al., 2014). In our previous research, *Ixodes canisuga*, *Ixodes kaiseri*, *Haemaphysalis erinacei*, and *Dermacentor marginatus* were detected in red foxes (Liu et al., 2021). Further research is needed to determine the distribution of this two pathogens in domestic dogs and wild canine, and it is also important to identify the transmission route.

## 5. Conclusion

To our best knowledge, this is the first study to report the molecular characterization of *B. rochalimae* and *H. canis* in wild red foxes from China. This work has particular importance for the continued and extended surveillance of wild red foxes as potential sources of canine vector-borne pathogens.

## Authors' contributions

SW, NC and ZL conceived and designed the study, and wrote the manuscript. NW, GL, SZ and CL performed the experiments and analyzed the data. SW and YW contributed to study design and edited the manuscript. All authors have read and agreed to the published version of the manuscript

## Funding

The authors thank the Forestry and grassland resources monitoring center of Xinjiang Production and Construction Corps, Shihezi University, for contributions. This work was supported in part by the Natural Science Foundation of China (822260399 and 822260410), the National Key Research & Development Program of China (2022YFC2304000), Scientific research project of Shihezi University (CXPY202104).

## Data availability

DNA sequences were deposited in GenBank; accession numbers: OQ834668, OQ868002, OQ859899.

## Consent to participate

Informed verbal and written consent were obtained from each study participant.

## Ethical approval

All procedures performed in this study involving wild animals were in accordance with the ethical standards of the Animal Ethics Committee of Shihezi University (Approval No. AECSU2015-11).

## Declaration of competing interest

The authors declare that they have no conflict of interest.

## Acknowledgments

The author would like to thank all the veterinarians who participated in the study as well as all the colleagues who contributed to sample collecting and sample preparation.

## References

- Allen, K.E., Johnson, E.M., Little, S.E., 2011. *Hepatozoon* spp infections in the United States. *Vet Clin North Am Small Anim Pract* 41, 1221–1238.
- Altay, K., Erol, U., Sahin, O.F., Aydin, M.F., Aytmirzakizi, A., Dumanli, N., 2023. First molecular evidence of *Babesia vogeli*, *Babesia vulpes*, and *Theileria ovis* in dogs from Kyrgyzstan. *Pathogens* 15, 1046.
- Baneth, G., 2011. Perspectives on canine and feline hepatozoonosis. *Clin. Microbiol. Rev.* 181, 3–11.
- Checa, R., López-Beceiro, A.M., Montoya, A., Barrera, J.P., Ortega, N., Gálvez, R., Marino, V., González, J., Olmeda, Á.S., 2018. *Babesia microti*-like piroplasm (syn. *Babesia vulpes*) infection in red foxes (*Vulpes vulpes*) in NW Spain (Galicia) and its relationship with *Ixodes hexagonus*. *Vet. Parasitol.* 252, 22–28.
- Chomel, B.B., Boulouis, H.J., Maruyama, S., Breitschwerdt, E.B., 2006. *Bartonella* spp. in pets and effect on human health. *Emerg. Infect. Dis.* 12, 389–394.
- Dall'Agno, B., Souza, U.A., Weck, B., Trigo, T.C., Jardim, M.M.A., Costa, F.B., Labruna, M.B., Peters, F.B., Favari, M.O., Mazim, F.D., 2018. *Rickettsia parkeri* in

- free-ranging wild canids from Brazilian Pampa. *Transbound Emerg Dis* 65, e224–e230.
- Davoust, B., Mary, C., Marié, J.L., 2014. Detection of Leishmania in red foxes (*Vulpes vulpes*) from southeastern France using real-time quantitative PCR. *J. Wildl. Dis.* 50, 130–132.
- Eremeeva, M.E., Gerns, H.L., Lydy, S.L., Goo, J.S., Ryan, E.T., Mathew, S.S., Ferraro, M. J., Holden, J.M., Nicholson, W.L., 2007. Bacteremia, fever, and splenomegaly caused by a newly recognized *bartonella* species. *N. Engl. J. Med.* 356, 2381–2387.
- Farkas, R., Solymosi, N., Takács, N., Hornyák, Á., Hornok, S., Nachum-Biala, Y., Baneth, G., 2014. First molecular evidence of *Hepatozoon canis* infection in red foxes and golden jackals from Hungary. *Parasites Vectors* 7, 303.
- Fishman, Z., Gonen, L., Harrus, S., Strauss-Ayal, D., King, R., Baneth, G., 2004. A serosurvey of *Hepatozoon canis* and *Ehrlichia canis* antibodies in wild red foxes (*Vulpes vulpes*) from Israel. *Vet. Parasitol.* 119, 21–26.
- Gabriel, M.W., Henn, J., Foley, J.E., Brown, R.N., Kasten, R.W., Foley, P., Chomel, B.B., 2009. Zoonotic *Bartonella* species in fleas collected on gray foxes (*Urocyon cinereoargenteus*). *Vector Borne Zoonotic Dis.* 9, 597–602.
- Gerrikagoitia, X., Gil, H., García-Esteban, C., Anda, P., Juste, R.A., Barral, M., 2012. Presence of *Bartonella* species in wild carnivores of northern Spain. *Appl. Environ. Microbiol.* 78, 885–888.
- Giannelli, A., Lia, R.P., Annoscia, G., Buonavoglia, C., Lorusso, E., Dantas-Torres, F., Baneth, G., Otranto, D., 2017. *Rhipicephalus turanicus*, a new vector of *Hepatozoon canis*. *Parasitology* 144, 730–737.
- Greco, G., Zarea, A.A.K., Sgroi, G., Tempesta, M., D'Alessio, N., Lanave, G., Bezerra-Santos, M.A., Iatta, R., Veneziano, V., 2021. Zoonotic *Bartonella* species in Eurasian wolves and other free-ranging wild mammals from Italy. *Zoonoses Public Health* 68, 316–326.
- Guo, W.P., Xie, G.C., Xue, Z.Q., Yu, J.J., Jian, R., Du, L.Y., Li, Y.N., 2020. Molecular detection of *Hepatozoon canis* in dogs and ticks in Shaanxi province, China. *Comp. Immunol. Microbiol. Infect. Dis.* 72, 101514.
- Hamel, D., Silaghi, C., Lescai, D., Pfister, K., 2012. Epidemiological aspects on vector-borne infections in stray and pet dogs from Romania and Hungary with focus on *Babesia* spp. *Parasitol. Res.* 110, 1537–1545.
- Ji, N., Chen, X., Liu, G., Zhao, S., Tan, W., Liu, G., Zhang, J., Wang, Y., 2021. *Theileria*, *Hepatozoon* and *Taenia* infection in great gerbils (*Rhombomys opimus*) in northwestern China. *Int J Parasitol Parasites Wildl* 15, 79–86.
- Jiang, W., Liu, N., Zhang, G., Renqing, P., Xie, F., Li, T., Wang, Z., Wang, X., 2012. Specific detection of *Echinococcus* spp. from the Tibetan fox (*Vulpes ferrilata*) and the red fox (*V. vulpes*) using copro-DNA PCR analysis. *Parasitol. Res.* 111, 1531–1539.
- Kilpatrick, A.M., Randolph, S.E., 2012. Drivers, dynamics, and control of emerging vector-borne zoonotic diseases. *Lancet (London, England)* 380, 1946–1955.
- Liu, G., Zhao, S., Tan, W., Hornok, S., Yuan, W., Mi, L., Wang, S., Liu, Z., Zhang, Y., Hazihan, W., Gu, X., Wang, Y., 2021. Rickettsiae in red fox (*Vulpes vulpes*), marbled polecat (*Vormela peregusna*) and their ticks in northwestern China. *Parasites Vectors* 14, 204.
- Lledó, L., Serrano, J.L., Isabel Gegúndez, M., Giménez-Pardo, C., Saz, J.V., 2016. Antibodies to *Rickettsia* spp. and *Borrelia burgdorferi* in Spanish wild red foxes (*Vulpes vulpes*). *J. Wildl. Dis.* 52, 122–125.
- Mierzejewska, E.J., Dwuznik, D., Koczwarska, J., Stańczak, Ł., Opalińska, P., Krokowska-Paluszak, M., Wierzbicka, A., Górecki, G., Bajer, A., 2021. The red fox (*Vulpes vulpes*), a possible reservoir of *Babesia vulpes*, *B. canis* and *Hepatozoon canis* and its association with the tick *Dermacentor reticulatus* occurrence. *Ticks Tick Borne Dis.* 12, 101551.
- Najm, N.A., Meyer-Kayser, E., Hoffmann, L., Pfister, K., Silaghi, C., 2014. *Hepatozoon canis* in German red foxes (*Vulpes vulpes*) and their ticks: molecular characterization and the phylogenetic relationship to other *Hepatozoon* spp. *Parasitol. Res.* 113, 2679–2685.
- Otranto, D., Dantas-Torres, F., 2010. Canine and feline vector-borne diseases in Italy: current situation and perspectives. *Parasites Vectors* 3, 2.
- Otranto, D., Dantas-Torres, F., Breitschwerdt, E.B., 2009. Managing canine vector-borne diseases of zoonotic concern: part one. *Trends Parasitol.* 25, 157–163.
- Sang, C., Yang, Y., Dong, Q., Xu, B., Liu, G., Hornok, S., Liu, Z., Wang, Y., Hazihan, W., 2021. Molecular survey of *Babesia* spp. in red foxes (*Vulpes vulpes*), Asian badgers (*Meles leucurus*) and their ticks in China. *Ticks Tick Borne Dis.* 12, 101710.
- Wang, J., Li, L., Xu, Y., Mao, T., Ma, Y., Sun, X., Liu, X., Wang, Y., Duan, Z., 2022. Identification of a novel norovirus species in fox. *Infect. Genet. Evol.* 98, 105214.
- Wilke, A.B.B., Benelli, G., Beier, J.C., 2021. Anthropogenic changes and associated impacts on vector-borne diseases. *Trends Parasitol.* 37, 1027–1030.
- Wurm, R., Dobler, G., Peters, M., Kiessig, S.T., 2000. Serological investigations of red foxes (*Vulpes vulpes* L.) for determination of the spread of tick-borne encephalitis in Northrhine-Westphalia. *J. Vet Med B Infect Dis Vet Public Health* 47, 503–509.
- Xu, D., Zhang, J., Shi, Z., Song, C., Zheng, X., Zhang, Y., Hao, Y., Dong, H., Wei, L., El-Mahallawy, H.S., Kelly, P., Xiong, W., Wang, H., Li, J., Zhang, X., Gu, J., Wang, C., 2015. Molecular detection of vector-borne agents in dogs from ten provinces of China. *Parasites Vectors* 8, 501.
- Yakovchits, N.V., Adelshin, R.V., Zarva, I.D., Chupin, S.A., Melnikova, O.V., Andae, E.I., Shulpin, M.I., Metlin, A.E., Botvinkin, A.D., 2021. Fox rabies outbreaks in the republic of Buryatia: Connections with neighbouring areas of Russia, Mongolia and China. *Transbound Emerg Dis* 68, 427–434.
- Yin, X., Zhao, S., Yan, B., Tian, Y., Ba, T., Zhang, J., Wang, Y., 2019. *Bartonella rochalimae*, *B. grahamii*, *B. elizabethae*, and *Wolbachia* spp. in fleas from wild rodents near the China-Kazakhstan Border. *Kor. J. Parasitol.* 57, 553–559.