



Hybrid Assembly of the Quorum-Quenching Isolate *Variovorax paradoxus* VAI-C Genome Sequence

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ABSTRACT *Variovorax paradoxus* VAI-C was isolated due to its ability to utilize acyl-homoserine lactones (AHLs) as the sole source of carbon, energy, and nitrogen. Here, we present a hybrid assembly of the *V. paradoxus* VAI-C genome sequence, consisting of a primary chromosome, a secondary chromid, and a plasmid.

Variovorax paradoxus VAI-C was previously isolated (1) from soil adjacent to the Bowen Science Building at the University of Iowa, based on its ability to utilize the quorum-sensing signal molecule *N*-(3-oxohexanoyl)-L-homoserine lactone (3OC6-HSL; “*Vibrio fischeri* autoinducer-1”) as the sole source of carbon and energy. That study was also the first demonstration of quorum quenching (acyl-homoserine lactone [AHL] signal inactivation) by the acylase mechanism (1, 2). Other *V. paradoxus* strains also utilize AHLs (3), but not as rapidly as strain VAI-C does. Other species of bacteria also grow utilizing AHL substrates, albeit more slowly, via the same AHL acylase mechanism (4–6).

A pure culture of *V. paradoxus* VAI-C from a frozen stock was streaked onto a yeast extract (YE) agar plate (5 g/liter YE; Fisher Scientific). A culture derived from a single colony from this streak plate was grown at room temperature overnight in YE broth, and DNA was purified using the high-molecular-weight DNA protocol outlined for *Escherichia coli* (<https://www.protocols.io/view/ultra-long-read-sequencing-protocol-for-rad004-mrx57n>) (7). The genomic DNA quantity and quality were assessed spectrophotometrically using the NanoDrop 1 (Thermo Fisher). Moderate shearing of the DNA was performed using a sterile 26-gauge needle (Thermo Fisher); libraries were prepared using the rapid barcoding kit (catalog number SQK-RBK004) and sequenced using a MIN-106 flow cell (R9.4.1) in an Oxford Nanopore MinION instrument. Four sequencing runs were performed on barcoded libraries derived from the same genomic sample in separate flow cells. These data were combined for assembly after demultiplexing and base calling. For all subsequent data-processing steps, default parameters were used unless otherwise noted. MinION reads were base called in Guppy v2.3.1 using the Flipflop v1.1.0 (currently referred to as Flappie or high-accuracy base calling [HAC]) model and demultiplexed in Deepbiner v0.2.0 (8). Barcodes and adapters were removed using Porechop v0.2.4 (8). A total of 194,423 Nanopore reads were obtained for *V. paradoxus* VAI-C (average read length, 4,557.57 ± 5,302.50; coverage, 93.88×).

The same DNA sample with additional needle shearing was used to generate a 250- to 300-bp library with the Nextera DNA Flex library preparation kit (LPK), which was sequenced on the Illumina iSeq platform (2 × 150 bp). A total of 4,818,968 Illumina reads were obtained for *V. paradoxus* VAI-C (average read length, 132.20 ± 30.34; coverage, 67.49×) FastQC v0.11.8 was used for quality assessment of these data (9), and trimming was performed in Trimmomatic v0.38.0 (10). Assemblies of *V. paradoxus* VAI-C were created using a hybrid approach in Unicycler v0.4.8.0 (11) on the North America Galaxy hub (<http://usegalaxy.org>) (12). The final circularization of the genomes was completed using Unicycler.

The *V. paradoxus* strain VAI-C genomic DNA was assembled into three circular contigs, a 6,666,455-bp primary chromosome, a 2,479,635-bp chromid, and a 292,938-bp

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plasmid. The Prokaryotic Genome Annotation Pipeline (PGAP) by NCBI (13) identified 8,575 protein-coding open reading frames across the three contigs, along with 61 predicted RNA genes. Replication and partition machinery (ParAB or RepAB) was identified in all three contigs, and putative conjugal transfer machinery was identified near the replication locus on the chromid. The three contigs have the following G+C contents: 69.57% for the chromosome, 68.56% for the chromid, and 60.6% for the plasmid. The average G+C content for the overall genome is 69.0%. Three incomplete prophage elements and one questionable prophage-like element were identified using PHASTER (<https://phaster.ca/>) (14). Eight loci were annotated as encoding penicillin acylases with homology to previously identified AHL acylase proteins from *Pseudomonas aeruginosa* (PA0305, HacB; PA1032, QuiP; and PA2385, PvdQ). The availability of this genome sequence will permit further investigation of the variation in quorum-quenching activity among *Variovorax* isolates.

There is substantial diversity in genome structure in the genus *Variovorax* (3, 15). The reported strain VAI-C genome assembly is the largest finished *Variovorax paradoxus* genome sequence and is the only assembly that contains a putative chromid and plasmid.

Data availability. The assemblies and sequence data have been uploaded to the NCBI database. *Variovorax paradoxus* VAI-C can be found under BioProject number [PRJNA667957](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA667957), BioSample number [SAMN16392950](https://www.ncbi.nlm.nih.gov/biosample/SAMN16392950), and assembly numbers [CP063166](https://www.ncbi.nlm.nih.gov/assembly/CP063166) through [CP063168](https://www.ncbi.nlm.nih.gov/assembly/CP063168). The read data can be found under SRA accession numbers [SRX9260397](https://www.ncbi.nlm.nih.gov/sra/SRX9260397) and [SRX9260396](https://www.ncbi.nlm.nih.gov/sra/SRX9260396), including demultiplexed fastQ files with the barcodes removed for the MinION runs and paired fastQ files for the Illumina iSeq.

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