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Original article

Impact of plant growth regulators spray on fruit quantity and quality of pepper (Capsicum annuum L.) cultivars grown under plastic tunnels



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ABSTRACT

The study aims to investigate the effect of foliar spray with three plant growth regulators (PGRs) p-Chlorophenoxyacetic acid (CPA) at 20 and 40 ppm; Gibberellic acid (GA3) at 20 and 30 ppm, 1-Naphthaleneacetic acid (NAA) at 10 and 20 ppm on the response of fruit set, yield, and fruit quality of some hot pepper cultivars (Chillina, Parbirian, Shampion, and Hyffa) grown in sandy soil under plastic tunnels as compared to the control. Spraying Chillina cultivar GA3 at 30 ppm significantly increased the number of fruits/ plant and fruit set (%), yield/plant, and total yield/fad. In addition, the contents of TSS and Vit C, furthermore, maximum capsaicin content were observed in chili fruits in both seasons. However, the interaction between Chillina cultivar and spraying with GA3 at 20 ppm ranked second in yield and quality. The interaction between Parbirian cultivars and spraying with GA3 at 20 or 30 ppm increased the number of flowers/plants in both seasons. On the other hand, the interaction between Shampion cultivar and spraying with tap water (control) gave the lowest values of the number of flowers/ plants, the number of fruits/ plant and fruit set (%), yield, and its components, and fruit quality in both seasons.

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1. Introduction

Hot pepper (Capsicum annuum L) is one of the most essential medicinal and vegetable crops cultivated in Egypt. the plant has medicinal value. It is eaten as both fresh and dried spices (Bosland and Votava, 2000). Chili Pepper is the third most popular vegetable in the world after potatoes and tomatoes in production quantity. The total cultivated area of chilies and pepper (green) in Egypt in 2020 was 102,141 Feddan (fed.), which produced 790,525 tons with an average of 7.9 tonnes/fed. (Knoema, 2020)

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The active compound in chili pepper is capsaicin which had several activities. The therapeutic applications of capsaicin briefed in enhancing skin blood flow, treating rheumatism, sciatica, and pleurisy. In addition, diabetic headache and cluster headache is also used. It is an excellent source of vitamin B₂, potassium, phosphorus and calcium, besides, low calory. The pepper's high nutritional value results in high year-round demand on the market. Pepper fruits are used as a dried powder in salads, pickles, filling, sauces, and sauce (Mukul et al., 2018; Toyer, 2021).

Chili pepper production is not limited to genetic capacity, but also many environmental factors and cultivation practices. The production of chili decreases due to the decrement in flowers and fruits caused by the physiological and hormonal imbalance in plants as a result of nondurable conditions (Erickson and Markhart, 2001: Vega-Alfaro et al. 2021).

hot pepper cultivars showed significant differences for fruit setting (Chouhan et al 2017; Soreng and Kerketta 2017; Kesumawati et al., 2019). In addition to Productivity and fruit quality (Gungor and Yildirim, 2013, Chowdhury et al. 2015; Ibrahim et al. 2019).

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Plant regulators play a crucial role in various metabolic processes, such as cell division, differentiation and expansion, organogenesis, and germination and are being widely used to increase fruit quality. The quality of green pepper during harvest and post-harvest were maintained by applying gibberellic acid (GA₃) (dos Anjos et al, 2022). Spraying hot pepper with p-Chlorophenoxyacetic acid (CPA), Gibberellic acid (GA3) 1-Naphthaleneacetic acid (NAA) recorded the best results for enhancing fruit setting (Sreenivas et al. 2017; Akhter et al. 2018; Mahindre, et al. 2018). Yield (Patel et al. 2016; Shankhwar et al. 2017; Tapdiya et al. 2018) and fruit quality (Chaudhary et al. 2006; Deshmukh et al. 2010) than unsprayed plants.

The aim of this work to study the effect of GA₃, 4-CPA and NAA growth regulators foliar spray on fruit setting, productivity and fruit quality of some hot pepper cultivars grown under conditions of plastic tunnels.

2. Materials and methods

2.1. Materials

The source of chilli seeds for Chillina and Parbirian cultivars were obtained from Sand Valley Company, Shampion cultivar was from New Star Seed Company and Hyffa cultivar was from Enza zaden Company. The growth regulators (4-CPA, GA_3 and NAA) were purchased from Al Gomhouria Company, Zigzag , Sharkia , Egypt.

2.2. Chemical composition of soil

The soil was chemically analyzed according to Black (1982). The experimental soil was textured sandy, with a method of drip irrigation. The chemical analysis of soil at two seasons was organic matter, 0.07 and 0.09 %; N was 14.62 and 14.98 ppm, available P was 18.0 and 19.46 ppm, available K was 59.6 and 63.1 ppm, available pH 7.90 and 7.88 and E.C. 2.10 and 2.50 m mhos/cm, respectively.

2.3. Experimental design

During the two seasons of 2015/2016 and 2016/2017, the present study was performed at the private farm (The Experimental Farm of Sand Valley Company) at the Ismailia Governorate, Egypt. The response of flowering, yield and fruit quality of the pepper cultivars (Chillina, Parbirian, Shampion and Hyffa) affected by foliar spray of growth regulators treatments (CPA at 20 and 40 ppm, GA₃ at 20 and 30 ppm and NAA at 10 and 20 ppm were studied. Besides, unsprayed treatment (control) of hot chilli grown under tunnel plastic conditions.

This experiment included 28 treatments that were the combination of 4 cultivars and 3 plant growth regulators at two concentrations as compared to control. Such treatments were grouped with three replicates in a split plot in a complete randomized block design. Cultivars were arranged randomly in the main plots and some treatments of the growth regulators (PGRs) randomly distributed in the sub plots. Chili cultivar seeds were sown on 10th Oct. in speed trays under plastic tunnels in the 2015/2016 and 2016/2017 seasons, then were transplanted at 15th Nov. on both row with a width of 1.8 m and a length of 3.6 m with 0.4 m separation between each plant. The plot area was 6.48 m² which occupied by 20 plants. In both seasons, the tunnels were removed on 15th March, when temperatures were suitable for rising chili plants.

The plot was sprayed with 2L of 4-chlorophenoxyacetic acid (4-CPA), gibberellic acid (GA3); and naphthaleneacetic acid (NAA) after 20 days from sowing then weekly sprayed. The PGRs homog-

enized in aqueous solutions using spreading agent (reflecting material). The control plants were treated with tap water mixed with spreading agent. The following data were recorded Flowering characters: At flowering stage, the number of flowers per plant was recorded

2.4. Fruit yield

The cultivars Chillina, Parbirian, Shampion and Hyffa were harvested, respectively after 85, 88, 115 and 100 days from transplanting. Fruits were harvested at intervals of two days, at a length of 12–15 cm and the following data were recorded, Number of fruits/plant, fruit set % was determined by divided number of fruit setting /total flower per plant, fresh weight of fruit/ plant (gm), total yield / fad. was recorded during the harvesting period and relative increases in total yield were calculated (El-Saadony et al, 2021a).

2.5. Fruit quality

The quality parameters; vitamin C (mg /100 g FW) in chili fruits were estimated according to AOAC (2000). TSS (Brix ^o) were mesured by using Hand Refractometer (Saad et al., 2021a). Total capsaicin content in chili fruit (mg /100 g as dry weight) during the second season (2016/2017) only was determined as Popelka et al. (2017).

2.6. Statistical analysis:

The data were statistically analysis using the COSTAT program. The differences between values means were estimated by least significant value (L.S.D.) at 0.05 level of probability according to Snedecor and Cochran (1967).

3. Results

3.1. Number of flowers, number of fruits/ plant and fruit set

3.1.1. Effect of cultivars

There were significant differences among four cultivars respecting number of flowers, number of fruits/ plant and fruit set in both seasons (Tables 1 to 3).

Parbirian cultivar recorded maximum number of flowers/plant, followed by Chillina cultivar, whereas Hyffa cultivar recorded minimum values. On the other hand, Chillina cultivar recorded maximum number of fruits/ plant and fruit set followed by Hyffa cultivar, whereas Champion cultivar recorded minimum number of fruits/ plant and fruit set (%).

3.1.2. Effect of some PGRs

Foliar spray chili plants with 4-CPA, GA_3 and NAA at different concentrations increased number of flowers / plant, number of fruits/ plant and fruit set (%) in both seasons compared to control (spraying with tap water) as shown in Tables 1-3.

Spraying with GA₃ at 30 ppm increased number of flowers / plants, and number of fruits/ plant, followed by spraying with GA₃ at 20 ppm. as for fruit set (%), spraying with 4-CPA at 40 and 20 ppm increased fruit set followed by GA₃ at 30 and 20 ppm in both seasons. Spraying NAA at 10 ppm recorded minimum values of number of flowers, number of fruits/ plant and fruit set (%) in both seasons.

3.1.3. Effect of the interaction

Data in Tables 1-3 indicate that the interaction between Parbirian cultivar and spraying with GA_3 at 30 ppm increased number of flowers / plant in both seasons. However, spraying Chillina cul-

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on Number of flowers /plant of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Chilli cultivars	Plant growth i	regulators types	(ppm)					
	Control	4-CPA		GA ₃		NAA		Mean (C)
	0.0	20	40	20	30	10	20	
	2015/2016 sea	ason						
Chillina	35.33	37.00	48.00	56.33	60.33	42.00	49.00	46.86
Parbirian	36.67	51.33	64.00	72.00	74.33	47.00	50.67	56.57
Shampion	37.67	44.33	51.33	53.67	61.67	41.67	47.00	48.19
Hyffa	33.00	38.33	41.33	41.67	45.67	35.67	41.00	39.52
Mean (PGRs)	35.67	42.75	51.17	55.92	60.50	41.59	46.92	
LSD at 5%	(C) = 0.81		(P) = 1.52			(C × P) = 2.93		
	2016/ 2017 se	ason						
Chillina	41.33	48.33	53.67	65.00	66.67	49.67	54.33	54.14
Parbirian	41.67	51.67	53.67	66.33	69.67	53.00	55.67	55.95
Shampion	41.33	39.67	50.00	49.00	58.67	40.00	42.33	45.86
Hyffa	35.33	37.00	42.67	47.00	51.67	38.33	40.00	41.71
Mean (PGRs)	39.92	44.17	50.00	56.83	61.67	45.25	48.08	
LSD at 5%	(C) = 1.23		(P) = 1.44			(C × P) = 2.92		

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid

Table 2

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on Number of fruits /plant of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Chilli cultivars	Plant growth r	Plant growth regulators types (ppm)									
	Control	4-CPA	GA_3	NAA	Mean (C)						
	0.0	20	40	20	30	10	20				
2015/2016 season											
Chillina	24.00	23.33	25.67	29.67	33.67	23.67	26.33	26.62			
Parbirian	10.67	18.00	23.33	22.00	24.00	15.33	18.00	18.76			
Shampion	8.67	17.00	16.67	18.33	21.67	9.67	16.67	15.53			
Hyffa	11.67	19.00	22.67	23.33	25.33	14.33	18.67	19.29			
Mean (PGRs)	13.75	19.33	22.09	23.33	26.17	15.75	19.92				
LSD at 5%	(C) = 0.37	(P) = 0.83	(C × P) = 1.58								
2016/ 2017 season											
Chillina	21.00	27.00	32.33	35.33	42.67	26.33	31.33	30.86			
Parbirian	11.33	22.00	26.00	28.00	28.33	16.00	20.67	21.76			
Shampion	10.67	16.33	18.00	17.67	19.00	13.33	17.00	16.00			
Hyffa	13.00	19.67	23.67	25.00	26.67	18.33	21.67	21.14			
Mean (PGRs)	14.00	21.25	25.00	26.50	29.17	18.50	22.67				
LSD at 5%	(C) = 0.91		(P) = 1.05			(C × P) = 2.13					

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid

Table 3

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on fruit set (%) of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Chilli cultivars	Plant growth re	egulators types ((ppm)					
	Control	4-CPA		GA ₃	GA ₃		NAA	
	0.0	20	40	20	30	10	20	
	2015/2016 seas	son						
Chillina	67.97	63.07	53.49	52.64	55.86	56.36	53.87	57.61
Parbirian	29.21	36.07	36.50	30.57	32.28	32.72	35.56	33.27
Shampion	23.04	38.50	32.47	34.17	35.13	23.21	35.47	31.71
Hyffa	35.37	49.59	54.89	56.02	55.49	40.16	45.55	48.15
Mean (PGRs)	38.90	46.81	44.34	43.35	44.69	38.11	42.61	
LSD at 5%	(C) = 0.77		(P) = 2.21			(C × P) = 4.16		
	2016/ 2017 sea	ison						
Chillina	50.81	55.84	60.30	54.38	64.34	53.02	57.72	56.63
Parbirian	27.26	42.63	48.44	42.20	40.65	30.19	37.11	38.35
Shampion	25.79	41.27	36.00	36.06	32.38	33.41	40.02	34.99
Hyffa	36.79	53.20	55.48	53.24	51.62	47.82	54.21	50.34
Mean (PGRs)	35.16	48.24	50.06	46.47	47.25	41.11	47.27	
LSD at 5%	(C) = 1.62		(P) = 2.54			(C × P) = 4.92		

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on fruit yield /plant (g) of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Chilli cultivars	Plant growth regulators types (ppm)							
	Control	4-CPA		GA ₃	NAA			Mean (C)
	0.0	20	40	20	30	10	20	
	2015/2016 sea	ison						
Chillina	356.67	420.00	430.00	466.67	505.00	376.67	413.33	424.05
Parbirian	223.33	278.33	371.67	440.00	435.00	240.00	331.67	331.43
Shampion	83.33	106.67	101.67	118.33	140.00	96.67	93.33	105.71
Hyffa	186.67	263.33	316.67	335.00	383.33	221.67	265.00	281.67
Mean (PGRs)	212.50	267.08	305.00	340.00	365.83	233.75	275.83	
LSD at 5%	(C) = 6.64		(P) = 11.45			(C × P) = 22.19		
	2016/ 2017 se	ason						
Chillina	420.00	453.33	520.00	540.00	596.67	440.00	470.00	491.43
Parbirian	233.33	265.00	356.67	450.00	473.33	273.33	360.00	344.52
Shampion	73.33	111.67	96.67	110.00	153.33	88.33	95.00	104.05
Hyffa	195.00	305.00	303.33	336.67	378.33	250.00	281.67	292.86
Mean (PGS)	230.42	283.75	319.17	359.17	400.42	262.92	301.67	
LSD at 5%	(C) = 3.68		(P) = 12.58			(C × P) = 23.57		

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid

tivar with GA₃ at 30 ppm increased number of fruits/ plant in both seasons, followed by the effect of foliar spray of GA₃ (20 ppm) on Chillina. Regarding, fruit set, the interaction between Chillina treated with GA₃ spray (30 ppm) increased fruit set in the 2nd season. Moreover, the Shampion cultivar treated with tap water (control) gave the lowest values of number of flowers/ plant, number of fruits/ plant and fruit set (%) in both seasons.

3.2. Yield / plant and total yield/fad.

3.2.1. Effect of cultivars

There were significant differences among four cultivars in total fruit yield in both seasons (Tables 4-6). Chillina cultivar recorded the highest fruit yield / plant and total fruit yield /fad. followed by Parbirian cultivar, while Shampion cultivar recorded the minimum values in this respect.

The decreases in total fruit yield /fad were about 21.86 and 27.92 % for Parbirian cultivar, 75.08 and 74.38 % for Shampion cultivar and 33.6 and 15.21 % for Hyffa and Chillina cultivar in the 1st and 2nd seasons, respectively.

3.2.2. Effect of the PGRs

Spraying with some PGRs increased yield of fruits/ plant and total fruit yield /fad. compared to control (sprayed with tap water) shown in Tables 4-6. Spraying pepper cultivars with GA₃ at 30 ppm recorded maximum values of yield / plant and total fruit yield/fad. followed by GA₃ (20 ppm), whereas minimum chili yield was recorded with NAA (10 ppm) spraying followed by spraying with 4-CPA at 20 ppm in both seasons.

The increases in total fruit yield /fad. were about 72.18 and 73.79 % for GA₃ at 30 ppm , 60.00 and 58.89 % for GA₃ at 20 ppm and 43.52 and 38.52 % for 4-CPA at 40 ppm over the control (spraying with tap water) in the 1st and 2nd seasons, respectively.

3.2.3. Interaction effect

Data in Tables 4-6 show that, the interaction effect between pepper cultivars and PGRs. Fruit yield/plant and total fruit yield/fad were increased in Chillina cultivar treated with GA₃ (30 ppm) followed by Chillina sprayed with GA₃ (20 ppm) in both seasons, whereas, minimum values were observed in Shampion cultivar treated with tap water (control) in both seasons.

From the obtained results, it could be concluded that Parbirian cultivar recorded maximum number of flowers/ plant, followed by Chillina cultivar. Chillina cultivar recorded maximum number of fruits/ plant and fret set (%), followed by Hyffa cultivar. Chillina cultivar recorded maximum fruit weight / plant and total fruit yield /fad. followed by Parbirian cultivar, Shampion cultivar recorded minimum number of fruits/ pant, fruit set, yield / plant, early and total yield /fad.

Also, spraying with GA₃ at 30 ppm increased number of flowers/ plant, number of fruits/ plant and fret set (%), fruit weight / plant and total fruit yield /fad., followed by spraying with GA₃ at 20 ppm , whereas spraying with NAA at 20 ppm, followed by spraying with 4-CPA at 20 ppm gave the lowest values of these characters.

3.3. Fruit quality

3.3.1. TSS and vitamin C

3.3.1.1. Effect of cultivars. There were a significant differences among four cultivars in fruit quality, i.e., total soluble solids (TSS) and vitamin C (Vit.C) in fruits in both seasons (Tables 7 and 8). Chillina cultivar recorded maximum values of TSS and Vit. C in fruits compared to other cultivars.

3.3.1.2. Effect of the PGRs. Spraying chilli plants with the PGRs increased TSS and Vit C in fruits in both seasons compared to control (sprayed with tap water) as shown in (Tables 7 and 8). Spraying with GA₃ at 30 ppm increased TSS and Vit C in fruits, followed by spraying with GA₃ at 20 ppm in both seasons.

3.3.1.3. Effect of the interaction. Data in Tables 7 and 8 indicate that, the interaction between cultivars and spraying with the PGRs at different concentrations had significant effect on TSS and Vit.C in fruits in both season, where the interaction between Chillina cultivar and spraying with GA₃ at 30 ppm increased TSS and Vit. C in fruits, followed by Chillina cultivar sprayed with GA₃ at 20 ppm and Hyffa cultivar with GA₃ at 30 ppm in both seasons.

3.3.2. *Capsaicin content*

3.3.2.1. *Effect of cultivars*. There were significant differences among four cultivars in Capsaicin content in chilli fruits (Table 9). Chillina cultivar recorded maximum Capsaicin content in fruits compared to other cultivars.

3.3.2.2. Effect of the PGRs. Foliar spray with the PGRs at different concentrations increased capsaicin content in chili fruits compared to control (spraying with tap water) as shown in table (Table 9).

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on total fruit yield /fad. ton) of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Chilli cultivars	Plant growth regulators types (ppm)								
	Control	4-CPA		GA ₃		NAA		Mean (C)	
	0.0	20	40	20	30	10	20		
2015/2016 season									
Chillina	1.997	2.352	2.408	2.613	2.828	2.109	2.315	2.375	
Parbirian	1.521	1.559	2.081	2.464	2.436	1.344	1.857	1.895	
Shampion	0.467	0.597	0.569	0.663	0.784	0.541	0.523	0.592	
Hyffa	1.045	1.475	1.773	1.876	2.147	1.241	1.484	1.577	
Mean (PGRs)	1.258	1.496	1.708	1.904	2.049	1.309	1.545		
LSD at 5%	(C) = 0.037		(P) = 0.064			(C × P) = 0.124			
2016/ 2017 season									
Chillina	2.352	2.539	2.912	3.024	3.341	2.464	2.632	2.752	
Parbirian	1.307	1.484	1.997	2.520	2.651	1.531	2.016	1.929	
Shampion	0.411	0.625	0.541	0.616	0.859	0.495	0.532	0.583	
Hyffa	1.092	1.708	1.699	1.885	2.119	1.400	1.577	1.640	
Mean (PGRs)	1.291	1.589	1.787	2.011	2.243	1.473	1.689		
LSD at 5%	(C) = 0.021		(P) = 0.070			(C × P) = 0.132			

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid

Table 6

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on relative ± (%) in total fruit yield of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Plant growth regulators types (ppm)							
Control	Control 4-CPA		GA ₃		NAA		RYcv
0.0	20	40	20	30	10	20	
2015/2016 se	eason						
100.00	117.78	120.58	130.85	141.61	105.61	115.92	100.0
76.16	78.07	104.21	123.39	121.98	67.30	92.99	78.14
23.39	29.89	28.49	33.20	39.26	27.09	26.19	24.92
52.33	73.86	88.78	93.94	107.51	62.14	74.31	66.40
100.00	125.71	143.52	160.00	172.18	110.00	129.83	
2016/ 2017 s	eason						
100.00	107.95	123.81	128.57	142.05	104.76	111.90	100.0
55.57	63.10	84.91	107.14	112.71	65.09	85.71	72.08
17.47	26.57	23.00	26.19	36.52	21.05	22.62	84.79
46.43	72.62	72.24	80.14	90.09	59.52	67.05	59.59
100.0	123.25	138.52	155.89	173.79	114.10	130.93	
	Plant growth Control 0.0 2015/2016 se 100.00 76.16 23.39 52.33 100.00 2016/ 2017 s 100.00 55.57 17.47 46.43 100.0	Plant growth regulators types (p Control 4-CPA 0.0 20 2015/2016 season 100.00 100.00 117.78 76.16 78.07 23.39 29.89 52.33 73.86 100.00 125.71 2016/2017 season 100.95 55.57 63.10 17.47 26.57 46.43 72.62 100.0 123.25	Plant growth regulators types (ppm) Control 4-CPA 0.0 20 40 2015/2016 season 2015/2016 season 76.16 78.07 104.21 23.39 29.89 28.49 52.33 73.86 88.78 100.00 125.71 143.52 2016/ 2017 season 2016/ 2017 season 17.47 26.57 23.00 46.43 72.62 72.24 100.0 123.25 138.52	Plant growth regulators types (ppm) GA3 Control 4-CPA GA3 0.0 20 40 20 2015/2016 season 100.00 117.78 120.58 130.85 76.16 78.07 104.21 123.39 23.39 29.89 28.49 33.20 52.33 73.86 88.78 93.94 100.00 125.71 143.52 160.00 2016/2017 season 100.00 107.95 123.81 128.57 55.57 63.10 84.91 107.14 17.47 26.57 23.00 26.19 46.43 72.62 72.24 80.14 100.0 123.25 138.52 155.89	Plant growth regulators types (ppm) Control 4-CPA GA ₃ 0.0 20 40 20 30 2015/2016 season 100.00 117.78 120.58 130.85 141.61 76.16 78.07 104.21 123.39 121.98 23.39 29.89 28.49 33.20 39.26 52.33 73.86 88.78 93.94 107.51 100.00 125.71 143.52 160.00 172.18 2016/ 2017 season 7 123.81 128.57 142.05 55.57 63.10 84.91 107.14 112.71 17.47 26.57 23.00 26.19 36.52 46.43 72.62 72.24 80.14 90.09 100.0 123.25 138.52 155.89 173.79	Plant growth regulators types (ppm) GA3 NAA 0.0 20 40 20 30 10 2015/2016 season 20 30 10 20.0 10.0.0 117.78 120.58 130.85 141.61 105.61 76.16 78.07 104.21 123.39 121.98 67.30 23.39 29.89 28.49 33.20 39.26 27.09 52.33 73.86 88.78 93.94 107.51 62.14 100.00 125.71 143.52 160.00 172.18 110.00 2016/2017 season 107.95 123.81 128.57 142.05 104.76 55.57 63.10 84.91 107.14 112.71 65.09 17.47 26.57 23.00 26.19 36.52 21.05 46.43 72.62 72.24 80.14 90.09 59.52 100.0 123.25 138.52 155.89 173.79 114.10	$\begin{tabular}{ c c c c c } \hline Plant growth regulators types (ppm) \\ \hline \hline Control & 4-CPA & GA_3 & NAA \\ \hline \hline 0.0 & 20 & 40 & 20 & 30 & 10 & 20 \\ \hline $2015/2016 season$ & 130.85 & 141.61 & 105.61 & 115.92 \\ \hline 100.00 & 117.78 & 120.58 & 130.85 & 141.61 & 105.61 & 115.92 \\ \hline 76.16 & 78.07 & 104.21 & 123.39 & 121.98 & 67.30 & 92.99 \\ \hline 23.39 & 29.89 & 28.49 & 33.20 & 39.26 & 27.09 & 26.19 \\ \hline 52.33 & 73.86 & 88.78 & 93.94 & 107.51 & 62.14 & 74.31 \\ \hline 100.00 & 125.71 & 143.52 & 160.00 & 172.18 & 110.00 & 129.83 \\ \hline $2016/2017 season$ & $$21.65$ & 24.61 & 107.14 & 112.71 & 65.09 & 85.71 \\ \hline 100.00 & 107.95 & 123.81 & 128.57 & 142.05 & 104.76 & 111.90 \\ \hline 55.57 & 63.10 & 84.91 & 107.14 & 112.71 & 65.09 & 85.71 \\ \hline 17.47 & 26.57 & 23.00 & 26.19 & 36.52 & 21.05 & 22.62 \\ \hline 46.43 & 72.62 & 72.24 & 80.14 & 90.09 & 59.52 & 67.05 \\ \hline 100.0 & 123.25 $ 138.52 & 155.89 & 173.79 & 114.10 & 130.93 \\ \hline \end{tabular}$

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid.

Relative total yield %= Yield of treatment / yield of control x100.

Control of cultivars : Chillina, Control of PGRs = spraying with tap water.

Control of the interaction = Chillina \times spraying with tap water.

RYcv = Relative yield of cultivar.

RY PGRs. = Relative yield of PRGs.

Table 7

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on total soluble solids (TSS) of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Chilli cultivars	Plant growth regulators types (ppm)							
	Control	Control		4-CPA			NAA	Mean (C)
	0.0	20	40	20	30	10	20	
2015/2016 season								
Chillina	6.17	6.43	6.80	7.20	7.57	6.50	6.90	6.80
Parbirian	6.10	6.27	6.67	7.00	7.17	6.37	6.80	6.63
Shampion	5.93	6.03	6.37	6.43	6.87	6.07	6.37	6.30
Hyffa	6.03	6.37	6.67	6.83	7.07	6.30	6.77	6.58
Mean (PGS)	6.06	6.28	6.63	6.87	7.17	6.31	6.71	
LSD at 5%	(C) = 0.06		(P) = 0.08			(C × P) = 0.16		
2016/ 2017 season								
Chillina	6.33	6.27	6.97	7.07	7.67	6.73	6.83	6.84
Parbirian	6.20	6.33	6.83	6.93	7.30	6.37	6.70	6.67
Shampion	6.07	6.23	6.77	6.60	6.97	6.43	6.73	6.54
Hyffa	6.10	6.33	6.73	6.83	7.27	6.43	6.73	6.63
Mean (PGS)	6.18	6.29	6.83	6.86	7.30	6.49	6.75	
LSD at 5%	(C) = 0.07		(P) = 0.08			(C × P) = 0.16		

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid

Effect of different cultivars (C), plant growth regulators (P) types and their interaction (C × P) treatments on Vitamin C content (mg/100 g, f. w.) of chilli plant during 2015/2016 and 2016/2017 seasons under plastic tunnels conditions.

Chilli cultivars	Plant growth regulators types (ppm)							
	Control	4-CPA		GA ₃		NAA		Mean (C)
	0.0	20	40	20	30	10	20	
	2015/2016 se	eason						
Chillina	179.63	195.77	211.10	208.27	224.73	186.93	205.43	201.69
Parbirian	178.27	177.03	193.00	183.67	195.60	177.80	191.93	185.33
Shampion	172.57	172.27	183.03	174.10	179.47	173.53	182.07	176.72
Hyffa	180.17	185.00	198.43	197.90	209.23	185.33	195.10	193.02
Mean (PGRs)	177.66	182.52	196.39	190.99	202.26	180.90	193.63	
LSD at 5%	(C) = 1.75		(P) = 3.25			(C × P) = 6.	27	
	2016/2017 se	eason						
Chillina	182.33	183.27	205.43	219.17	232.00	190.60	199.10	201.70
Parbirian	174.77	177.63	195.43	198.50	208.43	177.57	194.07	189.49
Shampion	169.34	173.20	184.33	177.17	191.83	168.07	190.73	179.24
Hyffa	177.43	184.27	201.73	203.10	220.23	191.87	198.73	196.77
Mean (PGRs)	175.97	179.59	196.73	199.49	213.12	182.03	195.66	
LSD at 5%	(C) = 1.45		(P) = 1.85			(C × P) = 3.	71	

4- CPA = 4-chlorophenoxy acetic acid, GA₃ = gibberllic acid **and** NAA = naphthalene acetic acid.

Spraying with 4-CPA at 40 ppm or with GA₃ at 30 ppm increased capsaicin content in chili fruits.

3.3.2.3. Effect of the interaction. The foliar spray of 4-CPA at 40 or GA_3 at 30 ppm on Chillina cultivar or Champion cultivars recorded maximum Capsaicin content in chili fruits (Table 9).

4. Discussion

Plant growth regulators (PGRs) are organic compounds, which alter plant physiology. They play an important role in increasing plant growth and quality, stem elongation, and flower production, additionally, affect vegetative and fruit production (Ouzounidou et al., 2008; Leclerc et al., 2006). Growth regulators increased the number, size, and weight of sweet pepper fruits (Das et al. 2015). Spraying hot peppers with GA3, 4-CPA, and NAA showed better results for improving fruit set, quantity, and quality (Mahindre et al., 2018; Tapdiya et al., 2018; Deshmukh et al., 2010) compared to non-sprayed plants.

The variability in fruit yield may be due to the effect of temperature of the growing environment, associated features such as canopy diameter, which could restrict the number of branches. Moreover, as a number of major, secondary and tertiary divisions increased, there may be a possibility of growing the number of buds producing fruit that are the locations for fruit production (Delelegn, 2011).

Chili yield variability between different varieties may be due to difference in genetic make-up and prevailing soil environmental condition. Shashidhara (2003) stated that improvements in yield might be due to varieties' adaptability to local climatic and soil conditions. These results are agreeable with those reported by Hasanuzzaman et al. (2007), Dahanayake et al. (2012), Wahb-Allah (2013), Das et al. (2015), Bilal et al. (2019) and Ngullie and Biswas (2019) on chilli. They showed that the differences among cultivars, genotypes and varieties regarding total yield of chilli. In addition, Results are harmony with those obtained with Sarker et al. (2009) showed that spraying BARI Chilli-1 with NNA at 40 ppm significantly increased total yield per hectar than other interaction treatments.

Chowdhury et al. (2015), Chouhan et al. (2017), Soreng and Kerketta (2017) and Kesumawati et al. (2019) reported that number of lowers/ plant, number of fruits/ in chili plant and fruit set percentage significantly affected by different cultivars, genotypes and varieties.

The regulatory impact of exogenous application of PGRs affects early floral initiation, application of auxin at flowering time, and reduced flowers that improve fruit setting and lead to higher fruit setting percentage (Das et al. 2015).

Our Results are harmony with Chaudhary, et al. (2006), Sreenivas et al.(2017), Akhter et al. (2018), Arivazhagan et al. (2018) on Brinjal cv. Annamalai, Mahindre, et al. (2018) on chilli. They showed that spraying plants PGRs recorded the highest values of number of flowers/ plant, number of fruits/ plant and fruit set percentage, especially when sprayed with n GA₃ as compared to other PGRs or unsprayed plant.

Chaudhary, et al. (2006) showed that the interaction between Suryamukhi chilli cultivar and sprayed with NAA at 40 ppm recorded the highest values of number of fruits/ plant than other interaction treatments. In addition, Sarker et al. (2009) showed that spraying BARI Chilli-1 with NNA at 40 ppm significantly increased fruit set (%), number of fruits / plant, than other interaction treatments. In addition, Das et al. (2015) showed that the highest number of flowers and fruits /plant of chilli were recorded with spraying Lamuyo variety with 4-CPA than spraying BARI Misti morich-1 with tap water.

An improvement in fruit yield and its component characteristics due to the application of 4-CPA and GA₃ could be attributed to a more effective use of food for reproductive growth (flowering and fruit set), increased photosynthetic output and increased source for plant sinking, increased nutrient and water intake, reduced transpiration and breathing, increased translocation and accumulation of sugar and other metabolites (Chaudhary, et al. 2006).

Balraj et al. (2002), Natesh et al. (2005), Vandana and Verma, 2014, Sanjay, and Singh (2019) found that the highest early and total yield of chilli were obtained with the plants, which sprayed with PGRs, especially GA₃. On the other hand, the fruit quality affected with PGRs spraying where the different cultivars, genotypes and varieties showed significant differences concerning TSS and Vit. C in fruits according to Chaudhary, et al. (2006) Gungor and Yildirim, (2013) Chowdhury et al. (2015) and Ibrahim et al. (2019). The increase of ascorbic acid with GA₃ treatment may be due either to the promotion of ascorbic acid biosynthesis or to the defense of synthesized ascorbic acid from oxidation by the ascorbic acid oxidase enzyme.

Effect of different cultivars (C), plant growth regulators (P) types and their interaction ($C \times P$) treatments on total capsaicin content (mg/100 g as dry weight) in fruits of chilli plant during 2016/2017 season under plastic tunnels conditions.

Chilli cultivars	Plant growth regulators types (ppm)								
	Control		4-CPA	4-CPA			NAA	Mean (C)	
	0.0	20	40	20	30	10	20		
Chillina	129.24	137.44	140.97	137.59	140.03	130.51	134.91	135.81	
Parbirian	124.67	134.58	136.65	134.83	138.60	128.52	134.01	133.12	
Shampion	127.92	134.46	141.86	138.48	138.99	129.92	133.14	134.97	
Hyffa	126.84	132.45	137.43	133.96	137.50	131.51	135.60	133.61	
Mean (PGRs)	127.16	134.73	139.23	136.22	138.78	130.12	134.41		
LSD at 5%	(C) = 0.66			(P) = 1.09			(C × P) = 2.12		

4- CPA = 4-chlorophenoxy acetic acid, GA_3 = gibberllic acid **and** NAA = naphthalene acetic acid

Chaudhary et al. (2006) and Deshmukh et al. (2010) showed that fruit quality such as TSS and Vit. C in fruits were the highest with the plants, which sprayed with different PGRs as compared the other treatments. In this regard, Sarker et al. (2009) showed that spraying BARI Chilli-1 with NNA at 40 ppm significantly increased ascorbic acid content in fruits, while the interaction between spraying the same cultivar with tap water gave the highest values of TSS than other interaction treatments. capsaicin content (%) was varied based on cultivars, genotypes and varieties (Phimchan and Techawongstien, 2012).

Growth hormones, whether auxins or cytokinins improve plant growth, productivity, and quality, some natural substances acting the same role, i.e., herbal extracts (Saad et al, 2020a; El-Tarabily et al, 2021; Abdel-Moneim et al, 2022; El-Saadony et al, 2021b), peptides (Saad et al, 2020b; Saad et al, 2021b; El-Saadony et al, 2022; El-Saadony et al, 2021d), microorganisms (Desoky et al, 2020), and phenolic compounds extracted from agricultural wastes (Saad et al, 2021c; Saad et al, 2021d). Conventional growth regulators can also be mixed with natural materials to maximize yield. The mechanism of growth promoters is briefed in production of phenolics in chili peppers, i.e., ABA can act as a biochemical signaler and trigger the expression of genes that encode proteins involved in the biosynthesis and metabolism of phenylpropanoids, and the phenylpropanoid pathway is important in the secondary metabolism of vegetables, the main products produced include phytoalexins, phenolic acids, and important precursors, such as chalcones (Tan et al., 2016; Moreira et al., 2020), which are fundamental in the synthesis of flavonoids.

5. Conclusion

The foliar spray of PGRs on chili pepper significantly affected fruit quantity and quality where Plant growth regulators (PGRs) play a vital role in increasing plant growth, quality, stem elongation, and flower production, additionally, affect vegetative, and fruit production increased the number, size, and weight of sweet pepper fruits. Spraying hot peppers with GA3, 4-CPA, and NAA showed better results for improving fruit set, quantity, and quality. We recommended using GA₃ as PGRs in improving chili yield quality.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

- Abdel-Moneim, A.M.E., El-Saadony, M.T., Shehata, A.M., Saad, A.M., Aldhumri, S.A., Ouda, S.M., Mesalam, N.M., 2022. Antioxidant and antimicrobial activities of *Spirulina platensis* extracts and biogenic selenium nanoparticles against selected pathogenic bacteria and fungi. Saudi J. Biol. Sci. 29 (2), 1197–1209. https://doi. org/10.1016/j.sjbs.2021.09.046, accessed date: 17 Sep. 2021.
- Akhter, S., Mostarin, T., Khatun, K., Akhter, F., Parvin, A., 2018. Effects of plant growth regulator on yield and economic Benefit of sweet pepper (*Capsicum* annum L.). Agric. 16 (2), 58–64.
- AOAC, 2000. Association of Official Analytical Chemists.17th ED. Of AOAC International Published by AOAC International Maryland, USA, 1250pp.
- Arivazhagan, E., Kavitha, A., Kandasamy, R., 2018. Influence of plant growth regulators on yield and quality characters of brinjal (*Solanum melongena* L.) Cv. Annamalai. The Asian J. Hort. 13 (2), 45–49.
- Balraj, R., Kurdikeri, M., Revanappa, B., 2002. Effect of growth regulators on growth and yield of chilli (*Capsicum annuum*) at different pickings. Indian J. Hort. 59 (1), 84–88.
- Bilal, H., Aman, F., Ullah, I., Awan, A.A., Ullah, S., Khan, S., 2019. Response of chilli varieties to various sources of organic fertilizers. ARPN J. Agric. Biol. Sci. 13 (12), 15–24.
- Black, C.A., 1982. Methods of Soil Analysis. Part2. American Society of Agronomy, INC, Pub., Madison, Wisconsin, USA.
- Bosland, P.W., Votava, E.J., 2000. Peppers, vegetables and spices Capsicum. CABI Publishing, New York, p. 198p.
- Chaudhary, B.R., Sharma, M.D., Shakya, S.M., Gautam, D.M., 2006. Effect of plant growth regulators on growth, yield and quality of chilli (*Capsicum annuum* L.) at Rampur. Chitwan. J. Inst. Agric. Anim. Sci. 27, 65–68.
- Chouhan, K.S., Baghel, S.S., Mishra, K., Singh, A.K., Singh, V., 2017. Effect of varieties and integrated nutrient management on growth and yield of chilli (*Capsicum annuum* L.). Int. J. Pure App. Biosci. 5 (4), 2114–2120.
- Chowdhury, M.S.N., Hoque, F., Mehraj, H., Jamal Uddin, A.F.M., 2015. Vegetative growth and yield performance of four chilli (*Capsicum frutescens*) cultivars. American-Eurasian J. Agric. Environ. Sci. 15 (4), 514–517.
- Dahanayake, N., Madurangi, S.A.P., Ranawake, A.L., 2012. Effect of potting mixture on growth and yield of chilli varieties (*Capsicum* Spp) and microbial activity. Tropical Agric. Res. Extension 15 (3), 1–3.
- Das, S.K., Sarkar, M.D., Alam, M.J., Robbani, M.G., Kabir, M.H., 2015. Influence of plant growth regulators on yield contributing characters and yield of bell pepper (*Capsicum annum*) varieties. J. Plant Sci. 10 (2), 63–69.
- Delelegn, S., 2011. Evaluation of elite hot pepper varieties (*Capsicum species*) for growth, dry pod yield and quality under Jimma condition, south west Ethiopia. Jimma Univ, Ethiopia. M.Sc Thesis.
- Deshmukh, D.A., Telang, S.M., Patil, S.S., 2010. Influence of foliar application of growth hormones and fertilizers on the field and ascorbic content in chilli cv Parbhani Tajas. Asian J. Soil Sci. 5 (1), 114–115.
- Desoky, E.-S., Saad, A.M., El-Saadony, M.T., Merwad, A.-R., Rady, M.M., 2020. Plant growth-promoting rhizobacteria: Potential improvement in antioxidant defense system and suppression of oxidative stress for alleviating salinity stress in *Triticum aestivum* (L.) plants. Biocatal. Agric. Biotechnol. 30, 101878. https://doi.org/10.1016/j.bcab.2020.101878.
- dos Anjos, G.L., Moreira, G.C., Carneiro, C.N., Dias, F.D.S., 2022. Effect of phytoregulators on the composition of phenolic compounds in chili peppers (Capsicum frutescens) and exploratory analysis. Sci. Hortic. 292, 110660. https://doi.org/10.1016/j.scienta.2021.110660.

- El-Saadony, M.T., Abd El-Hack, M.E., Swelum, A.A., Al-Sultan, S.I., El-Ghareeb, W.R., Hussein, E.O., Nader, M.M., 2021a. Enhancing quality and safety of raw buffalo meat using the bioactive peptides of pea and red kidney bean under refrigeration conditions. Ital. J. Anim. Sci. 20 (1), 762–776.
- El-Saadony, M.T., S. F. Khalil, O., Osman, A., Alshilawi, M.S., Taha, A.E., Aboelenin, S. M., Shukry, M., Saad, A.M., 2021b. Bioactive peptides supplemented raw buffalo milk: Biological activity, shelf life and quality properties during cold preservation. Saudi J. Biol. Sci. 28 (8), 4581–4591.
- El-Saadony, M.T., Saad, A.M., Elakkad, H.A., El-Tahan, A.M., Alshahrani, O.A., Alshilawi, M.S., Ahmed, A.I., 2022. Flavoring and extending the shelf life of cucumber juice with aroma compounds-rich herbal extracts at 4° C through controlling chemical and microbial fluctuations. Saudi J. Biol. Sci. 29 (1), 346– 354.
- El-Saadony, M.T., Saad, A.M., Najjar, A.A., Alzahrani, S.O., Alkhatib, F.M., Shafi, M.E., Selem, E., Desoky, E.-S., Fouda, S.E.E., El-Tahan, A.M., Hassan, M.A.A., 2021d. The use of biological selenium nanoparticles to suppress *Triticum aestivum* L. crown and root rot diseases induced by Fusarium species and improve yield under drought and heat stress. Saudi. J. Biol. Sci. 28 (8), 4461–4471.
- El-Tarabily, K.A., El-Saadony, M.T., Alagawany, M., Arif, M., Batiha, G.E., Khafaga, A.F., Elwan, H.A.M., Elnesr, S.S., E. Abd El-Hack, M., 2021. Using essential oils to overcome bacterial biofilm formation and their antimicrobial resistance. Saudi Journal of Biological Sciences. 28 (9), 5145–5156.
- Erickson, A.N., Markhart, A.H., 2001. Flower production, fruit set and physiology of bell pepper during elevated temperature and vapor pressure deficit. J. American Soc. Hort. Sci. 126 (6), 697–702.
- Gungor, F., Yildirim, E., 2013. Effect of different growing media on quality, growth and yield of pepper (*Capsicum annuum* L.) under greenhouse conditions. Pak. J. Bot. 45 (5), 1605–1608.
- Hasanuzzaman, S.M., Hossain, S.M.M., Ali, M.O., Hossain, M.A., Hannan, A., 2007. Performance of different bell pepper (*Capsicum annuum* L.) genotypes in response to sythetic hormones. Int. J. Sustain. Crop Prod. 2, 78–84.
- Ibrahim, A., Abdel-Razzak, H., Wahb-Allah, M., Alenazi, M., Alsadon, A., Dewir, Y.H., 2019. Improvement in growth, yield, and fruit quality of three red sweet pepper cultivars by foliar application of humic and salicylic acids. Hortechnol. 29 (2), 170–178.
- Kesumawati, E., Apriyatna, D., Rahmawati, M., 2019. The effect of shading levels and varieties on the growth and yield of chili plants (*Capsicum annuum* L). IOP Conf. Ser. Earth Environ. Sci. 425 (1), 012080. https://doi.org/10.1088/1755-1315/ 425/1/012080.
- Knoema, 2020. <u>https://knoema.com/data/egypt+agriculture-indicators-production</u> <u>+chillies-and-peppers-green</u>, 2020 Leclerc, Mélanie, Caldwell, C.D., Lada, R.R., Norrie, J., 2006. Effect of plant growth
- Leclerc, Mélanie, Caldwell, C.D., Lada, R.R., Norrie, J., 2006. Effect of plant growth regulators on propagule formation in *Hemerocallis* spp. and Hosta spp. Hort Sci. 41 (3), 651–653.
- Mahindre, P.B., Jawarkar, A.K., Ghawade, S.M., Tayade, V.D., 2018. Effect of different concentration of plant growth regulators on growth and quality of green chilli, JPP.; SP1:3040-3042.
- Moreira, G.C., dos Anjos, G.L., Carneiro, C.N., Ribas, R.F., Dias, F.d.S., 2020. Phenolic compounds and photosynthetic activity in *Physalis angulata L.* (Solanaceae) in response to application of abscisic acid exogenous. Phytochem. Lett. 40, 96–100.
- Mukul, C., Kant, C., Negi, C., Chauhan, P., 2018. Blood circulation stimulation properties of cayenne pepper: a review. J. Appl. Chem. 11 (5), 78–83. Natesh, N., Vyakaranhal, B.S., Gouda, M.S., Deshpande, V.K., 2005. Influence of
- Natesh, N., Vyakaranhal, B.S., Gouda, M.S., Deshpande, V.K., 2005. Influence of growth regulators on growth, seed yield and quality of chilli cv ByadgiKaddi. Karnataka J. Agric. Sci. 18 (1), 36–38.
- Ngullie, R., Biswas, P.K., 2019. Vegetative growth and yield performance of four chilli (*Capsicum annuum* L) cultivars under mokokchung district of Nagaland. Inter. J. Agric. Sci. 11 (3), 7833–7835.
- Ouzounidou, G., Papadopoulou, P., Giannakoula, A., Ilias, I., 2008. Plant growth regulators treatments modulate growth, physiology and quality characteristics of *Cucumis melo* L. plants. Pak. J. Bot. 40, 1185–1193.
- Patel, V.P., Plal, E., John, S., 2016. Comparative study of the effect of plant growth regulators on growth, yield and physiological attributes of chilli (*Capsicum annum* L.) cv. Kashi Anmol. Int. J. Farm Sci. 6 (1), 199–204.
 Phimchan, P., Techawongstien, S., Chanthai, S., Bosland, P.W., 2012. Impact of
- Phimchan, P., Techawongstien, S., Chanthai, S., Bosland, P.W., 2012. Impact of drought stress on the accumulation of capsaicinoids in capsicum cultivars with different initial capsaicinoid levels. Hortscience 47 (9), 1204–1209.

- Popelka, P., Jevinov, P., Smejkal, K., Roba, P., 2017. Determination of capsaicin content and pungency level of different fresh and dried chilli peppers. Folia Veterinaria 61 (2), 11–16.
- Saad, A.M., Mohamed, A.S., El-Saadony, M.T., Sitohy, M.Z., 2021a. Palatable functional cucumber juices supplemented with polyphenols-rich herbal extracts. LWT - Food Sci. Technol. 148, 111668. https://doi.org/10.1016/j. lwt.2021.111668.
- Saad, A.M., Mohamed, A.S., Ramadan, M.F., 2020a. Storage and heat processing affect flavors of cucumber juice enriched with plant extracts. Int. J. Veg. Sci., 1–11
- Saad, A.M., Osman, A.O.M., Mohamed, A.S., Ramadan, M.F., 2020b. Enzymatic hydrolysis of *Phaseolus vulgaris* protein isolate: Characterization of hydrolysates and effect on the quality of minced beef during cold storage. Int. J. Pept. Res. Ther. 26 (1), 567–577.
- Saad, A.M., Sitohy, M.Z., Ahmed, A.I., Rabie, N.A., Amin, S.A., Aboelenin, S.M., Soliman, M.M., El-Saadony, M.T., 2021b. Biochemical and functional characterization of kidney bean protein alcalase-hydrolysates and their preservative action on stored chicken meat. Molecules 26 (15), 4690. https:// doi.org/10.3390/molecules26154690.
- Saad, A.M., El-Saadony, M.T., El-Tahan, A.M., Sayed, S., Moustafa, M.A.M., Taha, A.E., Taha, T.F., Ramadan, M.M., 2021c. Polyphenolic extracts from pomegranate and watermelon wastes as substrate to fabricate sustainable silver nanoparticles with larvicidal effect against Spodoptera littoralis. Saudi J. Biolog. Sci. 28 (10), 5674–5683.
- Saad, A.M., El-Saadony, M.T., Mohamed, A.S., Ahmed, A.I., Sitohy, M.Z., 2021d. Impact of cucumber pomace fortification on the nutritional, sensorial and technological quality of soft wheat flour-based noodles. Int. J. Food Sci. 56 (7), 3255–3268.
- Sanjay, S., Singh, T., 2019. Effect of gibberellic acid on growth, yield and quality parameters of chilli (*Capsicum annuum* L.). J. pharmacogn. phytochem. 8 (2), 2021–2023.
- Sarker, P., Hossain, T., Mia, M.A., Islam, R., Miah, M.N.A., 2009. Effect of NAA on growth, yield and quality of chilli (*Capsicum frutescence*). Bangladesh Res. Pub. J. 2 (3), 612–617.
- Shankhwar, B., Nigam, A.K., Vasure, N., Vishvakarma, D., 2017. Effect of different plant growth regulators on growth of chilli (*Capsicum annuum* L.) cv. PUSA JWALA. Agric. Update 12 (5), 1187–1189.
- Shashidhara G. B., 2003. Effect of dates of planting on yield of chili cultivars under rainfed conditions in Alfisols of Northern Transition zone of Karanataka. Kanrantaka. J. Agri. Sci. 16 (4).585-587.
- Snedecor, G.W., Cochran, W.G., 1967. Statistical Methods. Press, Amer., Iowa, USA, The Iowa State Univ.
- Soreng, M.K., Kerketta, N.S., 2017. Effect of organic manures on different plant varieties of chilli (*Capsicum annuum* L.) under subabul (*Leucaena leucocephala*) based Horti silviculture system. J. Med. Plants Stud. 5 (5), 273–276.
- Sreenivas, M., Sharangi, A.B., Raj, A.C., 2017. Evaluation of bio-efficacy and phytotoxicity of gibberellic acid on chilli. J. Crop and Weed 13 (3), 174–177.
- Tan, B.A., Daim, L.D.J., Ithnin, N., Ooi, T.E.K., Md-Noh, N., Mohamed, M., Mohd-Yusof, H., Appleton, D.R., Kulaveerasingam, H., 2016. Expression of phenylpropanoid and flavonoid pathway genes in oil palm roots during infection by Ganoderma boninense. Plant Gene 7, 11–20.
- Tapdiya, G.H., Gawande, P.P., Ulemale, P.H., Patil, R.K., Naware, M.S., 2018. Effect of growth regulators on quantitative characters of chilli (*Capsicum annuum* L.). Int. J. Curr. Microbiol. App. Sci. 6, 2151–2157.
- Toyer, C., 2021. Evaluation of chili pepper (*Capsicum Annuum*), Tiger nuts (Cyperus Esculentus), and Turmeric (Curcuma Longa) as Sources of Antioxidant Compounds for the Potential of AntiAging-like Activity. thesis.
- Vandana, P., Verma, L.R., 2014. Effect of spray treatment of growth substances at different stages on growth and yield of sweet pepper (*Capsicum annum* L) cv. Indra under green house. Int. J. Life Sci. Res. 2 (4), 235–240.
- Vega-Alfaro, A., Ramírez-Vargas, C., Chávez, G., Lacayo, F., Bethke, P.C., Nienhuis, J., 2021. Flowering Time and Productivity of Interspecific Grafts Between Pepper Species in Contrasting High Tunnel-sheltered and Open-field Production Environments in Costa Rica. HortTechnology 1 (aop), 1–10.
- Wahb-Allah, M.A., 2013. Responses of some bell-pepper (*Capsicum Annuum* L.) cultivars to salt stress under greenhouse conditions. J. Agric. Env. Sci. Dam. Univ. 12 (1):1–20.