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Environmental and Endogenous Acids Can Trigger Allergic-Type Airway Reactions

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Abstract: Inflammatory allergic and nonallergic respiratory disorders are spreading worldwide and often coexist. The root cause is not clear. This review demonstrates that, from a biochemical point of view, it is ascribable to protons (H^+) released into cells by exogenous and endogenous acids. The hypothesis of acids as the common cause stems from two considerations: (a) it has long been known that exogenous acids present in air pollutants can induce the irritation of epithelial surfaces, particularly the airways, inflammation, and bronchospasm; (b) according to recent articles, endogenous acids, generated in cells by phospholipases, play a key role in the biochemical mechanisms of initiation and progression of allergic-type reactions. Therefore, the intracellular acidification and consequent Ca^{2+} increase, induced by protons generated by either acid pollutants or endogenous phospholipases, may constitute the basic mechanism of the multimorbidity of these disorders, and environmental acidity may contribute to their spread.

Keywords: atmospheric acidity; air pollution; allergic reactions; mechanisms of allergy; allergic rhinitis; asthma; chronic; allergic multimorbidity; nonallergic; pseudo-allergic

1. Introduction

Inflammation and hypersensitivity of the airways and epidermis, whether allergic or nonallergic, acute or chronic, are pandemic illnesses and epidemiological studies show that they are growing faster in developing countries [1]. The increase has been attributed to several factors, both genetic [2] and environmental [3–5]; this work focuses on the latter.

Environmental factors that can affect the aetiology of these diseases, such as lifestyle, climate change, and air contaminants, have long been the subject of study and debate the world over [3–9]. The World Health Organization (WHO) has provided recommendations on how to reduce air pollution produced by household activities, one of these being to properly ventilate the home [9]. This is useful in rural areas but not in cities or industrial areas, where the outside air is often more polluted than the air indoors. Consequently, today, inflammatory allergic and nonallergic (also known as pseudoallergic) diseases are more widespread in urban than rural areas [10]. Authoritative research confirms that the higher prevalence in urban areas correlates with some outdoor air pollutants [3,10–13]. Immunological effects can be observed in both the upper and lower respiratory tract after exposure to diseal exhaust, and the short-term exposure to traffic-related nitrogen dioxide (NO₂), an acidic gas, has a direct effect on respiratory morbidity [13]. Furthermore, a relationship between allergic infant sensitization has been demonstrated [14]. The MeDALL (Mechanisms of the Development of Allergy) European study confirmed the relevance of environmental exposure [15,16]. Wherever possible, prevention by allergen avoidance remains the first measure [17]. Recent research has provided new data and technologies

for therapeutic improvements [18]. However, further studies are needed to discover the molecular determinants and to clarify the basic onset mechanisms of allergic and nonallergic diseases [17,19].

In addition to the relevance of environmental exposures, the MeDALL study highlighted that air pollution not only correlates with bronchitis, rhinitis, asthma, and even eczema [10–14], but these diseases often co-exist and share causal mechanisms [15,16,20,21].

While the mechanism of allergic response has been extensively studied and remains mainly an IgE/Fc&RI-based individual hypersensitivity reaction to specific allergens [1,6], there is no fully convincing biochemical explanation of the nonallergic response and the relationship between increasing allergic and nonallergic hypersensitivities, their multimorbidity, and air pollutants. IgE sensitization can no longer be considered the dominant causal mechanism of multimorbidity of such diseases [15,16,20], because allergic symptoms exist even in the absence of positive IgE tests. For these non-IgE-associated diseases, it is necessary to hypothesize other mechanisms, which should be investigated [6,16,20]. Some studies proved that this is in part attributable to genetic predisposition.

Regarding the consequences of environmental pollution, many studies have analyzed the toxic effects induced by air pollutants, in particular oxidative [22] and nitrosative [3,11] stress, and the causal relationship with allergies. Studies of acid stress began in the 1980s [23,24], without investigating the correlation between extra- and intracellular acidity. Acids can cause stress because they lower the physiological pH by the release of protons (H⁺).

The aim of this review is to highlight the chemistry of atmospheric acid pollutants, their irritating effects on the airways, and the existence of a possibly shared causal, proton-based mechanism, induced by both exogenous and endogenous acids, for the onset and spread of allergic and nonallergic inflammatory reactions.

Scientific literature available online from 1970–April 2020 was taken into consideration. The main databases, such as Embase, Medline, PubMed Central, Scopus, Web of Science, were searched and the most cited and most recent papers were selected. We analyzed the data and critically evaluated the fundamental biochemical concepts concerning the topic under study and their possible consequences on a cellular level.

2. Results

2.1. Outdoor Acid Air Pollutants: Chemical and Toxicological Characteristics

Polluting atmospheric acids damage surface water, buildings, and living organisms, either by direct reactions or through acid rain. Epidemiological studies on acute respiratory effects show that fine particulate matter (PM_{2.5}) and gaseous acid pollutants can have a major impact on the airways [8,11,25,26], because of their significant toxic potential. Given their small size, fine particles are able to penetrate deeply and reach the lower respiratory airways [13,27]. Furthermore, as a result of their low polarity and high liposolubility, the gases can spread quickly through biological membranes [13,28] and hence enter cells. Among these gases, NO and O₃ are known to cause nitrosative and oxidative stress, respectively. Recent studies have drawn attention to the health impacts of PM [13,14,26,27,29], NO₂ [3,11,13,14,25,30], and SO₂ [31–33].

It is known that PM from anthropogenic sources, such as heating systems, industrial plants, and motor vehicles, is mainly acid, since PM is associated with the anthropogenic acid pollutants NO₂ and SO₂ [11,13]. In addition, NO₂ and SO₂ can react with water and oxygen to give the corresponding acids: nitric acid (HNO₃), sulphurous acid (H₂SO₃), sulphuric acid (H₂SO₄), and their related acidic salts. The toxicity of acid compounds is mainly due to their ability to release protons (H⁺). Both HNO₃ and H₂SO₄ are strong acids, important sources of protons, and therefore are fiercely corrosive. While, at normal temperature and pressure, NO₂ and SO₂ are gases, the corresponding acids HNO₃, H₂SO₃, and H₂SO₄ are liquids, and easily soluble in water. Their acidic salts are water-soluble as well. Therefore, most air acidity is concentrated in the microscopic PM, suspended in the air itself, in the form of both

moist solid particles and watery droplets, known as acid aerosols. Notably, for its smaller particle size and its larger specific surface area, $PM_{2.5}$ is richer than PM_{10} in water and acids.

Around the year 1985, interest in the effects of acid aerosols increased as a result of the risk of high exposure levels in the US and Canada. Clinical studies were carried out to assess the toxicity of some atmospheric pollutants. The results showed that:

- (a) The bronchoconstrictor action of carbachol could be enhanced by acid sulphate aerosols [23,34], even though the sulphate is not itself toxic [34];
- (b) The biologically active portion of these compounds is H⁺ rather than sulphate and the potency is proportional to their acidity [34];
- (c) Titratable acidity appears to be a more important stimulus to bronchoconstriction than pH [35].

Consistently, it was shown that bronchoconstriction provoked by inhalation of sodium sulphite aerosols was caused by the released gaseous SO₂, or by bisulphite, but not by sulphite [35]. Combined exposures to acidic aerosols and pollutant gases have synergic effects.

The abovementioned PM, HNO₃, H_2SO_3 , and H_2SO_4 are the strongest acid components of acid aerosols. In addition, some other weaker acids are present, including carbonic acid, nitrous acid, and hydrogen sulphide, which essentially contribute to titratable acidity. All acids can contribute to the effects of total air acidity by releasing H^+s to different extents.

2.2. Biochemical Effects of Cellular Acidification in Epithelial Tissues

It has been shown that exogenous acids can cause irritation and the bronchoconstriction of the airways [23,24,34–36] in both asthmatic [24] and healthy subjects [36]. Moreover, they can stimulate both immune cells (mast cells [37], neutrophils [38–40], dendritic cells [41], eosinophils [42], Jurkat cells, and primary T cells [43–45]) and nonimmune cells (epithelial cells [46,47], fibroblasts [20], and smooth airway muscle cells [48–50]). It is reasonable to assume that the effects of limited exposure by healthy subjects are negligible, because air acidity can be entirely neutralized within a short time by the buffering capacity of airway surface liquid (ASL) [51]. On the contrary, major exposure, or for sensitive people even limited exposure, can overcome the ASL defense, giving rise to the transfer of H⁺ into cells as described below.

Regarding endogenous acids, it should be remembered that cells use intracellular enzymes such as phospholipase C (PLC) and messengers such as inositol 1,4,5-trisphosphate (IP₃) to increase the free Ca²⁺ concentration in cytosol ((Ca²⁺)_c). PLC and other phospholipases are powerful acidifying enzymes, because one H⁺ is released for each hydrolyzed ester bond [52–54]. The hydrolysis of phospholipid esters and the generation of endogenous acid molecules, such as arachidonic acid (AA), phosphatidic acid, and IP₃, are at the base of the production of allergic mediators. It is known that external acidification can cause mobilization of the segregated Ca²⁺ from intracellular stores [38,39,46,47,53,55–59], because protons can readily replace Ca²⁺ in its ligand locations [53,60–62]. Moreover, it is known that the increase of [Ca²⁺]_c is involved in many physiological processes [63–67], but also in the triggering of pathological manifestations such as allergic responses [65], airway hyper-responsiveness (AHR) [66], and abnormal contraction and remodeling of airway smooth muscle (ASM) [67].

The two paragraphs below give a more detailed description of the biochemical mechanisms by which intra- and extracellular acidification take place and foster allergic reactions.

2.3. Intracellular H⁺: Intracellular Acidification May Be Caused by the Action of Phospholipases in the Cytosol or by Protons Entering the Cell through the Plasma Membrane

The cells responsible for triggering the allergic response, such as mast cells [6,65,68-70] and basophils [6,69-71], have numerous receptors sensitive to various agonists. These receptors can be classified historically as IgE-dependent and non-IgE-dependent receptors, based on their positive or negative response to immunoglobulins E (IgE) [72]. The best example of an IgE-dependent receptor is the high-affinity IgE receptor (FccRI) [73,74]. Non-IgE-dependent receptors include the recently

discovered G-Protein-Coupled Receptors (GPCR), which respond to less specific agonists [69,71,75–78]. Furthermore, in the GPCR group are Mas-related G-protein coupled receptors (MRGR) [72,76], protease-activated receptors (PAR) [79], and purinergic receptors [80]. Given the sheer number of GPCRs, many combinations with different agonists are possible. For example, GPR4, GPR65, GPR68, and GPR132 receptors may be activated by extracellular protons [48,49,55,81,82]. Alternatively, muscarinic agonists may stimulate the G α q/11 subunits of the acetylcholine GPCRs [48]. So-called pseudoallergic agents also follow this route [83]. All individuals can respond via GPCR receptors to the stimulus of the agonist, but only sensitized individuals (the "truly" allergic) can respond via IgE/FccRI.

Contact between the agonist and the receptor triggers a PLC/IP₃-pathway-type complex chain reaction which, via the activation of numerous enzymes and the increase in the concentration of H⁺ and cytosolic Ca²⁺ (respectively (H⁺)_c and (Ca²⁺)_c), culminates with degranulation, by the exocytotic secretion of allergic mediators and the onset of an acute allergic response. The responses of the various agonist/receptor couples may differ [69,77,84], but depend in each case on the concentration and affinity of the agonist [74], and the fundamental steps in the basic biochemical mechanism of allergic reactions do not vary (Figure 1):

- (a) The stimulation of the receptor, both of the FcεRI and GPCR types, activates phospholipase C (PLC) [85–88] and hence the hydrolysis of phosphatidylinositol 4,5-biphosphate (PIP₂) on the inner wall of the plasma membrane, generating and releasing IP₃, a protonated acid salt [62,89], in the cytosol;
- (b) Through dissociation, the IP₃ releases H⁺ [62,89] and, via its IP₃R receptor, induces cell calcium release and store depletion, increasing (Ca²⁺)_c [62,90];
- (c) The increase in (Ca²⁺)_c activates numerous calcium-dependent enzymes, including phospholipase A₂ (PLA₂), which produces arachidonic acid (AA) [91,92], which in turn dissociates releasing more H⁺ and inducing the release of more Ca²⁺ [56,58,93]; from the AA hundreds of derivatives (eicosanoids cascade) are formed, including leukotrienes (LTs) and prostaglandins (PGs) [94,95]. Both leukotrienes and prostaglandins are known to play a pivotal role in inflammatory and allergic reactions;
- (d) The store depletion stimulates the entry of more Ca²⁺ from the extracellular space (calcium influx) via the mechanism known as Store Operated Calcium Entry (SOCE), in which, from the surface of the Endoplasmic Reticulum (ER), Stromal Interaction Molecule1 (STIM1) activates the opening of ORAI1 and Transient Receptor Potential Cation Canonical (TRPC) [96–99] channels on the plasma membrane;
- (e) The calcium influx further stimulates PLA₂ activity and fosters the maturing of the granules and subsequent degranulation and release [100–103] of mediators [94,104,105], including histamine, PGs, LTs, cytokine, tryptase, and chymase, which promote the acute phase of allergic inflammation. The cysteinyl LTs are thought to be responsible for the increase in the basal tone of the ASM and in bronchoconstriction in asthma [6,106].

In conclusion, as shown in Figure 1, the allergic and nonallergic responses differ only in the first step of agonist stimulation, which leads to PLC activation. The subsequent PLC/IP₃ pathway is the same for both responses and is characterized by the generation of acids, such as IP₃ and AA, and thus, H⁺ release by acid dissociation. Figure 1 shows two different steps in the intracellular generation of H⁺ through the action of phospholipase, the first dependent on the IP₃ produced by PLC, the second on the AA produced by PLA₂. A third step, not shown in Figure 1, can depend on the action of phosphatases which dephosphorilate the IP₃ on IP₃R [62]. Each of these three steps gives rise to a rapid transient increase in (H⁺)_c. This increase, as a result of the protons derived from the IP₃ and the AA, contributes to cell store depletion/calcium release and to the subsequent Ca²⁺ influx, via the activation of SOCE, with a consequent increase in (Ca²⁺)_c [39,47,56–59,62,107]. The rise in (H⁺)_c is transient because it is subject to feedback control and can be rapidly neutralized by the buffering capacity of cytosol and the calcium influx itself, which leads to the alkalization of the cytosol, because the extracellular pH

outweighs the intracellular pH. The influx may also be induced by a mechanism other than SOCE and independently of the reserves, known as Store-Independent Calcium Entry (SICE), by direct activation, via STIM1 and ORAI, due to the AA or LTs [108].



Figure 1. Basic biochemical mechanism of allergic-type response; PLC—Phospholipase C; PLA₂—Phospholipase A₂; AA—Arachidonic Acid; SICE—Store-Independent Calcium Entry; PGs—Prostaglandins; LTs—Leukotrienes; ASL—Airway Surface Liquid.

In addition to being generated in the cytosol by the phospholipases as described above, H^+ can penetrate the cell directly [28,38,46,109–114], passing through the epithelial barrier and plasma membrane, thanks to the acid loaders (Figure 2A). This is possible because the permeability of the epithelial barrier can vary as a result of the stimuli received from the cellular receptors [115], or the barrier itself may be destroyed [116,117]. Examples of acid loaders are the Cl⁻/HCO₃⁻ exchangers of the SLC4 and SLC26 type, [117–120] and the Na⁺-HCO₃⁻ cotransporter of the SLC4 type [118,120], which are chemically equivalent to a counterflux of H⁺ ions, the plasma membrane Ca²⁺ ATPase pump (PMCA), which exchanges H⁺ for Ca²⁺ [121], acid-sensing ion channels (ASICs) [53,82,122], ORAI [123], and some types of TRP channels [109–112]. Furthermore, H⁺ can be released into the cell after entry by passive transfer of reactive oxides coming from atmospheric pollution, such as CO₂, NO₂, and SO₂. CO₂ can react with water to release H⁺ much quicker than NO₂ and SO₂ due to the specific ubiquitous catalyst Carbonic Anhydrase (CA) [124]. Therefore, the CO₂/CA system is possibly an excellent means of transport for H⁺, as some researchers believe [38,120,124]. Lastly, the extracellular excess of protons may enter the cell by diffusion [125].



Figure 2. Biochemical routes for variation in pH of the Airway Surface Liquid (ASL). (**A**) Acid loaders: Cl⁻/HCO₃⁻ exchanger; Na⁺- HCO₃⁻ cotransporter; PMCA pump; TRP, ORAI and ASIC channels; CO₂/Carbonic Anhydrase system, etc. (**B**) Acid extruders: NHE exchanger; Hv1, TRP, and CFTR channels; ATPase pumps, etc. PLA₂—Phospholipase A₂; PLC—Phospholipase C; IP3—Inositol 1,4,5-trisphosphate; AA—Arachidonic Acid.

2.4. Extracellular H⁺: The Acidification of the Surfaces of the Respiratory Airways May Be Due to Environmental Acid Pollutants or Endogenous Acids

The outer surface of the epithelia of respiratory pathways is kept moist at all times by ASL. ASL plays a key role in the defense of the airways from pathogens and contains some phagocytes and a number of proteic and peptidic antimicrobials for this purpose. For optimum antimicrobial activity, both in the nose and lungs, ASL pH should be maintained within slightly acidic physiological values (circa pH = 6.80) [24,126,127] and a lowering can be counterproductive [128]. Interestingly, a decrease in ASL pH after exposure to airborne traffic pollutants has been detected in asthmatic [128] and healthy subjects [129], albeit to different extents.

Historically, endogenous acidification of the airway surfaces has been suggested as a way to measure airway disease [126]. In 2000, Hunt et al. found that the pH of Exhaled Breath Condensate (EBC) was over two log orders lower in patients with acute asthma than in healthy subjects. Hence, they suggested a possible causal relationship between endogenous airway acidification and the airflow limitation observed in acute asthma [130]. Similarly, in more recent years, low pH values of ASL have been observed by other authors in asthma, allergic rhinitis, atopic dermatitis [131], and even in nonallergic inflammatory diseases, such as bronchiectasis and Chronic Obstructive Pulmonary Disease (COPD) [117].

Figure 2B shows the possible causes of the lowering of ASL pH depending on endogenous or exogenous acidity. Extracellular and ASL acidification may be caused in four different ways:

- (a) H⁺ derived from the physiological process of restoring prestimulus conditions, carried out by all cells through the expulsion of excess protons, generated by acidifying enzymes, to return to the steady state; cells can use acid extruders as exchangers and channels to transfer H⁺ externally; the Na⁺-H⁺ exchanger (NHE) in some cells is the major acid-extruder, also the Cystic fibrosis transmembrane conductance regulator (CFTR) plays an important role in the acidification of the ASL [117]; in addition, the excess protons in the cytosol may exit the cell via voltage-gated proton channels (Hv1), TRP channels, plasma membrane vacuolar V-type H⁺-ATPase [126,132–136], and diffusion [125];
- (b) The degranulation of phagocytes, such as macrophages and granulocyte neutrophils and eosinophils [69,135,137,138], produced as a defensive inflammatory action [24,126] in response to the stimulus. This acidifying action may be significant and long lasting, and is therefore the basis for chronic disease;
- (c) The degranulation of mast cells and basophils, caused by the stimulus, the basis of the acute allergic response [77,78,80,84,138], as described above in Figure 1. It is known that, like phagocytes, basophils and mast cells [138] can produce and secrete acids and phospholipolytic enzymes with the contents of their cytosolic granules and vesicles. Examples of secreted acids are lactic, hypochlorous, uric, phosphoric acid, and fatty acids. Examples of enzymes are the cytosolic and secretory phospholipases A2, which produce fatty acids such as AA through hydrolysis of cellular triglycerides and phospholipids [139]. Each of the secreted acids can contribute to the release of protons and thus act as new stimuli for cellular responses;
- (d) In addition to the endogenous acids described above in point a, band c, which are transferred by the cells to the ASL by means of expulsion, extrusion, and/or degranulation, the acidification of the ASL may be due to exogenous acids, and hence, possibly, to the presence and direct action of atmospheric acid pollutants.

All four processes of acidification described in points a–d, and in Figure 2B, can contribute separately or simultaneously to lowering ASL pH. It is known that protons are allosteric modulators and protein structure modifiers [140]. The harmful consequences of the lowering of ASL pH, caused by either exogenous or endogenous acids, can include an increase in mucus viscosity, a decrease of ciliary beat frequency, recruitment of proinflammatory mediators, oedema, leukocyte infiltration, AHR,

tissue remodeling, and damage [24,40,50,136]. These consequences foster the origin and development of acute and chronic diseases of an allergic kind [24,37,40,66,72,130].

3. Discussion

3.1. Difficulties to Overcome

It is difficult to demonstrate if a common causal mechanism for the onset and increasing spread of inflammatory allergic and nonallergic diseases exists and how it works, but it is very important. To our knowledge, a great number of interesting publications are available in the literature on this theme, but none specifically takes into consideration inflammatory nonallergic manifestations from a biochemical point of view. The association of airway inflammation, bronchoconstriction, and/or asthma with acids [23,24,33–37,40,130], and more specifically, of allergic responses with some particular acids, such as sulphurous [33,35], and AA [6,58,141], has long been known. The association of environmental acids with allergic sensitization [37,142] and the hypothesis that these diseases might share a common mechanism [15,16,20] have been considered more recently. Some critical issues emerge from reading the existing toxicological studies. Important criticalities arise, above all, from the features of the proton (H⁺ ion) (small, very mobile and fast, able to interact with many molecular entities). These properties, which make it an ideal activation factor, are at the same time an obstacle to detection by normal instruments. In addition, the interdependence of the concentrations of H^+ and Ca^{2+} suggests that the latter varies rapidly and in parallel with the former. This depends on the intrinsic chemical properties of the two ions [62]. Even in simple aqueous solutions without biological structures, it can be observed that the addition of acids quickly solubilizes the bound calcium and therefore produces an increase in (Ca^{2+}) , whilst the addition of alkali causes it to deposit and therefore reduces (Ca^{2+}) .

Other criticalities arise from the difficulty in isolating a single pollutant, and in the case of PM, its nonspecific composition. There are a number of variables at play, some of which are hard to investigate. Studies often include different cells and different experimental conditions in terms of method and duration. It is therefore very difficult to evaluate and compare data and conclusions. This should be carefully considered in experimental studies. However, modern instruments and techniques can help. In particular, biosensors, which allow one to study subcellular H⁺ and Ca²⁺ dynamics simultaneously, in combination with electron cryomicroscopy and X-ray crystallography should give interesting results.

One question arises spontaneously: "If acids play an important role in asthma and allergies, how are the pathologic responses to basic compounds to be explained?" As recently pointed out [83], most so-called pseudoallergic compounds are basic. The answer, given by the same author, is that pseudoallergic compounds activate G proteins, directly or through GCPRs [83]. Accordingly, the subsequent steps follow the abovementioned PLC/IP₃ pathway, involving acid generation and thus release of H⁺.

Some other studies [123,143,144] showed that in vivo cellular alkalinization causes a substantial increase in $(Ca^{2+})_c$.

This is not in contradiction with what is reported here (Section 2.3, the intracellular H⁺ paragraph), because different events are involved. In their article, Yu et al. [123] describe studies of the regulatory not the activation mechanisms, and some authors think the increase in $(Ca^{2+})_c$ caused by the alkalinization may be due to influx [143] or to the inhibition of Ca^{2+} ATPase and influx [144]. Influx causes an increase in intracellular calcium, since $[Ca^{2+}]$ is normally much higher outside than inside the cell. In Section 2.3, the intracellular H⁺ paragraph, we describe the rapid and transient increase in $[Ca^{2+}]_c$, caused by the activation of phospholipase or the entry of extracellular H⁺. Both of these events occur before influx.

3.2. Possible Deductions

This review on the acidification/increase of (H^+) in external cells and epithelia highlights that the acidity of external epithelia can have both exogenous (environmental acidity) and endogenous

(phospholipase activation) origins (Figure 2B), and therefore, the cellular calcium homeostasis can be altered from both outside and inside.

By the release of protons in various ways via acid loaders and acid extruders through the plasma membrane, the acids increase $(Ca^{2+})_c$ and activate immune cells, inducing the inflammation of the airways and bronchospasm. The parallelism and interdependence of the concentrations of H⁺ and Ca²⁺, and the well-known ability of H⁺ to easily replace Ca²⁺ in its binding sites are the basic facts that suggest that the various means of intracellular acidification, of both exogenous and endogenous origin, have a common mechanism, with H⁺ acting as a stimulus for the increase in $(Ca^{2+})_c$.

Many enzymes at the basis of the allergic and nonallergic inflammatory response have catalytic activity, strictly dependent on pH and/or Ca^{2+} as cofactors. The PLC and PLA₂ themselves are Ca-dependent. Therefore, intracellular acidification, of both exogenous and endogenous origin, may induce acute inflammatory reactions and hypersensitivity through the activation of specific enzymes and the modulation of their action.

Furthermore, exogenous and/or endogenous acidification may favor the lowering of the ASL pH and the reiteration of the acid stimulus, triggering the recruitment of proinflammatory mediators and chronic disease. To sum up, it is possible that:

- (a) Environmental acidity increases the sensitivity of epithelial surfaces and promotes AHR;
- (b) Exogenous and endogenous acids contribute to both the decrease in ASL pH and the increase in ASM basal tone, thus favoring bronchoconstriction;
- (c) The excess of temporary intracellular acidification is at the origin of acute manifestations of an allergic kind;
- (d) Recurrent or continuous acidification is the biochemical basis of airway inflammation, hyper-responsiveness, tissue remodeling, and chronicity.

Accordingly, the impairment of H^+/Ca^{2+} homeostasis and particularly their abnormally high concentrations can constitute a powerful biochemical basis for the onset, continuation, and multimorbidity of disorders, such as inflammatory allergic and nonallergic acute and chronic reactions. The entry/exit pathways for the protons, as described above, are based on physiological activation mechanisms and therefore could be carried out either in healthy or sensitized subjects. The variety of possible paths to increase and control intracellular H^+ and its numerous interactions in the human organism require biomedical studies to explain the diversity of responses and existing situations.

A relation between oxidative or nitrosative stress and acid stress was also put forward [24]. Notoriously, protons can readily produce modifications in the conformation of proteic molecules [140]. Moreover, environmental pollutants have been associated with some asthma phenotypes through the mediation of IL-13 and DNA methylation [2]. DNA methylation is favored by heavy metals, which in turn are made available by acid mobilization. Certain metal constituents of PM_{2.5} were associated with circulating biomarkers of endothelial function [145]. Therefore, environmental acids might play a role in genetic/environmental interactions, by inducing epigenetic modifications with consequent allergic sensitization.

4. Conclusions

Acid pollutants can have toxic, cumulative effects on human epithelia via the release of protons. Protons can affect cellular homeostasis from both outside and inside. Therefore, it can be assumed that intracellular acidification, and the consequent increase in Ca²⁺ concentration induced by protons generated either by acid pollutants or endogenous phospholipases, may be at the basis of the shared causal mechanism of acid stress and multimorbidity of respiratory and hypersensitivity reactions. Moreover, acid environmental pollutants can contribute to the development and growing spread of inflammatory allergic and nonallergic reactions worldwide.

Further studies are required to clarify the specificity and the activation pathways of G proteins in general, and in relation to protons, considering the very high number of GPCRs discovered in recent

years. Similarly, further studies are required into the ability of ion channels to transfer H^+ into cells, together with an investigation of the permeability of plasma membranes to gaseous pollutants, such as NO, NO₂, SO₂, and particularly to CO₂, because CO₂ may have considerable influence on intracellular pH as well as on titratable acidity. Studies of acidity are often limited to measuring only pH, but the measurement of both pH and titratable acidity is indicated for better evaluation purposes.

Identifying and understanding the mechanisms of feedback and control of the different processes of cytosolic acidification, either of internal or external origin, temporary or lasting, and their consequences represents a major challenge for future research.

Reducing air acidity may be an important aim to limit the spread of the disorders taken into consideration in the present study, and to improve the health, especially in children and in frail subjects, of those more exposed to the risk of diseases. We believe our review calls attention to the fundamental importance of H^+/Ca^{2+} interdependence and hope it contributes to further studies into allergic reactions and the identification of the molecular causes of these disorders.

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References

- 1. Thomsen, S.F. Epidemiology and natural history of atopic diseases. *Eur. Clin. Respir. J.* 2015, 2, 506. [CrossRef]
- 2. Bønnelykke, K.; Ober, C. Leveraging gene-environment interactions and endotypes for asthma gene discovery. *J. Allergy Clin. Immunol.* **2016**, 137, 667–679. [CrossRef]
- 3. Bowatte, G.; Lodge, C.; Knibbs, L.D.; Lowe, A.; Erbas, B.; Dennekamp, M.; Marks, G.; Giles, G.G.; Morrison, S.; Thompson, B.; et al. Traffic-related air pollution exposure is associated with allergic sensitization, asthma, and poor lung function in middle age. *J. Allergy Clin. Immunol.* **2017**, *139*, 122–129.e1. [CrossRef] [PubMed]
- 4. Acevedo, N.; Zakzuk, J.; Caraballo, L. House Dust Mite Allergy Under Changing Environments. *Allergy Asthma Immunol. Res.* **2019**, *11*, 450–469. [CrossRef] [PubMed]
- Damialis, A.; Traidl-Hoffmann, C.; Treudler, R. Climate Change and Pollen Allergies. In *Biodiversity and Health in the Face of Climate Change*; Marselle, M., Stadler, J., Korn, H., Irvine, K., Bonn, A., Eds.; Springer Science and Business Media LLC: Cham, Switzerland, 2019; Chapter 3; pp. 47–66.
- Bousquet, J.; Khaltaev, N.; Cruz, A.A.; Denburg, J.; Fokkens, W.J.; Togias, A.; Zuberbier, T.; Baena-Cagnani, C.E.; Canonica, G.W.; van Weel, C.; et al. Allergic Rhinitis and its Impact on Asthma (ARIA) 2008 Update. *Allergy* 2008, 63, 8–160. [CrossRef] [PubMed]
- D'Amato, G.; Holgate, S.; Pawankar, R.; Ledford, D.; Cecchi, L.; Al-Ahmad, M.; Al-Enezi, F.; Al-Muhsen, S.; Ansotegui, I.; Baena-Cagnani, C.E.; et al. Meteorological conditions, climate change, new emerging factors, and asthma and related allergic disorders. A statement of the World Allergy Organization. *World Allergy Organ. J.* 2015, *8*, 1–52. [CrossRef] [PubMed]
- 8. Song, W.; Kang, M.-G.; Chang, Y.-S.; Cho, S.-H. Epidemiology of adult asthma in Asia: Toward a better understanding. *Asia Pac. Allergy* **2014**, *4*, 75–85. [CrossRef]
- 9. World Health Organization. WHO Guidelines for indoor air quality. In *WHO Housing and Health Guidelines;* World Health Organization: Geneva, Switzerland, 2018; pp. 90–96, ISBN-13:978-92-4-155037-6.
- 10. Nicolaou, N.; Siddique, N.; Custovic, A. Allergic disease in urban and rural populations: Increasing prevalence with increasing urbanization. *Allergy* **2005**, *60*, 1357–1360. [CrossRef] [PubMed]

- 11. Rojas-Rueda, D.; Turner, M.C. Commentary: Diesel, Cars, and Public Health. *Epidemiology* **2016**, *27*, 159–162. [PubMed]
- 12. Gehring, U.; Gruzieva, O.; Agius, R.M.; Beelen, R.; Custovic, A.; Cyrys, J.; Eeftens, M.; Flexeder, C.; Fuertes, E.; Heinrich, J.; et al. Air Pollution Exposure and Lung Function in Children: The ESCAPE Project. *Environ. Health Perspect.* **2013**, *121*, 1357–1364. [CrossRef] [PubMed]
- WHO (World Health Organization), WHO Regional Office for Europe. Review of Evidence on Health Aspects of Air Pollution–REVIHAAP Project; Technical Report; WHO: Geneva, Switzerland, 2013; Available online: http: //www.euro.who.int/__data/assets/pdf_file/0004/193108/REVIHAAP-Final-technical-report.pdf (accessed on 27 June 2020).
- Bowatte, G.; Lodge, C.; Lowe, A.J.; Erbas, B.; Perret, J.; Abramson, M.J.; Matheson, M.C.; Dharmage, S.C. The influence of childhood traffic-related air pollution exposure on asthma, allergy and sensitization: A systematic review and a meta-analysis of birth cohort studies. *Allergy* 2015, *70*, 245–256. [CrossRef] [PubMed]
- Bousquet, J.; Anto, J.M.; Akdis, M.; Auffray, C.; Keil, T.; Momas, I.; Postma, D.; Valenta, R.; Wickman, M.; Cambon-Thomsen, A.; et al. Paving the way of systems biology and precision medicine in allergic diseases: The MeDALL success story. Mechanisms of the Development of Allergy. *Allergy* 2016, 71, 1513–1525. [CrossRef]
- Anto, J.M.; Bousquet, J.; Akdis, M.; Auffray, C.; Keil, T.; Momas, I.; Postma, D.S.; Valenta, R.; Wickman, M.; Cambon-Thomsen, A.; et al. Mechanisms of the Development of Allergy (MeDALL): Introducing novel concepts in allergy phenotypes. *J. Allergy Clin. Immunol.* 2017, 139, 388–399. [CrossRef] [PubMed]
- 17. D'Amato, G.; Ortega, O.P.M.; Annesi-Maesano, I.; D'Amato, M. Prevention of Allergic Asthma with Allergen Avoidance Measures and the Role of Exposome. *Curr. Allergy Asthma Rep.* **2020**, *20*, 8. [CrossRef]
- Simon, D. Recent Advances in Clinical Allergy and Immunology. Int. Arch. Allergy Immunol. 2018, 177, 324–333. [CrossRef] [PubMed]
- 19. Adams, O.J.; von Gunten, S. Recent Advances in Experimental Allergy. *Int. Arch. Allergy Immunol.* **2018**, 177, 281–289. [CrossRef] [PubMed]
- 20. Pinart, M.; Benet, M.; Annesi-Maesano, I.; von Berg, A.; Berdel, D.; Carlsen, K.C.L.; Carlsen, K.-H.; Bindslev-Jensen, C.; Eller, E.; Fantini, M.P.; et al. Comorbidity of eczema, rhinitis, and asthma in IgE-sensitised and non-IgE-sensitised children in MeDALL: A population-based cohort study. *Lancet Respir. Med.* **2014**, *2*, 131–140. [CrossRef]
- Apel, K.; Costet, N.; Chapron, A.; Cordier, S.; Monfort, C.; Chevrier, C.; Pelé, F. Home environment: Respiratory and allergic phenotypes from birth to age six in the PELAGIE cohort. *NPJ Prim. Care Respir. Med.* 2019, 29, 1–8. [CrossRef]
- 22. Liu, L.; Poon, R.; Chen, L.; Frescura, A.M.; Montuschi, P.; Ciabattoni, G.; Wheeler, A.; Dales, R. Acute Effects of Air Pollution on Pulmonary Function, Airway Inflammation, and Oxidative Stress in Asthmatic Children. *Environ. Health Perspect.* **2009**, 117, 668–674. [CrossRef]
- 23. Balmes, J.R.; Fine, J.M.; Gordon, T.; Sheppard, D. Potential Bronchoconstrictor Stimuli in Acid Fog. *Environ. Health Perspect.* **1989**, *79*, 163–166. [CrossRef]
- 24. Ricciardolo, F.L.; Gaston, B.; Hunt, J. Acid stress in the pathology of asthma. *J. Allergy Clin. Immunol.* 2004, 113, 610–619. [CrossRef] [PubMed]
- Héroux, M.E.; Anderson, H.R.; Atkinson, R.; Brunekreef, B.; Cohen, A.; Forastiere, F.; Hurley, F.; Katsouyanni, K.; Krewski, D.; Krzyzanowski, M.; et al. Quantifying the health impacts of ambient air pollutants: Recommendations of a WHO/Europe project. *Int. J. Public Health* 2015, *60*, 619–627. [CrossRef] [PubMed]
- Zhang, Y.; Wang, S.G.; Ma, Y.X.; Shang, K.Z.; Cheng, Y.F.; Li, X.; Ning, G.C.; Zhao, W.J.; Li, N.R. Association between Ambient Air Pollution and Hospital Emergency Admissions for Respiratory and Cardiovascular Diseases in Beijing: A Time Series Study. *Biomed. Environ. Sci.* 2015, *28*, 352–363. [PubMed]
- 27. Paulin, L.M.; Hansel, N. Particulate air pollution and impaired lung function. *F1000Research* **2016**, *5*, F1000. [CrossRef] [PubMed]
- 28. Endeward, V.; Al-Samir, S.; Itel, F.; Gros, G. How does carbon dioxide permeate cell membranes? A discussion of concepts, results and methods. *Front. Physiol.* **2014**, *4*, 382. [CrossRef]
- 29. Ye, Q.; Zhang, T.; Mao, J. Haze facilitates sensitization to house dust mites in children. *Environ. Geochem. Health* **2019**. [CrossRef]

- 30. Koehler, C.; Paulus, M.; Ginzkey, C.; Hackenberg, S.; Scherzad, A.; Ickrath, P.; Hagen, R.; Kleinsasser, N. The Proinflammatory Potential of Nitrogen Dioxide and Its Influence on the House Dust Mite Allergen Der p 1. *Int. Arch. Allergy Immunol.* **2016**, *171*, 27–35. [CrossRef]
- 31. Reno, A.L.; Brooks, E.G.; Ameredes, B.T. Mechanisms of Heightened Airway Sensitivity and Responses to Inhaled SO2 in Asthmatics. *Environ. Health Insights* **2015**, *9*, 13–25. [CrossRef]
- 32. Yorifuji, T.; Suzuki, E.; Kashima, S. Hourly differences in air pollution and risk of respiratory disease in the elderly: A time-stratified case-crossover study. *Environ. Health* **2014**, *13*, 67. [CrossRef]
- Vally, H.; Misso, N.L.A. Adverse reactions to the sulphite additives. *Gastroenterol. Hepatol. Bed Bench* 2012, 5, 16–23.
- 34. Schlesinger, R.B.; Chen, L.C.; Finkelsein, I.; Zelikoff, J.T. Comparative potency of inhaled acidic sulphates: Speciation and the role of hydrogen ion. *Environ. Res.* **1990**, *52*, 210–224. [CrossRef]
- 35. Fine, J.M.; Gordon, T.; Sheppard, D. The Roles of pH and Ionic Species in Sulfur Dioxide- and Sulfite-Induced Bronchoconstriction. *Am. Rev. Respir. Dis.* **1987**, *136*, 1122–1126. [CrossRef]
- Mirić, M.; Plavec, D. Risk of Acute Bronchospasm and Bronchial Hyperreactivity from Inhaled Acid Aerosol in 79 Healthy Subjects: Randomized, Double-blind Controlled Trial. *Croat. Med. J.* 2004, 45, 709–714. [PubMed]
- 37. Kamide, Y.; Ishizuka, T.; Tobo, M.; Tsurumaki, H.; Aoki, H.; Mogi, C.; Nakakura, T.; Yatomi, M.; Ono, A.; Koga, Y.; et al. Acidic environment augments FcεRI-mediated production of IL-6 and IL-13 in mast cells. *Biochem. Biophys. Res. Commun.* 2015, 464, 949–955. [CrossRef] [PubMed]
- Trevani, A.S.; Andonegui, G.; Giordano, M.; López, D.H.; Gamberale, R.; Minucci, F.; Geffner, J. Extracellular Acidification Induces Human Neutrophil Activation. J. Immunol. 1999, 162, 4849–4857. [PubMed]
- Herrmann, J.M.; Kantarci, A.; Long, H.; Bernardo, J.; Hasturk, H.; Wray, L.V., Jr.; Simons, E.R.; Van Dyke, T.E. Simultaneous measurements of cytoplasmic Ca²⁺ responses and intracellular pH in neutrophils of localized aggressive periodontitis (LAP) patients. *J. Leukoc. Biol.* 2005, *78*, 612–619. [CrossRef]
- Martinez, D.; Vermeulen, M.; Trevani, A.; Ceballos, A.; Sabatté, J.; Gamberale, R.; Alvarez, M.E.; Salamone, G.; Tanos, T.; Coso, O.A.; et al. Extracellular Acidosis Induces Neutrophil Activation by a Mechanism Dependent on Activation of Phosphatidylinositol 3-Kinase/Akt and ERK Pathways. *J. Immunol.* 2006, 176, 1163–1171. [CrossRef]
- 41. Vermeulen, M.; Giordano, M.; Trevani, A.S.; Sedlik, C.; Gamberale, R.; Fernández-Calotti, P.; Salamone, G.; Raiden, S.; Sanjurjo, J.; Geffner, J.R. Acidosis improves uptake of antigens and MHC class I-restricted presentation by dendritic cells. *J. Immunol.* **2004**, *172*, 3196–3204. [CrossRef]
- 42. Kottyan, L.C.; Collier, A.R.; Cao, K.H.; Niese, K.A.; Hedgebeth, M.; Radu, C.G.; Witte, O.N.; Hershey, G.K.K.; Rothenberg, M.E.; Zimmermann, N. Eosinophil viability is increased by acidic pH in a cAMP- and GPR65-dependent manner. *Blood* **2009**, *114*, 2774–2782. [CrossRef]
- 43. Smith-Garvin, J.E.; Koretzky, G.A.; Jordan, M.S. T Cell Activation. *Annu. Rev. Immunol.* **2009**, 27, 591–619. [CrossRef]
- 44. Wang, X.; Hatatani, K.; Sun, Y.; Fukamachi, T.; Saito, H.; Kobayashi, H. TCR Signaling via ZAP-70 Induced by CD 3 Stimulation is More Active Under Acidic Conditions. *J. Cell Sci. Ther.* **2012**, *S15*, 002.
- 45. Huber, V.; Camisaschi, C.; Berzi, A.; Ferro, S.; Lugini, L.; Triulzi, T.; alTuccitto, A.; Tagliabue, E.; Castelli, C.; Rivoltini, L. Cancer acidity: An ultimate frontier of tumor immune escape and a novel target of immunomodulation. *Semin. Cancer Biol.* **2017**, *43*, 74–89. [CrossRef]
- 46. Narita, A.; Yawata, K.; Nagata, M.; et al. Effects of Extracellular Acidification on Intracellular pH and ATP-Induced Calcium Mobilization in Rabbit Lens Epithelial Cells. *Yonago Acta Med.* **1999**, *42*, 51–59.
- Chin, W.-C.; Quesada, I.; Nguyen, T.; Verdugo, P. Oscillations of pH inside the secretory granule control the gain of Ca²⁺ release for signal transduction in goblet cell exocytosis. *Novartis Found. Symp.* 2002, 248, 132–141. [PubMed]
- Ichimonji, I.; Tomura, H.; Mogi, C.; Sato, K.; Aoki, H.; Hisada, T.; Dobashi, K.; Ishizuka, T.; Mori, M.; Okajima, F. Extracellular acidification stimulates IL-6 production and Ca²⁺ mobilization through proton-sensing OGR1 receptors in human airway smooth muscle cells. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2010, 299, L567–L577. [CrossRef]

- Saxena, H.; Deshpande, D.A.; Tiegs, B.C.; Yan, H.; Battafarano, R.J.; Burrows, W.M.; Damera, G.; Panettieri, R.; Duboes, T.D., Jr.; An, S.; et al. The GPCR OGR1 (GPR68) mediates diverse signalling and contraction of airway smooth muscle in response to small reductions in extracellular pH. *Br. J. Pharmacol.* 2012, *166*, 981–990. [CrossRef]
- 50. Prakash, Y.S. Airway smooth muscle in airway reactivity and remodeling: What have we learned? *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2013**, *305*, L912–L933. [CrossRef] [PubMed]
- 51. Kim, D.; Liao, J.; Hanrahan, J.W. The buffer capacity of airway epithelial secretions. *Front. Physiol.* **2014**, *5*, 188. [CrossRef] [PubMed]
- 52. Wilton, D.C. Phospholipases. In *Biochemistry of Lipids, Lipoproteins and Membranes,* 5th ed.; Vance, D.E., Vance, J.E., Eds.; Elsevier Science: Amsterdam, The Netherlands, 2008; Chapter 11; pp. 305–329.
- 53. Hu, Y.L.; Mi, X.; Huang, C.; Wang, H.F.; Song, J.-R.; Shu, Q.; Ni, L.; Chen, J.-G.; Wang, F.; Hu, Z.-L. Multiple H⁺ sensors mediate the extracellular acidification-induced [Ca²⁺]_i elevation in cultured rat ventricular cardiomyocytes. *Sci. Rep.* 2017, 7, 44951. [CrossRef]
- 54. Huang, J.; Liu, C.H.; Hughes, S.A.; Postma, M.; Schwiening, C.J.; Hardie, R.C. Activation of TRP channels by protons and phosphoinositide depletion in Drosophila photoreceptors. *Curr. Biol.* **2010**, *20*, 189–197. [CrossRef]
- 55. Smith, J.B.; Dwyer, S.D.; Smith, L. Lowering Extracellular pH Evokes Inositol Polyphosphate Formation and Calcium Mobilization. *J. Biol. Chem.* **1989**, *264*, 8723–8728. [PubMed]
- 56. Tsunoda, Y.; Matsuno, K.; Tashiro, Y. Cytosolic acidification leads to Ca²⁺ mobilization from intracellular stores in single and populational parietal cells and platelets. *Exp. Cell Res.* **1991**, *193*, 356–363. [CrossRef]
- 57. Donoso, P.; Beltrán, M.; Hidalgo, C. Luminal pH regulated calcium release kinetics in sarcoplasmic reticulum vesicles. *Biochemistry* **1996**, *35*, 13419–13425. [CrossRef] [PubMed]
- Chen, W.-H.; Chen, C.-R.; Yang, K.-T.; Chang, W.-L.; Su, M.-J.; Wu, C.-C.; Wu, M.-L. Arachidonic acid-induced H⁺ and Ca²⁺ increases in both the cytoplasm and nucleoplasm of rat cerebellar granule cells. *J. Physiol.* 2001, 537 (*Pt 2*), 497–510. [CrossRef]
- Bates, R.C.; Fees, C.P.; Holland, W.L.; Winger, C.C.; Batbayar, K.; Ancar, R.; Bergren, T.; Petcoff, D.; Stith, B. Activation of Src and release of intracellular calcium by phosphatidic acid during Xenopus laevis fertilization. *Dev. Biol.* 2014, 386, 165–180. [CrossRef]
- 60. Busa, W.B.; Nuccitelli, R. Metabolic regulation via intracellular pH. *Am. J. Physiol.* **1984**, 246, R409–R438. [CrossRef]
- 61. Iida, S.; Potter, J.D. Calcium binding to calmodulin. Cooperativity of the calcium-binding sites. *J. Biochem.* **1986**, *99*, 1765–1772. [CrossRef]
- 62. Molinari, G.; Nervo, E. Role of Protons in Calcium Signaling. Preprints 2020, 2020030274. [CrossRef]
- 63. Berridge, M.J.; Bootman, M.D.; Roderick, H.L. Calcium signalling: Dynamics, homeostasis and remodelling. *Nat. Rev. Mol. Cell. Biol.* **2003**, *4*, 517–529. [CrossRef]
- 64. Lee, R.J.; Foskett, J.K. Ca²⁺ signaling and fluid secretion by secretory cells of the airway epithelium. *Cell Calcium* **2014**, *55*, 325–336. [CrossRef] [PubMed]
- 65. Ma, H.-T.; Beaven, M.A. Regulators of Ca²⁺ signaling in mast cells: Potential targets for treatment of mast-cell related diseases? *Adv. Exp. Med. Biol.* **2011**, *716*, 62–90. [PubMed]
- 66. Chapman, D.G.; Irvin, C.G. Mechanisms of airway hyper-responsiveness in asthma: The past, present and yet to come. *Clin. Exp. Allergy* **2015**, *45*, 706–719. [CrossRef] [PubMed]
- 67. Ozier, A.; Allard, B.; Bara, I.; Girodet, P.-O.; Trian, T.; Marthan, R.; Berger, P. The Pivotal Role of Airway Smooth Muscle in Asthma Pathophysiology. *J. Allergy* **2011**, 742710. [CrossRef]
- 68. Galli, S.J.; Tsai, M. IgE and mast cells in allergic disease. Nat. Med. 2012, 18, 693–704. [CrossRef] [PubMed]
- 69. Vines, C.M.; Prossnitz, E.R. Mechanisms of G protein-coupled receptor-mediated degranulation. *FEMS Microbiol. Lett.* **2004**, 236, 1–6. [CrossRef]
- 70. Maurer, M.; Pucillo, C. What we know (and don't know) about the biology and functions of mast cells and basophils. *Immunol. Rev.* **2018**, *282*, 5–7. [CrossRef]
- 71. Miyake, K.; Karasuyama, H. Emerging roles of basophils in allergic inflammation. *Allergol. Int.* **2017**, *66*, 382–391. [CrossRef]
- 72. Olivera, A.; Beaven, M.A.; Metcalfe, D.D. Mast cells signal their importance in health and disease. *J. Allergy Clin. Immunol.* **2018**, 142, 381–393. [CrossRef]

- 73. Gilfillan, A.M.; Peavy, R.D.; Metcalfe, D.D. Amplification mechanisms for the enhancement of antigen mediated mast cell activation. *Immunol. Res.* **2009**, *43*, 15–24. [CrossRef]
- 74. Suzuki, R. The Emerging Picture of Mast Cell Activation: The Complex Regulatory Network of High-Affinity Receptor for Immunoglobulin E Signaling. *Biol. Pharm. Bull.* **2017**, *40*, 1828–1832. [CrossRef]
- 75. Kuehn, H.S.; Gilfillan, A.M. G protein-coupled receptors and the modification of FcεRI-mediated mast cell activation. *Immunol. Lett.* **2007**, *113*, 59–69. [CrossRef] [PubMed]
- 76. Subramanian, H.; Gupta, K.; Ali, H. Roles of MAS-related G protein coupled receptor-X2 (MRGPRX2) on mast cell-mediated host defense, pseudoallergic drug reactions and chronic inflammatory diseases. *J. Allergy Clin. Immunol.* 2016, 138, 700–710. [CrossRef] [PubMed]
- 77. Gaudenzio, N.; Sibilano, R.; Marichal, T.; Starkl, P.; Reber, L.L.; Cenac, N.; McNeil, B.D.; Dong, X.; Hernandez, J.D.; Sagi-Eisenberg, R.; et al. Different activation signals induce distinct mast cell degranulation strategies. *J. Clin. Invest.* **2016**, *126*, 3981–3998. [CrossRef]
- Gao, Z.-G.; Jacobson, K.A. Purinergic Signaling in Mast Cell Degranulation and Asthma. *Front. Pharmacol.* 2017, *8*, 947. [CrossRef] [PubMed]
- 79. Jairaman, A.; Maguire, C.H.; Schleimer, R.P.; Prakriya, M. Allergens stimulate store-operated calcium entry and cytokine production in airway epithelial cells. *Sci. Rep.* **2016**, *6*, 32311. [CrossRef] [PubMed]
- Nakano, M.; Ito, K.; Yuno, T.; Soma, N.; Aburakawa, S.; Kasai, K.; Nakamura, T.; Takami, H. UDP/P2Y6 receptor signaling regulates IgE-dependent degranulation in human basophils. *Allergol. Int.* 2017, 66, 574–580. [CrossRef] [PubMed]
- 81. Aoki, H.; Mogi, C.; Okajima, F. Ionotropic and Metabotropic Proton-Sensing Receptors Involved in Airway Inflammation in Allergic Asthma. *Mediators Inflamm.* **2014**, 2014, 712962. [CrossRef] [PubMed]
- Alexander, S.P.H.; Fabbro, R.; Kelly, E.; Marrion, N.V.; A Peters, J.; Faccenda, E.; Harding, S.D.; Pawson, A.J.; Sharman, J.L.; Southan, C.; et al. The concise guide to pharmacology 2017/18: G protein-coupled receptors. *Br. J. Pharmacol.* 2017, 174, S17–S129. [CrossRef]
- 83. Seifert, R. How do basic secretagogues activate mast cells? *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2015**, *388*, 279–281. [CrossRef]
- Chen, Y.-C.; Chang, Y.-C.; Chang, H.-A.; Lin, Y.-S.; Tsao, C.-W.; Shen, M.-R.; Chiu, W.-T. Differential Ca²⁺ mobilization and mast cell degranulation by FcεRI- and GPCR-mediated signaling. *Cell Calcium* 2017, 67, 31–39. [CrossRef]
- 85. Gilfillan, A.M.; Tkaczyk, C. Integrated signalling pathways for mast-cell activation. *Nat. Rev. Immunol.* **2006**, *6*, 218–230. [CrossRef] [PubMed]
- 86. Itsuki, K.; Imai, Y.; Hase, H.; Okamura, Y.; Inoue, R.; Mori, M.X. PLC-mediated PI(4,5)P2 hydrolysis regulates activation and inactivation of TRPC6/7 channels. *J. Gen. Physiol.* **2014**, *143*, 183–201. [CrossRef] [PubMed]
- 87. Cocco, L.; Follo, M.Y.; Manzoli, L.; Suh, P.-G. Phosphoinositide-specific phospholipase C in health and disease. *J. Lip. Res.* **2015**, *53*, 1853–1860. [CrossRef] [PubMed]
- Nakamura, Y.; Fukami, K. Regulation and physiological functions of mammalian phospholipase C. J. Biochem. 2017, 161, 315–321. [CrossRef]
- 89. Molinari, G. Is hydrogen ion (H(+)) the real second messenger in calcium signalling? *Cell. Signal.* **2015**, *27*, 1392–1397. [CrossRef]
- 90. Berridge, M.J. The inositol trisphosphate/calcium signaling pathway in health and disease. *Physiol. Rev.* **2016**, *96*, 1261–1296. [CrossRef]
- 91. Fujishima, H.; Sanchez Mejia, R.O.; Bingham, C.O.; Lam, B.K.; Sapirstein, A.; Bonventre, J.V.; Austen, K.F.; Arm, J.P. Cytosolic phospholipase A₂ is essential for both the immediate and the delayed phases of eicosanoid generation in mouse bone marrow-derived mast cells. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 4803–4807. [CrossRef]
- 92. Leslie, C.C. Cytosolic phospholipase A₂: Physiological function and role in disease. *J. Lipid Res.* **2015**, *56*, 1386–1402. [CrossRef]
- Damron, D.S.; Van Wagoner, D.R.; Moravec, C.S.; Bond, M. Arachidonic acid and endothelin potentiate Ca²⁺ transients in rat cardiac myocytes via inhibition of distinct K⁺ channels. *J. Biol. Chem.* **1993**, *268*, 27335–27344.
- 94. Dennis, E.A.; Norris, P.C. Eicosanoid Storm in Infection and Inflammation. *Nat. Rev. Immunol.* 2015, 15, 511–523. [CrossRef]
- 95. Hanna, V.S.; Hafez, E.A.A. Synopsis of arachidonic acid metabolism: A review. J. Adv. Res. 2018, 11, 23–32. [CrossRef]

- Bodnar, D.; Chung, W.Y.; Yang, D.; Hong, J.H.; Jha, A.; Muallem, S. STIM-TRP Pathways and Microdomain Organization: Ca2+ Influx Channels: The Orai-STIM1-TRPC Complexes. *Adv. Exp. Med. Biol.* 2017, 993, 139–157. [CrossRef]
- 97. Parenti, A.; De Logu, F.; Geppetti, P.; Benemei, S. What is the evidence for the role of TRP channels in inflammatory and immune cells? *Br. J. Pharmacol.* **2016**, *173*, 953–969. [CrossRef]
- 98. Ambudkar, I.S.; de Souza, L.B.; Ong, H.L. TRPC1, Orai1, and STIM1 in SOCE: Friends in tight spaces. *Cell Calcium* **2017**, *63*, 33–39. [CrossRef] [PubMed]
- 99. Fahrner, M.; Stadlbauer, M.; Muik, M.; Rathner, P.; Stathopulos, P.; Ikura, M.; Mueller, N.; Romanin, C. A dual mechanism promotes switching of the Stormorken STIM1 R304W mutant into the activated state. *Nat. Commun.* **2018**, *9*, 825. [CrossRef] [PubMed]
- 100. Nicaise, G.; Maggio, K.; Thirion, S.; Horoyan, M.; Keicher, E. The calcium loading of secretory granules. A possible key event in stimulus-secretion coupling. *Biol. Cell* **1992**, *75*, 89–99. [CrossRef]
- Wernersson, S.; Pejler, G. Mast cell secretory granules: Armed for battle. *Nat. Rev. Immunol.* 2014, 14, 478–494.
 [CrossRef] [PubMed]
- 102. Murakami, M.; Yamamoto, K.; Miki, Y.; Murase, R.; Sato, H.; Taketomi, Y. The Roles of the Secreted Phospholipase A₂ Gene Family in Immunology. *Adv. Immunol.* **2016**, *132*, 91–134.
- 103. Pejler, G.; Frisk, J.M.; Sjöström, D.; Paivandy, A.; Ohrvik, H. Acidic pH is essential for maintaining mast cell secretory granule homeostasis. *Cell Death Dis.* **2017**, *8*, 22785. [CrossRef]
- 104. Galli, S.J.; Tsai, M.; Marichal, T.; Tchougounova, E.; Reber, L.L.; Pejler, G. Approaches for Analyzing the Roles of Mast Cells and Their Proteases In Vivo. *Adv. Immunol.* **2015**, *126*, 45–127.
- 105. Kormelink, T.G.; Arkesteijn, G.J.A.; van de Lest, C.H.A.; Geerts, W.J.C.; Goerdayal, S.S.; Altelaar, M.; Redegeld, F.A.; Nolte-'t Hoen, E.N.M.; Wauben, M.H.M. Mast Cell Degranulation Is Accompanied by the Release of a Selective Subset of Extracellular Vesicles That Contain Mast Cell–Specific Proteases. *J. Immunol.* 2016, 197, 3382–3392. [CrossRef] [PubMed]
- 106. Abdulkhaleq, L.A.; Assi, M.A.; Abdullah, R.; Zamri-Saad, M.; Taufiq-Yap, Y.H.; Hezmee, M. The crucial roles of inflammatory mediators in inflammation: A review. *Vet. World* **2018**, *11*, 627–635. [CrossRef]
- 107. Swietach, P.; Youm, J.B.; Saegusa, N.; Leem, C.H.; Spitzer, K.W.; Vaughan-Jones, R.D. Coupled Ca²⁺/H⁺ transport by cytoplasmic buffers regulates local Ca²⁺ and H⁺ ion signaling. *Proc. Natl. Acad. Sci. USA* 2013, 110, E2064–E2073. [CrossRef] [PubMed]
- 108. Zhang, X.; Gueguinou, M.; Trebak, M. Store-Independent Orai Channels Regulated by STIM. In *Calcium Entry Channels in Non-Excitable Cells*; Kozak, J.A., Putney, J.W., Jr., Eds.; Taylor & Francis Group: Abingdon, UK, 2018; Chapter 11; pp. 197–213.
- 109. Hellwig, N.; Plant, T.D.; Janson, W.; Schäfer, M.; Schultz, G.; Schaefer, M. TRPV1 acts as proton channel to induce acidification in nociceptive neurons. *J. Biol. Chem.* **2004**, *279*, 34553–34561. [CrossRef] [PubMed]
- 110. Jiang, J.; Li, M.; Yue, L. Potentiation of TRPM7 inward currents by protons. *J. Gen. Physiol.* **2005**, *126*, 137–150. [CrossRef]
- 111. White, J.P.M.; Cibelli, M.; Urban, L.; Nilius, B.; McGeown, J.G.; Nagy, I. TRPV4: Molecular Conductor of a Diverse Orchestra. *Physiol. Rev.* **2016**, *96*, 911–973. [CrossRef]
- 112. Starkus, J.G.; Fleig, A.; Penner, R. The calcium-permeable non-selective cation channel TRPM2 is modulated by cellular acidification. *J. Physiol.* **2010**, *588* (*Pt 8*), 1227–1240. [CrossRef]
- 113. Cairns, S.P.; Westerblad, H.; Allen, D.G. Changes in myoplasmic pH and calcium concentration during exposure to lactate in isolated rat ventricular Myocytes. *J. Physiol.* **1993**, *464*, 561–574. [CrossRef]
- 114. Marin, M.; Sellier, C.; Paul-Antoine, A.F.; Cailliau, K.; Browaeys-Poly, E.; Bodart, J.F.; Vilain, J.P. Calcium Dynamics During Physiological Acidification in Xenopus Oocyte. J. Membr. Biol. 2010, 236, 233–245. [CrossRef]
- 115. Lee, S.E.; Lee, S.H. Skin Barrier and Calcium. Ann. Dermatol. 2018, 30, 265–275. [CrossRef]
- Brune, K.; Frank, J.A.; Schwingshackl, A.; Finigan, J.; Venkataramana, K.S. Pulmonary epithelial barrier function: Some new players and mechanisms. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2015, 308, L731–L745. [CrossRef]
- 117. De Rose, V.; Molloy, K.; Gohy, S.; Pilette, C.; Greene, C.M. Airway Epithelium Dysfunction in Cystic Fibrosis and COPD. *Mediat. Inflamm.* **2018**, *8*, 1309746. [CrossRef] [PubMed]
- 118. Alka, K.; Casey, J.R. Bicarbonate Transport in Health and Disease. IUBMB Life 2014, 66, 596-615. [CrossRef]

- Liu, X.; Li, T.; Tuo, B. Physiological and Pathophysiological Relevance of the Anion Transporter Slc26a9 in Multiple Organs. *Front. Physiol.* 2018, *9*, 1197. [CrossRef] [PubMed]
- Lee, D.; Hong, J.H. The Fundamental Role of Bicarbonate Transporters and Associated Carbonic Anhydrase Enzymes in Maintaining Ion and pH Homeostasis in Non-Secretory Organs. *Int. J. Mol. Sci.* 2020, 21, 339. [CrossRef]
- 121. Brini, M.; Carafoli, E. The plasma membrane Ca²⁺ ATPase and the plasma membrane sodium calcium exchanger cooperate in the regulation of cell calcium. *Cold Spring Harb Perspect Biol.* **2011**, *3*, a004168. [CrossRef] [PubMed]
- 122. Boscardin, E.; Alijevic, O.; Hummler, E.; Frateschi, S.; Kellenberger, S. The function and regulation of acid-sensingion channels (ASICs) and the epithelial Na⁺ channel (ENaC): IUPHAR Review 19. *Br. J. Pharmacol.* **2016**, *173*, 2671–2701. [CrossRef] [PubMed]
- 123. Yu, A.S.; Yue, Z.; Feng, J.; Yue, L. Regulation of Orai/STIM Channels by pH. In *Calcium Entry Channels in Non-Excitable Cells*; Kozak, J.A., Putney, J.W., Jr., Eds.; CRC Press/Taylor & Francis Group: Boca Raton, FL, USA; Abingdon, UK , 2018; Chapter 9; pp. 161–176.
- 124. DeCoursey, T.E. Voltage-Gated Proton Channels and Other Proton Transfer Pathways. *Physiol. Rev.* 2003, *83*, 475–579. [CrossRef]
- 125. Amdursky, N.; Lin, Y.; Aho, N.; Groenhof, G. Exploring fast proton transfer events associated with lateral proton diffusion on the surface of membranes. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 2443–2451. [CrossRef]
- 126. Fischer, H.; Widdicombe, J.H. Mechanisms of Acid and Base Secretion by the Airway Epithelium. *J. Membr. Biol.* **2006**, *211*, 139–150. [CrossRef]
- 127. Jayaraman, S.; Song, Y.; Vetrivel, L.; Shankar, L.; Verkman, A.S. Noninvasive in vivo fluorescence measurement of airway-surface liquid depth, salt concentration, and pH. *J. Clin. Investig.* 2001, 107, 317–324. [CrossRef] [PubMed]
- 128. McCreanor, J.; Cullinan, P.; Nieuwenhuijsen, M.J.; Stewart-Evans, J.; Malliarou, E.; Jarup, L.; Harrington, R.; Svartengren, M.; Han, I.-K.; Ohman-Strickland, P.; et al. Respiratory Effects of Exposure to Diesel Traffic in Persons with Asthma. N. Engl. J. Med. 2007, 357, 2348–2358. [CrossRef] [PubMed]
- 129. Brant, T.C.; Yoshida, C.T.; Carvalho, T.S.; Nicola, M.L.; Martins, J.A.; Braga, L.M.; De Oliveira, R.C.; Leyton, V.; De André, C.S.; Saldiva, P.H.N.; et al. Mucociliary clearance, airway inflammation and nasal symptoms in urban motorcyclists. *Clinics* 2014, 69, 867–870. [CrossRef]
- Hunt, J.F.; Fang, K.; Malik, R.; Snyder, A.; Malhotra, N.; Platts-Mills, T.A.E.; Gaston, B. Endogenous Airway Acidification Implications for Asthma Pathophysiology. *Am. J. Respir. Crit. Care Med.* 2000, 161, 694–699. [CrossRef]
- Brunetti, L.; Francavilla, R.; Tesse, R.; Strippoli, A.; Polimeno, L.; Loforese, A.; Miniello, V.L.; Armenio, L. Exhaled breath condensate pH measurement in children with asthma, allergic rhinitis and atopic dermatitis. *Pediatr. Allergy Immunol.* 2006, 17, 422–427. [CrossRef]
- 132. Bartoszewski, R.; Matalon, S.; Collawn, J.F. Ion channels of the lung and their role in disease pathogenesis. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2017**, *313*, L859–L872. [CrossRef] [PubMed]
- 133. Seifter, J.L.; Chang, H.-Y. Extracellular Acid-Base Balance and Ion Transport Between Body Fluid Compartments. *Physiology* **2017**, *32*, 367–379. [CrossRef] [PubMed]
- 134. DeCoursey, T.E. Voltage and pH sensing by the voltage-gated proton channel, HV1. *J. R. Soc. Interface.* **2018**, *15*, 20180108. [CrossRef]
- 135. Sedlyarov, V.; Eichner, R.; Girardi, E.; Essletzbichler, P.; Goldmann, U.; Nunes-Hasler, P.; Srndic, I.; Moskovskich, A.; Heinz, L.X.; Kartnig, F.; et al. The Bicarbonate Transporter SLC4A7 Plays a Key Role in Macrophage Phagosome Acidification. *Cell Host Microbe* **2018**, *23*, 766–774.e5. [CrossRef]
- 136. Hollenhorst, M.I.; Richter, K.; Fronius, M. Ion Transport by Pulmonary Epithelia. *J. Biomed. Biotechnol.* **2011**, 2011, 174306. [CrossRef]
- Weller, P.F.; Spencer, L.A. Functions of tissue-resident eosinophils. *Nat. Rev. Immunol.* 2017, 17, 746–760. [CrossRef] [PubMed]
- Stone, K.D.; Prussin, C.; Metcalfe, D.D. IgE, Mast Cells, Basophils, and Eosinophils. J. Allergy Clin. Immunol. 2010, 125, S73–S80. [CrossRef] [PubMed]
- Granata, F.; Nardicchi, V.; Loffredo, S.; Frattini, A.; Staiano, R.I.; Agostini, C.; Triggiani, M. Secreted phospholipases A₂: A proinflammatory connection between macrophages and mast cells in the human lung. *Immunobiology* 2009, 214, 811–821. [CrossRef] [PubMed]

- 140. Onufriev, A.V.; Alexov, E. Protonation and pK changes in protein-ligand binding. *Q. Rev. Biophys.* **2013**, *46*, 181–209. [CrossRef] [PubMed]
- 141. Liu, M.; Yokomizo, T. The role of leukotrienes in allergic diseases. *Allergol. Int.* **2015**, *64*, 17–26. [CrossRef] [PubMed]
- 142. Gold, M.J.; Hiebert, P.R.; Park, H.Y.; Stefanowicz, D.; Le, A.; Starkey, M.R.; Deane, A.; Brown, A.C.; Liu, G.; Horvat, J.C.; et al. Mucosal production of uric acid by airway epithelial cells contributes to particulate matter-induced allergic sensitization. *Mucosal Immunol.* **2016**, *9*, 809–820. [CrossRef]
- 143. Lindeman, K.S.; Croxton, T.L.; Lande, B.; Hirshman, C.A. Hypocapnia-induced contraction of porcine airway smooth muscle. *Eur. Respir. J.* **1998**, *12*, 1046–1052. [CrossRef]
- 144. Li, S.; Hao, B.; Lu, Y.; Yu, P.; Lee, H.-C.; Yue, J. Intracellular Alkalinization Induces Cytosolic Ca²⁺ Increases by Inhibiting Sarco/Endoplasmic Reticulum Ca²⁺-ATPase (SERCA). *PLoS ONE* **2012**, *7*, e31905. [CrossRef]
- 145. Wu, S.; Yang, D.; Pan, L.; Shan, J.; Li, H.; Wei, H.; Wang, B.; Huang, J.; Baccarelli, A.A.; Shima, M.; et al. Chemical constituents and sources of ambient particulate air pollution and biomarkers of endothelial function in a panel of healthy adults in Beijing, China. *Sci. Total Environ.* **2016**, *560–561*, 141–149. [CrossRef]



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