



Cow's Milk Processing—Friend or Foe in Food Allergy?

Sabine Geiselhart⁺, Aleksandra Podzhilkova⁺ and Karin Hoffmann-Sommergruber^{*}

Department of Pathophysiology and Allergy Research, Medical University of Vienna, 1090 Vienna, Austria; sabine.geiselhart@muv.ac.at (S.G.); aleksandra.podzhilkova@muv.ac.at (A.P.)

* Correspondence: karin.hoffmann@muv.ac.at

+ These authors contributed equally to this work.

Abstract: Cow's milk (CM) is an integral part of our daily diet starting in infancy and continuing throughout our lifetime. Its composition is rich in proteins with a high nutritional value, bioactive components, milk minerals including calcium, and a range of immunoactive substances. However, cow's milk can also induce a range of immune-mediated diseases including non-IgE-mediated food allergies and IgE-mediated food allergies. Cow's milk allergens have been identified and characterized and the most relevant ones can be assigned to both, the whey and casein fraction. For preservation a range of processing methods are applied to make cow's milk and dairy products safe for consumers. However, these methods affect milk components and thus alter the overall immunogenic activity of cow's milk. This review summarizes the current knowledge on cow's milk allergens and immunoactive substances and the impact of the different processes up- or downregulating the immunogenicity of the respective proteins. It highlights the gaps of knowledge of the related disease mechanisms and the still unidentified beneficial immunomodulating compounds of cow's milk.

check for updates

Citation: Geiselhart, S.; Podzhilkova, A.; Hoffmann-Sommergruber, K. Cow's Milk Processing—Friend or Foe in Food Allergy?. *Foods* **2021**, *10*, 572. https://doi.org/10.3390/ foods10030572

Academic Editor: Michelle Colgrave

Received: 7 February 2021 Accepted: 2 March 2021 Published: 9 March 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Keywords: cow's milk allergens; whey; casein; food processing; IgE-mediated food allergy

1. Introduction

Milk consumption by humans has been tightly connected with settledness and agriculture from the very beginning. Recent findings provide evidence that humans were already drinking milk at least 6000 years ago [1]. Collecting, processing, and consuming milk from animals enabled one of the most profound revolutions in human diet and for centuries, cow's milk has been an integral part of our diet. Animal milk is a renewable food source rich in proteins, fat, and micronutrients. For centuries, it has been used as alternative for breast milk and enabled early weaning with significant demographic implications. Since then strategies for preserving milk have been developed and continuously improved—for example, the generation of yogurt, butter, and cheese are old traditional methods. Nowadays, highly refined technologies are in place to offer safe and convenient consumption. However, these methods may also have a negative effect on the micronutrients which are present in natural milk products. Furthermore, the food and pharmaceuticals industries use individual milk components in many different applications.

It is also known that milk can induce immune-mediated diseases. Although milk is regarded as a healthy food with high nutritional value it can be harmful to some individuals with a predisposition to develop immune-mediated adverse reactions to foods. Food allergy prevalence rates have increased, including cow's milk allergy. Changes in lifestyle and dietary habits may account for this, since milk consumption is no longer only regarded as part of a healthy diet for infants and children—nowadays it is consumed throughout one's lifetime. Moreover, exposure to milk-derived components (milk protein, sugars, lipids) has increased since they are present in a range of highly processed convenience food products, cosmetics, and pharmaceutical drugs.

This review is dedicated to summarizing the current knowledge of milk allergens and the food processing techniques which may modulate their immunogenicity.

2. Brief Overview on Immune-Mediated Diseases Caused by Milk Proteins

2.1. IgE- and Cell-Mediated Allergies

Adverse reactions to foods can induce a variety of immune-mediated reactions, ranging from mild to severe symptoms, some of which persist throughout one's lifetime while others resolve. For a restricted number of food-related diseases the causative foods are known, including milk and dairy products. After birth newborns are usually breastfed and in due course exposed to cow's milk and/or related milk sources, representing one of the earliest encounters with foreign (nonself) dietary antigens. Therefore, most immune-mediated diseases caused by milk intake start early in life and resolve within early childhood, while in adolescence and adulthood only a comparatively reduced number of diseases persist or develop.

Adverse immune reactions to foods can be assigned to IgE-mediated, mixed IgEand cell-mediated, and cell-mediated groups based on the underlying mechanisms [2]. Currently, food protein-induced enterocolitis syndrome (FPIES), food protein-induced enteropathy (FPE), and food protein-induced allergic proctocolitis (FPIAP) are listed under "non-IgE-mediated gastrointestinal food allergic disorders (non IgE-GI-FAs)" [3]. These diseases start early in infancy, are relatively rare and with a good prognosis resolving after 1–3 years. The majority of patients experience symptoms until the age of 5 years [3]. For these diseases cow's milk proteins seem to be a relevant trigger, although other food sources such as soy, wheat, and egg have been reported. For all these foods no specific immunogenic proteins have been identified so far; it is generally accepted that the cow's milk protein fraction per se is causative.

Food protein-induced allergic proctocolitis (FPIAP) is a cell-mediated immune disease caused by cow's milk, soy, egg, and wheat in the maternal diet when breastfed, and milk and soy formulas [3]. Usually, the symptoms are bloody intermittent stools in otherwise healthy and thriving babies [3]. Although the underlying pathomechanism is not well understood, increased eosinophils have been identified in intestinal endoscopies. Diagnosis is based on characteristic symptoms and elimination diet for diagnostic purposes can be performed. After 4–8 weeks, reintroduction of the causative foods is possible based on symptom amelioration. Usually FPIAP resolves after some months.

Food protein-induced enterocolitis syndrome (FPIES) starts within the first year of life. The clinical picture includes malabsorption, anemia, diarrhea, and vomiting and can lead to failure to thrive. Similarly to FPIAP the pathogenesis of FPIES is not well understood but studies provided evidence of T cells secreting inflammatory cytokines and the role of neuroendocrine pathways in this disease was discussed. Moreover, neutrophilia and thrombocytosis are frequently identified in patients [4]. Milk was in most cases the causative food and children resolving the disease within 5 years had no detectable milk specific IgE antibodies, whereas patients with milk positive IgE had consistent milk adverse reactions [5].

Food protein enteropathy (FPE) usually presents with chronic diarrhea which does not result in severe dehydration. Cow's milk protein is also the most important food trigger for this disease. The histology of FPE patients provides damage to the villi, and the presence of eosinophils and cow's milk-specific Th2 cells and sometimes local IgE production.

In summary, non-IgE-GI-FAs share the same food(s) that trigger/induce symptoms, their individual pathomechanisms are not well understood and a potential risk of underdiagnosed cases is possible. However, the majority of cases experience tolerance induction within the first year [6]. The question remains whether cow's milk proteins are the causative trigger of the diseases mentioned above or whether it is the first encounter between dietary proteins and an as yet immature digestive tract which causes the symptoms.

Another immune-mediated disease that is triggered by certain foods including milk is eosinophilic esophagitis (EoE). This disease is diagnosed by a high number of eosinophilic infiltrates and dysfunction of the esophagus resulting in difficulties swallowing food and food impaction. This fairly recently identified disease has been diagnosed in both children and adults. Although food-specific IgE antibodies have been determined in a subgroup of patients, it remains to be established if the presence of IgE antibodies is an epiphenomenon or linked with the pathomechanism which is still poorly understood for EoE [7].

2.2. IgE-Mediated Milk Allergy

2.2.1. Prevalence

Cow's milk is one of the first nutritional sources for infants and in atopic individuals IgE-mediated allergic symptoms can be diagnosed soon after first exposure. Symptoms range from acute cutaneous reactions e.g., urticaria, atopic dermatitis, immediate breathing problems, gastrointestinal problems, and asthma attacks up to anaphylactic reactions. In a systematic review prevalence rates of 6.0% for self-reported cow's milk allergy were published, compared to 0.6% for an objectively verified cow's milk allergy [8]. Higher prevalence data are known for infants and children as compared to adolescents and adults, which is due to tolerance induction in the majority of children when reaching school age.

The underlying pathomechanism of an IgE-mediated allergy is based on the production of allergen-specific IgE antibodies from B cells together with the favoring cytokine milieu provided by T helper cells type 2 in genetically predisposed individuals. When reexposure to the same allergen source occurs the allergen is recognized by IgE antibodies that are bound to receptors on mast cells and basophils and upon cross-linking an immediate reaction is triggered by the release of active substances and cytokines. So far, a number of cow's milk allergens have been identified and their immunogenic activity can be tested in cellular tests and in vitro diagnostic tests. A detailed overview of the currently known cow's milk allergens is presented below.

Diagnosis is based on taking patient history including reported symptoms, a skin prick test and an in vitro test for allergen-specific IgE antibodies. In addition, an oral food challenge either in an open or blinded setting can be performed. For in vitro tests the detection of serum-derived milk allergen-specific IgE is identified in assays that use either total extracts or panels of individual allergens. In the later assay the specific immune response to single allergens provides a detailed analysis of the allergens causing the immune response in each patient.

In cellular tests basophils are used and incubated with the patient's serum-derived IgE antibodies. Upon the addition of milk allergens and/or milk protein extract cross-linking takes place, providing a positive test result.

For food challenges different protocols have been developed including the approach using "baked milk". This approach is applied to investigate whether there is an increased risk of persisting milk allergy compared to tolerance induction to milk proteins during childhood. The underlying rationale is that heat treatment of milk proteins can affect their structure and thus their presentation of IgE epitopes. If patients tolerate baked milk whereas they react to untreated milk extract, the probability of developing a tolerance to milk products is very likely (see also Section 5.1).

2.2.2. Treatmen

The avoidance of milk and dairy products is the method of choice for treatment. For infants peptide based milk formulas can be offered. These peptides or amino acid formulas are unable to trigger the cross-linking of IgE and are thus safe for consumption by those with milk allergies. For a subgroup of people with milk allergies, heat-treated milk can also be tolerated (see also heat processing and effect on allergen structures). The use of goat's milk or sheep's milk as an alternative nourishment to breastfeeding and cow's milk is not recommended. Although in some cases amelioration of symptoms is observed, the effect does not last and the allergic symptoms reoccur. This is due to the presence of highly homologous allergens in the milk products of these closely related species.

As mentioned above, the majority of milk allergic children gain tolerance and upon a confirmative negative food challenge milk can then be reintroduced to the daily diet. Currently, no approved immunotherapy for milk allergy is available; however, in a number of specialized centers, a rush immunotherapy can be offered to highly sensitive patients so that they reach tolerance to minute amounts of milk proteins.

2.2.3. Prevention

Regarding the prevention of IgE-mediated cow's milk allergy, the introduction of cow's milk into the diet between months 4–6 is recommended, followed by regular cow's milk ingestion, based on recent studies [9].

3. Milk and Its Components

The milk of herbivorous species (cows, sheep, goats, etc.) comprises homologous proteins with similar structural, functional, and biological properties. Along with proteins, milk contains nitrogenous compounds of a nonprotein nature: free amino acids, peptides, urea, ammonia and uric acid, etc. Furthermore, a huge amount of bioactive components such as casein hydrolysates, lactoferrin, lactoperoxidase, lysozyme, glycomacropeptide or caseinomacropeptide, whey protein hydrolysate, milk minerals including calcium and magnesium [10], α -lactalbumin, galacto-oligosaccharide (GOS), conjugated linoleic acid (CLA), are present in milk [11]. All these constituents have a particular biological impact on human health.

Milk includes more than 40 proteins (30–35 g per liter), comprising 80% casein and 20% milk serum (whey) [12]. Cow's milk contains allergens and homologues of those are also present in all ruminant species (Table 1). They share structural, functional and biological properties [13,14]. In addition, milk contains a range of immunoactive substances such as osteopontin, cytokines (e.g., TGF-beta, IL-10), alkaline-phosphatase and vitamin D. According to the authors, leucine is the major amino acid in casein and in whey, as shown in Figure 1 [14] and Figure 2 [15,16]. In Figure 2, caseins and whey proteins from different species are presented. As we can see, in human milk β -lactoglobulin and the caseins as1 and as2 are absent.

Milk Proteins		Conc. (g/L)	Molecular Mass (kDa)	Biological Function	Amino Acid No.	Allergenic Activity *
Whey proteins $(20\% \approx 7 \text{ g/L})$	α-lactalbumin Bos d 4	1.2–1.5	14.2	Contributes to lactose synthesis	123	Major
	β-lactoglobulin Bos d 5	3–4	18.3	Binds to numerous hydrophobic and amphiphilic ligands (defined biological function still unclear)	162	Major
	BSA Bos d 6	0.4	67	Binds to fatty acids, flavors, metal ions	583	Minor
	Immunoglobulins Bos d 7	0.47	76.2	Antibacterial and antiviral activities		Minor
Caseins (80% ≈ 29 g/L) Bos d 8	αs1-casein Bos d 9	12–14	23.6	Calcium binding	199	Major
	αs2-casein Bos d 10	3.75–4	25.2	Calcium binding	207	Minor
	β-casein Bos d 11	10.5–12	24.0	Calcium binding	209	Major
	к-casein Bos d 12	3.75–4	19.0	Stabilization and coagulation of milk	169	Minor

Table 1. Characteristics of main cow's milk allergens [13,17] and IUIS Allergen Nomenclature.

* Allergenic activity is presented as major versus minor allergen.



Adapted from Rafiq et al., 2015

Figure 1. (a) Essential amino acids in caseins from different species. (b) Essential amino acids in whey from different milk species. (adapted from [14]).



Adapted from Borkova et al., 2005, Guo et al., 2008

Figure 2. (a) Representation of caseins in human, raw bovine, ewe, and goat's milk. (b) Composition and content of whey proteins in human, skimmed cow, ewe and goat's milk adapted from [15,16].

4. Cow's Milk Allergens

So far, eleven individual IgE-binding allergens from *Bos domesticus* have been accepted by the official allergen nomenclature committee (www.allergen.org) (accessed on 27 January 2021). Out of those, eight are present in milk (Table 1).

4.1. Whey

Milk whey proteins are characterized by high nutritional value, have the ability to emulsify lipids and to bind and retain water, which improves the structural and organoleptic properties of food products [18,19]. According to Monaci et al., the quality of whey proteins is well-suited to their unique amino acid composition, which is better balanced than caseins [13]. The majority of whey is globular proteins, α -lactalbumin and β -lactoglobulin, which are produced in the mammary glands. Other proteins, such as bovine serum albumin (BSA), lactoferrin and immunoglobulins derive from blood.

4.1.1. Beta-Lactoglobulin (Bos d 5)

Beta-lactoglobulin (Bos d 5)-a small protein with a molecular mass of 18.3 kDa, makes up to 50% of all whey proteins and 10% of whole milk proteins but it is essentially absent in human milk [20,21]. Beta-lactoglobulin, a dimeric protein from the lipocalin family, is one of the best characterized lipid-binding proteins. In addition, the protein is also efficiently binding many hydrophobic molecules, suggesting a role in their transport [13]. It is already known that Bos d 5 is one of the major allergenic proteins in cow's milk (Table 1). Bos d 5 contains about 8% of α -helices, 45% of β -sheets and 47% of random coils [14]. Some of the β -sheets comprise a "calyx" with two different sheets from β -A to β -D and from β -E to β -I [22]. It consists of 162 amino acids and occurs as three variants (A, B and C) with two disulfide bonds and one free sulfhydryl group buried within the protein structure [22]. These three variants A, B, and C, which were recently studied, contain two different point mutations [22]. Although the structure of the A and B variants is almost identical, they differ in amino acid residues: Asp64 in A is changed to Gly in B, and Val118 in A is changed to Ala in B [17,22]. Based on the results of the experimental studies, these two amino acid exchanges account for the different intensity in and duration of the IgE response [13]. Moreover, it was suggested, that the structure of variant A is more flexible compared to variant B [23]. However, variant B is more thermally stable than variants A and C.

Under certain conditions (including pH, temperature, ionic strength and protein concentration), the individual β -lactoglobulin isoforms are present in different oligomeric states. For example, after increasing acidic conditions we can observe that genetic variant A (dimer) associates into octomers with 144 kDa, while variants B and C do not oligomerize to octomers [22]. Different conformational changes and reversible dissociations appear at 60 °C. Some irreversible conformational changes in monomers can happen after heating up to 70–80 °C.

As mentioned above, β -lactoglobulin is a major allergen, recognized by specific IgE in more than 50% of milk allergic patients. The molecule contains several IgE epitopes, which are located (exposed) on its surface. It was previously shown that patients with IgE- mediated cow's milk allergy had seven IgE and six IgG binding epitopes, while in younger patients only three of these IgE binding epitopes were recognized [24]. In that case, a large number of β -lactoglobulin epitopes may be a marker of persistent cow's milk allergy. While much is known about the allergenic activity of Bos d 5 less is known about unexpected exposure to milk allergens. For example, β -lactoglobulin was also detected in house dust and cosmetics [21].

Interestingly, β -lactoglobulin can be used as a transporter for drugs in cancer treatment because of the physicochemical properties of the protein and its ability to bind a wide range of different ligands [25–27].

4.1.2. Alpha-Lactalbumin (Bos d 4)

Alpha-lactalbumin ranks second after β -lactoglobulin in whey proteins regarding abundancy. It represents 20–25% of whey proteins, or 2–5% of the total protein [14]. Alpha-lactalbumin is a 14.2 kDa (pH 4–5) monomeric globular protein [28]. The protein regulates the production of lactose in the milk of almost all mammalian species and it is found in considerable quantity in human breast milk.

Bos d 4 has two Ca²⁺ binding sites [17,20,28] and therefore many researchers use this protein as a calcium-binding model. Alpha-lactalbumin has intermediate molten folded globule-like states, which is of relevance for food processing strategies [29]. Moreover, it was recently shown that certain α -lactalbumin variants might induce apoptosis in tumors. A lower number of α -lactalbumin molecules binding oleic acid as a cofactor and thus can induce cytolysis of several types of malignant cells [18].

The primary structures of bovine and human α -lactalbumin share 72% sequence similarity. It has antibacterial and immunostimulating properties, which makes it a protein with high nutritional value in general and especially for babies [30].

Native α -lactalbumin from different species (humans, cows, camels and goats) consists of 123 amino acids. Furthermore, the protein contains a large number of essential amino acids (Trp, Val, Lys, Ile, Leu, Thr, Met, Phe and His) ensuring its excellent nutritional value. The amino acid composition is dominated (milligrams of amino acid per gram of protein) by leucine (108), lysine (109) and isoleucine (60) [31]. It should also be noted that four cysteine residues (48), allow the formation of disulfide bridges, and it can be released by digestion and appear in the blood as either the disulfide cysteine or free cysteine [31]. Bos d 4 contains two structural regions: a large α -helical domain and a small β -sheet domain, which are separated from each other by a deep cleft [29] but they are held together due to the cysteine bridges that form the Ca²⁺ binding loop. Ostrovsky et al. provided information about the structural changes of Bos d 4 after binding Ca²⁺. They observed that this binding might lead to both a tryptophan fluorescence blue shift and a decrease in fluorescence quantum yield, respectively [32]. While α -lactalbumin in cow's milk represents up to only 25% of the whey fraction, the human α -lactalbumin increases up 40% [18] (Figure 2b).

4.1.3. Serum Albumin (Bos d 6)

Serum albumin is the main protein of mammalian blood, present in milk and meat. It is present in milk up to 5% of total whey proteins (67 kDa). Bos d 6 is described as a protein with strong ligand binding capacity. It not only binds fatty acids, but also flavor compounds and metal ions [18]. As mentioned above, the concentration of Bos d 6 in milk is low, and BSA has little effect on the physicochemical properties of whey protein concentrates and whey protein isolates [33]. Bovine serum albumin contains 584 amino acids. The protein consists of nine loops connected by 17 disulfide bonds [12]. Bos d 6 is a minor allergen, affecting <50% of cow's milk allergic patients. Interestingly, it was described that beef allergic patients sensitized to Bos d 6 develop cross-reactivity to milk [19].

Moreover, patients with milk allergy and sensitization to cow's serum albumin are at risk of developing sensitivity to animal dandruff, which can be a cause of rhinoconjunctivitis and/or bronchial asthma [14].

4.2. Caseins (Bos d 8; Bos d 9-12)

Caseins (Bos d 8) form the main protein fraction of cow's milk (80%) and consist of: α S1- (Bos d 9), α S2- (Bos d 10), β - (Bos d 11), and κ -caseins (Bos 12) representing 40%, 13%, 37%, and 10%, respectively [13].

Caseins play a significant role in human health. The biological function of caseins is to [34,35] (1) carry calcium and phosphate (preventing the calcification of the mammary gland); (2) to provide immunological protection to infants; and (3) containing high content of amino acids, minerals, and lipids [36].

It was shown [37] that caseins aggregate into particles with micellar structure with colloidal calcium phosphate in fresh milk. The molecules form a casein micelle with a hydrophobic central part and a hydrophilic peripheral layer [14]. The size of casein micelles ranges from 0.01 to 0.3 μ m. This micelle formation can be applied to deliver various bioactive food ingredients [38].

Caseins are usually phosphate-conjugated, and form calcium phosphate-micelle complexes with mainly α S1-, α S2-, β - and, κ -caseins.

Alpha S1-casein is the main fraction of casein (40%) and consists of one major and one minor subunit [39]. According to Chianesea et al., bovine α s1-casein contains two common isoforms (A, B) with one amino acid exchange of Leu 178 (A) \rightarrow Ser 178 (B) [40]. IgE sensitization is especially frequent against α S1-casein, inducing strong immediate or delayed allergic reactions [41].

Alpha S1-casein consists of 199 amino acids with a high amount of proline residues and a lack of disulfide bonds [42]. In that case, all IgE epitopes are linear. Cocco et al. reported that denatured α -casein can bind IgE with the same binding capacity compared to native α -casein [42].

Alpha S2-casein, representing up to 13% of the caseins in cow's milk, is hydrophobic and the most phosphorylated casein fraction [43]. It has been shown that α S2-casein could form amyloid fibrils at 37 °C, however, only under nonreducing conditions [34].

Beta-casein represents about 35–37% (209 amino acids) of caseins. Plasmin can degrade β -casein into γ 1- γ 2- γ 3-casein fragments [13]. Beta-casein consists of two isoforms: A1 and A2, which differ in amino acid residue 67 (Histidine in A1 and Proline in A2) [44]. Chatchatee et al. identified six major and three minor IgE-binding epitopes using sera from 15 milk allergic patients [24].

Kappa-casein represents only 10% of caseins [13]. Nine different isoforms (A to J) of κ -casein were found [45]. The main isoforms (A and B) differ at position 136 (Ile \rightarrow Thr) and 148 (Ala \rightarrow Asp). Kappa-casein contributes to the stability of milk due to its ability to provide steric and electrostatic repulsion [46]. So far, eight major IgE epitopes have been identified by Chatchatee et al. [24].

It has been shown that even after heating caseins do not undergo significant structural changes [13]. However, caseins are sensitive to degradation by various proteinases. Caseins have different primary structures and functional properties. For example, three of them— α S1-, α S2-, and β -caseins—are calcium-sensitive, while κ -casein is not [13].

The biological function of caseins is to transfer nutritional components from the mother to the newborn [47]. Due to their colloidal properties, they are also added to a large number of food products, cosmetics and drugs, such as infant food and protein cocktails [47].

Interestingly, caseins can be used as a carrier of different drugs and pharmaceutical compounds [47]. Gandhi and colleagues showed that casein nanoparticles with doxorubicin (1.29 μ g/mg of casein nanoparticles) could release 90% of the drug doxorubicin under acidic pH. In that case, these nanoparticles can act as a drug release vehicles that enable successful drug delivery into the stomach [47].

4.3. IgE Cross-Reactivity of Milk Proteins from Different Species

It is well known that people with cow's milk allergies can develop symptoms when consuming milk from other species. In a study performed by Restani et al., serum samples from patients with cow's milk allergy were tested for IgE cross-reactivity to milk from other species [48]. It was shown that specific IgE antibodies recognized proteins from buffalo's, goat's, and ewe's milk. Cross-reactivity between cow's milk's components and other mammalian species' (ewe and buffalo) is evident for caseins (especially for α S1, α S2-caseins) and for β -lactoglobulin [48]. Most interestingly, children with cow's milk allergy did not show cross-reactivity to camel's milk. This was confirmed by another study from Ehlayel et al. showing that almost 80% of cow's milk allergic patients tolerated camel's milk and had negative skin-prick test results [49].

Bellioni-Businco et al. investigated, in vitro and in vivo, the allergenicity of goat's milk in children with cow's milk allergy [50]. The authors concluded that goat's milk is not a recommended substitution for children with cow's milk allergy. In another study Businco showed that 96% of children with IgE-mediated cow's milk allergy (n = 25) tolerated consumption of mare's milk [51].

Summarizing, camel and mare's milk might be a promising substitute for cow's milk for allergic children; however, these studies need to be performed in larger cohorts.

A range of different processing techniques for milk have been developed and the following chapter will provide a summary on the currently applied methods and their impact on the individual cow's milk allergens.

5. Food Processing: Applied Techniques and Effect on the Allergenicity of Individual Cow's Milk Proteins

Cow's milk is consumed daily worldwide with annual total numbers of 81 billion tons for India, 33.4 billion tons for the EU and 21.2 billion tons for the United States in 2019 according to Statista (https://www.statista.com) (accessed on 27 January 2021). In industrialized countries fresh cow's milk is usually extensively processed to be safe for human consumption, to meet consumer requirements and also to prolong its shelf life. After harvesting, milk is immediately cooled and transported to milk factories.

The raw milk is subjected to different processes to inactivate pathogenic microorganisms such as bacteria, spores, yeast, molds, and viruses, which can cause health problems in humans. Mostly heat-treatments are applied, although microfiltration processes are also put in place [52]. Additional processes employed include conventional techniques such as homogenization, fermentation to produce yogurts, vacuum evaporation to obtain condensed milk, or spray drying for milk powders. Newer methods such as irradiation, ultrasound processing, or cold plasma treatment are also applied.

Different treatments may affect milk proteins and induce modifications. The ratio in which the different modifications occur depends on the processes that are employed and their combination to obtain specific products (liquid and dry dairy products). Furthermore, chemical reactions occur between proteins and fat and sugars of the food matrix.

These methods can be categorized into two processing types, thermal and nonthermal. In the first case, food can be thermally processed by using moist heat or dry heat. Each of these steps induces profound changes in the quality of the milk, resulting in altered health properties.

5.1. Thermal Processing

Differences in time and temperature are the most crucial factors during heat-treatment. In industry, three principal categories of moist heat-processing are commonly used: pasteurization, sterilization, and ultrahigh temperature (UHT) processing.

5.1.1. Pasteurization

For pasteurization, the conditions vary from 65 $^{\circ}$ C for a few seconds to more than 80 $^{\circ}$ C for up to several minutes. Even mild conditions are appropriate to destroy pathogens while other microorganisms are significantly reduced (depending on the temperature and the time) but can still cause spoilage.

The degree of the structural changes of the proteins occurring during heat-treatment depends on the thermal conditions and time of treatment as well as on the type of protein and the presence of other food components such as lipids and carbohydrates, known as the "matrix effect" [53].

Caseins are stable to heat-treatment because they show very few secondary and tertiary structures (as described above). Morisawa and colleagues showed that heat treatment alone without subsequent enzymatic digestion did not alter the allergenicity of α -casein [54]. In line with this, Bloom et al. demonstrated that when using sera from milk allergic patients, IgE binding to heat-treated casein (90 °C) persists regardless of heating time. Interestingly, the presence of wheat during heating resulted in the decreased binding of the specific IgE to milk proteins. In contrast, in a study by Xu et al., the allergenicity of α -casein and β -casein showed varied changes, but was generally lower than in the untreated samples. When heating up to 65–70 °C, the allergenicity of α -casein decreased, whereas the allergenicity of β -casein severely increased, leading to the conclusion that different proteins show different sensitivities under heat treatment [55]. A recent study in a Moroccan population on the allergenicity of caseins after heating revealed reduced binding of specific human IgE [56].

Whey proteins are unstable when heated, leading to alterations in structure and thus allergenicity. It has been shown that the antigenicity of α -lactalbumin and β -lactoglobulin increases in parallel with temperature from 50–90 °C, with the highest antigenicity of α -lactalbumin and β -lactoglobulin detected at 90 °C. However, above 90 °C the antigenicity of both proteins showed a remarkable decrease.

5.1.2. Sterilization

When milk is sterilized (120 °C for 20 min), the antigenicity of α -lactalbumin decreased below the initial value of the untreated sample [57]. Heat-treatment also reduced the IgE-binding capacity of β -lactoglobulin [58]. Xu et al. obtained comparable results when heating cow's milk allergens to 65–100 °C for up to 30 min. The antigenicity of α -lactalbumin significantly decreased, whereas that of β -lactoglobulin showed an increase up to 85 °C but decreased significantly at higher temperatures [55].

In line with this, thermal treatment of β -lactoglobulin (80–100 °C) reduced its ability to induce histamine release from sensitized human basophils [54]. Thus, it seems that β -lactoglobulin presents new epitopes upon heating, but at temperatures above 85 °C it builds aggregates via covalent and noncovalent interactions, thus masking conformational

epitopes. Linear epitopes also become inaccessible in this compact structure, resulting in decreased allergenicity [59].

In vivo studies in mice revealed that oral sensitization to raw milk showed fewer acute allergic symptoms upon intradermal administration compared to processed milk. Allergen-specific IgE levels and Th2 cytokines were also significantly lower in mice sensitized to raw milk. This showed that raw milk and native whey proteins have a lower allergenic potential than their processed counterparts. These results were supported by a pilot study where milk allergic children tolerated up to 50 mL raw milk but only 8.6 mL processed "shop milk" [60].

Contradicting results came from a study in brown Norway rats where heat-treated whey proteins showed reduced intraperitoneal sensitizing capacity. Interestingly, heat-treatment did not influence the oral sensitizing capacity but significantly reduced the eliciting capacity compared to unmodified whey upon oral challenge. Heat-treatment did not reduce the tolerogenic properties of whey, as it equally prevented sensitization in naïve rats. Another interesting finding of this study was that heat-treated whey protein was less absorbed via the epithelium but more into the Peyer's patches. The authors concluded that the route of the uptake in the digestive tract may affect protein allergenicity [61].

5.1.3. Ultrahigh Temperature (UHT) Processing

UHT processing is done at higher temperatures compared to pasteurization and sterilization. During UHT, milk is exposed to a temperature of at least 135 °C to destroy bacterial and fungal spores, but only for a few seconds. As mentioned above, the heating of milk proteins leads to protein denaturation as well as extensive chemical modification (Maillard reaction). However, under these conditions the level of chemical modifications is much lower compared to lower temperatures for extended times [62], making UHT processing popular for industry. Unfortunately, little is known about the effect of UHT processing in the context of allergy. One study exists evaluating the immunogenicity of UHT treated milk by skin prick test in children with cow's milk allergy. However, UHT treated milk does not behave significantly different from other forms of cow's milk in this setting [63].

5.1.4. Baking Milk

There is emerging evidence that the properties of allergenic proteins are more complex than being stable to heat-processing or not and that the food matrix plays an important role. The majority of milk allergic children tolerate products containing baked milk. It has been shown that milk allergenicity is changed by the baking process in muffins. Baked milk within a matrix such as wheat is less likely to cause allergic reactions [64]. It seems that major allergens are destroyed during baking. The eliciting dose in children tolerant to baked milk was also higher [65,66]. Furthermore, in a follow up study it was shown that a diet including baked milk is associated with progressive immunomodulation compared to strict avoidance of milk products [67].

5.1.5. Spray Drying

Milk powder has a much longer shelf life and also lower transportation costs than liquid milk. Currently, it is produced through spray drying, where the liquid material is vaporized quickly with hot air. Lactose is a reducing sugar present in milk that under certain heating conditions reacts with free amino acid side chains, mainly from lysine, to form glycation products. These modifications have been shown to alter the allergenicity of proteins. However, little is known about the effect of spray drying on the allergenicity of milk allergens. A recent study investigated the changes of milk proteins after simulated industrial processing [68]. The authors could show that the degree of glycation after spray drying (170 °C) was increased, although only slightly. Another study investigated the IgG and IgE binding capacity of β -lactoglobulin after spray drying. At a drying temperature of 120 °C no changes were found, whereas when β -lactoglobulin was spray dried with 180 °C

under the presence of lactose, aggregation occurred and 7 lysine side chains were modified and the IgG/IgE binding capacity decreased [69]. A study examining the differential effects of the dry vs. wet heating of β -lactoglobulin revealed that dry heating requires the presence of lactose to show increased IgE recognition in most individuals tested [70].

5.2. Nonthermal Processing

5.2.1. Homogenization

Heat-treatment is often followed by homogenization to prevent phase separation, improve emulsion and also to increase shelf life. For homogenization, milk is forced through narrow pipes causing a sharp compression in the fluid flow. This leads to disruption of the relatively large and polydisperse fat globules into a large number of lipid droplets that are much smaller and show a homogeneous size range. In total, these lipid droplets have a much larger surface area adsorb mainly caseins and to a minor extent whey proteins, resulting in fat globules loaded with protein. Conflicting data exist concerning the effect of homogenization on the allergenicity of milk proteins. Using a murine model, Poulsen and colleagues could show that increasing the fat contents in combination with homogenization resulted in an increase in the ability of the milk to induce anaphylactic reactions [71]. In line with this, a double-blind placebo-controlled study in milk allergic children revealed an increased ability of homogenized/pasteurized milk to evoke allergic reactions [72]. However, this could not be confirmed by other studies [73–75]. Further studies in humans did not show differences in the tolerance to homogenized and unhomogenized milk, respectively [76]. A review by Paschke et al. indicated that homogenization does not alleviate the potency of cow's milk allergens [77].

5.2.2. High Pressure Homogenization (HPH)

HPH (150 MPa) has been proposed as a substitute for the thermal processing of food. The binding capacity of casein to IgE before and after high pressure homogenization was reduced, as shown by ELISA [78]. However, more research is needed to determine the precise effect of homogenization on the allergenicity of milk allergens.

5.2.3. Ultrasonic Treatment

Ultrasonic technology is used in food industry to improve food quality efficiently and also to develop new products with unique functions. Ultrasound improves the foaming and emulsifying properties and also casein stability [79] and can be done through a bath or an ultrasound probe. Two main differences exist when comparing these two possibilities: first, the ultrasonic probe is immersed directly into the solution, where the sonication takes place; and second, the ultrasonic power provided by the probe is much greater than the one supplied by the bath. The application of the high intensity focused ultrasound allows for the accelerated digestion of the proteins. This is of special interest when it comes to allergenicity. Depending on the intensity, different degrees of hydrolysis can be obtained. Decreased allergenicity has been reported for casein via colloid formation [80]. In line with this, a marked decrease in allergenicity was observed for β -lactoglobulin [81]. However, this technique has some disadvantages as it leads to the formation of free radicals [82].

5.2.4. Enzymatic Processes

Enzymatic hydrolysis is a method used to break down proteins into smaller peptides or amino acids, resulting in a loss of structure and thus the removal of the conformational epitopes that are recognized by specific IgE. This method is applied to generate hypoallergenic milk formulas for milk allergic babies. These products contain only short peptides, which are unable to induce an allergic reaction. In a mouse model of sensitization, Duan and colleagues could show that mice sensitized by hydrolysates of β -lactoglobulin showed a significantly lower spleen lymphocyte proliferation level than those sensitized by intact β -lactoglobulin suggesting that enzymatic hydrolysis reduces its allergenicity [83]. Other studies showed retained or even enhanced allergenicity after enzymatic proteolysis. The digestion of milk proteins could also unmask epitopes leading to the increased binding of specific IgE. This disagreement is most likely due to the use of different enzymes [84,85]. Moreover, it was shown that peptides with a molecular mass smaller than 3 kDa remained allergenic [86]. It seems that choice of an appropriate enzyme to effectively decrease residual antigenicity is of great importance. Other studies showed that the combination of enzymatic digestion with heat-processing led to increased reduction of antigenicity [87,88]. The effect of enzymatic hydrolysis is reviewed in detail by [89].

The cross-linking of food proteins is often used to enhance food stability. Several highly specific enzymes (e.g., transglutaminase, horse radish peroxidase, laccase, tyrosinase) are used as additives in food industry to improve texture and functional properties. Currently, the only enzyme approved by the European Union for food marketing is transglutaminase. The treatment of whey proteins with transglutaminase seems to hide important epitopes, thus reducing their allergenic potential [90]. However, when using other enzymes, the treatment may also show unwanted effects such as increased allergenicity [91]. Another important factor is the pretreatment of proteins to make proteins accessible for enzymatic digestion. Chemical or thermal pretreatments are usually applied to improve the accessibility for enzymatic digestion. Therefore, all cross-linked proteins have to be tested for their capacity to bind specific IgE [92].

5.2.5. Fermentation

During microbial fermentation, lactic acid bacteria (LAB) produce proteolytic enzymes causing the degradation of milk proteins to peptides and amino acids. This not only decreases the allergenicity of milk allergens due to the breakdown of IgE epitopes [93,94], but also leads to the production of bioactive peptides. The grade of the reduction in allergenicity is dependent on the proteases and peptidases of the LAB strains used [95]. Several studies exist showing increased tolerability of fermented milk products such as yogurt and cheese. Alessandri and colleagues showed that 58% of patients clinically reactive to cow's milk tolerated fully maturated Parmigiano-Reggiano [96]. Yogurt is tolerated by the majority of children with cow's milk allergy [97].

5.3. Novel Techniques

5.3.1. Irradiation

Gamma irradiation has been proven to be an effective and safe method for the sterilization of certain products. However, this treatment does not only inhibit microorganisms but also alters the structure of the molecules targeted by this process. Thus, irradiation can improve the quality of milk, but can also introduce modifications. Beta-lactoglobulin (in solution) subjected to gamma radiation led to changes in the secondary and tertiary structure leading to protein aggregation [98]. Several studies have indicated that ionizing radiation could reduce allergenicity by the destruction of human IgE-binding epitopes in milk allergens depending on the dose of irradiation [99]. A study assessing the allergenicity of irradiated dairy products in BALB/c mice revealed that gamma irradiation influenced the epitopes of the major milk proteins and was associated with lower allergenicity of lyophilized irradiated milk [100]. In line with this, another study, investigating ultrasoundassisted irradiation for reducing the allergenicity of β -lactoglobulin, revealed that IgE binding capacity and release of inflammatory mediators from human basophil cells were reduced significantly [69].

5.3.2. Microwave Treatment

Microwave treatment has been shown to be a good alternative to conventional heat treatments. Microwaves are electromagnetic waves and heat is generated following the absorption of microwave energy by water, organic molecules, or ions. However, phenomena that cannot be explained by the increase in temperature also occur and there is evidence that microwaves contribute to the existence of the nonthermal effects of microwave treatment [101]. Therefore, it is important to investigate changes in the structure

and allergenicity of the proteins, especially if microwave treatment is performed in combination with other treatments. A study by Izquierdo and colleagues showed that microwave irradiation accelerates enzymatic treatments and also increases the degree of hydrolysis [87]. Another study, comparing the effects of microwave heating and conventional heating on the secondary and tertiary structures of β -lactoglobulin, revealed a substantial enhanced unfolding and exposure of buried amide groups. Thus, microwave processing could be a future alternative to produce hydrolysates with lower allergenicity [102,103].

5.3.3. Cold Plasma Treatment

Atmospheric cold plasma appeared to be a promising novel technology to induce structural modifications of proteins [104]. It has been reported that treatment with cold plasma induces structural modifications and alters the antigenic response of the bovine milk allergens. In a recent study, casein, β -lactoglobulin and α -lactalbumin were analyzed before and after plasma treatment. The results revealed alterations in the secondary structure of the protein and decreased antigenicity of the casein and α -lactalbumin, whereas β -lactoglobulin showed increased antigenicity [105].

6. Concluding Remarks

For thousands of years, milk has been part of the human diet. In ancient times milk was consumed either raw or fermented, but since the late 19th century, milk has regularly been heat-treated to destroy pathogenic microorganisms to be safe for human consumption. In the recent past, milk has become more and more processed, not only for safety reasons but also to prolong shelf life and fulfil consumer expectations (convenient food products). However, intensive processing can also lead to the reduction of the bioactive compounds, which become degraded.

Cow's milk also causes immune-mediated diseases such as non-IgE-mediated gastrointestinal food allergic disorders, which start early in life, and the total protein fraction seems to be a relevant trigger. Currently there is a lack of clear understanding of the different pathomechanisms. However, since most of these diseases resolve early in childhood, maturation and tolerance induction in the gastrointestinal tract may also be supportive of the resolution of the symptoms. IgE-mediated cow's milk allergy also starts early in infancy and the majority of patients experience cure within 4–5 years, while a minority will also suffer from cow's milk allergy in adulthood. Relevant IgE binding allergens have been identified and the diagnosis and treatment of cow's milk allergy has improved due to our knowledge on the individual immunoactive proteins. However, there is only limited knowledge on how different processing methods affect the allergenicity of individual milk proteins. The thermal processing of proteins can affect their 3D structure and thus destroy IgE-binding epitopes or expose those that have been buried inside the structure. For whey proteins, different temperature ranges affect them, either up- or downregulating their allergenicity. This was also found for caseins, which showed different sensitivities for the individual caseins. The homogenization of milk results in a large number of lipid droplets adsorbing caseins and whey proteins. This can lead to an increase in allergic reactions in patients. Ultrasonic treatment facilitates colloid formation, which in turn can reduce allergenicity, as shown for β -lactoglobulin. It is generally accepted that enzymatic degradation breaks down proteins into peptides, resulting in hypoallergenic products. However, this mainly depends on the enzymes applied and the detailed analysis of the size and sequence of the obtained peptides. In the case of the cross-linking of food proteins via transglutaminase, epitopes may be hidden, as shown for whey proteins. Applying microbial fermentation for the production of yogurt and cheese can be beneficial for milk allergic patients since the allergens are degraded. For other processing techniques such as irradiation, microwave treatment, and cold plasma treatment, reports showed that these methods can affect the structure of the individual allergens and thus their allergenicity. However, at present only a low number of studies have been performed, with sometimes conflicting evidence. More evidence is needed to obtain a better understanding of which methods are relevant for

reducing the allergenicity of the individual proteins present in milk, while preserving the bioactive ingredients of this precious food as part of the human diet.

Author Contributions: S.G., A.P., and K.H.-S. performed the literature survey, wrote the manuscript and read and agreed to the published version of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: Work on allergen processing, including writing this review, is financially supported by a grant from the Austrian Science Fund (FWF), P 33582-B and by a grant from the Danube Allergy Research Cluster, DARC-07.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Bleasdale, M.; Richter, K.K.; Janzen, A.; Brown, S.; Scott, A.; Zech, J.; Wilkin, S.; Wang, K.; Schiffels, S.; Desideri, J.; et al. Ancient proteins provide evidence of dairy consumption in eastern Africa. *Nat. Commun.* 2021, 12, 1–11. [CrossRef]
- Muraro, A.; Werfel, T.; Hoffmann-Sommergruber, K.; Roberts, G.A.; Beyer, K.; Bindslev-Jensen, C.; Cardona, V.; Dubois, A.; Dutoit, G.; Eigenmann, P.; et al. EAACI Food Allergy and Anaphylaxis Guidelines: Diagnosis and management of food allergy. *Allergy* 2014, 69, 1008–1025. [CrossRef]
- Caubet, J.-C.; Szajewska, H.; Shamir, R.; Nowak-Węgrzyn, A. Non-IgE-mediated gastrointestinal food allergies in children. *Pediatr. Allergy Immunol.* 2017, 28, 6–17. [CrossRef] [PubMed]
- 4. Mehr, S.; Kakakios, A.; Frith, K.; Kemp, A.S. Food Protein-Induced Enterocolitis Syndrome: 16-Year Experience. *Pediatrics* 2009, 123, e459–e464. [CrossRef]
- Caubet, J.C.; Ford, L.S.; Sickles, L.; Järvinen, K.M.; Sicherer, S.H.; Sampson, H.A.; Nowak-Węgrzyn, A. Clinical features and resolution of food protein–induced enterocolitis syndrome: 10-year experience. *J. Allergy Clin. Immunol.* 2014, 134, 382–389.e4. [CrossRef] [PubMed]
- Kaya, A.; Toyran, M.; Civelek, E.; Misirlioglu, E.; Kirsaclioglu, C.; Kocabas, C.N. Characteristics and Prognosis of Allergic Proctocolitis in Infants. J. Pediatr. Gastroenterol. Nutr. 2015, 61, 69–73. [CrossRef]
- Calvani, M.; Anania, C.; Cuomo, B.; D'Auria, E.; Decimo, F.; Indirli, G.; Marseglia, G.; Mastrorilli, V.; Sartorio, M.; Santoro, A.; et al. Non–IgE- or Mixed IgE/Non–IgE-Mediated Gastrointestinal Food Allergies in the First Years of Life: Old and New Tools for Diagnosis. *Nutrients* 2021, 13, 226. [CrossRef]
- Nwaru, B.I.; Hickstein, L.; Panesar, S.S.; Roberts, G.; Muraro, A.; Sheikh, A.; the EAACI Food Allergy and Anaphylaxis Guidelines Group. Prevalence of common food allergies in Europe: A systematic review and meta-analysis. *Allergy* 2014, 69, 992–1007. [CrossRef] [PubMed]
- 9. Abrams, E.M.; Sicherer, S.H. Cow's milk allergy prevention. Ann. Allergy Asthma Immunol. 2021. [CrossRef] [PubMed]
- 10. Cazzola, R.; Della Porta, M.; Manoni, M.; Iotti, S.; Pinotti, L.; Maier, J.A. Going to the roots of reduced magnesium dietary intake: A tradeoff between climate changes and sources. *Heliyon* **2020**, *6*, 05390. [CrossRef]
- 11. Mudgil, D.; Barak, S. Dairy-Based Functional Beverages. Milk-Based Beverages 2019, 67–93. [CrossRef]
- 12. Wal, J.-M. Cow's milk proteins/allergens. Ann. Allergy Asthma Immunol. 2002, 89, 3–10. [CrossRef]
- 13. Monaci, L.; Tregoat, V.; Van Hengel, A.J.; Anklam, E. Milk allergens, their characteristics and their detection in food: A review. *Eur. Food Res. Technol.* **2006**, 223, 149–179. [CrossRef]
- 14. Rafiq, S.; Huma, N.; Pasha, I.; Sameen, A.; Mukhtar, O.; Khan, M.I. Chemical Composition, Nitrogen Fractions and Amino Acids Profile of Milk from Different Animal Species. *Asian-Australas. J. Anim. Sci.* **2016**, *29*, 1022–1028. [CrossRef]
- 15. Borková, M.; Snášelová, J. Possibilities of different animal milk detection in milk and dairy products—A review. *Czech J. Food Sci.* **2005**, *23*, 41–50. [CrossRef]
- 16. Guo, M. Chemistry and Biological Properties of Human Milk. Curr. Nutr. Food Sci. 2008, 4, 305–320. [CrossRef]
- Hochwallner, H.; Schulmeister, U.; Swoboda, I.; Spitzauer, S.; Valenta, R. Cow's milk allergy: From allergens to new forms of diagnosis, therapy and prevention. *Methods* 2014, 66, 22–33. [CrossRef] [PubMed]
- 18. Deeth, H.; Bansal, N. Whey Proteins. Whey Proteins 2019, 1–50. [CrossRef]
- Sánchez, L.; Pérez, M.D.; Parrón, J.A. HPP in dairy products: Impact on quality and applications. In *Present and Future of High Pressure Processing*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 245–272.
- Krunic, T.; Rakin, M.; Bulatovic, M.; Zaric, D. The Contribution of Bioactive Peptides of Whey to Quality of Food Products. *Food Process. Increased Qual. Consum.* 2018, 251–285. [CrossRef]
- Witteman, A.M.; Van Leeuwen, J.; Van Der Zee, J.S.; Aalberse, R.C. Food Allergens in House Dust. *Int. Arch. Allergy Immunol.* 1995, 107, 566–568. [CrossRef]

- 22. Qin, B.Y.; Jameson, G.B.; Bewley, M.C.; Baker, E.N.; Creamer, L.K. Functional implications of structural differences between variants A and B of bovine β-lactoglobulin. *Protein Sci.* **2008**, *8*, 75–83. [CrossRef]
- 23. Sawyer, L. β-Lactoglobulin; Springer International Publishing: Berlin/Heidelberg, Germany, 2012; pp. 211–259.
- Chatchatee, P.; Jarvinen, K.M.; Bardina, L.; Vila, L.; Beyer, K.; Sampson, H.A. Identification of IgE and IgG binding epitopes on βand κ-casein in cow's milk allergic patients. *Clin. Exp. Allergy* 2001, *31*, 1256–1262. [CrossRef]
- Izadi, Z.; Divsalar, A.; Saboury, A.A.; Sawyer, L. β-lactoglobulin-pectin Nanoparticle-based Oral Drug Delivery System for Potential Treatment of Colon Cancer. *Chem. Biol. Drug Des.* 2016, *88*, 209–216. [CrossRef]
- 26. Ball, G.; Shelton, M.J.; Walsh, B.J.; Hill, D.J.; Hosking, C.S.; Howden, M.E.H. A major continuous allergenic epitope of bovine beta-lactoglobulin recognized by human IgE binding. *Clin. Exp. Allergy* **1994**, *24*, 758–764. [CrossRef] [PubMed]
- Ghalandari, B.; Divsalar, A.; Eslami-Moghadam, M.; Saboury, A.A.; Haertlé, T.; Amanlou, M.; Parivar, K. Probing of the Interaction Between β-Lactoglobulin and the Anticancer Drug Oxaliplatin. *Appl. Biochem. Biotechnol.* 2014, 175, 974–987. [CrossRef] [PubMed]
- Caira, S.; Pizzano, R.; Picariello, G.; Pinto, G.; Cuollo, M.; Chianese, L.; Addeo, L.C.A.F. Allergenicity of Milk Proteins. *Milk Protein* 2012. [CrossRef]
- 29. Permyakov, E.A.; Berliner, L.J. α-Lactalbumin: Structure and function. *FEBS Lett.* 2000, 473, 269–274. [CrossRef]
- Heine, W.E.; Klein, P.D.; Reeds, P.J. The Importance of oc-Lactalbumin in Infant Nutrition1. Available online: https://academic. oup.com/jn/article-abstract/121/3/277/4754512 (accessed on 27 January 2021).
- Layman, D.K.; Lönnerdal, B.; Fernstrom, J.D. Applications for α-lactalbumin in human nutrition. *Nutr. Rev.* 2018, 76, 444–460. [CrossRef] [PubMed]
- Ostrovsky, A.V.; Kalinichenko, L.P.; Emelyanenko, V.I.; Klimanov, A.V.; Permyakov, E.A. Environment of tryptophan residues in various conformational states of α-lactalbumin studied by time-resolved and steady-state fluorescence spectrosc. *Biophys. Chem.* **1988**, *30*, 105–112. [CrossRef]
- 33. Goulding, D.; Fox, P.; O'Mahony, J. Milk proteins: An overview. Milk Proteins 2020, 21–98. [CrossRef]
- 34. Thorn, D.C.; Ecroyd, H.; Carver, J.A. The two-faced nature of milk casein proteins: Amyloid fibril formation and chaperone-like activity. *Aust. J. Dairy Technol.* **2009**, *6*, 34–40.
- 35. Holt, C.; Carver, J.A.; Ecroyd, H.; Thorn, D.C. Invited review: Caseins and the casein micelle: Their biological functions, structures, and behavior in foods. *J. Dairy Sci.* 2013, *96*, 6127–6146. [CrossRef]
- 36. Bhat, M.Y.; Dar, T.A.; Singh, L.R. Casein Proteins: Structural and Functional Aspects. In *Milk Proteins—From Structure to Biological Properties and Health Aspects*; InTech: Rijeka, Croatia, 2016. [CrossRef]
- 37. Dalgleish, D.G.; Corredig, M. The Structure of the Casein Micelle of Milk and Its Changes During Processing. *Annu. Rev. Food Sci. Technol.* **2012**, *3*, 449–467. [CrossRef] [PubMed]
- Zhuang, F.; Li, X.; Hu, J.; Liu, X.; Zhang, S.; Tang, C.; Zhou, P. Effects of casein micellar structure on the stability of milk protein-based conjugated linoleic acid microcapsules. *Food Chem.* 2018, 269, 327–334. [CrossRef] [PubMed]
- 39. Farrell, H.; Jimenez-Flores, R.; Bleck, G.; Brown, E.; Butler, J.; Creamer, L.; Hicks, C.; Hollar, C.; Ng-Kwai-Hang, K.; Swaisgood, H. Nomenclature of the Proteins of Cows' Milk—Sixth Revision. *J. Dairy Sci.* **2004**, *87*, 1641–1674. [CrossRef]
- Chianese, L.; Quarto, M.; Pizzolongo, F.; Calabrese, M.G.; Caira, S.; Mauriello, R.; De Pascale, S.; Addeo, F. Occurrence of genetic polymorphism at the αs1-casein locus in Mediterranean water buffalo milk. *Int. Dairy J.* 2009, *19*, 181–189. [CrossRef]
- Archila, L.D.; Khan, F.S.; Bhatnagar, N.; Robinson, D.; Farrington, M.L.; Kwok, W.W. αS1-Casein elucidate major T-cell responses in cow's milk allergy. J. Allergy Clin. Immunol. 2017, 140, 854–857.e6. [CrossRef]
- 42. Cocco, R.R.; Järvinen, K.-M.; Sampson, H.A.; Beyer, K. Mutational analysis of major, sequential IgE-binding epitopes in αs1-casein, a major cow's milk allergen. J. Allergy Clin. Immunol. 2003, 112, 433–437. [CrossRef]
- 43. Farrell, H. Milk Proteins | Casein Nomenclature, Structure, and Association. In *Encyclopedia of Dairy Sciences*; Elsevier: Amsterdam, The Netherlands, 2011; pp. 765–771.
- 44. De Gaudry, D.K.; Lohner, S.; Schmucker, C.; Kapp, P.; Motschall, E.; Hörrlein, S.; Röger, C.; Meerpohl, J.J. Milk A1 β-casein and health-related outcomes in humans: A systematic review. *Nutr. Rev.* **2019**, *77*, 278–306. [CrossRef]
- 45. Miranda, G.; Anglade, P.; Mahé, M.F.; Erhardt, G. Biochemical characterization of the bovine genetic K-casein C and E variants. *Anim. Genet.* **2009**, *24*, 27–31. [CrossRef] [PubMed]
- Bonfatti, V.; Chiarot, G.; Carnier, P. Glycosylation of κ-casein: Genetic and nongenetic variation and effects on rennet coagulation properties of milk. J. Dairy Sci. 2014, 97, 1961–1969. [CrossRef]
- 47. Gandhi, S.; Roy, I. Doxorubicin-loaded casein nanoparticles for drug delivery: Preparation, characterization and in vitro evaluation. *Int. J. Biol. Macromol.* 2019, 121, 6–12. [CrossRef] [PubMed]
- 48. Restani, P.; Gaiaschi, A.; Plebani, A.; Beretta, B.; Cavagni, G.; Fiocchi, A.; Poiesi, C.; Velonà, T.; Ugazio, A.G.; Galli, C.L. Cross-reactivity between milk proteins from different animal species. *Clin. Exp. Allergy* **1999**, *29*, 997–1004. [CrossRef] [PubMed]
- 49. Ehlayel, M.S.; Abu Hazeima, K.; Al-Mesaifri, F.; Bener, A. Camel milk: An alternative for cow's milk allergy in children. *Allergy Asthma Proc.* **2011**, *32*, 255–258. [CrossRef]
- 50. Bellioni-Businco, B.; Paganelli, R.; Lucenti, P.; Giampietro, P.G.; Perbornc, H.; Businco, L. Allergenicity of goat's milk in children with cow's milk allergy. *J. Allergy Clin. Immunol.* **1999**, *103*, 1191–1194. [CrossRef]
- Businco, L.; Giampietro, P.G.; Lucenti, P.; Lucaroni, F.; Pini, C.; Di Felice, G.; Iacovacci, P.; Curadi, C.; Orlandi, M. Allergenicity of mare's milk in children with cow's milk allergy. J. Allergy Clin. Immunol. 2000, 105, 1031–1034. [CrossRef] [PubMed]

- 52. Griep, E.R.; Cheng, Y.; Moraru, C.I. Efficient removal of spores from skim milk using cold microfiltration: Spore size and surface property considerations. *J. Dairy Sci.* 2018, 101, 9703–9713. [CrossRef] [PubMed]
- 53. Nowak-Wegrzyn, A.; Fiocchi, A. Rare, medium, or well done? The effect of heating and food matrix on food protein allergenicity. *Curr. Opin. Allergy Clin. Immunol.* 2009, *9*, 234–237. [CrossRef]
- Morisawa, Y.; Kitamura, A.; Ujihara, T.; Zushi, N.; Kuzume, K.; Shimanouchi, Y.; Tamura, S.; Wakiguchi, H.; Saito, H.; Matsumoto, K. Effect of heat treatment and enzymatic digestion on the B cell epitopes of cow's milk proteins. *Clin. Exp. Allergy* 2009, 39, 918–925. [CrossRef]
- 55. Xu, Q.; Shi, J.; Yao, M.; Jiang, M.; Luo, Y. Effects of heat treatment on the antigenicity of four milk proteins in milk protein concentrates. *Food Agric. Immunol.* **2015**, *27*, 401–413. [CrossRef]
- Azdad, O.; Mejrhit, N.; El Kabbaoui, M.; Chda, A.; Ouahidi, I.; Tazi, A.; Bencheikh, R.; Aarab, L. Effect of heating and enzymatic hydrolysis on casein cow milk sensitivity in Moroccan population. *Food Agric. Immunol.* 2017, 29, 424–433. [CrossRef]
- 57. Bu, G.; Luo, Y.; Zheng, Z.; Zheng, H. Effect of heat treatment on the antigenicity of bovine α-lactalbumin and β-lactoglobulin in whey protein isolate. *Food Agric. Immunol.* **2009**, *20*, 195–206. [CrossRef]
- Bloom, K.A.; Huang, F.R.; Bencharitiwong, R.; Bardina, L.; Ross, A.; Sampson, H.A.; Nowak-Wegrzyn, A. Effect of heat treatment on milk and egg proteins allergenicity. *Pediatr. Allergy Immunol.* 2014, 25, 740–746. [CrossRef] [PubMed]
- 59. Rahaman, T.; Vasiljevic, T.; Ramchandran, L. Effect of processing on conformational changes of food proteins related to allergenicity. *Trends Food Sci. Technol.* **2016**, *49*, 24–34. [CrossRef]
- Abbring, S.; Kusche, D.; Roos, T.C.; Diks, M.A.P.; Hols, G.; Garssen, J.; Baars, T.; Esch, B.C.A.M.V. Milk processing increases the allergenicity of cow's milk-Preclinical evidence supported by a human proof-of-concept provocation pilot. *Clin. Exp. Allergy* 2019, 49, 1013–1025. [CrossRef]
- Graversen, K.B.; Ballegaard, A.R.; Kræmer, L.H.; Hornslet, S.E.; Sørensen, L.V.; Christoffersen, H.F.; Jacobsen, L.N.; Untersmayr, E.; Smit, J.J.; Bøgh, K.L. Cow's milk allergy prevention and treatment by heat-treated whey—A study in Brown Norway rats. *Clin. Exp. Allergy* 2020, *50*, 708–721. [CrossRef]
- 62. Deeth, H.; Datta, N. Ultra-high temperature treatment (UHT) | Heating Systems. Encycl. Dairy Sci. 2002, 2642–2652. [CrossRef]
- 63. Shadur, B.; Fong, A.; Altavilla, B.; Saad, R.A.; Wainstein, B.K. Skin testing with ultra-heat-treated (UHT) cow's milk in children with cow's milk allergy. *Ann. Allergy Asthma Immunol.* **2020**, *124*, 185–189. [CrossRef] [PubMed]
- 64. Sopo, S.M.; Greco, M.; Monaco, S.; Bianchi, A.; Cuomo, B.; Liotti, L.; Iacono, I. Matrix effect on baked milk tolerance in children with IgE cow milk allergy. *Allergol. Immunopathol.* **2016**, *44*, 517–523. [CrossRef]
- 65. Nowak-Węgrzyn, A.; Bloom, K.A.; Sicherer, S.H.; Shreffler, W.G.; Noone, S.; Wanich, N.; Sampson, H.A. Tolerance to extensively heated milk in children with cow's milk allergy. *J. Allergy Clin. Immunol.* **2008**, 122, 342–347.e2. [CrossRef]
- Goldberg, M.R.; Nachshon, L.; Appel, M.Y.; Elizur, A.; Levy, M.B.; Eisenberg, E.; Sampson, H.A.; Katz, Y. Efficacy of baked milk oral immunotherapy in baked milk-reactive allergic patients. *J. Allergy Clin. Immunol.* 2015, 136, 1601–1606. [CrossRef] [PubMed]
- Nowak-Węgrzyn, A.; Lawson, K.; Masilamani, M.; Kattan, J.; Bahnson, H.; Sampson, H.A. Increased Tolerance to Less Extensively Heat-Denatured (Baked) Milk Products in Milk-Allergic Children. J. Allergy Clin. Immunol. Pract. 2018, 6, 486–495.e5. [CrossRef] [PubMed]
- 68. Liu, Y.; Zhang, W.; Zhang, L.; Hettinga, K.; Zhou, P. Characterizing the changes of bovine milk serum proteins after simulated industrial processing. *LWT* **2020**, *133*, 110101. [CrossRef]
- Yang, F.; Zou, L.; Wu, Y.; Wu, Z.; Yang, A.; Chen, H.; Li, X. Structure and allergenicity assessments of bovine β-lactoglobulin treated by sonication-assisted irradiation. *J. Dairy Sci.* 2020, *103*, 4109–4120. [CrossRef] [PubMed]
- 70. Zenker, H.E.; Ewaz, A.; Deng, Y.; Savelkoul, H.F.J.; Van Neerven, R.J.; De Jong, N.W.; Wichers, H.J.; Hettinga, K.A.; Teodorowicz, M. Differential Effects of Dry vs. Wet Heating of β-Lactoglobulin on Formation of sRAGE Binding Ligands and sIgE Epitope Recognition. *Nutrients* 2019, *11*, 1432. [CrossRef] [PubMed]
- 71. Poulsen, O.M.; Hau, J.; Kollerup, J. Effect of homogenization and pasteurization on the allergenicity of bovine milk analysed by a murine anaphylactic shock model. *Clin. Exp. Allergy* **1987**, *17*, 449–458. [CrossRef] [PubMed]
- 72. Høst, A.; Samuelsson, E.-G. Allergic reactions to raw, pasteurized, and homogenized/pasteurized cow milk: A comparison. *Allergy* **1988**, *43*, 113–118. [CrossRef]
- 73. Michalski, M.-C. On the supposed influence of milk homogenization on the risk of CVD, diabetes and allergy. *Br. J. Nutr.* 2007, 97, 598–610. [CrossRef]
- 74. Michalski, M.-C.; Januel, C. Does homogenization affect the human health properties of cow's milk? *Trends Food Sci. Technol.* 2006, 17, 423–437. [CrossRef]
- 75. Mulder, H.; Walstra, P. The milk fat globule. In *Emulsion Science as Applied to Milk Products and Comparable Foods*; Commonwealth Agricultural Bureau: Bucks, UK, 1974; pp. 101–128.
- Paajanen, L.; Tuure, T.; Poussa, T.; Korpela, R. No difference in symptoms during challenges with homogenized and unhomogenized cow's milk in subjects with subjective hypersensitivity to homogenized milk. J. Dairy Res. 2003, 70, 175–179. [CrossRef]
- 77. Paschke, A.; Besler, M. Stability of bovine allergens during food processing. *Ann. Allergy Asthma Immunol.* 2002, *89*, 16–20. [CrossRef]
- 78. Han, T.; Wang, M.; Wang, Y.; Tang, L. Effects of high-pressure homogenization and ultrasonic treatment on the structure and characteristics of casein. *LWT* **2020**, *130*, 109560. [CrossRef]

- Jiang, Z.; Wang, C.; Li, T.; Sun, D.; Gao, H.; Gao, Z.; Mu, Z. Effect of ultrasound on the structure and functional properties of transglutaminase-crosslinked whey protein isolate exposed to prior heat treatment. *Int. Dairy J.* 2019, *88*, 79–88. [CrossRef]
- Wang, C.; Xie, Q.; Wang, Y.; Fu, L. Effect of Ultrasound Treatment on Allergenicity Reduction of Milk Casein via Colloid Formation. J. Agric. Food Chem. 2020, 68, 4678–4686. [CrossRef] [PubMed]
- Liu, G.-X.; Tu, Z.-C.; Yang, W.; Wang, H.; Zhang, L.; Ma, D.; Huang, T.; Liu, J.; Li, X. Investigation into allergenicity reduction and glycation sites of glycated β-lactoglobulin with ultrasound pretreatment by high-resolution mass spectrometry. *Food Chem.* 2018, 252, 99–107. [CrossRef]
- 82. Santos, H.; Capelo, J. Trends in ultrasonic-based equipment for analytical sample treatment. *Talanta* **2007**, *73*, 795–802. [CrossRef] [PubMed]
- Duan, C.-C.; Li, A.-L.; Yang, L.-J.; Zhao, R.; Fan, W.-G.; Huo, G.-C. Comparison of immunomodulating properties of Betalactoglobulin and its hydrolysates. *Iran. J. Allergy Asthma Immunol.* 2014, 13, 26–32.
- 84. Ena, J.M.; Beresteijn, E.C.H.; Robben, A.J.P.M.; Schmidt, D.G. Whey Protein Antigenicity Reduction by Fungal Proteinases and a Pepsin/Pancreatin Combination. *J. Food Sci.* **1995**, *60*, 104–110. [CrossRef]
- Sélo, I.; Clément, G.; Bernard, H.; Chatel, J.; Créminon, C.; Peltre, G.; Wal, J. Allergy to bovine β-lactoglobulin: Specificity of human IgE to tryptic peptides. *Clin. Exp. Allergy* 1999, 29, 1055–1063. [CrossRef]
- Puerta, A.; Diez-Masa, J.C.; De Frutos, M. Immunochromatographic determination of β-lactoglobulin and its antigenic peptides in hypoallergenic formulas. *Int. Dairy J.* 2006, 16, 406–414. [CrossRef]
- 87. Izquierdo, F.J.; Peñas, E.; Baeza, M.L.; Gomez, R. Effects of combined microwave and enzymatic treatments on the hydrolysis and immunoreactivity of dairy whey proteins. *Int. Dairy J.* 2008, *18*, 918–922. [CrossRef]
- El Mecherfi, K.E.; Curet, S.; Lupi, R.; Larré, C.; Rouaud, O.; Choiset, Y.; Rabesona, H.; Haertlé, T. Combined microwave processing and enzymatic proteolysis of bovine whey proteins: The impact on bovine β-lactoglobulin allergenicity. *J. Food Sci. Technol.* 2019, 56, 177–186. [CrossRef]
- 89. Bu, G.; Luo, Y.; Chen, F.; Liu, K.; Zhu, T. Milk processing as a tool to reduce cow's milk allergenicity: A mini-review. *Dairy Sci. Technol.* **2013**, *93*, 211–223. [CrossRef]
- Villas-Boas, M.B.; Vieira, K.P.; Trevizan, G.; Zollner, R.D.L.; Netto, F.M. The effect of transglutaminase-induced polymerization in the presence of cysteine on β-lactoglobulin antigenicity. *Int. Dairy J.* 2010, 20, 386–392. [CrossRef]
- 91. Stojadinovic, M.; Pieters, R.; Smit, J.; Velickovic, T.C. Cross-Linking of -Lactoglobulin Enhances Allergic Sensitization Through Changes in Cellular Uptake and Processing. *Toxicol. Sci.* 2014, 140, 224–235. [CrossRef]
- 92. Fernando, A.L. Control of Milk Allergenicity. J. Dairy Vet. Sci. 2017, 2. [CrossRef]
- 93. Yao, M.; Xu, Q.; Luo, Y.; Shi, J.; Li, Z. Study on reducing antigenic response and IgE-binding inhibitions of four milk proteins of Lactobacillus casei 1134. *J. Sci. Food Agric.* 2014, 95, 1303–1312. [CrossRef]
- Shi, J.; Luo, Y.; Xiao, Y.; Li, Z.; Xu, Q.; Yao, M. Effects of fermentation by Lactobacillus casei on the antigenicity and allergenicity of four bovine milk proteins. *Int. Dairy J.* 2014, 35, 75–80. [CrossRef]
- 95. Kazemi, R.; Taheri-Kafrani, A.; Motahari, A.; Kordesedehi, R. Allergenicity reduction of bovine milk β-lactoglobulin by proteolytic activity of lactococcus lactis BMC12C and BMC19H isolated from Iranian dairy products. *Int. J. Biol. Macromol.* 2018, 112, 876–881. [CrossRef]
- 96. Alessandri, C.; Sforza, S.; Palazzo, P.; Lambertini, F.; Paolella, S.; Zennaro, D.; Rafaiani, C.; Ferrara, R.; Bernardi, M.L.; Santoro, M.; et al. Tolerability of a Fully Maturated Cheese in Cow's Milk Allergic Children: Biochemical, Immunochemical, and Clinical Aspects. *PLoS ONE* 2012, 7, e40945. [CrossRef]
- 97. Monaco, S.; Russo, G.; Romano, A.; Liotti, L.; Verga, M.; Sopo, S.M. Yogurt is tolerated by the majority of children with IgE-mediated cow's milk allergy. *Allergol. Immunopathol.* **2019**, *47*, 322–327. [CrossRef] [PubMed]
- De La Hoz, L.; Netto, F.M. Structural modifications of β-lactoglobulin subjected to gamma radiation. *Int. Dairy J.* 2008, 18, 1126–1132. [CrossRef]
- Lee, J.-W.; Kim, J.-H.; Yook, H.-S.; Kang, K.-O.; Lee, S.-Y.; Hwang, H.-J.; Byun, M.-W. Effects of Gamma Radiation on the Allergenic and Antigenic Properties of Milk Proteins. J. Food Prot. 2001, 64, 272–276. [CrossRef]
- Miteva, D.; Solak, A.; Dyankova, S.; Nacheva, I.; Dimov, K. Assessment of allergenicity of irradiated dairy products in a Balb/c mice model. *Pharmacia* 2020, 67, 129–133. [CrossRef]
- Banik, S.; Bandyopadhyay, S.; Ganguly, S. Bioeffects of microwave—A brief review. *Bioresour. Technol.* 2003, 87, 155–159.
 [CrossRef]
- Gomaa, A.I.; Nsonzi, F.; Sedman, J.; Ismail, A.A. Enhanced Unfolding of Bovine β-Lactoglobulin Structure Using Microwave Treatment: A Multi-Spectroscopic Study. *Food Biophys.* 2016, 11, 370–379. [CrossRef]
- 103. Gomaa, A.; Sedman, J.; Ismail, A. An investigation of the effect of microwave treatment on the structure and unfolding pathways of β-lactoglobulin using FTIR spectroscopy with the application of two-dimensional correlation spectroscopy (2D-COS). *Vib. Spectrosc.* **2013**, *65*, 101–109. [CrossRef]
- 104. Bourke, P.; Ziuzina, D.; Boehm, D.; Cullen, P.J.; Keener, K. The Potential of Cold Plasma for Safe and Sustainable Food Production. *Trends Biotechnol.* **2018**, *36*, 615–626. [CrossRef] [PubMed]
- 105. Ng, S.W.; Lu, P.; Rulikowska, A.; Boehm, D.; O'Neill, G.; Bourke, P. The effect of atmospheric cold plasma treatment on the antigenic properties of bovine milk casein and whey proteins. *Food Chem.* **2021**, *342*, 128283. [CrossRef] [PubMed]