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Supplemental information

Single-cell chromatin accessibility landscape

identifies tissue repair program

in human regulatory T cells

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Figure S1. Single-cell ATAC landscape identifies repair signature in murine tissues and spleen. This is an extension of **Figure 1**. (**A**) Sort gates and post-sort QC for CD4⁺ T cells from murine spleen, colon, VAT, lung and skin for scATAC-Seq. (**B**) Sort gates and post-sort QC for CD25⁺ T cells from murine spleen for scATAC-Seq. (**C**) Total number of QC-passed and processed cells ("Barcode #") per tissue and cell type as well as mean # of fragments per cell. (**D**) UMAP of tissue T cells as in **Figure 1A** (left) with sort gate (CD4⁺ or CD25⁺, middle) and peak count (right). (**E**) Sample contribution to UMAP as in **Figure 1A**, with Colon CD4⁺, VAT CD4⁺, Lung CD4⁺, Skin CD4⁺, Spleen CD4⁺ and Spleen CD25⁺. (**F**) UMAP of scATAC-seq data with chromatin accessibility of the *Klrg1*, *Pparg*, *Areg*, and *ll10* gene loci, with blue=low and red=high accessibility.



Figure S2. Repair signature in scATAC landscape of T cells from germ-free animals. This is an extension of Figure 2. (A) Sort gates and post-sort QC for CD4⁺ T cells from murine spleen, colon, VAT and skin for scATAC-Seq. (B) Sort gates and post-sort QC for CD25⁺ T cells from murine spleen for scATAC-Seq. (C) Total number of QC-passed and processed cells ("Barcode #") per tissue and cell type as well as mean # of fragments per cell. (D) Sample contribution to UMAP as in Figure 2A, with Colon CD4⁺, VAT CD4⁺, CD4⁺, Skin CD4⁺, Spleen CD4⁺, Spleen CD25⁺ and peak count. (E) UMAP of scATACseq data where a bulk ATAC signature of VAT tisTregST2 an early tisTregST2 precursor signature were overlaid, with blue=low and yellow=high overlap. The contribution of each signature to the clusters is shown in a pie chart. (F) UMAP of scATAC-seq data with chromatin accessibility of the KIrg1, Pparg, Areg, and II10 gene loci, with blue=low and red=high accessibility. (G) Flow cytometry of VAT, skin, colon and spleen T cells from SPF vs gnotobiotic mice. Top, gating strategy. Bottom, Klrg1 expression in SPF vs gnotobiotic mice. (H) Intracellular expression of Areg and IL-10 in Klrg1+ Treg cells from skin of SPF vs gnotobiotic mice (n=5). (I) UMAP of scATAC-data with colon T where a bulk ATAC signature of colon tisTregST2 was overlaid, with blue=low and yellow=high overlap. The outlined fraction shows the localization of tisTregST2.



Figure S3. Single-cell ATAC and gene expression landscape of human blood, fat and skin CD4⁺ T cells. This is an extension of **Figure 3**. (**A**) Sort gates and post-sort QC for CD25-enriched T cells from human peripheral blood for scATAC-Seq. (**B**) Sort gates and post-sort QC of CD25-enriched T cells from human peripheral blood for scATAC-Seq of CD127⁻CD25⁺ Treg cells. (**C**) Sort gates and post-sort QC for CD4⁺ and CD127⁻CD25⁺ T cells from human fat tissue, including a Foxp3 staining control to verify Treg identity. (**D**) Sort gates and post-sort QC for CD4⁺ T cells from human skin tissue. (**E**) Total number of QC-passed and processed cells ("Barcode #") per tissue and cell type as well as mean # of fragments per cell. (**F**) Sort gates for CD127⁻CD25⁺ T cells from human skin and fat tissue for bulk RNA-seq. (**G**) Tissue contribution to UMAP as in **Figure 3A**, with fat, blood and skin, and peak count. (**H**) UMAP of scATAC-seq data with chromatin accessibility of the *ENTPD1*, *CTLA4*, *TIGIT*, *BHLHE40*, *KLRG1* and *ZC3H12C* gene loci. (**I**) Sort gates and post-sort QC of CD4-enriched T cells from human peripheral blood for bulk RNA-seq.



Figure S4. Species-conserved tissue Treg peakset identifies CCR8⁺ Treg population in human blood. This is an extension of Figure 4. (A-D) Left, chromatin accessibility of BACH2, CCR2, NR 125406 (CCR5) and TNFRSF9 in human Treg clusters 01, 03 and 07 with BATF ChIP-Seg data (GSM803538). Right, chromatin accessibility of Bach2, Ccr2, Ccr5 and Tnfrsf9 in murine Treg clusters 00, 11 and 16 combined with publicly available Batf ChIP-Seg data (GSE54191). Cluster datasets group-normalized to maximum peak height. (E) CCR8⁺ Treg cells (CD4⁺CD127⁻CD25⁺CD45RA⁻CCR8⁺, red), CCR8- Treg cells (CD4⁺CD127⁻CD25⁺CD45RA⁻CCR8⁻, blue), naive Treg cells (CD4⁺CD127⁻CD25⁺CD45RA⁺, green) and Tconv cells (CD4⁺CD127⁺CD25⁻, blue) from human blood have been sorted followed by fixation, permeabilization and BATF/FOXP3 staining. Cells have been re-acquired and color code indicates cell type. (F) Gating strategy to identify Treg cells in *Nfil3*(GFP), *Areg*(GFP) reporter mice or wildtype animals. (G) CCR8 and GFP expression in wildtype animals. (H) Gating strategy to identify Treg cells in Areg(GFP) x Foxp3(huCD2) animals and percentage of Areg(GFP)⁺CCR8⁺ cells of Treg cells (left, n=5, 2x repeated). (I) Percentage of Nfil3(GFP)⁺CCR8⁺ cells of Treg cells (right, n=4, 2x repeated).



Cluster 9 (287 incl lsotype ctrl and blanks)

Figure S5. Surface protein, transcriptional and epigenetic analysis of human CCR8⁺ **Treg cells in blood.** This is an extension of **Figure 5.** (**A**) Five populations (CD3⁺CD4⁺CD127⁺CD25⁻CD45RO⁻ antigen-naive Tconv, CD3⁺CD4⁺CD127⁺CD25⁺CD45RO⁺ memory Tconv, CD3⁺CD4⁺CD127⁻CD25⁺CD45RO⁻ antigen-naive Treg, CD3⁺CD4⁺CD127⁻CD25⁺CD45RO⁺CCR8⁺ and CD3⁺CD4⁺CD127⁻CD25⁺CD45RO⁺CCR8⁺ antigen-experienced Treg were gated and the expression of PE was analyzed. Data were subjected to "pheatmap" with 9 rows, yielding 9 groups. Isotype control staining for 5 groups shown below. (**B**) Group 8 and Group 9, as in **Figure 5**, with protein labels. (**C**) Chromatin accessibility of human *HLADRA*, *HLADRB3* and *HLADRB6*. Data are derived from antigen-naive Treg cells (Cluster 07), memory Treg cells (01) and fat and skin Treg cells (03). BATF ChIP-Seq signal below (GSM803538). (**D**) Gating scheme to identify Treg cells in human fat and skin tissue for CCR8 and HLA-DR measurement. (**E**) UMAP of scATAC-seq data with chromatin accessibility of the *FOXP3* and *KLRB1* gene. Expression of CD161 on skin and fat Treg cells displayed to the right.



Figure S6. Single-cell RNA and TCR landscape of donor-matched human blood, fat and skin CD4⁺ T cells. This is an extension of Figure 6. (A) Sort gates and post-sort QC for CD25 pre-enriched blood Treg (CD4⁺CD127⁻CD25⁺), blood memory Treg (CD4⁺CD127⁻CD25⁺CD45RA⁻CD45RO⁺), and blood memory CCR8 Treg (CD4⁺CD127⁻ CD25⁺CD45RA⁻CD45RO⁺CCR8⁺) from human peripheral blood for scRNA/TCR-Seq. (B) Sort gates and post-sort QC for CD4⁺ T cells from human fat tissue. (C) Sort gates and post-sort QC for CD4⁺ T cells from human skin tissue. (D) Total number of QC-passed and processed cells ("Barcode #") per tissue and cell type (data based on CellRanger output) as well as total # of linked alpha and beta TCRs per cell (data after additional internal QC filtering steps). (E) UMAP of scRNA-seg data derived from FACS-sorted CD4⁺ T cell populations of human peripheral blood, skin and fat of one individual donor (Donor 7). Top, cells color-coded based on sort strategy. Bottom, cells clustered and color-coded according to cluster. (F) Gene expression of FOXP3, CCR8 and HLA-DRB1 plotted on UMAP. Color code indicates expression strength. Bottom right, TCRs derived from all skin Treg cells (blue) and all fat Treg cells (yellow) were extracted and highlighted. (G) Tracking of skin Treg TCR clones in different sorted populations of peripheral blood of the same donor. Percentage indicates fraction of detected clones, with total number of clones shown below. (H) Clonality of all cells in samples donor 6 and donor 7. All TCR sequences listed in Table S4.



Figure S7. Treg cells in wound healing and cancer. This is an extension of Figure 7. (A) Flow cytometry of Treg cells (CD4⁺CD25⁺CD127⁻) from human blood, fat and skin tissue with CD279 (PD-1) expression (n=4-5, one-way ANOVA with Tukey post-test). (B) Naive Treg cells treated for six days with IL-2 or IL-2, IL-12, IL-21, IL-23 and TGF- β invitro, followed by washing, overnight re-stimulation and flow cytometry for BATF, BCL-6, FOXP3 and cell count analysis. Expression analysis to the right (n=4). (C) Suppression Assay for *in-vitro* generated IL-2 or Tfh-like Tregs. Left: Division index of CD8 T cells cocultured with Tfh-like (red) or IL-2 (blue) Tregs. Right: Flow Cytometry data of CFSE stained CD8 T cells cocultured with Tfh-like (red) or IL-2 (blue) Tregs, indicating division. (D) Peaks from tissue Treg, Tfh-like Treg and tumor Tfh signatures annotated with their closest gene. Overlaps between unique genes in these signatures counted. (E) Tfh-like Treg cells or IL-2 treated Treg cells of several donors stimulated overnight and supernatant collected and used in a wound healing assay with HaCaT cells (n=3, 1+3 dilution, 1+7 dilution and 1+15 dilution of supernatant). (F) Tfh-like Treg cells or IL-2 treated Treg in a wound healing assay with HaCaT cells (1+7 dilution, individual donors shown). Bottom right, different dilutions of cytokine medium (Tfh mix) in wound healing assay. (G) Histological staining of human epidermal reconstruction model treated with Tfh-like Treg supernatant or IL-2 Treg supernatant. Top, unprocessed image. Bottom, processed image with Stratum corneum shown in green. Box in center = magnified area of interest. (H) UMAP of scATAC-seq data derived from CD4⁺ T cell populations of a murine HER2-transgenic breast carcinoma model. Far left plot indicates cell type by sort gate, left plot shows murine core tisTregST2 signature. Right; chromatin accessibility of the *lkzf2* and *Ccr8* gene locus is shown. (I) Gating scheme for CCR8 vs CD45RA expression of Treg cells in skin, fat, NAT and liver tumor tissue from Figure 7I. (J)

Expression of BCL6 and BATF in all samples from **Figure 7J**. (**K**) UMAP of scATAC-seq data derived from FACS-sorted CD4⁺ T cell populations of human peripheral blood, skin and fat of individual female donors as in **Figure 3**. A tumor Treg signature (940 peaks) was overlaid and clusters 01 (memory Treg), 03 (fat and skin tissue Treg) and 07 (naive Treg) labeled.