



High-Quality Draft Single-Cell Genome Sequence Belonging to the Archaeal Candidate Division SA1, Isolated from Nereus Deep in the Red Sea

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ABSTRACT Candidate division SA1 encompasses a phylogenetically coherent archaeal group ubiquitous in deep hypersaline anoxic brines around the globe. Recently, the genome sequences of two cultivated representatives from hypersaline soda lake sediments were published. Here, we present a single-cell genome sequence from Nereus Deep in the Red Sea that represents a putatively novel family within SA1.

Environmental surveys of prokaryotic communities have revealed the existence of several novel uncultivated 16S rRNA gene lineages in deep hypersaline anoxic brines (DHABs [1]). Among these is the archaeal candidate division SA1, which branches basal to Haloarchaea (2). SA1 is believed to harbor clues to the evolutionary history of methanogenic and halophilic archaea (1). Recently, two methyl-reducing halophilic methanogenic strains from hypersaline soda lake sediments were discovered and proposed to be a new euryarchaeal class within SA1 called Methanonatronarchaeia (3), which provided first insights into the role of SA1 in DHABs.

These two SA1 strains diverge by up to 10% at the 16S rRNA gene level but affiliate with one of the two discrete phylogenetic clusters originally identified in the Shaban Deep brine pool in the Red Sea (2, 3). Here, we report a nearly complete single-cell genome sequence (that of SCG-AAA382-B04) of a novel SA1 clade member from Nereus Deep in the Red Sea.

Brine water samples were collected at a depth of 2,445 meters below sea level (mbsl) from Nereus Deep (23°11'53" N, 37°25'09" E) in November 2011. The sample site conditions were as follows: 30.1°C, 22.4% salinity, pH 5.5, and 2.8 μM dissolved oxygen (4). Single cells were sorted and amplified using the REPLI-g kit (Qiagen) at the Single Cell Genomics Center in the Bigelow Laboratory for Ocean Sciences, as described by Ngugi et al. (4).

A paired-end sequence library (2 × 101 bp) of the single-cell amplified genome (SAG) was prepared using the TruSeq DNA library kit and sequenced using an Illumina HiSeq sequencer at the Bioscience Core Laboratory at the King Abdullah University of Science and Technology (KAUST). Thirty-two million reads were quality trimmed using Trimmomatic version 0.32 (5) and assembled into contigs with SPAdes version 3.9.0 (6), applying the error correction and the single-cell mode. Genome completeness was estimated using CheckM (7).

The SAG is composed of 132 contigs totaling 1.42 Mbp (N_{50} , 74.2 kbp; 720× coverage), with a G+C content of 35.5%. The SAG is of high quality, at 94.6% completeness (~4.6% contamination), based on operational standards for SAGs (8), and contains 1,646 protein-coding genes plus 40 RNA-coding genes annotated with PGAP (9) and the Rapid Annotations using Subsystems Technology (RAST) server (10). The SAG contained a 16S rRNA gene sequence with 99% identity to environmental SA1 sequences from DHABs but only 93% identity to Methanonatronarchaeia representa-

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tives (3). Also, all three exhibit a relatively low degree of gene conservation (44 to 52%), syntenic orthologs (30 to 40%), and average amino acid identities (55 to 58%), suggesting that they likely encompass organisms from two families, considering the genomic threshold for this taxonomic rank (11, 12). The SAG has a proteome with a circumneutral isoelectric point and lacks osmolyte biosynthesis pathways, but it encodes potassium uptake systems, suggesting a reliance on potassium for osmoregulation. The methyl coenzyme M reductase complex implicated in methanogenesis (3) is absent, however, signifying the potential for a “nonmethanogenic” methylotrophic lifestyle for other SA1 lineages. We are currently reconstructing the full metabolism of these SA1 subclades.

Accession number(s). The whole-genome shotgun project has been deposited in GenBank under accession number [PZKD00000000](https://ncbi.nlm.nih.gov/nucl/PZKD00000000). The version described here is PZKD01000000.

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REFERENCES

1. Antunes A, Ngugi DK, Stingl U. 2011. Microbiology of the Red Sea (and other) deep-sea anoxic brine lakes. *Environ Microbiol Rep* 3:416–433. <https://doi.org/10.1111/j.1758-2229.2011.00264.x>.
2. Eder W, Schmidt M, Koch M, Garbe-Schönberg D, Huber R. 2002. Prokaryotic phylogenetic diversity and corresponding geochemical data of the brine-seawater interface of the Shaban Deep, Red Sea. *Environ Microbiol* 4:758–763. <https://doi.org/10.1046/j.1462-2920.2002.00351.x>.
3. Sorokin DY, Makarova KS, Abbas B, Ferrer M, Golyshin PN, Galinski EA, Ciordia S, Mena MC, Merkel AY, Wolf YI, van Loosdrecht MCM, Koonin EV. 2017. Discovery of extremely halophilic, methyl-reducing euryarchaea provides insights into the evolutionary origin of methanogenesis. *Nat Microbiol* 2:17081. <https://doi.org/10.1038/nmicrobiol.2017.81>.
4. Ngugi DK, Blom J, Alam I, Rashid M, Ba-Alawi W, Zhang G, Hikmawan T, Guan Y, Antunes A, Siam R, Dorry El H, Bajic V, Stingl U. 2015. Comparative genomics reveals adaptations of a halotolerant thaumarchaeon in the interfaces of brine pools in the Red Sea. *ISME J* 9:396–411. <https://doi.org/10.1038/ismej.2014.137>.
5. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshtkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
7. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
8. Bowers RM, Kyrpides NC, Stepanauskas R, Harmon-Smith M, Doud D, Reddy TBK, Schulz F, Jarett J, Rivers AR, Eloie-Fadrosch EA, Tringe SG, Ivanova NN, Copeland A, Clum A, Becraft ED, Malmstrom RR, Birren B, Podar M, Bork P, Weinstock GM, Garrity GM, Dodsworth JA, Yooshep S, Sutton G, Glöckner FO, Gilbert JA, Nelson WC, Hallam SJ, Jungbluth SP, Ettema TJG, Tighe S, Konstantinidis KT, Liu W-T, Baker BJ, Rattei T, Eisen JA, Hedlund B, McMahon KD, Fierer N, Knight R, Finn R, Cochrane G, Karsch-Mizrachi I, Tyson GW, Rinke C, Genome Standards Consortium, Lapidus A, Meyer F, Yilmaz P, Parks DH, et al. 2017. Minimum information about a single amplified genome (MISAG) and a metagenome-assembled genome (MIMAG) of bacteria and archaea. *Nat Biotechnol* 35:725–731. <https://doi.org/10.1038/nbt.3893>.
9. Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of standard operating procedures (SOPs) for (meta)genomic annotation. *OmicS* 12:137–141. <https://doi.org/10.1089/omi.2008.0017>.
10. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
11. Yarza P, Yilmaz P, Pruesse E, Glöckner FO, Ludwig W, Schleifer K-H, Whitman WB, Euzéby J, Amann R, Rosselló-Móra R. 2014. Uniting the classification of cultured and uncultured bacteria and archaea using 16S rRNA gene sequences. *Nat Rev Microbiol* 12:635–645. <https://doi.org/10.1038/nrmicro3330>.
12. Konstantinidis KT, Tiedje JM. 2007. Prokaryotic taxonomy and phylogeny in the genomic era: advancements and challenges ahead. *Curr Opin Microbiol* 10:504–509. <https://doi.org/10.1016/j.mib.2007.08.006>.