

LETTER TO EDITOR

Effect and application of cryopreserved three-dimensional microcardiac spheroids in myocardial infarction therapy

To the Editor:

Heart failure remains a leading cause of death worldwide. It exerts a severe social burden, particularly in a global ageing population. Despite the clinical significance of heart failure, currently available therapeutic strategies, including heart transplantation and left ventricular assist device, are not realistic owing to several limitations and drawbacks.^{1,2} Therefore, in the past few decades, cell-based cardiac regeneration therapy has emerged as a promising alternative form of intervention.³ Among several potential cell candidates, cardiomyocytes derived from human pluripotent stem cells (hPSC-CMs) have been suggested as the next most promising cell type to treat heart diseases, as they are the most similar to primary human CMs. hPSC-CMs display a distinct cardiac phenotype featuring spontaneous contraction, coupling of cardiac excitation and contraction, and the expressions of cardiac-specific genes, ion channels and structural proteins. However, major issues, including the heterogeneity of hPSC-CMs, may cause tumourigenicity and arrhythmia may affect the long-term storage of hPSC-CMs, and cause poor survival, retention and engraftment of hPSC-CMs in vivo following implantation into diseased hearts. These issues must be resolved before pursuing clinical application.⁴ Here, we aimed to develop a comprehensive protocol in order to overcome the aforementioned obstacles.

First, we identified the cluster of differentiation 71 (CD71; also known as transferrin receptor protein 1, TFR-1) surface protein⁵ as a putative marker to generate a homogeneous population of functional hPSC-CMs. To achieve this, we manually collected contracting green fluorescent protein (GFP)-positive CMs derived from the NKX2.5^{eGFP/w} reporter human embryonic stem cell (hESC) line⁶ (Figure S1A,B, Supporting Live Image S1). They express GFP signal upon microscopically evident expression of NKX2.5 (NK2 homeobox 5), an established marker for cardiac progenitor cells (Figure S1C,D). The hESC cell line has been widely used as a reporter cell line

to discover surface protein markers, including signal regulatory protein alpha (SIRPA).⁷ Subsequently, several contracting GFP⁺ hESC-CMs and hiPSC-CMs were used to perform a single-cell array and finally identify CD71/TFR-1 (Figure S1E-I, Supporting Live Images S2-S4). Studies have verified the functional roles of CD71 in the heart.⁸ Here, a series of characterisation experiments revealed that CD71⁺ hESC and hiPSC-CMs (Figure 1A,B and Figure S1A-C, Supporting Live Images S5-S7) expressed significant amounts of CM-specific markers, including alpha-actinin and cardiac troponin T, and exhibited the ventricular type of action potential ($\geq 65\%$) (Figure 1C and Figure S2C-F). Importantly, CD71⁺ hiPSC-CMs displayed mature phenotypes when cultured for more than 70 days (Figure S2G,H, Supporting Live Images S8 and S9).

Next, to develop an effective strategy for the long-term preservation of hiPSC-CMs for further clinical applications, we examined a form of hiPSC-CM aggregate that we named 'cryopreservable microcardiac spheroids'. Considering that the retention and engraftment of hiPSC-CM aggregates were substantially higher in the hearts in which myocardial infarction (MI) was induced, we assumed that microcardiac spheroids could be an optimal system to induce higher survival following the prolonged storage of hiPSC-CMs and eventual implantation in MI hearts. We observed that the microcardiac spheroids were uniform in size (100–200 μm diameter) and exhibited significantly higher survival rates in 3% hypoxic conditions lasting 7 days compared with single hiPSC-CMs or larger hiPSC-CM aggregates ($>300 \mu\text{m}$ in diameter) (Figure 1D,E and Figure S3A-C). Furthermore, an analysis of post-thaw survival revealed that 80% of the frozen microcardiac spheroids ($<300 \mu\text{m}$ in diameter) remained viable after 6 months of cryopreservation in liquid nitrogen (Figure 1F-H, Supporting Live images S10-S13). We speculate that the substantial expression of connexin 43 protein within microcardiac spheroids (due to the proximal intracellular contact of hiPSC-CMs) to maintain the integrity of the

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. *Clinical and Translational Medicine* published by John Wiley & Sons Australia, Ltd on behalf of Shanghai Institute of Clinical Bioinformatics

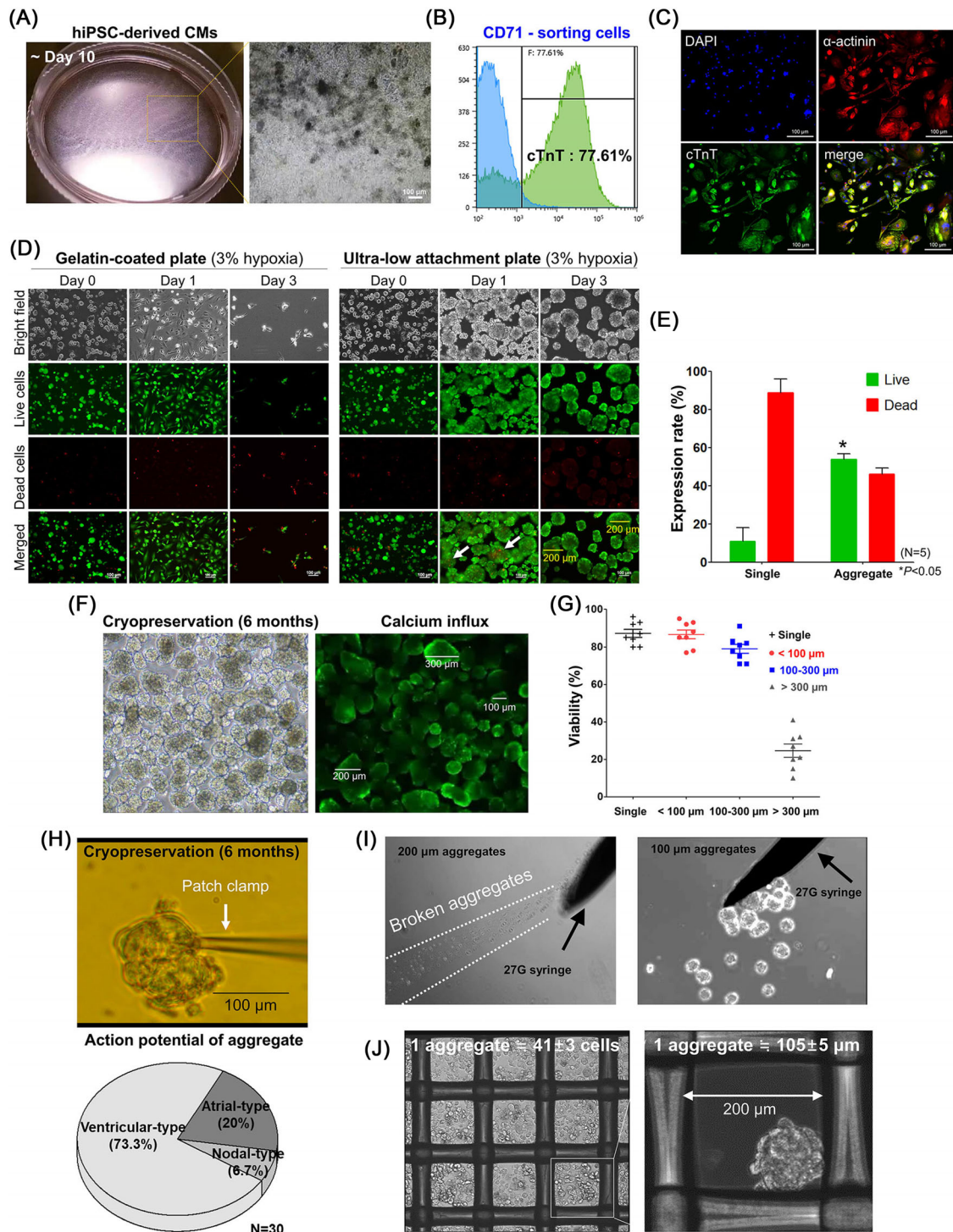


FIGURE 1 Generation of cardiomyocytes derived from human induced pluripotent stem cells. (A) Morphology of hiPSC-CMs. (B) Flow cytometric analysis for the expression of cTnT in CD71⁺ cells. (C) Immunocytochemistry for cTnT and α -actinin on CD71⁺ cells. Scale bars: 100 μ m. (D–H) Hypoxic conditions, and cryopreservation cardiomyocyte aggregates characterisation. (D) Survival analysis of single cardiomyocytes cultured for 3 days in gelatin-coated and ULA dish. White arrow indicates dead aggregates on day 3. Scale bars: 100 μ m. (E) Cell survival rate on day 3. Data are represented as the mean \pm SEM. $n = 5$ biologically independent samples per group. (F) hiPSC-CMs cryopreservation and thawing after 6 months. Staining of different sizes of aggregates by calcium influx staining. (G) After thawing, >300 μ m-sized aggregates had the lowest survival rate (approximately 20%). (H) Electrophysiological analysis of thawed aggregates. (I) Morphology of 100- μ m aggregates was maintained while passage through a 27G syringe (left panel). The 200- μ m aggregates disintegrated while passing through a 27G syringe (right panel). (J) Formation of homogeneous aggregates using spheroid forming dish. 105 ± 5 μ m aggregates were formed by plating 41 ± 3 single cells

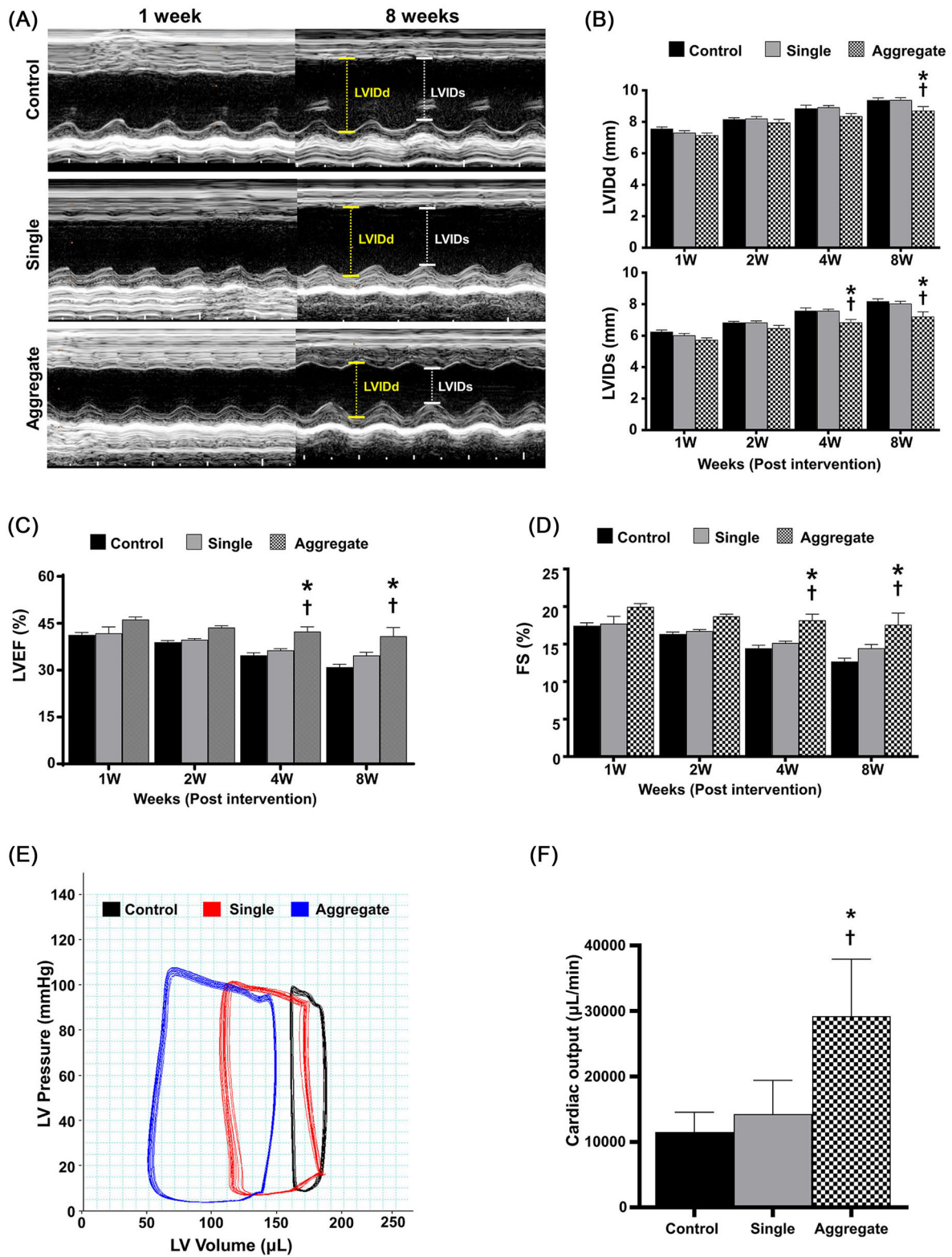


FIGURE 2 Therapeutic effects of human induced pluripotent stem cell (hiPSC)-derived cardiomyocytes (CMs) and aggregated CMs in MI. (A) Representative images (M-mode) of all groups at 1 and 8 weeks after transplantation. (B–D) Echocardiography was used to measure. (B) The left ventricular internal diastolic dimension (LVIDd) and left ventricular internal systolic dimension (LVISd). (C) Left ventricular ejection fraction and (D) fractional shortening. (E) Representative image of the haemodynamic pressure and volume measured using pressure volume (PV) curve at 8 weeks. (F) Haemodynamic cardiac function as measured by the cardiac output (CO). Data are expressed as the mean \pm SEM. * $p < .05$ compared to control group, † $p < .05$ compared to the single CMs treated group; two-way analysis of variance (ANOVA) followed by multiple comparisons with the Tukey method ($n = 6$ –10 per experimental group)

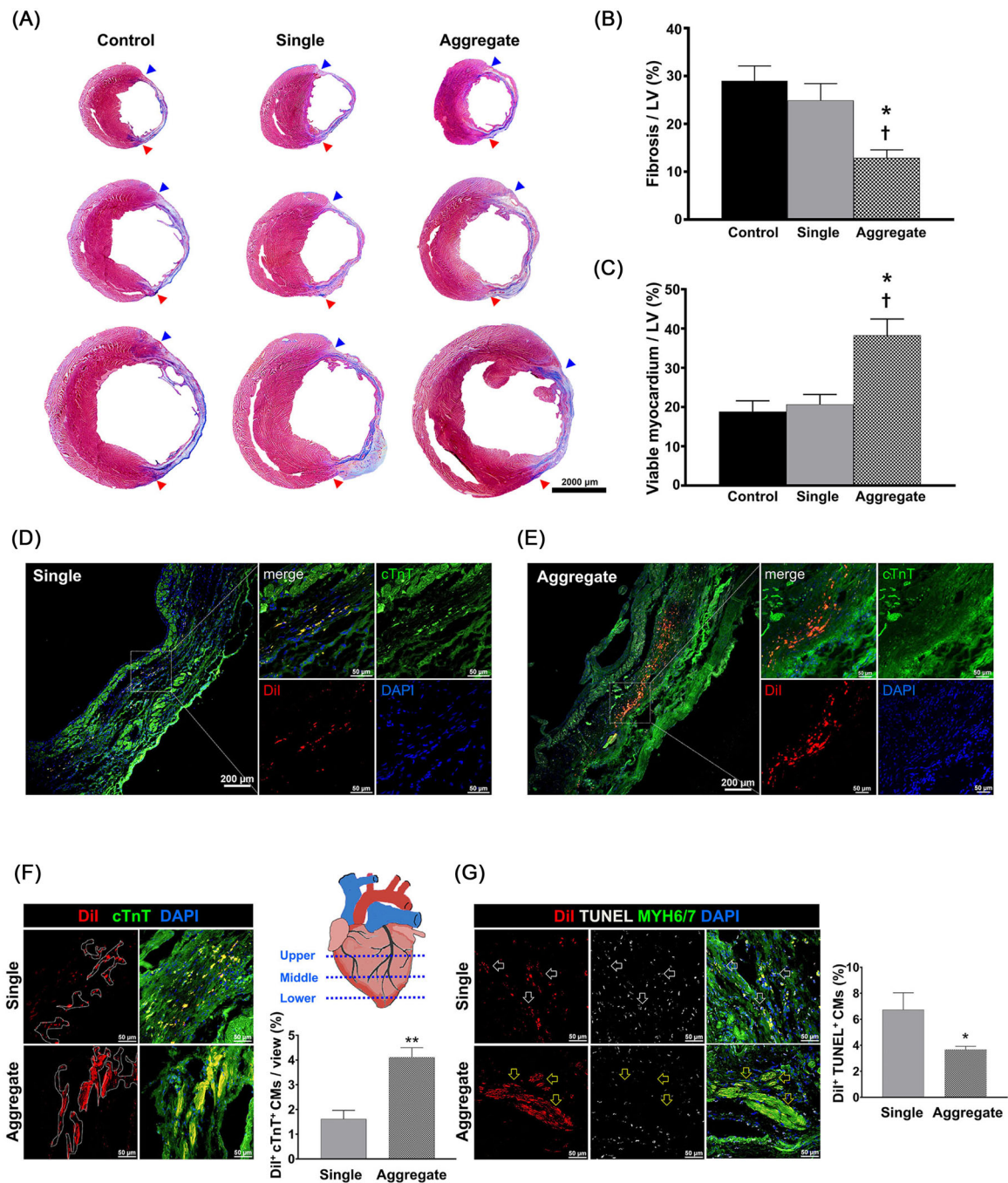


FIGURE 3 Transplantation of aggregates to MI heart improves cardiac function, cell retention and survival rate. (A) Representative images of Masson's trichrome staining from three groups show cardiac fibrosis in the heart tissues collected at 8 weeks after MI. (B) Quantitative result of fibrosis percentage in the left ventricle. (C) Quantification of the area of viable myocardium to the total area of the left ventricle. The area of fibrosis and viable tissues in the left ventricle was calculated from three different levels of each heart. One-way ANOVA followed by Tukey's method was used for statistical analyses. Data are expressed as the mean \pm SEM. * $p < .05$ versus control group; † $p < .05$ versus single group ($n = 3$ per group). (D and E) Representative immunostaining images of the injected DiI-labelled CMs (red) with cTnT (green) in both single cell- and aggregate-transplanted group; scale bar: 200 μm in low-magnification panels and 50 μm in high-magnified panels. (F) Quantification of DiI-labelled hPSC-CMs in the infarcted area. The area positive for both DiI (red) and cTnT (green) per five randomly selected fields from three different levels from the apex to top were quantified. ** $p < .01$ versus single-type group; statistical significance was evaluated using the Student's t -test ($n = 3$ per group). (G) Representative images of TUNEL assay and the quantitative result of TUNEL assay. The single-cell transplanted group showed a few TUNEL-positive DiI-labelled CMs (black arrows), whereas most of the DiI-labelled CMs in the aggregate transplanted group were negative for TUNEL (yellow arrows). The TUNEL-positive DiI⁺ CMs on five randomly selected areas were counted. Scale bar: 50 μm . Student's t -test was used for statistical analyses. Data are expressed as the mean \pm SEM. * $p < .05$, ** $p < .01$ versus single-type group ($n = 3$ per group)

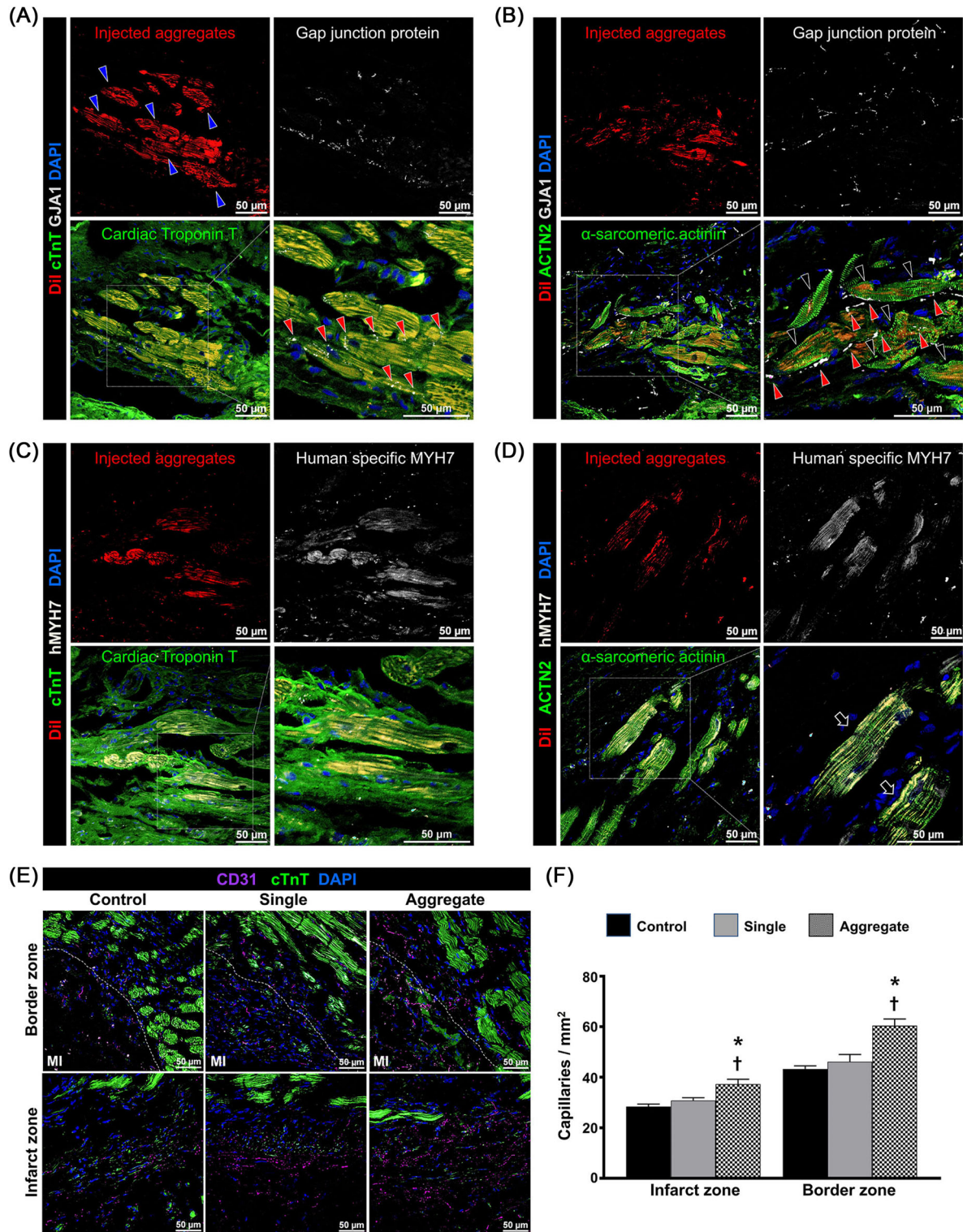


FIGURE 4 Aggregates improve engraftment and functional capacity. (A and B) Immunostaining images with DiI-labelled CMs (red), cardiac-specific markers (cTnT and ACTN2; green) and GJA1 (grey) showed that CA formed a gap junction with the host myocardium. Scale bar: 50 μ m. (C and D) Representative immunostaining images with DiI-labelled CMs (red), cTnT (green), ACTN2 (green) and hMYH7 (grey) verified the identity of injected hPSC-CMs as being human origin. Scale bar: 50 μ m. (E) Representative images of the capillary density. Cardiac tissue was stained with anti-CD31 (purple) antibody to visualise the vessels in both infarct and border area at 8 weeks post cell implantation. Scale bar: 50 μ m. (F) Quantitative result of capillary density. Capillaries on five randomly selected fields from three different levels were counted and the data are expressed as the number of capillaries per square millimetre. * $p < .05$ versus control group; † $p < .05$ versus single group ($n = 3$ per group). Statistical difference was analysed via one-way ANOVA followed by multiple comparisons with Tukey method

spheroids as a whole is the underlying mechanism for the higher survival rate in a hypoxic environment and during the physical stress of freezing.

Finally, the therapeutic efficacy of cryopreserved microcardiac spheroids (Figure 1I,J and Figure S4 Supporting Live Images S14–S17) was analysed in a rat model of MI^{8,9} induced by ligating the left anterior descending artery (Figure S5, number of injected cells: 1×10^6). Echocardiography demonstrated that cardiac function in rats treated using microcardiac spheroids was significantly higher than that in the MI control or in rats injected solely with hiPSC-CM, as determined by ejection fraction and fractional shortening (Figure 2A–D). The results of several parameters for cardiac remodelling,¹⁰ such as left ventricular internal diastolic dimension and posterior wall thickness, suggested that the overall adverse cardiac remodelling in the aggregate group was significantly reduced compared with that in the other experimental groups (Figure S5A–D). Haemodynamic parameters, such as cardiac output and stroke volume, were increased in the aggregate group (Figure 2E,F and Figures S6 and S7). Similarly, Masson's trichrome staining of the cardiac tissue harvested at 8 weeks showed an area of fibrosis (%) that was substantially smaller in the aggregate group than in the other experimental groups (Figure 3A–C). Notably, transplanted microcardiac spheroids exhibited exceptional retention, engraftment and functional stability, all of which contributed to the improvement in heart function. Unexpectedly, histological analysis revealed the maturation of microcardiac spheroids in vivo in relation to the shape, structure and specific protein expression (Figure 3D–G). Maturation was not observed in single-cell transplants, suggesting that intracellular contact and the subsequent cell–cell communication in aggregates are important for survival and drive the maturation of transplanted cells (Figure 4). Evidently, uniformity in size is also essential to induce maturation, which reduces the risk of arrhythmia.

In conclusion, the approach described in this study is simple and effective for the production of readily available hPSC-CMs in clinical settings. We first enriched a pure population of contractile hPSC-CMs based on CD71 expression. The 100- μ m microcardiac spheroids generated using CD71⁺ hiPSC-CMs demonstrated increased survival in both hypoxia and cryopreservation, and enhanced cardiac recovery following transplantation compared to single cells. It is worth noting that the microcardiac spheroids established in this study can be preserved and stored; they exhibit superior viability under hypoxic conditions and in vivo. Thus, they are readily available for direct implantation into patients in emergencies. Despite several meaningful results, future studies examining more sophisticated implantation methods, such as the 27G NOGA catheter

system⁹ and applicability to large animals, considered limitations of this study, will warrant more effective cell-based cardiac regenerative therapy.

ACKNOWLEDGEMENTS

We thank the Murdoch Children's Research Institute (MCRI) for providing the NKX2.5 eGFP/w hESC line. The human stem cell line CMC-011-iPSC was provided by the National Stem Cell Bank of Korea (Korea National Institute of Health). This study was supported by Ministry of Health & Welfare, Republic of Korea HI20C0184; Korea Fund for Regenerative Medicine (KARM), Republic of Korea 21A0403L1 & 21A0104L1-11; National Research Foundation of Korea (NRF) 2021R1A4A3031875; The CityU Applied Research Grant 9667200 and Hong Kong Research Grants Council 21100818.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

Soon-Jung Park^{1,2}
 Hyeok Kim^{3,4}
 Sunghun Lee⁵
 Jongsoo Kim^{1,6}
 Taek-Hee Jung^{1,2}
 Seong Woo Choi⁷
 Bong-Woo Park^{3,4}
 Sun-Woong Kang⁸
 David A. Elliott⁹
 Edouard G. Stanley⁹
 Andrew G. Elefanty⁹
 Kiwon Ban⁵
 Hun-Jun Park^{3,4,10}
 Sung-Hwan Moon^{1,2} 

¹ Department of Medicine, Konkuk University School of Medicine, Seoul, Korea

² Research Institute, T&R Biofab Co. Ltd, Siheung, Korea

³ Department of Medical Life Science, College of Medicine, The Catholic University of Korea, Seoul, Korea

⁴ Division of Cardiology, Department of Internal Medicine, Seoul St. Mary's Hospital, The Catholic University of Korea, Seoul, Korea

⁵ Department of Biomedical Sciences, City University of Hong Kong, Kowloon, Hong Kong SAR

⁶ Department of Surgery, Wexner Medical Center, Ohio State University, Columbus, Ohio, USA

⁷ Department of Physiology, Department of Biomedical Sciences, College of Medicine, Seoul National University, Seoul, Korea

⁸ Research Group for Biomimetic Advanced Technology, Korea Institute of Toxicology, Daejeon, Korea

⁹ Monash Immunology and Stem Cell Laboratories,
Monash University, Clayton, Victoria, Australia

¹⁰ Cell Death Disease Research Center, College of
Medicine The Catholic University of Korea, Seoul, Korea

Correspondence

Kiwon Ban, Department of Biomedical Sciences, City
University of Hong Kong, 83 Tat Chee Ave, Kowloon,
Hong Kong SAR.

Email: kiwonban@cityu.edu.hk

Hun-Jun Park, Seoul St. Mary's Hospital, The Catholic
University of Korea, 222 Banpo-daero, Seocho-gu, Seoul,
Korea.

Email: cardioman@catholic.ac.kr

Sung-Hwan Moon, Research Institute, T&R Biofab Co.
Ltd, Siheung, Korea.

Email: safe33msh@gmail.com

Soon-Jung Park, Hyeok Kim and Sunghun Lee
contributed equally to this work.

ORCID

Sung-Hwan Moon  <https://orcid.org/0000-0002-6642-5699>

REFERENCES

1. Kretzschma K, Post Y, Bannier-Hélaouët M, et al. Profiling proliferative cells and their progeny in damaged murine hearts. *Proc Natl Acad Sci*. 2018;115(52):E12245-E12254.
2. Kamihata H, Matsubara H, Nishiue T, et al. Implantation of bone marrow mononuclear cells into ischemic myocardium

enhances collateral perfusion and regional function via side supply of angioblasts, angiogenic ligands, and cytokines. *Circulation*. 2001;104:1046-1052.

3. Uemura R, Xu M, Ahmad N, Ashraf M. Bone marrow stem cells prevent left ventricular remodeling of ischemic heart through paracrine signaling. *Circ Res*. 2006;98:1414-1421.
4. Nguyen PK, Neofytou E, Rhee JW, Wu JC. Potential strategies to address the major clinical barriers facing stem cell regenerative therapy for cardiovascular disease: a review. *JAMA Cardiol*. 2016;1:953-962.
5. Xu W, Barrientos T, Mao L, Rockman HA, Sauve AA, Andrews NC. Lethal cardiomyopathy in mice lacking transferrin receptor in the heart. *Cell Rep*. 2015;13:533-545.
6. Elliott DA, Braam SR, Koutsis K, et al. NKX2-5(eGFP/w) hESCs for isolation of human cardiac progenitors and cardiomyocytes. *Nat Methods*. 2011;8:1037-1040.
7. Dubois NC, Craft AM, Sharma P, et al. SIRPA is a specific cell-surface marker for isolating cardiomyocytes derived from human pluripotent stem cells. *Nat Biotechnol*. 2011;29:1011-1018.
8. Moon SH, Kang SW, Park SJ, et al. The use of aggregates of purified cardiomyocytes derived from human ESCs for functional engraftment after myocardial infarction. *Biomaterials*. 2013;34:4013-4026.
9. Bolli R, Hare JM, March KL, et al. Rationale and design of the CONCERT-HF trial (combination of mesenchymal and c-kit(+) cardiac stem cells as regenerative therapy for heart failure). *Circ Res*. 2018;122:1703-1715.
10. Park SJ, Kim RY, Park BW, et al. Dual stem cell therapy synergistically improves cardiac function and vascular regeneration following myocardial infarction. *Nat Commun*. 2019;10:3123.

SUPPORTING INFORMATION

Additional supporting information may be found in the online version of the article at the publisher's website.