

Review Article

Exosomes in Human Immunodeficiency Virus Type I Pathogenesis: Threat or Opportunity?

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Nanometre-sized vesicles, also known as exosomes, are derived from endosomes of diverse cell types and present in multiple biological fluids. Depending on their cellular origins, the membrane-bound exosomes packed a variety of functional proteins and RNA species. These microvesicles are secreted into the extracellular space to facilitate intercellular communication. Collective findings demonstrated that exosomes from HIV-infected subjects share many commonalities with Human Immunodeficiency Virus Type I (HIV-1) particles in terms of proteomics and lipid profiles. These observations postulated that HIV-resembled exosomes may contribute to HIV pathogenesis. Interestingly, recent reports illustrated that exosomes from body fluids could inhibit HIV infection, which then bring up a new paradigm for HIV/AIDS therapy. Accumulative findings suggested that the cellular origin of exosomes may define their effects towards HIV-1. This review summarizes the two distinctive roles of exosomes in regulating HIV pathogenesis. We also highlighted several additional factors that govern the exosomal functions. Deeper understanding on how exosomes promote or abate HIV infection can significantly contribute to the development of new and potent antiviral therapeutic strategy and vaccine designs.

1. Introduction

The membrane-bound exosomes are present in a wide range of human fluids such as urine [1], plasma [2], saliva [3], ascites [4], breast milk [5], semen [6], bronchoalveolar lavage liquid [7], amniotic fluid [8], and cerebrospinal fluid [9]. These microvesicles are secreted from various types of immune cells such as dendritic cells (DCs) [10], macrophages [11], T cells [12], and B cells [13], as well as tumor cells from various cancers [14, 15]. Exosomes are mainly responsible for cell-cell communication processes such as cell proliferation [15], cell invasion [16], and immune and gene regulation [17, 18]. It is known that exosomes are derived from cellular endosomes, where the inward budding takes place on the endosomal multivesicular bodies (MVBs) to form the intraluminal vesicles (ILVs) [19]. The subsequent molecular mechanism

then determines the fate of ILVs, entering the lysosomal degradation pathway or released extracellularly as exosomes upon fusion of MVB membrane with the plasma membrane [20].

Accumulative findings have demonstrated that exosomes highly resembled HIV particles in many aspects, from their physical properties to composition [21–24]. This has given rise to two models that explain these similarities [24]. First, the Trojan exosome hypothesis proposed that retroviruses are originated from exosomes following the evolution involving *gag* gene mutation [25]. This explained the ability of virus to exploit the preexisting exosome biogenesis pathway for viral dissemination and be able to infect cells in Env- and receptor-independent manner [26, 27]. The second model, however, is not in line with the evolutionary theory of the virus. Instead, the “crosstalk” or “hijacker” hypothesis

suggested that the retroviruses have evolved to hijack the intercellular communication pathway of the host to promote HIV pathogenesis [28]. Although both models differ from each other, the similarity of the compositions (i.e., lipids, proteins, carbohydrates, and RNAs) between viral particles and exosomes suggests that exosomes may play an indispensable role in HIV pathogenesis.

Recently, several reports have demonstrated that exosomes contain internal cargoes that can inhibit HIV infection and replication [29–31]. These antiviral exosomes were mostly found in the body fluids such as semen and breast milk. However, the inhibitory action of exosomes is not well described compared to its viral infection enhancement effects. This may be due to the high abundance of HIV pathogenesis promoting molecules within the composition of exosomes, which may mask the existing antiviral effects, if any. By far, collective findings have shown that exosomes can either promote or inhibit HIV infection, with little understanding upon the critical factors and/or the exact mechanisms that determine the exosomal effects in viral infection. In general, the source (i.e., from different cell types and biological fluids) and the composition of exosomes may exert the decisive role in contribution to HIV/AIDS pathogenesis. More effort is required to thoroughly understand the exosomal function in HIV infection in order to benefit the development of new-era HIV/AIDS therapy and vaccine designs.

2. Morphological and Biological Properties of Exosomes and HIV Particles

Exosomes share several common structural and molecular properties with HIV. Physically, their size and density range from 50 to 150 nm in diameter [32] and 1.13 to 1.21 g/mL [33], respectively, and both are surrounded by a lipid bilayer. In addition to morphological similarities, they possess similar composition such as lipids (i.e., cholesterol and glycosphingolipids) [13], carbohydrates (i.e., high mannose and complex N-linked glycans) [34], proteins (i.e., tetraspanins, MHC molecules, actin, and TSG101) [35, 36], and RNA species [24]. Exosomes from HIV-infected cells are also enriched with viral proteins such as Nef [37] and viral RNAs [18, 38]. Due to these similarities, HIV-1 is believed to be generated by the same pathway of exosome biogenesis [24, 39]. Moreover, a substantial amount of the host component (e.g., MHC-II) can enter the viral particles [25]. This can be one of the mechanisms that is exploited by viruses to evade the host immune surveillance.

Despite sharing most of the biochemical features, HIV particles have a few principal differences in comparison with exosomes. First, HIV has more organized and uniform structures regardless of the type of infected cells while the structure of exosomal vesicles varies depending on the parental cell after the membrane budding [40]. Second, the exosomal contents are highly diverse from different sources while the biochemical content of HIV virions is steadily consistent. These differences allow the purification methods based on

iodixanol density gradients and immunoaffinity isolation to efficiently harvest exosome-free HIV virions [40, 41].

3. Distinct Functions of Exosomes from Different Sources

The composition of exosomes derived from biological fluids is highly variable from one to the other, which suggest the composition may define the distinct effects of exosomes (either promoting or inhibiting viral pathogenesis). Cumulative evidence suggested that the exosomal effect on HIV mainly depends on their cellular origins [42]. In most cases but not all, exosomes derived from HIV-infected cells are more virulent and enhance infection, while exosomes from uninfected cells have protective properties. In this section, we discuss the exosomal functions and their effects on HIV pathogenesis based on their sources or origins (summarized in Table 1).

3.1. Blood/Plasma/Serum. Human blood is where the HIV virions reside and is the main biofluid responsible for HIV transmission. The blood also contains various types of cells of both HIV-susceptible and uninfected cells that secrete exosomes. It has been postulated that HIV hijacks the exosome biogenesis pathway which carries various viral proteins and RNAs for viral dissemination process [24]. A general review on how exosomes enhance the spread of various infections has been recently reported [23, 42]. Exosomes secreted from HIV-infected cells had been found to contain chemokine receptors, CCR5 and CXCR4, that were delivered to recipient cells to facilitate HIV establishment and spreading [43, 44]. Exosomes from HIV-1 infected macrophages had also been known to facilitate viral transfer to uninfected cells [45]. Exosomes that contain HIV Nef protein have multiple pathogenic effects such as induction of T-cell apoptosis [37], inhibition of RNA interference [46], and downmodulation of cell surface molecules (i.e., MHC-I and CD4) for immune evasion [47]. Nef protein also induces exosomal secretion [37], thereby contributing to HIV/AIDS pathogenesis. Unlike the transfer of CCR5 and/or CXCR4 which primarily direct HIV infection, Nef proteins promote HIV infection by activating the uninfected cells.

Other viral components that are usually found in exosomes are HIV Gag [39], viral mRNA/miRNA [18], and pathogen-associated RNAs such as HIV trans-activation response (TAR) RNA [48] that could enhance the viral infection and replication in the recipient cells. Various host surface molecules (CD45, CD86, and MHC-II) have also been exported from HIV-infected cells via exosomes to silence the immune response [49]. Additionally, exosomes derived from infected dendritic cells (DCs) have a profound enhancing effect on CD4+ T cell infection [50]. More recently, Nef-induced exosome-associated ADAMI7 (ADAM metalloproteinase domain 17) had rendered resting CD4+ T cells permissive to HIV-1 infection [51], whereas ADAMI7 along with TNF- α has been known to synergistically activate the latent HIV-1 in primary CD4+ T lymphocytes and macrophages [52]. HIV particles are known to incorporate

TABLE I: Dual effects of body fluids-derived exosomes against HIV infection.

Viral/antiviral effect	Exosomal source	Active component	Reference	
Promote HIV infection	Blood/plasma/serum	CCR5 and CXCR4	[43, 44]	
		Nef	[37, 46, 47]	
		Gag	[39]	
		Viral mRNA/miRNA	[18]	
		TAR RNA	[48]	
		CD45, CD86, and MHC-II	[49]	
		ADAM17, TNF- α	[51, 52]	
		Undefined (from DCs)	[50]	
		APOBEC3G	[29, 53, 54]	
		CD4	[59]	
Inhibit HIV infection	Blood/plasma/serum	Interferon-alpha (IFN- α)	[63]	
		Interferon-beta (IFN- β)	[64]	
		Tumor necrosis factor (TNF- α)	[61, 65]	
		Interleukins	[61, 62]	
		Undefined (from CD8+ T cells)	[60]	
		Lewis X	[66]	
		Bile lipase	[67]	
		Breast milk	IgA and IgG antibodies	[68]
			Mucin 1 (MUC-1)	[69]
			Oligosaccharides	[70]
Semen	Undefined	[31]		
	Mucin 6	[72]		
	Undefined	[30, 71]		
Promote/inhibit HIV infection (unexplored)	Urine	Unexplored	Unexplored	
	Saliva			
	Ascites			
	Bronchoalveolar lavage liquid (BAL)			
	Amniotic fluid			
	Cerebrospinal fluid			
	Vaginal fluid			

various host cell components that enable the viruses to evade the immune system. The exchange of these components could be facilitated by the existing exosomes that are enriched with the host components. While extensive studies have been conducted on exosomes derived from HIV-infected cells, little is known of the role of exosomes derived from uninfected cells in viral pathogenesis. The increased exosomal secretion may have increased the susceptibility of uninfected cells to HIV; this however warrants future investigation.

Although exosomes are mainly enriched with viral components that promote HIV/AIDS pathogenesis, a few reports have shown that exosomes may potentially inhibit HIV infection. APOBEC3G, a host cellular protein, has been known to transfer from cell to cell through exosomes to protect the recipient cell from HIV infection [29, 53, 54]. APOBEC3G, the most prominent member of APOBEC3 (A3) proteins, is a cellular cytidine deaminase that restricts HIV replication by both DNA-editing and editing-independent activities [55, 56]. In order to function, the APOBEC3G must be incorporated into the virions [57]. The extensive

role of APOBEC3G in antiviral immunity has been recently reviewed [58]. The restrictive effect is more prominent in *Vif*-deficient HIV-1 than the wild-type strain as the *Vif* proteins target and counteract APOBEC3G for polyubiquitination and degradation by the 26S proteasome [29]. APOBEC3G proteins have been detected in human mammary tissues and were packaged into milk-borne virions and subsequently restricted HIV-1 infectivity [53]. Besides, it has also been shown that APOBEC3G inhibited viral replication by blocking the function of HIV-1 reverse transcriptase [54].

Although T lymphocytes are the primary reservoir of HIV infection, exosomes released from T lymphocytes had been found to have inhibitory effects against HIV. When compared to CD4-depleted exosomes from CD4+ T cells, CD4-containing exosomes efficiently inhibited HIV-1 infection [59]. This may be due to the masking of HIV-1 envelope proteins by the exosomal CD4 that has subsequently blocked HIV infection. Similarly, exosomes secreted from CD8+ T cells were able to suppress HIV transcription within the infected cells [60]. Other components from

exosomes that inhibit viral infections are interleukins [61, 62], interferon-alpha [63], interferon-beta [64], and tumor necrosis factor (TNF- α) [61, 65].

3.2. Breast Milk. While exosome's dual-functions were seen in the blood-derived exosomes, current findings on milk-derived exosomes are skewed towards their antiviral effects, probably due to its role in providing natural passive immunity for infants. Exosomes were found to contain several components such as Lewis X [66], bile lipase [67], antibodies [68], mucin 1 (MUC-1) [69], and oligosaccharides [70] that inhibit the DC-mediated HIV transmission to CD4+ T lymphocytes. More recently, the antiviral effect of exosomes has been shown to be specifically derived from the milk since no HIV-1 inhibition was seen in plasma-derived exosomes when experimented in parallel [31]. In this study, the milk exosomes bound to monocyte-derived dendritic cells (MDDCs) and inhibited HIV-1 infection of MDDCs and the subsequent viral transfer to CD4+ T cells. Cumulative work showed that milk exosomes have a strong inhibitory effect; these protective effects may be transferred to the uninfected cells of newborns via breastfeeding as part of the passive antiviral immunity. This may also be a reason why the HIV-1 transmission via breastfeeding is rare. More efforts are currently in progress to evaluate the potential of utilizing milk exosomes in the antiviral therapy [42].

3.3. Semen. HIV-1 infection is notoriously known to be transmitted through sexual intercourse. Surprisingly, the semen-derived exosomes mainly possess antiviral effects compared to mediating HIV infection. It has been shown that exosomes purified from healthy individuals inhibit the HIV-1 replication in various cell types by blocking the postentry viral RNA reverse transcription [30]. The same group has also shown that exosomes in human semen were able to restrict the HIV transmission *in vivo* in LP-BM5-infected mice model [71]. Another group has also reported that the mucin-containing exosomes were able to prevent the HIV-1 transfer from DCs to CD4+ T cells [72]. Similar to the milk exosomes, these exosomes may presumably be transferred from one to another to exert the antiviral or protective effects. Yet, this seems to be inefficient as the number of HIV cases due to sexual transmission increases every year. Although several reports have highlighted the protective role of semen-derived exosomes, the exact mechanism that is involved is still unknown. This is important in order to control or prevent the HIV spreading, particularly through unprotected sexual intercourse.

3.4. Urine, Saliva, Ascites, and Other Biological Fluids. While the anti-HIV action of exosomes has been shown in human blood, semen, and breast milk, its antiviral potential in other sources such as saliva and ascites is yet to be determined. Since the composition of exosomes is heterogeneous depending on the origins, it is of crucial importance to understand the factors that may eventually lead to its viral or antiviral effects. More efforts need to be done to reveal the activity of

exosomes and the mechanisms in these biofluids as they may serve as an important source for antiviral therapies.

4. Decisive Factors for Exosome Functions in HIV Infection

It is interesting that the exosome nanovesicles packed with a very complex composition displayed two contradictory functions towards HIV infection (Table 1). However, little is known about the criteria that drive the ultimate role of exosomes in HIV/AIDS pathogenesis. Cumulative findings have demonstrated that the function of exosomes is mainly directed by their cellular origin and composition. Exosomes derived from HIV-infected T cells, monocyte/macrophage, and dendritic cells contain several components that abate viral infection [21, 24, 45]. These immune cells may release immune-regulatory factors that possess antiviral property such as interferon-alpha (IFN- α) [63], interferon-beta (IFN- β) [64], tumor necrosis factor-alpha (TNF- α) [65, 66], interleukins (ILs) [66, 67], and APOBEG3G [29, 53, 54], which are exported by exosomes from the cells. However, this antiviral implication might be masked due to the lytic replication of HIV that resulted in the pathogenic effects. To note, some of the viral molecules such as Nef and viral TAR RNA have been detected in exosomes which may further enhance the infection [37, 48]. Similarly, the antiviral activity of exosomes has been found in biological fluids (i.e., semen and breast milk) that are rich in immunological molecules [30, 31]. This indicates that the function of exosomes partly depends on the cellular origin. However, the exact components and their underlying mechanism that contributes to the antiviral action are left to be discovered.

In addition to the cellular origin of exosomes, the target or recipient cells in which exosomes are delivered also play a pivotal role in viral pathogenesis. Exosomes are known to transport from HIV-infected CD4+ T cells among themselves [51] as well as to other recipients cells such as dendritic cells and macrophages [45, 73]. The dendritic cells were also known to capture exosomes from the exosome-producer cells or infected CD4+ T cells and export to HIV-susceptible cells for the infection. These processes were termed as *trans*-infection and *trans*-dissemination, respectively [73, 74]. During the process, viral proteins such as chemokine receptors and Nef may be delivered to the recipient cells, thereby enhancing HIV infection and replication. Indeed, the ultimate functions of exosomes in the HIV-infected individuals largely rely on the composition of the nanosized exosomes. When both surface molecules and internal cargoes of exosome have net pathogenic effects, there is more likelihood that exosomes would contribute to viral pathogenesis and *vice versa*. The components of exosomes may also alter the intracellular signalling pathways [75, 76], thereby affecting HIV-1 infection.

It is well-known that the content of exosomes largely varies depending on their origins. However, it is intriguing that exosomes derived from the same source (i.e., blood-derived exosomes) have opposing effects in HIV pathogenesis. While individual components within the exosomes appear to have opposing effects on HIV pathogenesis, the

net effect of the exosomes may depend on the relative strength of the effects. In addition to the net pathogenic effect of exosomal contents, it is noteworthy that variation in testing conditions such as culture conditions, exosome or virions preparation, cell infection status, and exosomal transfer or delivery status may also play a role in determining the outcome of exosomes in viral infection. For example, the profile of exosomes generated *in vitro* from cultured cells may differ drastically from exosomes that are freshly harvested from biofluids. The variation of exosomes derived from tumor cells cultured *in vitro* and *in vivo* had been previously reported [77]. In this case, the experiments utilizing exosomes prepared *in vitro* may not represent the *in vivo* exosomal effects on viral infection. Second, the different methods used in exosome or virions preparation may also complicate the data interpretation. Exosomes prepared *in vitro* might be contaminated with HIV virions during the purification, thereby affecting the outcome of exosomes on viral pathogenesis. Third, the efficiency of exosomal transfer from the exosome producer cells to the recipient cells may also play a key role in viral pathogenesis. For instance, exosomes derived from cultured cells *in vitro* may not be as potent in the exchange of exosomal contents (i.e., virulence factors and/or antiviral components) as compared with *in vivo* transfer [42]. Moreover, the level of exosome secretion is dependent on the HIV infection status as Nef proteins mediate the exosome release [37]. All of these factors have to be taken into consideration by researchers before concluding the study outcome.

5. Development of a New-Era HIV Therapeutic Strategy

Cumulative findings demonstrate that the cell-encoded exosome pathways enhance HIV infection, supported by the Trojan exosome hypotheses [25] and the envelope protein- and receptor-independent viral dissemination without involving fusion events [26]. On the other hand, exosomes derived from the breast milk and semen exhibit modest antiviral effects [30, 31]. Both of these contradictory functions of exosomes pose a major impact to the existing strategy of anti-HIV drug development and urge the discovery of new-era therapeutics. For instance, the Env-targeting therapeutics may contribute to a low extent or none at all to the exosome-mediated infection that can take place without Env-receptor fusion. Hence, a more comprehensive strategy is needed to control the infection. The exosomal pathway in which HIV was hijacked for the viral dissemination can be targeted by newly designed inhibitors to target exosome biogenesis and exosome uptake in both Env-dependent and independent infections. Potential exosomal targets are cellular enzymes (RNases, proteases, and lipase), cytoplasmic proteins (TSG101, cyclophilins, MHC-II, and tetraspanins), and HIV-related proteins (e.g., Nef, CCR5, and CXCR4) [21, 23]. However, the specificity of the therapy must be monitored as the host-derived exosomes may trigger deleterious side effects. Several reports have shown that exosomal inhibitors have reduced the overall Env-dependent infection [30, 31] and these suggest the applicability of

inclusion of exosome inhibitors in the adjunctive HIV therapy. Exosomes that exhibit antiviral activity can be purified from the particular sources and developed into a potential therapy. Indeed, more promising action is envisioned when the exact components and mechanisms are revealed. It would also be interesting to determine whether the relative amount of HIV promoting versus HIV inhibiting exosomes could affect disease progression and thereby serve as potential biomarkers for prognosis. While this review focuses on the role of exosomes in HIV infection, these nanovesicles have also been shown to contribute to pathogenesis of other viral classes such as Hepatitis B and C virus [21, 78] and Herpes Simplex virus (HSV) [22]. Continuous efforts must be made to enhance the understanding of the exosomal function in these viral infections and their potential use in the development of antiviral therapies.

6. Conclusions

To summarize, the host-derived exosomes enhance HIV infection by several routes, including intercellular dissemination of viral components and immune evasion whereas only a limited number of reports have shown the antiviral function of exosomes. Numerous factors, such as the cellular origins, recipient cells, and the intracellular signalling that are affected by exosomes, are currently thought to contribute to the final outcome of exosome in the infection. Variation in exosome preparation and testing conditions may significantly affect the outcome of exosomes. Compared to other fields, the exosomal functions in HIV and other types of viral infections are apparently underexplored. More research is anticipated to improve the understanding of the association between exosomes and viral infections in order to reveal the potential of exosomes in the development of anti-HIV therapy.

Abbreviations

ADAMI7:	ADAM metallopeptidase domain 17
AIDS:	Acquired immunodeficiency syndrome
CD:	Cluster of differentiation
DCs:	Dendritic cells
Env:	Envelope protein
HIV:	Human immunodeficiency virus
ILVs:	Intraluminal vesicles
MDDCs:	Monocyte-derived dendritic cells
MHC-II:	Major histocompatibility complex class II
MUC:	Mucin
MVBs:	Multivesicular bodies
TAR:	trans-Activation response.

Conflict of Interests

The authors declare no conflict of interests.

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References

- [1] T. Pisitkun, R.-F. Shen, and M. A. Knepper, "Identification and proteomic profiling of exosomes in human urine," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 36, pp. 13368–13373, 2004.
- [2] M.-P. Caby, D. Lankar, C. Vincendeau-Scherrer, G. Raposo, and C. Bonnerot, "Exosomal-like vesicles are present in human blood plasma," *International Immunology*, vol. 17, no. 7, pp. 879–887, 2005.
- [3] A. Michael, S. D. Bajracharya, P. S. T. Yuen et al., "Exosomes from human saliva as a source of microRNA biomarkers," *Oral Diseases*, vol. 16, no. 1, pp. 34–38, 2010.
- [4] F. Andre, N. E. C. Schartz, M. Movassagh et al., "Malignant effusions and immunogenic tumour-derived exosomes," *The Lancet*, vol. 360, no. 9329, pp. 295–305, 2002.
- [5] C. Admyre, S. M. Johansson, K. R. Qazi et al., "Exosomes with immune modulatory features are present in human breast milk," *The Journal of Immunology*, vol. 179, no. 3, pp. 1969–1978, 2007.
- [6] R. Sullivan, F. Saez, J. Girouard, and G. Frenette, "Role of exosomes in sperm maturation during the transit along the male reproductive tract," *Blood Cells, Molecules, and Diseases*, vol. 35, no. 1, pp. 1–10, 2005.
- [7] C. Admyre, J. Grunewald, J. Thyberg et al., "Exosomes with major histocompatibility complex class II and co-stimulatory molecules are present in human BAL fluid," *European Respiratory Journal*, vol. 22, no. 4, pp. 578–583, 2003.
- [8] S. Keller, C. Rupp, A. Stoeckl et al., "CD24 is a marker of exosomes secreted into urine and amniotic fluid," *Kidney International*, vol. 72, no. 9, pp. 1095–1102, 2007.
- [9] J. M. Street, P. E. Barran, C. L. Mackay et al., "Identification and proteomic profiling of exosomes in human cerebrospinal fluid," *Journal of Translational Medicine*, vol. 10, article 5, 2012.
- [10] A. E. Morelli, A. T. Larregina, W. J. Shufesky et al., "Endocytosis, intracellular sorting, and processing of exosomes by dendritic cells," *Blood*, vol. 104, no. 10, pp. 3257–3266, 2004.
- [11] D. G. Nguyen, A. Booth, S. J. Gould, and J. E. K. Hildreth, "Evidence that HIV budding in primary macrophages occurs through the exosome release pathway," *The Journal of Biological Chemistry*, vol. 278, no. 52, pp. 52347–52354, 2003.
- [12] A. M. Booth, Y. Fang, J. K. Fallon et al., "Exosomes and HIV Gag bud from endosome-like domains of the T cell plasma membrane," *Journal of Cell Biology*, vol. 172, no. 6, pp. 923–935, 2006.
- [13] R. Wubbolts, R. S. Leckie, P. T. M. Veenhuizen et al., "Proteomic and biochemical analyses of human B cell-derived exosomes: potential implications for their function and multivesicular body formation," *The Journal of Biological Chemistry*, vol. 278, no. 13, pp. 10963–10972, 2003.
- [14] J. Wolfers, A. Lozier, G. Raposo et al., "Tumor-derived exosomes are a source of shared tumor rejection antigens for CTL cross-priming," *Nature Medicine*, vol. 7, no. 3, pp. 297–303, 2001.
- [15] B. S. Hong, J.-H. Cho, H. Kim et al., "Colorectal cancer cell-derived microvesicles are enriched in cell cycle-related mRNAs that promote proliferation of endothelial cells," *BMC Genomics*, vol. 10, article 556, 2009.
- [16] J. N. Higginbotham, M. Demory Beckler, J. D. Gephart et al., "Amphiregulin exosomes increase cancer cell invasion," *Current Biology*, vol. 21, no. 9, pp. 779–786, 2011.
- [17] A. Clayton and M. D. Mason, "Exosomes in tumour immunity," *Current Oncology*, vol. 16, no. 3, pp. 46–49, 2009.
- [18] H. Valadi, K. Ekström, A. Bossios, M. Sjöstrand, J. J. Lee, and J. O. Lötvall, "Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells," *Nature Cell Biology*, vol. 9, no. 6, pp. 654–659, 2007.
- [19] W. Stoorvogel, M. J. Kleijmeer, H. J. Geuze, and G. Raposo, "The biogenesis and functions of exosomes," *Traffic*, vol. 3, no. 5, pp. 321–330, 2002.
- [20] D. J. Katzmann, G. Odorizzi, and S. D. Emr, "Receptor down-regulation and multivesicular-body sorting," *Nature Reviews Molecular Cell Biology*, vol. 3, no. 12, pp. 893–905, 2002.
- [21] D. G. Meckes Jr. and N. Raab-Traub, "Microvesicles and viral infection," *Journal of Virology*, vol. 85, no. 24, pp. 12844–12854, 2011.
- [22] T. Wurdinger, N. N. Gatsenberger, L. Balaj, B. Kaur, X. O. Breakefield, and D. M. Pegtel, "Extracellular vesicles and their convergence with viral pathways," *Advances in Virology*, vol. 2012, Article ID 767694, 12 pages, 2012.
- [23] J. S. Schorey, Y. Cheng, P. P. Singh, and V. L. Smith, "Exosomes and other extracellular vesicles in host-pathogen interactions," *EMBO Reports*, vol. 16, pp. 24–43, 2015.
- [24] N. Izquierdo-Useros, M. C. Puertas, F. E. Borràs, J. Blanco, and J. Martínez-Picado, "Exosomes and retroviruses: the chicken or the egg?" *Cellular Microbiology*, vol. 13, no. 1, pp. 10–17, 2011.
- [25] S. J. Gould, A. M. Booth, and J. E. K. Hildreth, "The Trojan exosome hypothesis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 19, pp. 10592–10597, 2003.
- [26] Y.-H. Chow, D. Yu, J.-Y. Zhang et al., "gp120-independent infection of CD4⁺ epithelial cells and CD4⁺ T-cells by HIV-1," *Journal of Acquired Immune Deficiency Syndromes*, vol. 30, no. 1, pp. 1–8, 2002.
- [27] D. Marras, L. A. Bruggeman, F. Gao et al., "Replication and compartmentalization of HIV-1 in kidney epithelium of patients with HIV-associated nephropathy," *Nature Medicine*, vol. 8, no. 5, pp. 522–526, 2002.
- [28] A. Pelchen-Matthews, G. Raposo, and M. Marsh, "Endosomes, exosomes and Trojan viruses," *Trends in Microbiology*, vol. 12, no. 7, pp. 310–316, 2004.
- [29] A. K. Khatua, H. E. Taylor, J. E. K. Hildreth, and W. Popik, "Exosomes packaging APOBEC3G confer human immunodeficiency virus resistance to recipient cells," *Journal of Virology*, vol. 83, no. 2, pp. 512–521, 2009.
- [30] M. N. Madison, R. J. Roller, and C. M. Okeoma, "Human semen contains exosomes with potent anti-HIV-1 activity," *Retrovirology*, vol. 11, article 102, 2014.
- [31] T. I. Näslund, D. Paquin-Proulx, P. T. Paredes, H. Vallhov, J. K. Sandberg, and S. Gabrielsson, "Exosomes from breast milk inhibit HIV-1 infection of dendritic cells and subsequent viral transfer to CD4⁺ T cells," *AIDS*, vol. 28, no. 2, pp. 171–180, 2014.
- [32] J. Condeelis, E. Rodriguez-Suarez, N. Embade et al., "Characterization and comprehensive proteome profiling of exosomes secreted by hepatocytes," *Journal of Proteome Research*, vol. 7, no. 12, pp. 5157–5166, 2008.
- [33] C. Théry, M. Boussac, P. Véron et al., "Proteomic analysis of dendritic cell-derived exosomes: a secreted subcellular compartment distinct from apoptotic vesicles," *The Journal of Immunology*, vol. 166, no. 12, pp. 7309–7318, 2001.

- [34] L. Krishnamoorthy, J. W. Bess Jr., A. B. Preston, K. Nagashima, and L. K. Mahal, "HIV-1 and microvesicles from T cells share a common glycome, arguing for a common origin," *Nature Chemical Biology*, vol. 5, no. 4, pp. 244–250, 2009.
- [35] C. Théry, L. Zitvogel, and S. Amigorena, "Exosomes: composition, biogenesis and function," *Nature Reviews Immunology*, vol. 2, no. 8, pp. 569–579, 2002.
- [36] D. E. Ott, "Cellular proteins detected in HIV-1," *Reviews in Medical Virology*, vol. 18, no. 3, pp. 159–175, 2008.
- [37] M. Lenassi, G. Cagney, M. Liao et al., "HIV Nef is secreted in exosomes and triggers apoptosis in bystander CD4⁺ T cells," *Traffic*, vol. 11, no. 1, pp. 110–122, 2010.
- [38] D. M. Pegtel, K. Cosmopoulos, D. A. Thorley-Lawson et al., "Functional delivery of viral miRNAs via exosomes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 14, pp. 6328–6333, 2010.
- [39] Y. Fang, N. Wu, X. Gan, W. Yan, J. C. Morrell, and S. J. Gould, "Higher-order oligomerization targets plasma membrane proteins and HIV gag to exosomes," *PLoS Biology*, vol. 5, no. 6, article e158, 2007.
- [40] R. Cantin, J. Diou, D. Bélanger, A. M. Tremblay, and C. Gilbert, "Discrimination between exosomes and HIV-1: purification of both vesicles from cell-free supernatants," *Journal of Immunological Methods*, vol. 338, no. 1–2, pp. 21–30, 2008.
- [41] E. Chertova, O. Chertov, L. V. Coren et al., "Proteomic and biochemical analysis of purified human immunodeficiency virus type 1 produced from infected monocyte-derived macrophages," *Journal of Virology*, vol. 80, no. 18, pp. 9039–9052, 2006.
- [42] M. Madison and C. Okeoma, "Exosomes: implications in HIV-1 pathogenesis," *Viruses*, vol. 7, no. 7, pp. 4093–4118, 2015.
- [43] M. Mack, A. Kleinschmidt, H. Brühl et al., "Transfer of the chemokine receptor CCR5 between cells by membrane-derived microparticles: a mechanism for cellular human immunodeficiency virus 1 infection," *Nature Medicine*, vol. 6, no. 7, pp. 769–775, 2000.
- [44] T. Rozmyslowicz, M. Majka, J. Kijowski et al., "Platelet- and megakaryocyte-derived microparticles transfer CXCR4 receptor to CXCR4-null cells and make them susceptible to infection by X4-HIV," *AIDS*, vol. 17, no. 1, pp. 33–42, 2003.
- [45] I. Kadiu, P. Narayanasamy, P. K. Dash, W. Zhang, and H. E. Gendelman, "Biochemical and biologic characterization of exosomes and microvesicles as facilitators of HIV-1 infection in macrophages," *Journal of Immunology*, vol. 189, no. 2, pp. 744–754, 2012.
- [46] M. Aqil, A. R. Naqvi, A. S. Bano, and S. Jameel, "The HIV-1 Nef protein binds argonaute-2 and functions as a viral suppressor of RNA interference," *PLoS ONE*, vol. 8, no. 9, Article ID e74472, 2013.
- [47] L. R. Gray, D. Gabuzda, D. Cowley et al., "CD4 and MHC class I down-modulation activities of nef alleles from brain- and lymphoid tissue-derived primary HIV-1 isolates," *Journal of NeuroVirology*, vol. 17, no. 1, pp. 82–91, 2011.
- [48] A. Narayanan, S. Iordanskiy, R. Das et al., "Exosomes derived from HIV-1-infected cells contain trans-activation response element RNA," *The Journal of Biological Chemistry*, vol. 288, no. 27, pp. 20014–20033, 2013.
- [49] M. T. Esser, D. R. Graham, L. V. Coren et al., "Differential incorporation of CD45, CD80 (B7-1), CD86 (B7-2), and major histocompatibility complex class I and II molecules into human immunodeficiency virus type 1 virions and microvesicles: implications for viral pathogenesis and immune regulation," *Journal of Virology*, vol. 75, no. 13, pp. 6173–6182, 2001.
- [50] R. D. Wiley and S. Gummuluru, "Immature dendritic cell-derived exosomes can mediate HIV-1 trans infection," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 3, pp. 738–743, 2006.
- [51] C. Arenaccio, C. Chiozzini, S. Columba-Cabezas et al., "Exosomes from human immunodeficiency virus type 1 (HIV-1)-infected cells license quiescent CD4⁺ T lymphocytes to replicate HIV-1 through a Nef- and ADAM17-dependent mechanism," *Journal of Virology*, vol. 88, no. 19, pp. 11529–11539, 2014.
- [52] C. Arenaccio, S. Anticoli, F. Manfredi, C. Chiozzini, E. Olivetta, and M. Federico, "Latent HIV-1 is activated by exosomes from cells infected with either replication-competent or defective HIV-1," *Retrovirology*, vol. 12, article 87, 2015.
- [53] C. M. Okeoma, A. L. Huegel, J. Lingappa, M. D. Feldman, and S. R. Ross, "APOBEC3 proteins expressed in mammary epithelial cells are packaged into retroviruses and can restrict transmission of milk-borne virions," *Cell Host & Microbe*, vol. 8, no. 6, pp. 534–543, 2010.
- [54] X. Wang, Z. Ao, L. Chen, G. Kobinger, J. Peng, and X. Yao, "The cellular antiviral protein APOBEC3G interacts with HIV-1 reverse transcriptase and inhibits its function during viral replication," *Journal of Virology*, vol. 86, no. 7, pp. 3777–3786, 2012.
- [55] R. K. Holmes, F. A. Koning, K. N. Bishop, and M. H. Malim, "APOBEC3F can inhibit the accumulation of HIV-1 reverse transcription products in the absence of hypermutation. Comparisons with APOBEC3G," *Journal of Biological Chemistry*, vol. 282, no. 4, pp. 2587–2595, 2007.
- [56] A. J. Schumacher, G. Haché, D. A. MacDuff, W. L. Brown, and R. S. Harris, "The DNA deaminase activity of human APOBEC3G is required for Tyl, MusD, and human immunodeficiency virus type 1 restriction," *Journal of Virology*, vol. 82, no. 6, pp. 2652–2660, 2008.
- [57] T. M. Alce and W. Popik, "APOBEC3G is incorporated into virus-like particles by a direct interaction with HIV-1 gag nucleocapsid protein," *The Journal of Biological Chemistry*, vol. 279, no. 33, pp. 34083–34086, 2004.
- [58] S. Stavrou and S. R. Ross, "APOBEC3 proteins in viral immunity," *Journal of Immunology*, vol. 195, no. 10, pp. 4565–4570, 2015.
- [59] J. V. De Carvalho, R. O. De Castro, E. Z. M. Da Silva et al., "Nef neutralizes the ability of exosomes from CD4⁺ T cells to act as decoys during HIV-1 infection," *PLoS ONE*, vol. 9, no. 11, Article ID e113691, 2014.
- [60] A. Tumne, V. S. Prasad, Y. Chen et al., "Noncytotoxic suppression of human immunodeficiency virus type 1 transcription by exosomes secreted from CD8⁺ T cells," *Journal of Virology*, vol. 83, no. 9, pp. 4354–4364, 2009.
- [61] R. T. Bailer, B. Lee, and L. J. Montaner, "IL-13 and TNF- α inhibit dual-tropic HIV-1 in primary macrophages by reduction of surface expression of CD4, chemokine receptors CCR5, CXCR4 and post-entry viral gene expression," *European Journal of Immunology*, vol. 30, no. 5, pp. 1340–1349, 2000.
- [62] D. Creery, W. Weiss, G. Graziani-Bowering et al., "Differential regulation of CXCR4 and CCR5 expression by interleukin (IL)-4 and IL-13 is associated with inhibition of chemotaxis and human immunodeficiency virus (HIV) type 1 replication but not HIV entry into human monocytes," *Viral Immunology*, vol. 19, no. 3, pp. 409–423, 2006.

- [63] J. Li, K. Liu, Y. Liu et al., "Exosomes mediate the cell-to-cell transmission of IFN- α -induced antiviral activity," *Nature Immunology*, vol. 14, no. 8, pp. 793–803, 2013.
- [64] S. D. Barr, J. R. Smiley, and F. D. Bushman, "The interferon response inhibits HIV particle production by induction of TRIM22," *PLoS Pathogens*, vol. 4, no. 2, Article ID e1000007, 2008.
- [65] B. R. Lane, D. M. Markovitz, N. L. Woodford, R. Rochford, R. M. Strieter, and M. J. Coffey, "TNF- α inhibits HIV-1 replication in peripheral blood monocytes and alveolar macrophages by inducing the production of RANTES and decreasing C-C chemokine receptor 5 (CCR5) expression," *The Journal of Immunology*, vol. 163, no. 7, pp. 3653–3661, 1999.
- [66] M. A. Naarding, I. S. Ludwig, F. Groot et al., "Lewis X component in human milk binds DC-SIGN and inhibits HIV-1 transfer to CD4⁺ T lymphocytes," *Journal of Clinical Investigation*, vol. 115, no. 11, pp. 3256–3264, 2005.
- [67] M. A. Naarding, A. M. Dirac, I. S. Ludwig et al., "Bile salt-stimulated lipase from human milk binds DC-SIGN and inhibits human immunodeficiency virus type 1 transfer to CD4⁺ T cells," *Antimicrobial Agents and Chemotherapy*, vol. 50, no. 10, pp. 3367–3374, 2006.
- [68] M. Requena, H. Bouhlal, N. Nasreddine et al., "Inhibition of HIV-1 transmission in trans from dendritic cells to CD4⁺ T lymphocytes by natural antibodies to the CRD domain of DC-SIGN purified from breast milk and intravenous immunoglobulins," *Immunology*, vol. 123, no. 4, pp. 508–518, 2008.
- [69] E. Saeland, M. A. W. P. de Jong, A. A. Nabatov, H. Kalay, T. B. H. Geijtenbeek, and Y. van Kooyk, "MUC1 in human milk blocks transmission of human immunodeficiency virus from dendritic cells to T cells," *Molecular Immunology*, vol. 46, no. 11-12, pp. 2309–2316, 2009.
- [70] P. Hong, M. R. Ninonuevo, B. Lee, C. Lebrilla, and L. Bode, "Human milk oligosaccharides reduce HIV-1-gp120 binding to dendritic cell-specific ICAM3-grabbing non-integrin (DC-SIGN)," *British Journal of Nutrition*, vol. 101, no. 4, pp. 482–486, 2009.
- [71] M. N. Madison, P. H. Jones, and C. M. Okeoma, "Exosomes in human semen restrict HIV-1 transmission by vaginal cells and block intravaginal replication of LP-BM5 murine AIDS virus complex," *Virology*, vol. 482, pp. 189–201, 2015.
- [72] M. J. Stax, T. van Montfort, R. R. Sprenger et al., "Mucin 6 in seminal plasma binds DC-SIGN and potentially blocks dendritic cell mediated transfer of HIV-1 to CD4⁺ T-lymphocytes," *Virology*, vol. 391, no. 2, pp. 203–211, 2009.
- [73] N. Izquierdo-Useros, M. Naranjo-Gómez, J. Archer et al., "Capture and transfer of HIV-1 particles by mature dendritic cells converges with the exosome-dissemination pathway," *Blood*, vol. 113, no. 12, pp. 2732–2741, 2009.
- [74] N. Izquierdo-Useros, M. Naranjo-Gómez, I. Erkizia et al., "HIV and mature dendritic cells: Trojan exosomes riding the Trojan horse?" *PLoS Pathogens*, vol. 6, no. 3, Article ID e1000740, 2010.
- [75] L. Abraham and O. T. Fackler, "HIV-1 Nef: a multifaceted modulator of T cell receptor signaling," *Cell Communication and Signaling*, vol. 10, no. 1, article 39, 2012.
- [76] L. Urbanelli, A. Magini, S. Buratta et al., "Signaling pathways in exosomes biogenesis, secretion and fate," *Genes*, vol. 4, no. 2, pp. 152–170, 2013.
- [77] X. Xiang, Y. Liu, X. Zhuang et al., "TLR2-mediated expansion of MDSCs is dependent on the source of tumor exosomes," *The American Journal of Pathology*, vol. 177, no. 4, pp. 1606–1610, 2010.
- [78] T. N. Bukong, F. Momen-Heravi, K. Kodys, S. Bala, and G. Szabo, "Exosomes from hepatitis C infected patients transmit HCV infection and contain replication competent viral RNA in complex with Ago2-miR122-HSP90," *PLoS Pathogens*, vol. 10, no. 10, Article ID e1004424, 2014.