

Confocal Laser Endomicroscopy in a Critically III COVID-19 Patient

Olesya Danilevskaya, M.D., Ph.D.^{1,2}, Dmitry Sazonov, M.D., Ph.D.¹, Tatyana Klypa, M.D., Ph.D.³, Fedor Zabozlaev, M.D., Ph.D.⁴ and Elena Popova, M.D.²

¹Department of Endoscopy, FSBI Federal Research and Clinical Center for Specialized Types of Medical Care and Medical Technologies of the Federal Medical Biological Agency of Russia, Moscow, ²Laboratory of Clinical Pulmonology, FSBI Pulmonology Scientific Research Institute under Federal Medical Biological Agency of Russia, Moscow, ³Intensive Care Unit, FSBI Federal Research and Clinical Center for Specialized Types of Medical Care and Medical Technologies of the Federal Medical Biological Agency of Russia, Moscow, ⁴Department of Pathology, FSBI Federal Research and Clinical Center for Specialized Types of Medical Care and Medical Technologies of the Federal Medical Biological Agency of Russia, Moscow, ⁸Pulssia

Probe-based confocal laser endomicroscopy (pCLE) is a minimally invasive technique for *in vivo* real-time microscopic imaging of proximal and distal airways during bronchoscopy. Using the laser beam with a wavelength of 488 nm, it allows detection of autofluorescent alveolar, vascular, cellular, and noncellular microstructures with an optical area of 600 μm and a focus depth of 50 μm^1 . The role of pCLE has not been clearly established in patients with coronavirus disease 2019 (COVID-19), especially correlating with high-resolution computed tomography data, bronchoalveolar lavage, and traditional histopathology.

The patient gave written informed consent for pCLE on the day of admission to the clinic. The study protocol was approved by local ethics committee of clinical center.

An age 70 male who never smoked was admitted COVID-19 positive on the 3rd day of illness. The patient's respiratory failure progressed rapidly. pCLE and high-resolution computed tomography were performed the next day after intubation. pCLE has been done in four right and three left lung segments (Figure 1). In alveolar spaces, pCLE showed large highly fluorescent floating structures $50\!\times\!240$ μm . Alveolar structure exhibited increased elastin fiber thickening, dystelectatic abnormalities. Disorganized microvessel growth was also observed.

The patient died 15 days after admission due to multiple organ failure.

On autopsy, protein agglomerates with increased expression of surfactant-associated protein A, macrophages, and desquamated alveolocytes were found (Figure 2A) in the corresponding zones with floating structures detected by pCLE (Figure 1B). In areas with partial loss of alveolar structure and filling of the alveoli lumen with moderately fluorescent elements (Figure 1A), histological examination revealed the growth of granulation tissue (Figure 2B).

Large floating complexes in the alveoli lumen were very similar to pCLE imaging in pulmonary alveolar proteinosis². However, in the context of acute respiratory distress syndrome such structures might be polymorphonuclear neutrophils and macrophages. But the fluorescence intensity of them in our study is visually much higher than presented in the images of Lesur et al.³, and these cells in our images look like they contain protein-lipid components inside.

In COVID-19 lung tissue, we found a combination of focal alveolar collapse with diffuse hyperplasia of alveolar epithelial type II cells, which express the angiotensin converting enzyme-2 receptor⁴. Apparently, severe acute respiratory syndrome coronavirus 2 primarily damaging pneumocytes, can lead to degradation of the sur-

https://doi.org/10.4046/trd.2022.0023 ISSN: 1738-3536(Print)/ 2005-6184(Online) Tuberc Respir Dis 2022:85:273-275



Copyright © 2022 The Korean Academy of Tuberculosis and Respiratory Diseases

Address for correspondence Olesya Danilevskaya, M.D., Ph.D.

Laboratory of Clinical
Pulmonology, FSBI Pulmonology
Scientific Research Institute
under Federal Medical Biological
Agency of Russia, Orekhovy blvd.,
28, Moscow 115682, Russia
Phone 8-10-7-495-395-05-11
Fax 8-10-7-495-395-63-93
E-mail danless@mail.ru
Received Mar. 1, 2022
Revised Mar. 18, 2022
Accepted Mar. 22, 2022
Published online Mar. 23, 2022



⊗ It is identical to the Creative Commons Attribution Non-Commercial License (http:// creativecommons.org/licenses/ by-nc/4.0/).

Figure 1. Endomicroscopic images of distal airways in active coronavirus disease 2019 patient (on the 9th day of hospital admission) show the loss of organization, increased connective tissue fibers and structures, which may correspond to areas of granulation tissue on histological image (B), only single alveoli (green arrows) are visualized (A). Highly fluorescent large floating structures (green arrowheads), alveolar macrophages (yellow arrowheads) (B) are present in the alveolar compartment. Signs of dystelectasis with an increase in the number of connective tissue structures in the field of view (C) and disorganized and abundant microvessel growth (red arrows)—probably predominantly through a mechanism of intussusceptive angiogenesis—can also be observed (D).

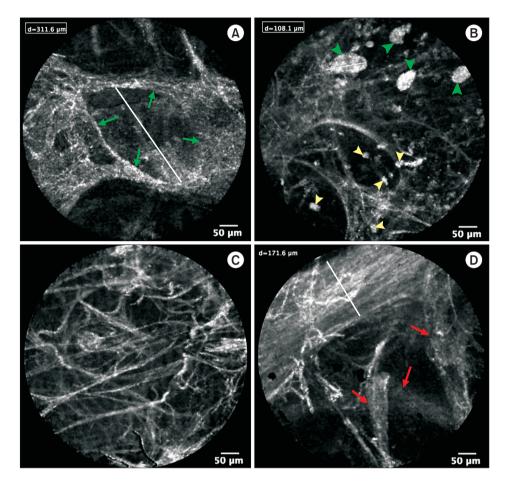
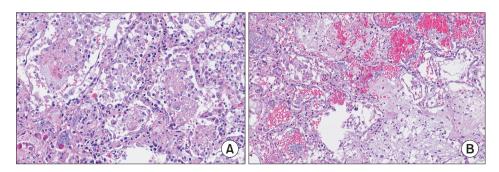


Figure 2. Protein agglomerates in the lumen of the alveoli with pulmonary macrophages, desquamated type II alveolocytes with the formation of single multinuclear structures (A) (H&E, ×200). Areas of deformed alveolar parenchyma with ingrowth of granulation tissue into the alveolar ducts and alveoli. Features of intraalveolar hemorrhage (B) (H&E, ×100).



factant layer and alveolar atelectasis⁵. In this way, pCLE data may prove the impairment of surfactant production and utilization.

These pathological changes can explain the positive effects of endobronchial surfactant treatment in critically ill covid-19 patients, demonstrated in separate cases^{6,7}.

Authors' Contributions

Conceptualization: Danilevskaya O, Sazonov D, Klypa T. Investigation: Danilevskaya O, Popova E. Writing original draft preparation: Danilevskaya O, Zabozlaev F. Writing - review and editing: Danilevskaya O, Zabozlaev F. Approval of final manuscript: all authors.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

We thank professor Alexander Averyanov, Evgeny Ilyushnikov, Dmitry Panchenkov and Bychinin Mikhail for their help in organizing of the study.

Funding

No funding to declare.

REFERENCES

- 1. Thiberville L, Salaun M, Lachkar S, Dominique S, Moreno-Swirc S, Vever-Bizet C, et al. Human *in vivo* fluorescence microimaging of the alveolar ducts and sacs during bronchoscopy. Eur Respir J 2009;33:974-85.
- 2. Salaun M, Roussel F, Hauss PA, Lachkar S, Thiberville L. *In vivo* imaging of pulmonary alveolar proteinosis using confocal endomicroscopy. Eur Respir J 2010;36:451-3.
- Lesur O, Chagnon F, Lebel R, Lepage M. In vivo endomicroscopy of lung injury and repair in ARDS: potential added value to current imaging. J Clin Med 2019;8:1197.
- 4. Zhang H, Penninger JM, Li Y, Zhong N, Slutsky AS. Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target. Intensive Care Med 2020;46:586-90.
- Cattel F, Giordano S, Bertiond C, Lupia T, Corcione S, Scaldaferri M, et al. Use of exogenous pulmonary surfactant in acute respiratory distress syndrome (ARDS): Role in SARS-CoV-2-related lung injury. Respir Physiol Neurobiol 2021;288:103645.
- **6.** Heching M, Lev S, Shitenberg D, Dicker D, Kramer MR. Surfactant for the treatment of ARDS in a patient with COVID-19. Chest 2021;160:e9-12.
- 7. Busani S, Dall'Ara L, Tonelli R, Clini E, Munari E, Venturelli S, et al. Surfactant replacement might help recovery of low-compliance lung in severe COVID-19 pneumonia. Ther Adv Respir Dis 2020;14:1753466620951043.