

Comparing the Efficacy of Three Different Nano-scale Bone Substitutes: *In vivo* Study

Abstract

Background: Synthetic biocompatible bone substitutions have been used widely for bone tissue regeneration as they are safe and effective. The aim of this animal study is to compare the effectiveness of three different biocompatible bone substitutes, including nano-hydroxyapatite (nano-HA) nano-bioglass (nano-BG) and forstrite scaffolds. **Materials and Methods:** In this interventional and experimental study, four healthy dogs were anesthetized, and the first to fourth premolars were extracted in each quadrant. After healing, the linear incision on the crestal ridge from molar to anterior segment prepared in each quadrant and 16 defects in each dog were prepared. Nano-HA, nano-BG, and forstrite scaffold was prepared according to the size of defects and placed in the 12 defects randomly, four defects remained as a control group. The dogs were sacrificed in four time intervals (15, 30, 45, and 60 days after) and the percentage of different types of regenerated bones (lamellar and woven) and connective tissue were recorded in histological process. The data were analyzed using Mann–Whitney test ($\alpha = 0.05$). **Results:** The difference in nano-HA and nano-BG with the control group was significant in three-time intervals regarding the amount of bone formation ($P < 0.01$). After 15 days, the nano-HA showed the highest amount of woven and lamellar bone regeneration (18.37 ± 1.06 and 30.44 ± 0.54). **Conclusion:** Nano-HA and nano-BG groups showed a significant amount of bone regeneration, especially after 30 days, but paying more surveys and observation to these materials as bone substitutes seem to be needed.

Keywords: Bone regeneration, nano-bioglass, nano-hydroxyapatite, forstrite

Introduction

Bone or massive tissue defects (caused by trauma, tumors, or degenerative disease, etc.) is still a major problem which needs sufficient bone with acceptable quality and quantity to be repaired.^[1,2] As tissue engineering developed, many opportunity has been made for reconstruction bone defects. There are many patients who are just now discovering that they can use implants if their implant sites reconstructed.^[3]

To achieve that goal, autogenously bone grafts have been examined for reconstructing bone defects. However, limited bone stocks, imposing additional surgical intervention, and following complications at the donor site are some of the major obstacles.^[1,4] To conquer those limitations bone graft substitutes were introduced. Biocompatibility, bio-restorability, and osteogenicity are the most characteristic of an appropriate substitute.^[5] Hence, allogeneic grafts was

tested as alternatives such as irradiated bone from cadavers (bone mineral from bone banks) and demineralized bone matrix (bone mineral dissolved away using acids), but probably immunological cross-actions might have happened.^[6]

To advance the properties, synthetic biocompatible bone substitutions have been used. they seem to be a safe and effective option in procedures such as filling with fracture defects and resolution of long bone nonunions but still some doubts remain.^[5,7] They mostly consist of calcium, phosphate, ceramics, and organic materials which can provide a stable situation for supporting bone responses and reconstructions.^[8] Although they fulfill the goal of repairing smaller defects, they are not appropriate for repairing extended bone defects.^[9]

The examples are bioactive glass (BG) ceramics and calcium phosphate ceramics such as hydroxyapatite (HA), β tricalcium phosphate, or biphasic calcium phosphate.^[10]

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How to cite this article: Razavi SM, Rismanchian M, Jafari-pozve N, Nosouhian S. Comparing the Efficacy of Three Different Nano-scale Bone Substitutes: *In vivo* Study. *Adv Biomed Res* 2017;6:64.

Received: November, 2014. **Accepted:** August, 2015.

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Access this article online

Website: www.advbiores.net

DOI: 10.4103/2277-9175.192627

Quick Response Code:



HA is an important component of the bone structure. Due to its biocompatibility and crystallographic structure HA is used to produce a synthetic bone substitute.^[11,12] The efficacy of HA in the bone regeneration approved by studies.^[13,14] Dense HA could not be degraded and remodeled in the host which was an obvious flaw for a suitable biomaterial^[4,15] so nano-HA was introduced as a favorable bone substitute. Compared with micro-HA, this material is highly biocompatible and biodegradable and could be rapidly substituted by the host.^[16-18]

BG is another popular bone substitute which has been emerged since 1971.^[19] The capability of BG in stimulating osseointegration and forming new bone has been confirmed in different studies.^[20-22] Their well surface characteristics induce the osteoblasts to generate an amorphous calcium phosphate layer which can turn to a biological hydroxycarbonate apatite and cause interfacial bonding.^[19]

Gatti *et al.* examined the clinical use of BG in treating dental extraction sites before loading implants. New bone formation and biodegradation of the BG was observed.^[23]

In recent years, researchers have declared that nano-bioglass (Nano-BG) has showed superiority over micro-bioglass in repairing bone defects.^[24]

Using a porous scaffold with interconnected spaces provides sufficient room for cell migration and adhesion and the ingrowth of new bone tissue.^[25] An ideal scaffold for bone regeneration would share loads with surrounded bone. They cannot do this when the bone defect is subjected to cyclic loads if they are brittle.^[26] Forstrite (Mg_2SiO_4) is one of them. It is well biocompatible nano-composite with high bonding strength, fracture toughness and when combined with the polycaprolactone could result in a scaffold structure useful for bone tissue repair.^[27-30] Tavangarian evaluated the bioactivity of forstrite, and their result revealed that Forstrite's mechanical properties make a good situation for load bearing applications in bone implant materials.^[27] Based on Diba *et al.* conclusion the forstrite scaffold provides the most suitable morphology with 30% weight and steady structure.^[29]

There are not enough animal studies with conclusive results regarding the efficacy of nano-particles. As forstrite is a newly invented biomaterial with nano-structural properties, and no study has compared the biological effect of forstrite with both nano-hydroxyapatite (Nano-HA) and nano-BG, this study was designed to compare the efficacy of nano-HA, nano-BG, and forstrite scaffold in repairing bone defects.

Materials and Methods

Surgical procedure

In this interventional and experimental study, four healthy male dogs aged 1–4 and weighed 32–46 kg were anesthetized initially with 10 mg/kg ketamine (ketamine

HCL, Alfasan, Woerden, Holland) and 0.15 mg/kg rampone. Maintenance followed under general anesthesia by 5% halothane (Halothane, Bp, Nicholas Piramal India Limited, India) and N_2O . First to fourth premolars were extracted respectfully in each quadrant with the preservation of surrounding bone, according to Helsinki roles. Parallel periapical radiographs were taken with extension cone paralleling film holders (Rinn Co., USA) to evaluate the healing of tooth extraction sites in further 3 months.

After appropriate bone healing phase, 3.6 ml lidocain (Darou Pakhsh Pharmaceutical Mfg. Co. Tehran, Iran) was derived in the mucobuccal fold. Linear incision was prepared on the crestal ridge from molar to anterior segment and a full thickness mucoperiosteal flap was elevated by a mucoperiosteal elevator. Two defects with 5 mm depth and 5 mm diameter on the crestal ridge and two similar defects on the buccal surface of the ridge were surgically created using trephine #5. As a result, four defects in each quadrant and 16 defects in each dog were created overall. These 16 defects were randomly divided into four groups and filled with:

- i. Nano-BG ($CaNaO_6PSi$) (SCHOTT Vitryxx, Mainz, Germany) amorphous powder form with <250 nm (BET) particle size
- ii. Nano-HA ($Ca_5(OH)(PO_4)_3$) (Sigma–Aldrich, NY, USA) amorphous powder form with <200 nm (BET) particle size, >9.4 m²/g surface area
- iii. Forstrite scaffold (Mg_2SiO_4) (New Nano, Isfahan University of Technology, Isfahan, Iran) with porosity of <83%, mean pore size of <25–45 nm
- iv. Control group.

All the mentioned biomaterials were purchased from their exclusive company at least 1-year before the expiration date. The above nano-HA and nano-BG powders were mixed with distilled water according to the manufacturers' instructions of each material; the blocks of forstrite scaffold prepared according to the size of defects and rinsed with distilled water too. Finally, all of those materials were placed separately in different defects. Therefore, 16 defects in each dog were filled randomly by nano-HA, nano-BG and forstrite scaffold and four defects were left as a control group. Then, all the defects were covered by a nonresorbable membrane (PTFE Whatman, Kent, UK).

The dogs were sacrificed at four-time intervals (15, 30, 45, and 60 days, each dog at each time point). Fifteen days after the surgery, a lethal injection of 40 ml pentobarbital sodium at 100 mg/ml in 290 g/1000 ml spiritus fortis, 100 mg/kg was given to one of the dogs. All 16 samples were extracted using trephine #8 with sufficient amount of surrounding bone. These procedures were also carried out in 30, 45, and 60 days after the surgery.

Histological analysis

Extracted specimens were kept in glutaraldehyde solution for 6 h. Longitudinal ground sections were prepared using Microtome (Accutom-50, Stuers, Copenhagen, Denmark). The samples were stained by Hematoxylin and Eosin stain (H and E) and mounted on the histological lams. All stained specimens were investigated under an optical microscope ($\times 40$) (Zeiss, Germany), and percentage of different types of regenerated bones (lamellar and woven), and connective tissue were recorded. The cross sections of specimens were surveyed by Adobe Photoshop 7.0 (SanJose, CA, USA) and the amount of the regenerated bone was re-evaluated to confirm data. Data analysis were performed by SPSS software version 15 (IBM Corporation, NY, USA) and Mann–Whitney test ($\alpha = 0.05$).

Ethics

This study was approved by the Animal Department of Torabinejad Dental Research Center and Local Ethical Committee of the Isfahan University of Medical Science.

Results

Using the Mann–Whitney test showed that the difference in nano-HA and nano-BG with the control group was significant in three-time intervals regarding the amount of bone formation ($P < 0.01$). Table 1 represents the hard tissue responses to the tested biomaterials at different intervals.

After 15 days, both nano-HA and nano-BG besides forsterite scaffold showed the significant difference with the control group in amount of woven bone production;

Table 1: The hard tissue regenerative response to different tested biomaterials at different intervals

Intervals	Groups	Mean \pm SD of woven bone	Mean \pm SD of lamellar bone	Mean \pm SD of connective tissue
15	Nano-HA	18.37 \pm 1.06	30.44 \pm 0.54	51.81 \pm 0.36
	Nano-BG	16.53 \pm 1.07	26.84 \pm 6.27	56.66 \pm 6.37
	Forstrite scaffold	15.87 \pm 0.83	25.22 \pm 3.84	58.97 \pm 4.18
	Control	15.37 \pm 0.50	21.59 \pm 0.66	63.03 \pm 0.83
30	Nano-HA	19.87 \pm 3.22	29.53 \pm 1.41	50.59 \pm 2.65
	Nano-BG	23.09 \pm 2.12	28.53 \pm 1.16	48.37 \pm 1.18
	Forstrite scaffold	18.28 \pm 2.35	26.56 \pm 6.68	54.78 \pm 8.63
	Control	16.50 \pm 0.71	22.37 \pm 3.44	61.12 \pm 3.81
45	Nano-HA	16.69 \pm 1.08	27.94 \pm 2.02	55.44 \pm 2.64
	Nano-BG	16.00 \pm 2.79	23.44 \pm 3.32	60.57 \pm 3.82
	Forstrite scaffold	16.37 \pm 1.24	25.22 \pm 3.28	58.37 \pm 2.61
	Control	16.81 \pm 1.64	25.81 \pm 2.75	57.37 \pm 1.47
60	Nano-HA	15.84 \pm 1.49	22.44 \pm 1.60	60.72 \pm 2.93
	Nano-BG	19.66 \pm 5.98	29.62 \pm 2.60	50.72 \pm 8.00
	Forstrite scaffold	14.53 \pm 1.31	26.66 \pm 4.25	58.19 \pm 5.66
	Control	18.78 \pm 2.48	25.44 \pm 5.27	57.03 \pm 5.70

SD: Standard deviation, Nano-HA: Nano-hydroxyapatite, Nano-BG: Nano-bioglass

also the results of nano-BG and forstrite scaffold had significant difference with nano-HA too ($P < 0.05$). The amount of lamellar was noticeable in forstrite and nano-HA with the control group ($P < 0.05$). The nano-HA showed the highest amount of woven and lamellar bone regeneration (18.37 \pm 1.06 and 30.44 \pm 0.54) [Table 1]. The mean percentage of connective tissue was the highest in control group (63.03 \pm 0.83) and the difference between other groups was not significant ($P > 0.05$) except forstrite and control group with nano-HA ($P < 0.05$).

After 30 days of the experiment, both nano-BG and nano-HA revealed a significant difference in the amount of connective tissue, woven and lamellar bone regeneration with the control group ($P < 0.05$). Another significance is between forstrite scaffold and nano-BG ($P < 0.05$). The nano-BG showed the highest rate of woven bone (23.09 \pm 2.12) and nano-HA showed the highest rate of lamellar bone regeneration (29.53 \pm 1.41). The percentage of connective tissue was the highest in the control group (61.12 \pm 3.81) and the lowest in nano-BG (48.37 \pm 1.18) [Table 1].

After 45 days, there was a significant lamellar bone difference between nano-HA and nano-BG groups ($P < 0.05$). The amount of lamellar bone production was the highest in nano-HA (27.94 \pm 2.02) but the amount of connective tissue was the highest in nano-BG (60.57 \pm 3.82) [Table 1]. The nano-HA reflected a significant difference with nano-BG and forstrite ($P < 0.05$), but the significance between nano-BG and control group should not be overlooked ($P < 0.05$).

After 60 days, the difference in woven bone production was significant among forstrite and nano-HA with the control group ($P < 0.05$) however they (forstrite and nano-HA) had discrepancies with each other ($P < 0.05$). Another significance was among forstrite and nano-BG with the nano-HA group in lamellar bone regeneration ($P < 0.05$). The amount of woven and lamellar bone was the highest in nano-BG (19.66 \pm 5.93 and 29.62 \pm 2.60). Also, the rate of connective tissue was the highest in nano-HA (60.72 \pm 8.00) [Table 1].

It should be noted that the mean amount of woven regenerated bone reached to its highest rate in nano-BG group after 30 days. Furthermore, the highest rate of lamellar bone regeneration was after 60 days by seeding nano-BG. Furthermore, Figures 1 and 2 show the microscopic appearance of regenerated woven and lamellar bones in four groups after 15 and 45 days [Figures 1 and 2].

Discussion

Tissue engineering techniques currently have offered great fortune to meet the clinical need for bone substitutes. Besides that, there has been growing interest in using three-dimensional scaffolds for supporting bone regeneration.^[6,31-33]

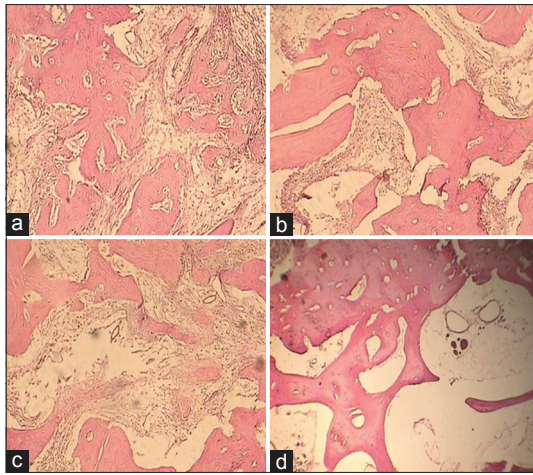


Figure 1: Histopathology of the samples after 15 days. (a) nano-bioglass, (b) nano-hydroxyapatite, (c) forstrite, (d) control

Forstrite is a nano-crystal bioceramic made of talc, alumina, and magnesium carbonate with not only lots of macropores but also plentiful micropores (on the scale of 1–10 μm) on the macroporous walls.^[29]

Zamet, Froum, and Nandi studies approved the positive effects of bioglass on bone formation.^[14,34,35]

Maybe it is needed to mention that in the initial stage of bone reconstruction, the matrix proteins can be attracted by nano-bone granules so a vascular rich protein matrix would be formed. In consequences, the osteogenesis happens on this matrix, and the final bone is regenerated. As nano-scale structures provide a larger surface area, the amount of attracted proteins would be much more. This can be an explanation why the use of nano-materials can accelerate the bone regeneration. On the other hand, these larger surface areas pretend as biological materials.^[36] Hence, the mechanical reliability and osseointegration could be improved, and the osteogenic differentiation of stem cells would be induced.^[37,38] For instance, Huang *et al.* claimed in their results that the size (micro/nano) of HA is an important factor in stimulation of the osteogenic differentiation.^[39]

Nano-hydroxyapatite

Based on present results, application of nano-HA provided promising results after 15, 30, and 45 days of reconstruction of bone defects which was in accordance to some studies. Götz *et al.* concluded that nano-porous HA materials show osteoconductive capacity and have the capability to integrate with the host bone.^[40] In Schwarz study, different cases with intrabony defects around dental implants were investigated. They applied nano-crystalline hydroxyapatite and bovine-derived xenograft in combination with a collagen membrane (BDX + BG). The final result was repair of all bone defects repaired and reduction of pocket depth.^[41] This result can be explained by osseointegration and biocompatibility of nano-HA bone material, and is in

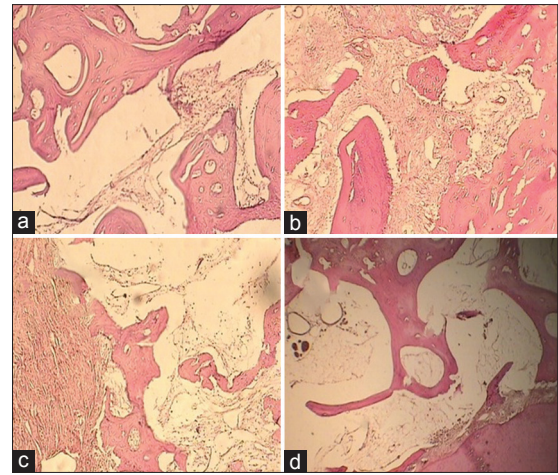


Figure 2: Histopathology of the samples after 45 days. (a) nano-bioglass, (b) nano-hydroxyapatite, (c) forstrite, (d) control

agreement with the Welch *et al.*,^[42] den Boer *et al.*,^[43] and Kruse *et al.*^[44] studies. Also, the present study revealed that the use of nano-HA showed better result than nano-BG, which is somehow in accordance with Rodenas–Roshina J examination. They conclude that presence of bioactive particles is not necessary to reach the higher amount of bone regeneration.^[31]

Nano-bioglass

Based on current results, the induced regenerated bone tissue by nano-BG was at second place after 15, 30, and 60 days. The bioactivity of BG is initiated exactly after mixing with saline or blood, and silicon oxide bonds break down. As a result, the silicic acid can be aggregated on the surface of particles and form a negatively charged gel. As time lasts, calcium hydroxide would be formed on these surfaces to form a new apatite layer which initiates the bioactivity of BG.^[45] These can be a good reason why the formation of new bone revealed a significant difference in nano-BG compared to the control group. However, the means of regenerated bone tissue was not promising after 45 days which might be due to inappropriate implantation of the biomaterial. Azenha *et al.* concluded that new bone formation around the materials is influenced by their location within the bone.^[46]

Forstrite scaffold

The published *in vivo* research about the bioactivity of forstrite is not extensive as it has been newly invented. Based on obtained results, the highest amount of regenerated bone tissue was only observed after 45 days, and at the other three time intervals it was at the third place of biomaterials ranking. That might be due to its degenerative properties and apatite formation which are not remarkable. In an *in vitro* study, Ni and Chang compared degenerative properties and apatite formation of Mg_2SiO_4 , the major composition of forstrite, with two other biomaterials. Their result confirmed that the its mentioned properties were poor.^[47]

Conclusion

With the limitation of this study, the nano-HA group is the first choice which showed a significant amount of bone regeneration, especially in the first 30 days. The second recommended choice is nano-BG but paying more surveys to these materials as bone substitutes are needed.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

References

- El-Gendy R, Yang XB, Newby PJ, Boccaccini AR, Kirkham J. Osteogenic differentiation of human dental pulp stromal cells on 45S5 Bioglass® based scaffolds *in vitro* and *in vivo*. *Tissue Eng Part A* 2013;19:707-15.
- Li LJ, Liu N, Shi JG, Liu Q, Jia LS, Yuan W. Osteogenic scaffolds for bone reconstruction. *Biores Open Access* 2012;1:137-44.
- Sándor GKB, Lindholm TC, Clokie CM. Bone Regeneration of the Craniomaxillofacial and Dento-alveolar Skeletons in the Framework of Tissue Engineering. Finland: University of Oulu; 2003.
- Cottrell DA, Wolford LM. Long-term evaluation of the use of coralline hydroxyapatite in orthognathic surgery. *J Oral Maxillofac Surg* 1998;56:935-41.
- Kweon H, Lee KG, Chae CH, Balázs C, Min SK, Kim JY, *et al.* Development of nano-hydroxyapatite graft with silk fibroin scaffold as a new bone substitute. *J Oral Maxillofac Surg* 2011;69:1578-86.
- Jones JR, Lin S, Yue S, Lee PD, Hanna JV, Smith ME, *et al.* Bioactive glass scaffolds for bone regeneration and their hierarchical characterisation. *Proc Inst Mech Eng H* 2010;224:1373-87.
- Välimäki VV, Aro HT. Molecular basis for action of bioactive glasses as bone graft substitute. *Scand J Surg* 2006;95:95-102.
- Sculean A, Barbe G, Chiantella GC, Arweiler NB, Berakdar M, Brex M. Clinical evaluation of an enamel matrix protein derivative combined with a bioactive glass for the treatment of intrabony periodontal defects in humans. *J Periodontol* 2002;73:401-8.
- Henkel KO, Gerber T, Lenz S, Gundlach KK, Bienengraber V. Macroscopical, histological, and morphometric studies of porous bone-replacement materials in minipigs 8 months after implantation. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006;102:606-13.
- Le Guéhennec L, Layrolle P, Daculsi G. A review of bioceramics and fibrin sealant. *Eur Cell Mater* 2004;8:1-10.
- Hing KA, Revell PA, Smith N, Buckland T. Effect of silicon level on rate, quality and progression of bone healing within silicate-substituted porous hydroxyapatite scaffolds. *Biomaterials* 2006;27:5014-26.
- Williams RL, Brown SA, Merritt K. Electrochemical studies on the influence of proteins on the corrosion of implant alloys. *Biomaterials* 1988;9:181-6.
- Soccol AT, Bettega S, Noronha L, Sass S, Soccol VT, Scholz MR, *et al.* Defect repair in rat mandible with hydroxyapatite cement compared to small intestine submucosa. *Braz J Otorhinolaryngol* 2006;72:195-9.
- Nandi SK, Kundu B, Ghosh SK, De DK, Basu D. Efficacy of nano-hydroxyapatite prepared by an aqueous solution combustion technique in healing bone defects of goat. *J Vet Sci* 2008;9:183-91.
- Pohunková H, Adam M. Reactivity and the fate of some composite bioimplants based on collagen in connective tissue. *Biomaterials* 1995;16:67-71.
- Strietzel FP, Reichart PA, Graf HL. Lateral alveolar ridge augmentation using a synthetic nano-crystalline hydroxyapatite bone substitution material (Ostim): Preliminary clinical and histological results. *Clin Oral Implants Res* 2007;18:743-51.
- Webster TJ, Siegel RW, Bizios R. Osteoblast adhesion on nanophase ceramics. *Biomaterials* 1999;20:1221-7.
- Kim HW, Kim HE, Salih V. Stimulation of osteoblast responses to biomimetic nanocomposites of gelatin-hydroxyapatite for tissue engineering scaffolds. *Biomaterials* 2005;26:5221-30.
- Saino E, Grandi S, Quartarone E, Maliardi V, Galli D, Bloise N, *et al.* *In vitro* calcified matrix deposition by human osteoblasts onto a zinc-containing bioactive glass. *Eur Cell Mater* 2011;21:59-72.
- Brandão SM, Schellini SA, Moraes AD, Padovani CR, Pellizzon CH, Peitl O, *et al.* Biocompatibility analysis of bioglass® 45S5 and biosilicate® implants in the rabbit eviscerated socket. *Orbit* 2012;31:143-9.
- Chitsazi MT, Shirmohammadi A, Faramarzie M, Pourabbas R, Rostamzadeh AN. A clinical comparison of nano-crystalline hydroxyapatite (Ostim) and autogenous bone graft in the treatment of periodontal intrabony defects. *Med Oral Patol Oral Cir Bucal* 2011;16:e448-53.
- Stavropoulos A, Sima C, Sima A, Nyengaard J, Karring T, Sculean A. Histological evaluation of healing after transalveolar maxillary sinus augmentation with bioglass and autogenous bone. *Clin Oral Implants Res* 2012;23:125-31.
- Gatti AM, Simonetti LA, Monari E, Guidi S, Greenspan D. Bone augmentation with bioactive glass in three cases of dental implant placement. *J Biomater Appl* 2006;20:325-39.
- Fathi MD. Bioactive glass nanopowder and bioglass coating for biocompatibility improvement of metallic implant. *J Mater Process Technol* 2009;209:1385-91.
- Su J, Cao L, Yu B, Song S, Liu X, Wang Z, *et al.* Composite scaffolds of mesoporous bioactive glass and polyamide for bone repair. *Int J Nanomedicine* 2012;7:2547-55.
- Jones JR. Review of bioactive glass: From Hench to hybrids. *Acta Biomater* 2013;9:4457-86.
- Tavangarian F, Emadi R. Nanostructure effects on the bioactivity of forsterite bioceramic. *Mater Lett* 2011;65:740-3.
- Tavangarian F, Emadi R. Synthesis of nanocrystalline forsterite (Mg₂SiO₄) powder by combined mechanical activation and thermal treatment. *Mater Res Bull* 2011;45:388-91.
- Diba M, Fathi MH, Kharaziha M. Novel forsterite/polycaprolactone nanocomposite scaffold for tissue engineering applications. *Mater Lett* 2011;65:1931-4.
- Emadi R, Tavangarian F, Roohani Esfahani SI. Biodegradable and bioactive properties of a novel bone scaffold coated with nanocrystalline bioactive glass for bone tissue engineering. *Mater Lett* 2010;64:1528-31.
- Ródenas-Rochina J, Ribelles JL, Lebourg M. Comparative study of PCL-HAp and PCL-bioglass composite scaffolds for bone tissue engineering. *J Mater Sci Mater Med* 2013;24:1293-308.
- Deb S, Mandegaran R, Di Silvio L. A porous scaffold for bone tissue engineering/45S5 Bioglass derived porous scaffolds for co-culturing osteoblasts and endothelial cells. *J Mater Sci Mater Med* 2010;21:893-905.

33. Hafezi F, Hosseinnejad F, Fooladi AA, Mafi SM, Amiri A, Nourani MR. Transplantation of nano-bioglass/gelatin scaffold in a non-autogenous setting for bone regeneration in a rabbit ulna. *J Mater Sci Mater Med* 2012;23:2783-92.
34. Froum S, Cho SC, Rosenberg E, Rohrer M, Tarnow D. Histological comparison of healing extraction sockets implanted with bioactive glass or demineralized freeze-dried bone allograft: A pilot study. *J Periodontol* 2002;73:94-102.
35. Zamet JS, Darbar UR, Griffiths GS, Bulman JS, Brägger U, Bürgin W, *et al.* Particulate bioglass as a grafting material in the treatment of periodontal intrabony defects. *J Clin Periodontol* 1997;24:410-8.
36. Quinones CL. Utilization of a bioactive synthetic particulate for periodontal therapy and bone augmentation techniques. *Pract Periodontics Aesthet Dent* 1997;9:1-9.
37. Tadjoeidin ES, de Lange GL, Lyaruu DM, Kuiper L, Burger EH. High concentrations of bioactive glass material (BioGran®) vs. autogenous bone for sinus floor elevation. *Clin Oral Implants Res* 2002;13:428-36.
38. Liu Y, Wang G, Cai Y, Ji H, Zhou G, Zhao X, *et al.* *In vitro* effects of nanophase hydroxyapatite particles on proliferation and osteogenic differentiation of bone marrow-derived mesenchymal stem cells. *J Biomed Mater Res A* 2009;90:1083-91.
39. Huang Y, Zhou G, Zheng L, Liu H, Niu X, Fan Y. Micro-/nano-sized hydroxyapatite directs differentiation of rat bone marrow derived mesenchymal stem cells towards an osteoblast lineage. *Nanoscale* 2012;4:2484-90.
40. Götz W, Gerber T, Michel B, Lossdörfer S, Henkel KO, Heinemann F. Immunohistochemical characterization of nanocrystalline hydroxyapatite silica gel (NanoBone (r)) osteogenesis: A study on biopsies from human jaws. *Clin Oral Implants Res* 2008;19:1016-26.
41. Schwarz F, Bieling K, Latz T, Nuesry E, Becker J. Healing of intrabony peri-implantitis defects following application of a nanocrystalline hydroxyapatite (Ostim) or a bovine-derived xenograft (Bio-Oss) in combination with a collagen membrane (Bio-Gide). A case series. *J Clin Periodontol* 2006;33:491-9.
42. Welch RD, Berry BH, Crawford K, Zhang H, Zobitz M, Bronson D, *et al.* Subchondral defects in caprine femora augmented with *in situ* setting hydroxyapatite cement, polymethylmethacrylate, or autogenous bone graft: Biomechanical and histomorphological analysis after two-years. *J Orthop Res* 2002;20:464-72.
43. den Boer FC, Wippermann BW, Blokhuis TJ, Patka P, Bakker FC, Haarman HJ. Healing of segmental bone defects with granular porous hydroxyapatite augmented with recombinant human osteogenic protein-1 or autologous bone marrow. *J Orthop Res* 2003;21:521-8.
44. Kruse A, Jung RE, Nicholls F, Zwahlen RA, Hämmerle CH, Weber FE. Bone regeneration in the presence of a synthetic hydroxyapatite/silica oxide-based and a xenogenic hydroxyapatite-based bone substitute material. *Clin Oral Implants Res* 2011;22:506-11.
45. Gosain AK; Plastic Surgery Educational Foundation DATA Committee. Bioactive glass for bone replacement in craniomaxillofacial reconstruction. *Plast Reconstr Surg* 2004;114:590-3.
46. Azenha MR, Peitl O, Barros VM. Bone response to biosilicates with different crystal phases. *Braz Dent J* 2010;21:383-9.
47. Ni S, Chang J. *In vitro* degradation, bioactivity, and cytocompatibility of calcium silicate, dimagnesium silicate, and tricalcium phosphate bioceramics. *J Biomater Appl* 2009;24:139-58.