

Spacing out dual checkpoint inhibition improves antigen-specific T cell manufacture

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The adoptive transfer of antigen-specific T cells is a promising therapy for treating infection, cancer, and autoimmunity. To generate large numbers of antigen-specific T cells, however, most protocols rely on *in vitro* culture with cytokines and repeated antigenic stimulation. This leads to terminal T cell effector differentiation and exhaustion, limiting the utility of expanded T cells.^{1–3} In this issue of *Molecular Therapy – Methods and Clinical Development*, Lak et al. demonstrate that blocking both programmed death-ligand 1 (PD-L1) and T cell immunoglobulin and mucin-containing protein 3 (TIM-3) during the generation of antigen-specific CD8⁺ T cells led to improved T cell expansion without inducing T cell dysfunction, but only when blockade was induced with specific timing.⁴ This “delayed double blockade,” or DDB, did not seem to induce changes in gene expression or T cell receptor (TCR) repertoire in expanded T cells. These findings suggest a translatable strategy for improving the manufacturing of adoptively transferred antigen-specific T cells.

The efficacy of adoptively transferred T cells is often limited by the dysfunction induced by continuous T cell stimulation and stimulatory cytokine treatment during the expansion process.^{1–3} Therefore, determining conditions that minimize T cell dysfunction is crucial for the therapeutic potency of adoptive cell immunotherapy. T cell exhaustion is accompanied by the upregulation of cell surface inhibitory receptors known as immune checkpoint molecules. Blocking inhibitory receptor signaling using non-signaling antibodies, also known as immune checkpoint blockade, enables T cells to overcome these inhibitory mechanisms to mitigate dysfunction.⁵ Previously, it had been demonstrated that combined *in vivo* blockade of PD-1 and TIM-3,

both of which are upregulated in exhausted CD8⁺ T cells, led to greater reduction in tumor burden than single blockade alone.⁶ In the current work, the authors added anti-PD-L1 on day 1 of *ex vivo* expansion, followed by anti-TIM-3 on day 7. They demonstrated that DDB improved CD8⁺ T cell expansion and antigen-specific reactivity for both viral and tumor antigens, whereas single blockade of either PD-L1 or TIM-3 alone failed to achieve the same results. Interestingly, compared with double blockade at day 0, DDB showed improved T cell expansion and function. Intriguingly, DDB did not appear to change T effector differentiation or exhaustion marker expression, although DDB-expanded T cells were more proliferative and had improved cytotoxic activity compared with controls. The authors were also unable to identify differences in T cell transcription or in TCR clonotype evolution due to DDB, perhaps as a consequence of high interdonor variability. Moreover, the benefit conferred by DDB was not universal; for instance, neither short-term virus-specific T cell generation nor chimeric antigen receptor (CAR) T cell expansion benefitted from DDB. The authors hypothesize that the beneficial effects of DDB may be most pronounced in long-term culture settings with specific patterns of antigen stimulation; one additional notable difference may be the inclusion of monocyte-derived dendritic cells (DCs) in T cell expansion protocols benefitting from DDB. To explain the context-dependent benefit of DDB, more work is clearly needed to investigate the mechanistic effects of DDB across donors and clonotypes. Nonetheless, this work suggests that dual blockade of PD-L1 and TIM-3 may be useful for improving the *ex vivo* long-term expansion of some types of antigen-specific CD8⁺ T cells without inducing T cell dysfunction, identifying a

potentially translatable improvement in the manufacture of antigen-specific T cell therapies.

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AUTHOR CONTRIBUTIONS

Conception, H.W. and L.D.W.; writing, H.W.; editing, H.W. and L.D.W.

DECLARATION OF INTERESTS

The authors have no conflicts of interest to disclose relevant to the information presented in this Commentary.

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