



Whole-Genome Sequencing and Annotation of *Fibrobacter succinogenes* HC4, Isolated from the Horse Cecum

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ABSTRACT Fibrobacter succinogenes is a major cellulolytic bacterial species living in the large intestines of herbivores. This study reports the genome sequencing, assembly, and annotation of *F. succinogenes* HC4 (DSM 33656), a strain isolated from horse cecal contents. The genome comprised a total of 3.74 Mbp, with a G+C content of 48.96%.

F ibrobacter succinogenes plays a key role in host nutrition and health by degrading cellulose into short-chain fatty acids (SCFAs) (1–5) in the digestive ecosystem. To date, cultured representatives of *F. succinogenes* living in the large intestines of mammals are lacking.

To isolate cellulolytic strains of F. succinogenes, samples were collected from one cecumcannulated horse (6). Contents were stored at 38°C in CO₂-saturated containers. Decimal dilutions were inoculated in a specific medium (7, 8). After 7 days of incubation at 38°C under CO_{2r} the roll tube method (9) and the enrichment method (10) were used alternately to isolate strain HC4. The 16S rRNA gene was amplified by PCR using the universal primers 27f and 1492r. DNA amplification was achieved using the following program: denaturation for 4 min at 95°C; 35 cycles consisting of 1 min at 94°C, 1 min at 55°C, and 2 min at 72°C; and then 15 min at 72°C and 3 min at 30°C (11). PCR products were sequenced by Genewiz using the Sanger method. The 16S rRNA gene of strain HC4 (accession number OP018198) exhibited 99% sequence identity to those of F. succinogenes group V equine strains (10). For genome sequencing, DNA was extracted from a 48-h culture of F. succinogenes HC4 at 38°C under CO_2 in Lowe medium (12) containing cellulose filter paper (13). The culture sample was concentrated and subjected to physical lysis using a bead beater (3 min at 25 Hz) and chemical lysis using a lysis buffer. After centrifugation for 5 min at $16,000 \times q$ and treatment with 10 M ammonium acetate and 70% (v/v) ethanol, DNA was purified using the QIAamp stool minikit from Qiagen (catalog number 51504). The F. succinogenes genome was sequenced, assembled, and annotated by Genewiz. Libraries were constructed using the NEBNext Ultra II DNA library preparation kit for Illumina and sequenced using an Illumina NovaSeg sequencing platform (2 \times 150 bp). A total of 13,163,761 raw reads were obtained and filtered with bcl2fastq v2.20.0 (14). Sequences of <30 bp were discarded. De novo assembly, with 50 scaffolds with an N_{so} value of 149,151 bp, was carried out using SPAdes v3.10.0 (k-mer values of 21, 33, 55, and 77) (15). The de novo assembled genome was created with a minimum contig length of 1,000 bp and analyzed statistically using QUAST v4.6 (16). Gene prediction and annotation were performed using Prokka v1.12 (17) with ISfinder, the NCBI Bacterial Antimicrobial Resistance Reference Gene Database, and the UniProtKB database. Default parameters were used for all software unless otherwise specified. The F. succinogenes HC4 genome comprised a total of 3.74 Mbp, with 3,143 coding sequences. The G+C content of the genome was 48.96%.

The HC4 strain of *F. succinogenes* will allow better characterization of the physiology and ecology of cellulolytic bacteria inhabiting the large intestines of mammals.

Data availability. This whole-genome shotgun project has been deposited in DDBJ/ ENA/GenBank under the accession number JALCZV00000000 (BioProject accession number PRJNA812370), and the raw reads have been deposited in the NCBI Sequence Read Archive **Editor** David Rasko, University of Maryland School of Medicine

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REFERENCES

- Froidurot A, Julliand V. 2022. Cellulolytic bacteria in the large intestine of mammals. Gut Microbes 14:2031694. https://doi.org/10.1080/19490976.2022 .2031694.
- Flint HJ, Scott KP, Duncan SH, Louis P, Forano E. 2012. Microbial degradation of complex carbohydrates in the gut. Gut Microbes 3:289–306. https://doi.org/ 10.4161/qmic.19897.
- Blaak EE, Canfora EE, Theis S, Frost G, Groen AK, Mithieux G, Nauta A, Scott K, Stahl B, van Harsselaar J, van Tol R, Vaughan EE, Verbeke K. 2020. Short chain fatty acids in human gut and metabolic health. Benef Microbes 11:411–455. https://doi.org/10.3920/BM2020.0057.
- Dalile B, van Oudenhove L, Vervliet B, Verbeke K. 2019. The role of short-chain fatty acids in microbiota–gut–brain communication. Nat Rev Gastroenterol Hepatol 16:461–478. https://doi.org/10.1038/s41575-019-0157-3.
- von Engelhardt W, Bartels J, Kirschberger S, zu Düttingdorf HDM, Busche R. 1998. Role of short-chain fatty acids in the hind gut. Vet Q 20:52–59. https://doi.org/10.1080/01652176.1998.9694970.
- Grimm P, Combes S, Pascal G, Cauquil L, Julliand V. 2020. Dietary composition and yeast/microalgae combination supplementation modulate the microbial ecosystem in the caecum, colon and faeces of horses. Br J Nutr 123:372–382. https://doi.org/10.1017/S0007114519002824.
- Halliwell G, Bryant MP. 1963. The cellulolytic activity of pure strains of bacteria from the rumen of cattle. J Gen Microbiol 32:441–448. https://doi .org/10.1099/00221287-32-3-441.
- Julliand V, de Vaux A, Millet L, Fonty G. 1999. Identification of *Ruminococcus flavefaciens* as the predominant cellulolytic bacterial species of the equine cecum. Appl Environ Microbiol 65:3738–3741. https://doi.org/10.1128/AEM .65.8.3738-3741.1999.
- 9. Hungate RE. 1969. Chapter IV: a roll tube method for cultivation of strict

anaerobes. Methods Microbiol 3:117-132. https://doi.org/10.1016/S0580 -9517(08)70503-8.

- Neumann AP, McCormick CA, Suen G. 2017. *Fibrobacter* communities in the gastrointestinal tracts of diverse hindgut-fermenting herbivores are distinct from those of the rumen. Environ Microbiol 19:3768–3783. https://doi .org/10.1111/1462-2920.13878.
- 11. Rousseaux S, Hartmann A, Soulas G. 2001. Isolation and characterisation of new Gram-negative and Gram-positive atrazine degrading bacteria from different French soils. FEMS Microbiol Ecol 36:211–222. https://doi .org/10.1111/j.1574-6941.2001.tb00842.x.
- Lowe SE, Theodorou MK, Trinci APJ, Hespell RB. 1985. Growth of anaerobic rumen fungi on defined and semi-defined media lacking rumen fluid. Microbiology (N Y) 131:2225–2229. https://doi.org/10.1099/00221287-131-9-2225.
- Yu Z, Morrison M. 2004. Improved extraction of PCR-quality community DNA from digesta and fecal samples. Biotechniques 36:808–812. https:// doi.org/10.2144/04365ST04.
- Brown J, Pirrung M, Mccue LA. 2017. FQC Dashboard: integrates FastQC results into a web-based, interactive, and extensible FASTQ quality control tool. Bioinformatics 33:3137–3139. https://doi.org/10.1093/bioinformatics/btx373.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin A, Sirotkin A, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https:// doi.org/10.1089/cmb.2012.0021.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https:// doi.org/10.1093/bioinformatics/btt086.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.