

## Complete chloroplast genome sequence of *Eranthis byunsanensis* B.Y. Sun (Ranunculaceae), an endemic species in Korea

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### ABSTRACT

The Korean endemic *Eranthis byunsanensis* B.Y. Sun, 1993 (Ranunculaceae) is a rare plant distributed in the southwestern part of the Korean Peninsula. The complete chloroplast (cp) genome of *E. byunsanensis* was sequenced by next-generation sequencing (NGS) using an Illumina HiSeq X platform. The cp genome of *E. byunsanensis* is 160,324 bp in length with 37.9% GC content. It showed a typical quadripartite structure consisting of a pair of inverted repeats (IRs; 28,356 bp), a large single-copy region (LSC; 87,671 bp), and a small single-copy region (SSC; 15,941 bp). The cp genome comprises 130 genes including 85 protein-coding genes (PCGs), 37 tRNA genes, and eight rRNA genes. The molecular phylogenetic analysis indicates that *E. byunsanensis* is closely related to *Eranthis stellata*, both of which belong to the genus *Eranthis*.

### ARTICLE HISTORY

Received 7 February 2023  
Accepted 26 April 2023

### KEYWORDS

*Eranthis byunsanensis*;  
chloroplast genome;  
phylogeny; Ranunculaceae

### Introduction

The genus *Eranthis* Salisb., an early flowering perennial plant belonging to Ranunculaceae Juss. Tribe Cimicifugeae Torr. & A. Grey, has been a subject of interest in phylogenetic research because it is often considered the most primitive of herbaceous flowering plant (Tamura 1995). This genus comprises approximately 13 species that are mainly distributed in East Asia and Europe (Tamura 1995). Of these, three *Eranthis* species including *E. stellata* Maxim., 1859, *E. byunsanensis* B.Y.Sun, 1993, and *E. pungdoensis* B.U.Oh, 2009 inhabit limited areas in the Korean peninsula. In particular, *E. byunsanensis* and *E. pungdoensis* were recently described as new species (Sun et al. 1993; Oh and Ji 2009). However, the phylogenetic position of these species has been controversial due to their high similarities in taxonomic characteristics. The shapes of petals, leaves, and bracts showed only minor differences between these species (Oh and Oh 2019).

Chloroplast (cp) genome sequences are precious resources for phylogenetic study on closely related taxa (Bi et al. 2018). To date, the complete cp genome sequences of *E. stellata* have been only reported among 13 *Eranthis* species (Zhai et al. 2019). Here, we analyzed the complete cp genome sequence of *E. byunsanensis* and evaluated its phylogenetic

position within the family Ranunculaceae. Our results may provide valuable resources for future genetic and evolutionary studies on Ranunculaceae.

### Materials and methods

#### Plant sampling

The plant material of *E. byunsanensis* was collected from the mountain area of Wanju, Jeollabuk-do, South Korea (127°18'15.50"E, 36°07'17.20"N) on 22 March 2022 and photographed with a digital camera to record the natural habitat (Figure 1). Each specimen was morphologically identified by Joon Moh Park (<https://forest.jb.go.kr/>, [joonmoh@korea.kr](mailto:joonmoh@korea.kr)). The specimen voucher with number JFERI0020-1 was deposited in the Jeollabuk-do Forest Environment Research Institute.

#### Sequencing, assembling, and annotating the chloroplast genome

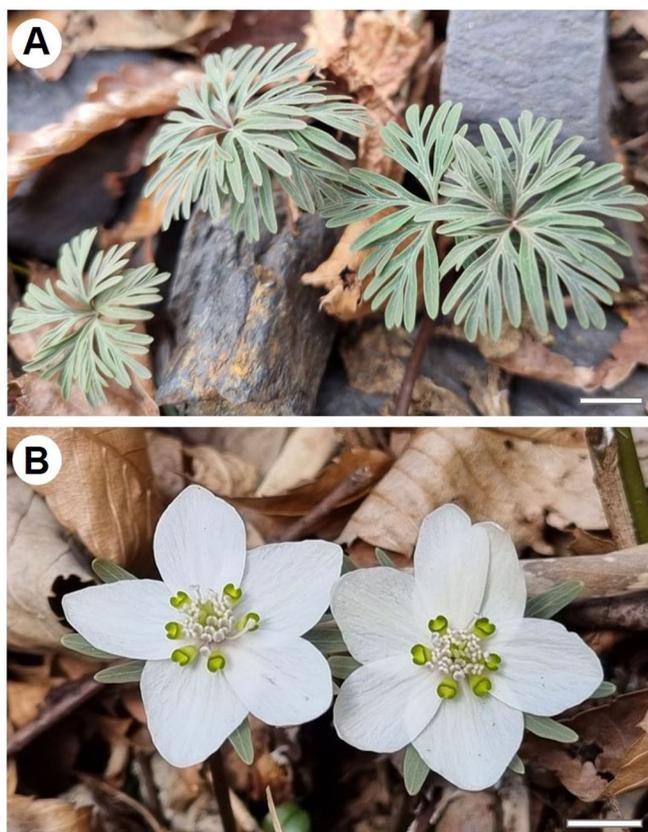
Total genomic DNA was isolated from the radical leaves of *E. byunsanensis* and deposited in the Jeollabuk-do

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 Supplemental data for this article can be accessed online at <https://doi.org/10.1080/23802359.2023.2209383>.

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**Figure 1.** *Eranthis byunsanensis* of Wanju, Republic of Korea. (A) Typical radical leaves with blades divided into several palmate segments. (B) White, compound floral structure with broad overlapping sepals and funnel-shaped petals. Scale bars represent 1 cm. Photographs were captured by Joon Moh Park using a digital camera.

Forest Environment Research Institute (voucher number JFERI-DNA0020-1; contact person, Joon Moh Park). The next-generation sequencing (NGS) sequencing library was constructed using TruSeq Nano DNA Kit (Illumina, San Diego, CA) and sequenced by paired-end sequencing using Illumina HiSeq X platform (Macrogen Inc., Seoul, Republic of Korea). Trimmomatic was used to eliminate adapter sequences and low-quality reads. De novo assembly was performed using NOVOPlasty v.4.3.1 (Dierckxsens et al. 2017). Annotation of cp genome was conducted by GeSeq v.1.59 (<https://chlorobox.mpimp-golm.mpg.de/geseq.html>). The cis- and trans-splicing genes were identified using CPGView software (Liu et al. 2023). A circular map of the complete cp genome was constructed by CPGView software.

### Phylogenetic analysis

The phylogenetic position of *E. byunsanensis* was investigated using 38 species of Ranunculaceae and two outgroup species. The nucleotide sequences of 77 shared non-redundant protein-coding genes (PCGs) were extracted from the complete cp genome sequences deposited in GenBank. The PCGs

were concatenated into a dataset using Geneious prime 2023 (Biomatters Ltd, Auckland, New Zealand). Multiple sequence alignment was performed using MAFFT (Kato and Standley 2013), and gaps and poorly aligned sequences were eliminated by TrimmAl. The resulting 75,566 bp of aligned sequences were used to generate a phylogenetic tree using two methods. The maximum-likelihood (ML) method was performed by RAxML (Stamatakis 2014) with 1000 replicates using the GTR + G model. Bayesian's inference (BI) method was performed using MrBayes (Ronquist et al. 2012) with the GTR + I + G model. Each analysis implemented four MCMC chains with 5,000,000 generations and sampled every 1000 iterations with the first 10% discarded.

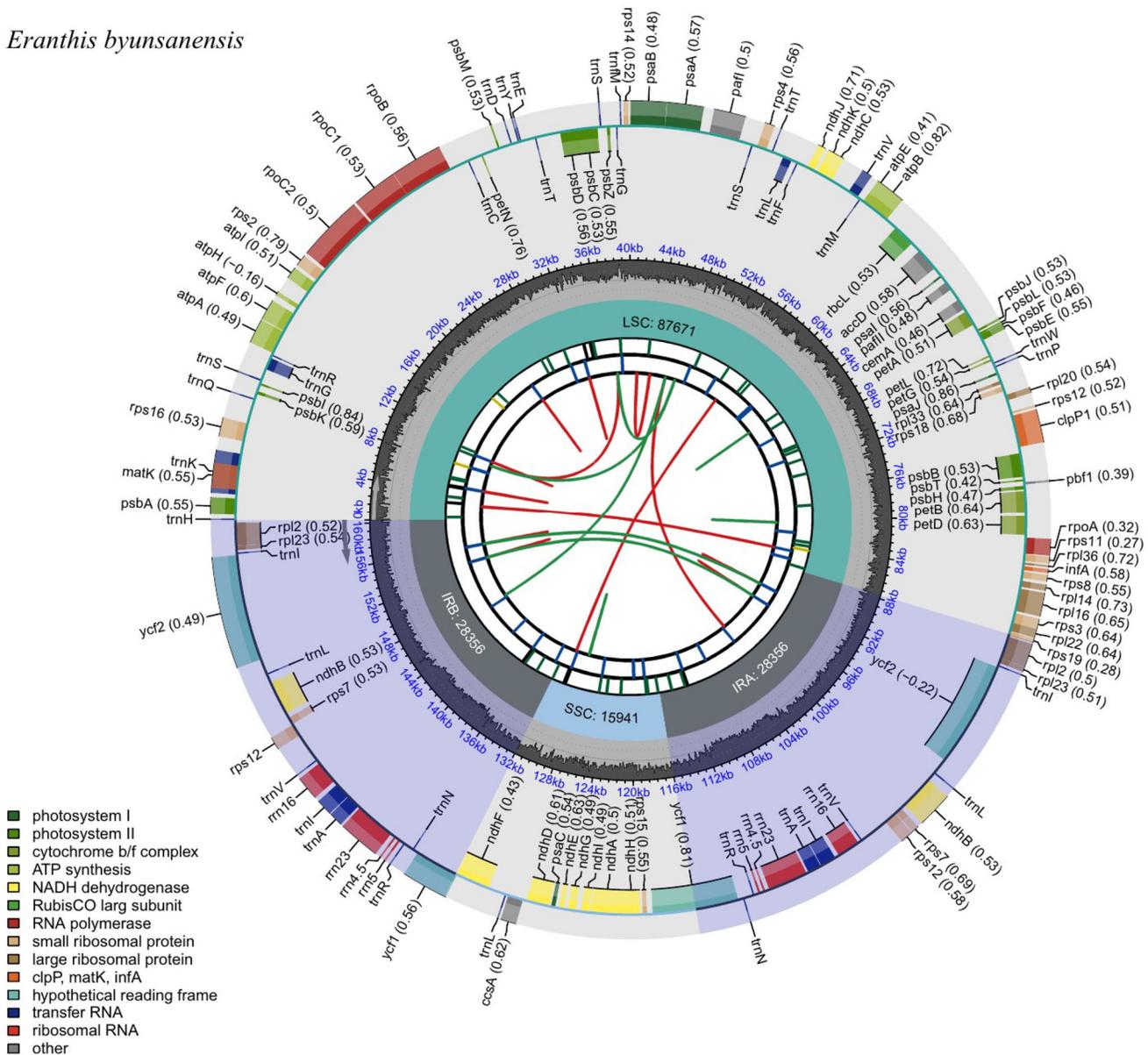
## Results

### Chloroplast genome features

The cp genome of *E. byunsanensis* was sequenced by paired-end sequencing. The 28,939,092 filtered sequencing reads were assembled into single contigs with an average read coverage of 1187 (Supplementary Figure S1). The assembly result was evaluated by depth of coverage using Bowtie2 and verified by comparing the synteny and sequence homology with the reference cp sequence (*Eranthis stellata*, MK569487) by NOVOPlasty v.4.3.1 (Supplementary Figure S1). PCR-based Sanger sequencing was performed to confirm the complete circular cp genome sequences (Supplementary Figure S2). A schematic representation of the plastome organization of *E. byunsanensis* is represented in Figure 2. The complete cp genome of *E. byunsanensis* (GenBank accession number ON564441) is 160,324 bp in length with an overall GC content of 37.9%. It showed a typical quadripartite structure comprising a pair of inverted repeats (IRA and IRB; 28,356 bp), a large single-copy region (LSC; 87,671 bp), and a small single-copy region (SSC; 15,941 bp). A total of 130 genes were identified, including 85 PCGs, 37 tRNA genes, and eight rRNA genes. Among the PCGs, 13 contain one or two introns (Supplementary Figure S3). The *rps12* is a trans-splicing gene with 5'-end and 3'-end exons located in the LSC and IRA regions, respectively.

### Phylogenetic analysis

Phylogenetic position of *E. byunsanensis* was analyzed using the 77 conserved PCGs in the complete cp genomes of 38 Ranunculaceae species including *E. byunsanensis* and two outgroups (Figure 3). Two phylogenetic trees were constructed using BI and ML methods. BI and ML trees showed identical topologies with 14 strongly supported clades, each of which corresponds to the defined tribe in previous studies (Zhai et al. 2019). In addition, the phylogenetic tree indicates that *E. byunsanensis* is most closely related to *E. stellata* with strong support (posterior probability = 1/boot-strap = 100).

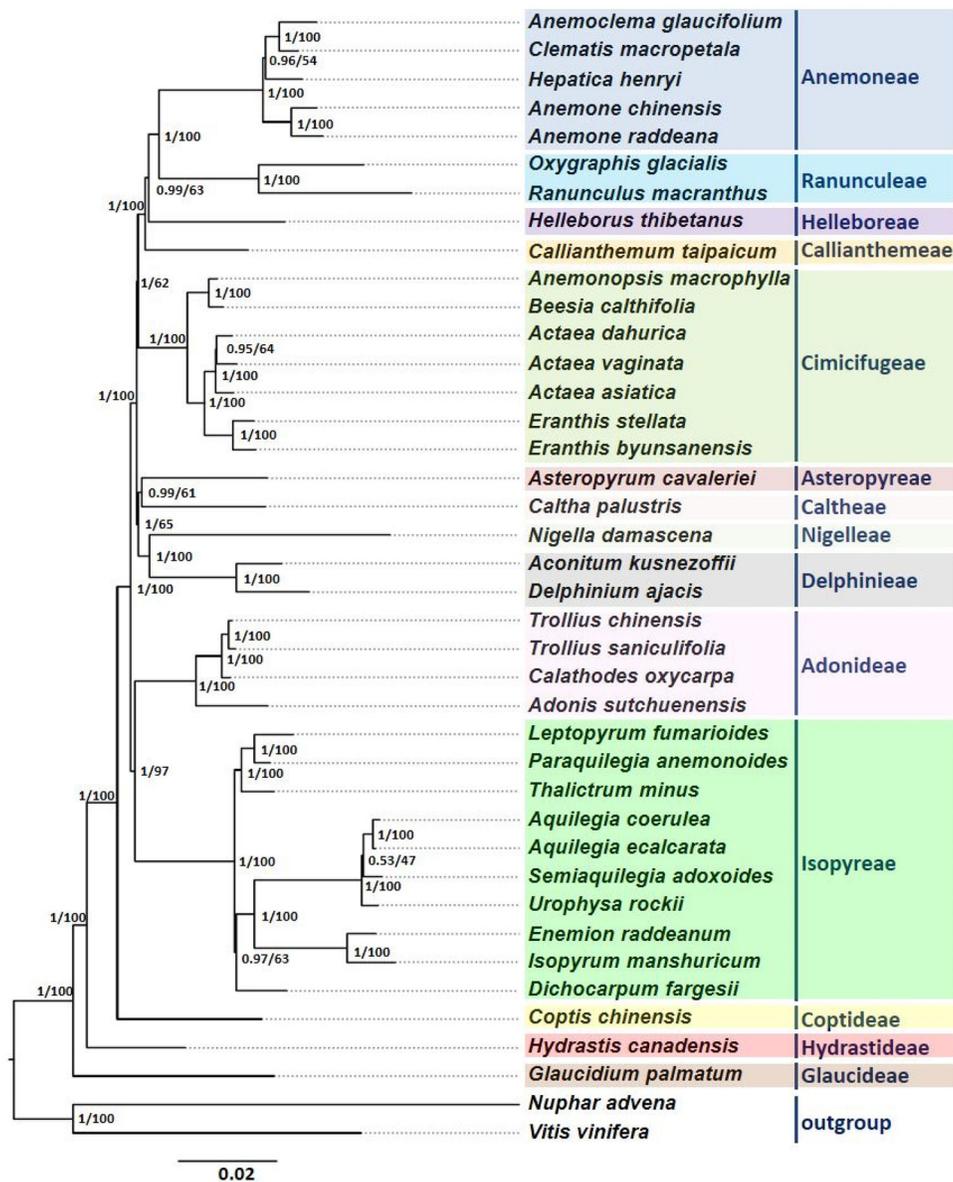
*Eranthis byunsanensis*

**Figure 2.** Chloroplast genome map of *E. byunsanensis*. A circular and complete cp genome map was generated by CPGview. Large single-copy, small single-copy, and inverted repeat are represented as LSC, SSC, and IR (IRA and IRB) on the fourth track, respectively. GC content is represented on the fifth track in dark gray. The genes are shown on the sixth track. Genes located on the inner and outer of circle are transcribed clockwise and anticlockwise, respectively. The functional classification of the genes is presented in the bottom left corner.

## Discussion and conclusions

The family Ranunculaceae has been the focus of systematic studies as a model due to its unstable position in flowering plants and diverse morphological characteristics (Tamura 1995). Recently, this family has received more attention as a model system for addressing crucial evolutionary questions regarding either primitive or derived morphological features that cannot be answered by other model plants (Kramer 2009). Numerous studies have attempted to determine the divergence times among the major lineages of Ranunculaceae, but no consensus has been achieved (Wang et al. 2016). In particular, the phylogenetic position of the genus *Eranthis* has been controversial due to their high

similarities in taxonomic characteristics. Recently, the cp genomes of 35 Ranunculaceae species representing 31 genera of the 14 tribes were phylogenetically analyzed, and inter-tribal relationships of the family Ranunculaceae were much clarified (Zhai et al. 2019). In the present study, the complete cp genome sequences of *E. byunsanensis* were determined and utilized to analyze its phylogenetic position within 38 Ranunculaceae species. The BI and ML phylogenetic trees strongly indicate that *E. byunsanensis* is most closely related to *E. stellata*, both of which belong to the genus *Eranthis*. Our data provide valuable information for further genetic and evolutionary studies on Ranunculaceae.



**Figure 3.** Phylogenetic relationships among 38 Ranunculaceae species based on 77 shared PCGs of the complete cp genome. The sequences used for tree construction are as follows: *Anemoclema glaucifolium* (MK569471; Zhai et al. 2019), *Anemone chinensis* (MK569491; Zhai et al. 2019), *Anemone raddeana* (MK569472; Zhai et al. 2019), *Clematis macropetala* (MK569482; Zhai et al. 2019), *Hepatica henryi* (MK569494; Zhai et al. 2019), *Oxygraphis glacialis* (MK569489; Zhai et al. 2019), *Ranunculus macranthus* (NC\_008796; Raubeson et al. 2007), *Helleborus thibetanus* (MK569493; Zhai et al. 2019), *Callianthemum taipaicum* (MK569479; Zhai et al. 2019), *Actaea asiatica* (MK569469; Zhai et al. 2019), *Actaea dahurica* (MK569481; Zhai et al. 2019), *Actaea vaginata* (MK569499; Zhai et al. 2019), *Anemonopsis macrophylla* (MK569473; Zhai et al. 2019), *Beesia calthifolia* (MK569477), *Eranthis stellata* (MK569487; Zhai et al. 2019), *Eranthis byunsanensis* (ON564441; this study), *Caltha palustris* (MK569480; Zhai et al. 2019), *Asteropyrum cavaleriei* (MK569476; Zhai et al. 2019), *Nigella damascena* (MK569488; Zhai et al. 2019), *Aconitum kusnezoffii* (MK569468; Zhai et al. 2019), *Delphinium ajacis* (MK569484; Zhai et al. 2019), *Adonis sutchuenensis* (MK569470; Zhai et al. 2019), *Calathodes oxycarpa* (MK569478; Zhai et al. 2019), *Trollius saniculifolia* (NC\_012615; Kim et al. 2009), *Trollius chinensis* (MK569501; Zhai et al. 2019), *Aquilegia coerulea* (MK569474; Zhai et al. 2019), *Aquilegia ecalcarata* (MK569475; Zhai et al. 2019), *Dichocarpum fargesii* (MK569485; Zhai et al. 2019), *Enemion raddeanum* (MK569486; Zhai et al. 2019), *Isopyrum manshuricum* (MK569496; Zhai et al. 2019), *Leptopyrum fumarioides* (MK569497; Zhai et al. 2019), *Paraquilegia anemonoides* (MK569490; Zhai et al. 2019), *Semiaquilegia adoxoides* (MK569498; Zhai et al. 2019), *Thalictrum minus* (MK569500; Zhai et al. 2019), *Urophysa rockii* (MK569502; Zhai et al. 2019), *Coptis chinensis* (MK569483; Zhai et al. 2019), *Hydrastis Canadensis* (MK569495; Zhai et al. 2019), *Glaucidium palmatum* (MK569492; Zhai et al. 2019), *Nuphar advena* (NC\_008788; Raubeson et al. 2007), and *Vitis vinifera* (NC\_007957; Jansen et al. 2006) were used as outgroups. The phylogenetic tree was constructed using BI and ML methods, and the trees showed identical topologies. The numbers on the nodes indicate the BI posterior probability and ML bootstrap value (%), respectively. The scale bar represents the number of substitutions per site.

## Author contributions

J.M.P., A.O., and J.K. carried out sample collection, experiments, data analysis, and data curation. Preparation of manuscript and project administration were performed by J.K. All authors have read and agreed to the published version of the manuscript.

## Ethical approval

The species used in this study does not need ethical approval or permissions to collect the sample. All procedures for the sampling and experiments in this article were conducted in compliance with the regulations of the Jeollabuk-do Forest Environment Research Institute.

## Disclosure statement

No potential conflict of interest was reported by the author(s).

## Funding

No funding was received.

## Data availability statement

The genome sequence data supporting the findings of this study are openly available as accession no. ON564441 at GenBank in NCBI (<https://www.ncbi.nlm.nih.gov>). The associated BioProject, SRA, and Bio-Sample numbers are PRJNA835661, SRR19213060, and SAMN28106645, respectively.

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