

Review

The effects of assisted reproduction technologies on metabolic health and disease[†]

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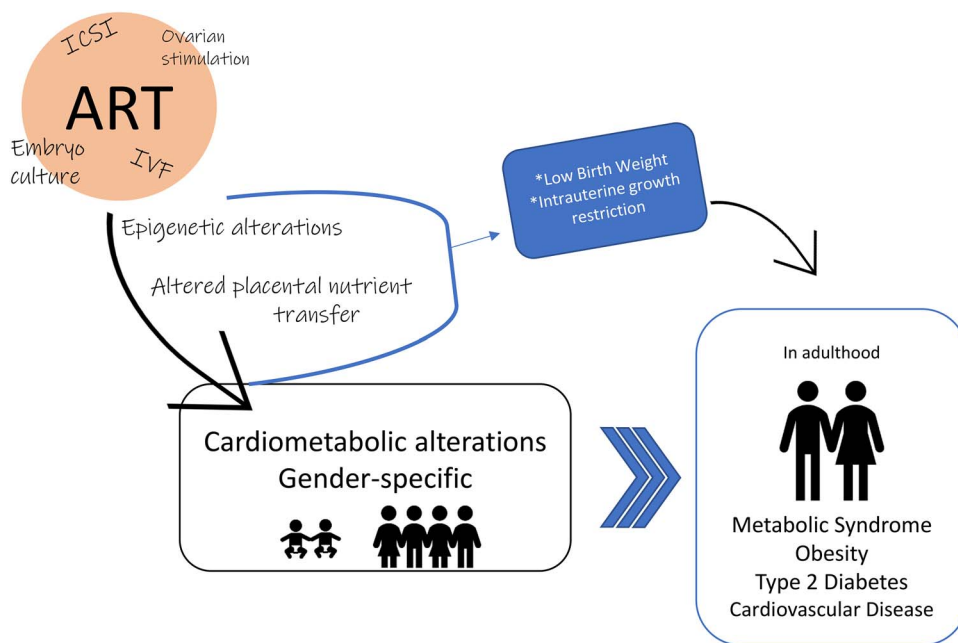
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Abstract

The increasing prevalence of metabolic diseases places a substantial burden on human health throughout the world. It is believed that predisposition to metabolic disease starts early in life, a period of great susceptibility to epigenetic reprogramming due to environmental insults. Assisted reproductive technologies (ART), i.e., treatments for infertility, may affect embryo development, resulting in multiple adverse health outcomes in postnatal life. The most frequently observed alteration in ART pregnancies is impaired placental nutrient transfer. Moreover, consequent intrauterine growth restriction and low birth weight followed by catch-up growth can all predict future obesity, insulin resistance, and chronic metabolic diseases. In this review, we have focused on evidence of adverse metabolic alterations associated with ART, which can contribute to the development of chronic adult-onset diseases, such as metabolic syndrome, type 2 diabetes, and cardiovascular disease. Due to high phenotypic plasticity, ART pregnancies can produce both offspring with adverse health outcomes, as well as healthy individuals. We further discuss the sex-specific and age-dependent metabolic alterations reflected in ART offspring, and how the degree of interference of a given ART procedure (from mild to more severe manipulation of the egg) affects the occurrence and degree of offspring alterations. Over the last few years, studies have reported signs of cardiometabolic alterations in ART offspring that are detectable at a young age but that do not appear to constitute a high risk of disease and morbidity per se. These abnormal phenotypes could be early indicators of the development of chronic diseases, including metabolic syndrome, in adulthood. The early detection of metabolic alterations could contribute to preventing the onset of disease in adulthood. Such early interventions may counteract the risk factors and improve the long-term health of the individual.

Graphical Abstract



Key words: metabolic syndrome, assisted reproductive technologies, epigenetics, cardiovascular disease, type 2 diabetes.

Introduction

In the past few decades, the incidence of metabolic diseases and metabolic syndrome in particular has increased dramatically in Western countries, becoming a global epidemic. Metabolic syndrome is a cluster of metabolic alterations characterized by obesity, insulin resistance, hypertension, altered glucose metabolism, and dyslipidaemia [1]. It is associated with increased risk of cardiovascular disease (CVD) and type 2 diabetes (T2D), among others [2]. The combination of at least three metabolic alterations forms the basis for metabolic syndrome diagnosis, whereas the presence of at least one alteration increases the risk of developing the syndrome later in life and furthermore represents a high risk of CVD [3]. Although metabolic syndrome in adulthood is undoubtedly caused by multiple factors, including modifiable lifestyle, fetal life may represent a critical window in which individuals are predisposed to metabolic syndrome later in life.

A strong relationship between adverse fetal environment and the onset of metabolic alterations has been extensively reported (reviewed in [4–6]). Human and animal studies have shown that both maternal and paternal undernutrition, obesity, stress, and exposure to endocrine disruptors can all induce metabolic alterations, leading to increased risk of T2D, CVD, and metabolic syndrome development.

The developmental origins of health and disease (DOHaD) hypothesis states that any insult during critical times of development (in utero or early life) has the ability to modify the individual's phenotype, which can lead to the onset of diseases during later life [7, 8]. This phenomenon, called developmental programming [9, 10], states that the influence of different factors—such as lifestyle, diet, environmental pollutants, medical and pharmaceutical interventions, and hormones, among others—during periods of phenotypic plasticity can affect the organism by altering its development [11]. Though

a variety of different insults are capable of programming fetal development, there is great similarity in the phenotypic outcome. The majority of these insults result in placental alterations, which may lead to intrauterine growth restriction (IUGR) and catch-up growth, which in later life, predisposes the offspring to adult disease [4].

Another example of an environmental insult is the use of assisted reproductive technologies (ART), on which we focus in this review. ART include various procedures of assisted conception used in cases of female or male infertility, and they have increased dramatically over the years. These procedures include various steps, including ovarian stimulation, oocyte/sperm recovery, gamete and embryo manipulation, cryopreservation, in vitro culture, and embryo transfer [12]. The degree of interference of each procedure also varies, and include exclusively hormonal treatment of the patient, or in vitro procedures: from mild interventions on gametes, like in vitro fertilization (IVF) to more severe interference as IntraCytoplasmic Sperm Injection (ICSI), or to the embryo, like trophectoderm or blastomere biopsy (BB). These steps are known to take place at critical times of development, when the genome is undergoing significant epigenetic remodeling and is vulnerable to environmental factors, and, if disturbed, could adversely influence developmental programming.

Little is known about the causes of developmental defects in ART-derived embryos occurring during early pregnancy, as most of them are detrimental and therefore, only the most robust and healthy embryos are able to develop to term. ART-derived neonates usually look healthy, though placental alterations of different degree and severity are frequently observed both in humans and in animals [13–17]. The authors' studies using sheep as a model showed defective placental vascularization and nutrient transport in early pregnancies obtained by ART [18, 19]. Temporal observations of ART-derived sheep embryos during early placentation demonstrated that their underdeveloped cardiovascular system was the main cause of

embryonic growth retardation and death [18]. These previous studies indicated that ART embryos with major defects are eliminated during placental vessel development. Placental vasculogenesis may therefore constitute a developmental bottleneck for ART embryos—in other words, a developmental life-or-death decision stage. High mortality of embryos with cardiovascular defects, could explain why only minor or nondetectable alterations (if any) are present at birth following ART pregnancies.

Even if a neonate looks healthy, the underdeveloped placental vascular network in ART pregnancies (caused by triggering less efficient catabolic removal and nutrient provision in fetal tissues) may cause developmental adaptations and may contribute to long-term health consequences. The placenta has a strong impact not only on fetal heart development but also on the offspring's future adulthood because it is a programming agent for cardiometabolic diseases. Increased blood pressure in ART-derived individuals [20, 21] could be the result of an underdeveloped vascular system. Altogether, the application of ART may induce impaired vasculogenesis early in gestation. This impairment may lead, in more severe circumstances, to embryo growth arrest and, in less severe ones, to fetal growth restriction throughout the pregnancy, programming the offspring to be susceptible to cardiometabolic disease.

Although the great majority of children born through ART are healthy, its use is associated with several adverse health outcomes, e.g., risk of gestational diabetes [22], pregnancy hypertension, and altered placental development and function [13, 23–25], as well as adverse perinatal outcomes, e.g., perinatal mortality, congenital defects, and epigenetic disorders [23, 26]. The most frequently observed ART complications include: (i) abnormalities and defects in the placenta (e.g., defects in nutrient transport, morphological abnormalities, impaired vasculogenesis, antioxidant defects) [14–17, 23]; (ii) low birth weight (human: [27, 28], rodents: [15, 16]; and (iii) IUGR [29–32]. IUGR and low birth weight, followed by catch-up growth has been associated with detrimental, long-term metabolic consequences and is known to predict obesity, insulin resistance, and CVD in adults [33, 34]. Thus, the adverse outcomes of ART during perinatal life have the potential to result in adverse metabolic health in postnatal life. Even if the influence of ART on a detrimental metabolic phenotype is still not completely clear, it has been suggested that children born following ART might be at higher risk of cardiovascular and metabolic alterations [25, 35].

ART and metabolic alterations

Evidence from human studies

Over the last several years, epidemiological studies have focused on the long-term metabolic health of ART-conceived children (see Figure 1). Studies have reported that young children conceived by both IVF and ICSI (the two most common ART) presented high systemic and diastolic blood pressure [20, 21, 36, 37]. Other authors have found increased blood pressure only in children conceived using hormonal induction of ovulation following ICSI or IVF [38]. Moreover, Scherrer et al. reported that ART-conceived children presented differences in systemic circulation, artery structure, and observed pulmonary hypertension in hypoxic conditions [39]. Along with this study, Chen et al. reported that IVF-conceived children, when challenged with a high-caloric diet, have increased blood pressure [36]. Overall, these alterations in the cardiovascular system could result in a high risk of cardiometabolic disease, which increases under stress situations.

Other signs of metabolic dysregulations have been observed in young children and adolescents conceived through ART. High fasting glucose levels have been reported in young ART-conceived children [21, 38]. In particular, Ceelen et al. found high glucose levels in pubertal IVF children independently of early life factors (e.g., birth weight and gestational age) or parental characteristics (e.g., parental age, subfertility, and body weight) [21]. Chen et al. noted only reduced peripheral insulin sensitivity in young adults conceived by IVF. When this cohort was challenged with a high-caloric diet, they presented high fasting glucose levels, glucose intolerance, and insulin resistance [36]. However, other studies have not found any alterations on glucose metabolism after ART [36, 37, 40]. Other studies have also reported increased peripheral adipose tissue in ART-conceived children [20, 21, 41]. Moreover, Belva et al. reported an increase in central adipose tissue in females conceived by ICSI, whereas young males showed an increase in peripheral adiposity when they reached advanced pubertal age [42]. In particular, Ceelen et al. observed a trend of increased, but not significant, body fat composition in late childhood; however at pubertal age, this high-fat deposition becomes more evident [41]. A few studies have also noted high levels of triglycerides [37]. Taken together, these studies show that ART could influence lipid and glucose metabolism in ART-conceived children, resulting in a high risk of T2D and metabolic syndrome. Moreover, the observed alterations seem to be more evident at pubertal age rather than in early childhood, suggesting that even if ART-conceived children seem healthy at a young age, is necessary to continue with follow-up studies, since metabolic alterations could arise later in life and influence the development of adult-onset disease.

Evidence from animal studies

Evidence from human studies are controversial because most ART-conceived children are still young and because it is problematic to elucidate whether the adverse outcomes are related to the procedures per se or to parental factors. Animal models have been advantageous, since you can “remove” the underlying parental infertility, as well as other confounding factors (e.g., ethnicity, genetics) and focus on the outcomes specifically correlated to the ART procedures. Alterations found in ART-conceived children have been confirmed and supported by animal models (see Figure 2). Several studies have demonstrated that mice conceived by various ART exhibit cardiovascular and metabolic abnormalities. Watkins et al. reported that embryo culture is able to induce increased blood pressure in mouse offspring, with more severe effects on female offspring, indicative of a risk of hypertension [43]. Other groups have found endothelial dysfunction and hypertension in IVF-produced mice offspring [44, 45], which could be associated with risk of premature cardiovascular morbidity. Moreover, ART-produced female offspring showed altered expression of the renin-angiotensin system, which regulates blood pressure and is known to play a role in the pathogenesis of CVD [46]. Several experimental studies in mouse have demonstrated that ART can alter glucose metabolism, from simple hyperglycaemia to impaired glucose tolerance, which can lead to hyperinsulinemia and in some cases, to insulin resistance [36, 47–49]. Due to sex-related developmental alterations observed in ART offspring, these results are apparently conflicting. Some studies have shown that female mouse offspring obtained by IVF or ICSI or under suboptimal embryo culture conditions displayed glucose intolerance and high glucose levels, whereas male offspring presented normal glucose homeostasis [47, 50]. Conversely, our studies showed that BB (an

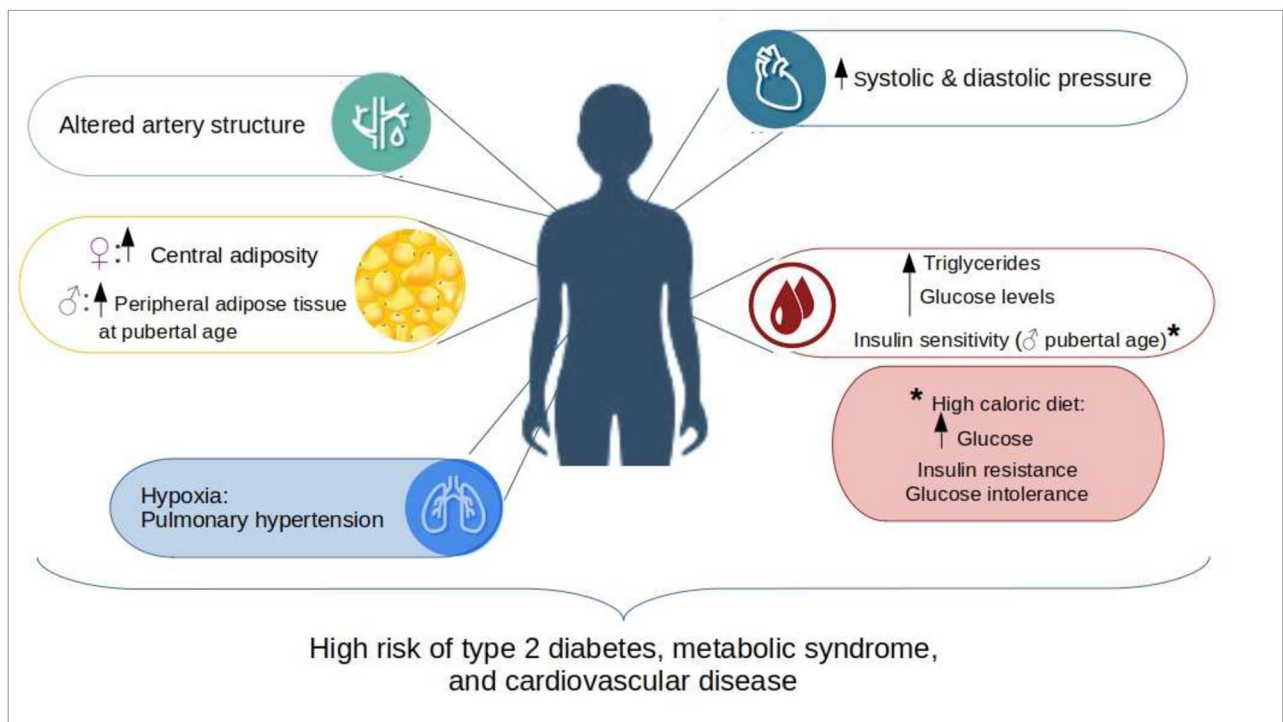


Figure 1. Evidence of cardiometabolic alterations from human studies. The illustration represents the main metabolic alterations found in ART-conceived children in the main organ systems involved in the pathogenesis of metabolic syndrome. The presence of at least one of the alterations represented in the figure increases the risk that these children will develop type 2 diabetes, metabolic syndrome, and cardiovascular disease later in life.

essential technique for performing preimplantation genetic diagnosis (PGD), a screening test that can detect genetic abnormalities of embryos before their transfer in utero) increased the body weight of the resulting male offspring as early as in the second week of life [51]. There is limited understanding of the postnatal consequences and safety of BB procedure; however, it cannot be excluded that such an invasive ART procedure affects the long-term postnatal developmental programming of male mice, suggesting that PGD could be a risk factor for late-onset, metabolic disease predisposition. Other studies have shown that male mouse offspring are more susceptible to displaying abnormal glucose metabolism [48, 52]. Sexual dimorphism has also been observed in other metabolic alterations. Feuer et al. reported that female IVF-produced offspring had increased body weight and a higher predisposition to fat deposition, as well as the altered expression of genes involved in lipid metabolism and elevated levels of reactive oxygen species and proinflammatory metabolites in adipose tissue [50]. This pattern of high postnatal body weight and increased fat deposition has been reported by other authors in mice offspring produced after in vitro embryo culture and IVF [29, 48]. These alterations are associated with accelerated neonatal growth (i.e., catch-up growth). In line with results from human studies, animal models have indicated that the alterations caused by ART depend on the specific procedure used (ICSI, IVF, or BB), and are more severe or become evident at later stages of life. Feuer et al. reported that IVF mouse offspring until 17 weeks of age presented a phenotype comparable to that of naturally conceived offspring; however, after this time, female offspring presented increased body weight, fat deposition, B cell dysfunction, and hyperinsulinemia [50]. Studies using rodent models have suggested that some metabolic or other alterations in ART-produced offspring present only later in postnatal life, usually when the offspring reaches adulthood [51].

Several researchers have studied the molecular mechanisms behind the observed adverse offspring outcomes. Rianudo et al. demonstrated that IVF mouse offspring had an altered liver and serum metabolomic profile, with the main changes corresponding to the pathways responsible for glucose metabolism [53]. Zheng et al. reported that the liver of mouse ART offspring had decreased activity of enzymes involved in glucose metabolism and that the insulin signaling pathway was altered [49], which could explain the altered glucose homeostasis. Another study reported that ART affects liver development, which results in altered lipid metabolism and the potential to develop fatty liver [54]. Overall, animal studies not only corroborate what has been observed in human studies but also provide evidence of lifelong observations of the alterations caused by ART to the cardiovascular system, adipose tissue, and liver, which are the main axes involved in the development of several metabolic diseases, including metabolic syndrome.

Epigenetics and ART

It has been well established that adverse environmental factors can induce permanent changes in the metabolic signaling pathways of fetal tissues, causing nonreversible effects on fetal growth and development that lead to the development of metabolic diseases in adult offspring (reviewed in [55, 56]). This concept is known as fetal metabolic programming. Even though the exact mechanism behind the onset of metabolic disease is still not clear, it has been postulated to be regulated by epigenetic alterations.

Epigenetics refers to the mechanisms that modify and regulate gene expression that do not involve changes in the DNA sequence

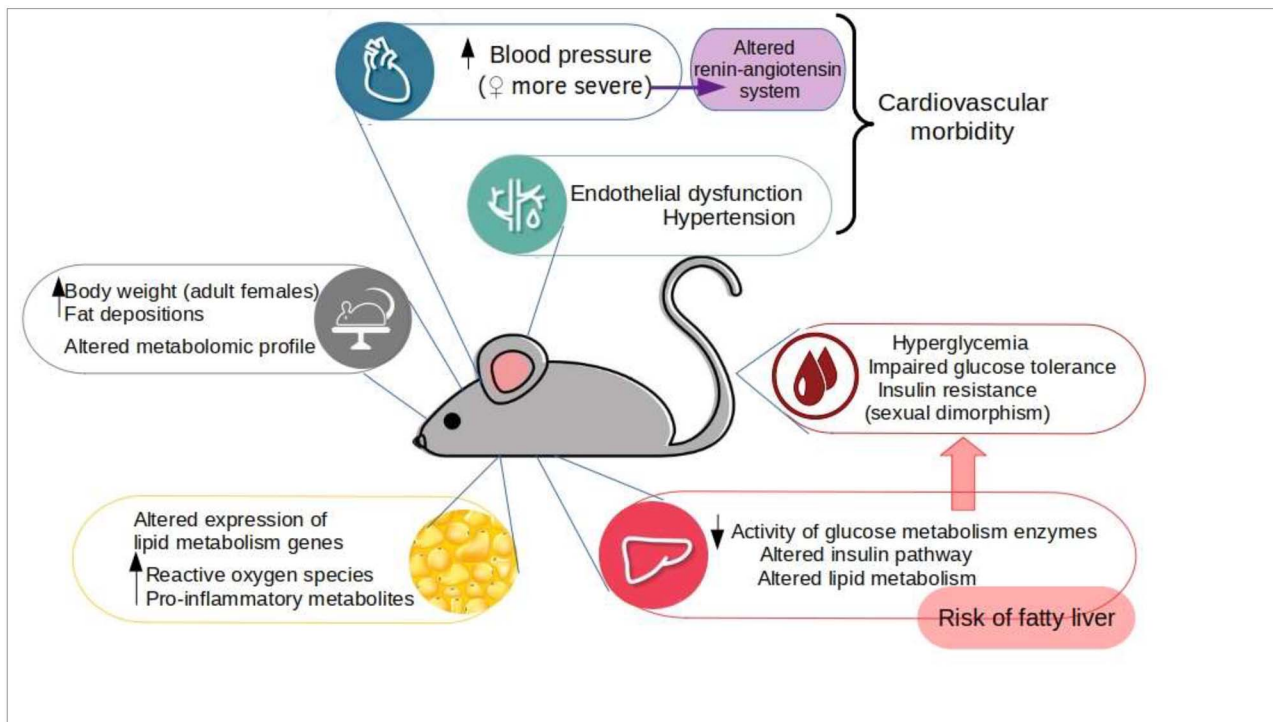


Figure 2. Evidence of cardiometabolic alterations from animal studies. The illustration summarizes the main metabolic alterations reported in ART-produced offspring from animal models and the possible molecular mechanisms behind the alterations found. Animal studies support the evidence of ART-induced alterations in human studies and provide evidence of alterations in the cardiovascular system, adipose tissue, and liver, which are the main axes involved in the development of metabolic syndrome.

[57]. Such mechanisms include chromatin structure and modifications, DNA methylation of cytosine bases, and histone protein modifications [58]. As a consequence, epigenetic modifications are responsible for phenotypic plasticity in the absence of genetic variability, which can be influenced by environmental factors. For example, maternal under- or over-nutrition is known to induce epigenetic modifications that affect gene expression in the pathways associated with a range of physiologic processes (e.g., abnormal fetal growth, energy balance regulation, adipocyte differentiation/maturity, and hepatic metabolism). These modifications lead to adverse metabolic phenotypes during adult life: obesity, insulin resistance, and cardiovascular risk, among others [59–61]. One of the most studied epigenetic mechanisms is DNA methylation, which is known to play a critical role in the genomic imprinting (parental-specific expression by silencing one allele) that occurs during gametogenesis; genomic imprinting is maintained throughout life and can affect long-term metabolic disease susceptibility [62, 63]. Several studies have reported that alterations in methylation of specific genes lead to increased risk of developing metabolic disease [56]. For instance, Xie et al. reported that IUGR leads to an insulin-resistant phenotype by altering the DNA methylation and transcriptional activity of PGC-1 α [60]. Moreover, epigenetics and imprinted genes play a critical role in the development and function of the placenta. Deletions or disruption of imprinted genes in mice show an altered balance of fetal and placental growth, evidenced by low efficiency of the placental nutrient supply [64, 65], which may lead to fetal growth restriction and thus constitutes a risk for metabolic alterations in later life. In our previous study on sheep, expression of demethyltransferase was reduced in the early placenta from in vitro-produced embryos, which is associated with growth arrest and subsequent death of the

embryos [66]. Moreover, aberrant methylation of imprinted genes such as IGF2 has been associated in human placenta with fetal growth retardation, indicative of poor nutrient supply [67]. Thus, any alteration can affect growth morphology and nutrient transfer, which can impact embryonic growth and development, leading to the adult onset of metabolic disease [68, 69]. Evidence of alterations in DNA methylation patterns associated with the development of several metabolic diseases has been described in human and animal studies. Epigenetic modifications of adiponectin and leptin genes, caused by a high-fat maternal diet, cause a metabolic syndrome-like phenotype in adult mice offspring [70]. Moreover, in obese mice, hypermethylation in the promoter region of adiponectin has been associated with an exacerbation of metabolic disease symptoms [59]. In particular, one of the most consistently observed epigenetic defects involves alterations in methylation of the imprinted regions H19/IGF2 [56, 71–73]. This imprinted gene plays key roles in placental nutrient transport, size, and morphology [74]. Changes in H19/IGF2 methylation patterns have been associated with IUGR and the subsequent development of obesity, T2D, and metabolic syndrome (reviewed in [56, 75]).

The genome undergoes several phases of epigenetic programming during development [76, 77]. Two main critical windows of epigenetic programming have been well described in murine models: (i) gametogenesis, where germinal cells undergo DNA remethylation of the genome, including imprinted regions; and (ii) early embryo development, which is characterized by global demethylation (with the exception of imprinted genes) [78, 79]. During these periods of epigenetic programming, the genome is particularly susceptible to environmentally induced epigenetic defects. The many procedures of ART coincide with these critical windows of epigenetic

programming, thus it is likely that perturbations of the environment associated with ART could lead to stable changes in the epigenome, resulting in adverse outcomes during later life.

Epidemiological studies in the past years have gathered information about imprinting disorders and epigenetic alterations in the cord blood and fetal and placental tissues of ART-conceived children. Several authors have reported, in different birth cohorts, differential methylation patterns not only in imprinted genes involved in placental development and nutrition [80–82] but also in genes involved in adipocyte differentiation and insulin signaling [82]. Most of these studies have found decreased methylation of the H19/IGF2 and SNRPN locus, associated with upregulation of gene expression [81, 83] in the placentas of IVF/ICSI pregnancies compared to naturally conceived ones [80, 83–85]. This result could explain the IUGR found in ART-conceived children and related metabolic alterations.

Some studies have suggested that ART procedures are also able to induce changes in methylation patterns in nonimprinted regions of the genome [82, 86, 87]. Moreover, this differential methylation occurs not only in the placenta but also in fetal tissues, and the changes appear to be tissue-specific [82, 85, 88, 89]. However, an important question—*are these alterations due to the ART procedures themselves or do they reflect the underlying infertility of the parents?*—remains difficult to answer due to confounding factors in human studies. Thus, researchers have turned to the use of animal models, in particular rodent models, to study the effects of ART procedures on epigenetic and imprinting alterations.

Animal studies support the findings in human studies: embryo culture and ART procedures (in the absence of infertility) can alter the epigenome and cause placental and fetal abnormalities. Studies in *in vitro*-produced sheep and cows exhibit abnormal placental and fetal development with alterations on the epigenetic control of imprinted genes, which often result in increased pregnancy loss and perinatal mortality [18, 66, 90, 91]. Hiendleder et al. reported that IVF in cows affects methylation levels in fetal tissue, in particular, hypermethylation of the fetal liver, that was found to be associated with fetal overgrowth [92]. A previous study from our group reported that embryonic growth arrest is associated with impaired placental expression and activity of DNA methyltransferase in *in vitro*-produced sheep embryos, which then results in hypomethylation and silencing of imprinted genes [66]. Similar findings in porcine embryos show that DNA methylation patterns were adversely affected by *in vitro* embryo production [93, 94]. Several authors have reported that different conditions of embryo culture in mouse can cause alterations in methylation and expression of imprinted genes involved in the growth and development of the embryo and placenta (*H19*, *Snrpn*, *Grb10*) [95–98]. A similar pattern of demethylation has been observed in the H19 locus with superovulation [99]. Furthermore, Li et al. showed that ART leads to decreased methylation levels at *H19*, *KvDMR1*, and *Snrpn* in the placenta, as well as altered expression of paternally imprinted genes involved in fetal growth [100]. Other authors have reported that IVF can alter methylation of genes involved in lipid metabolism in the placenta and can cause downregulation of these genes [30].

Altered epigenetic programming has been observed also in offspring somatic tissues (e.g., liver, brain) besides placenta and at different stages of life. Zheng et al. found differentially methylated genes related to liver development and associated with glucose (e.g., *Chrebp*, which has been associated with fatty liver and glucose intolerance) and insulin metabolism (e.g., *TNFA*, which can reduce insulin signaling) in 7.5 days postcoitum embryos [49]. These altered methylation patterns can influence abnormal glucose metabolism

and liver-derived insulin resistance in adulthood. Epigenetic alterations can also influence the development of alterations during adulthood. Rexhaj et al. reported that adult ART mice offspring present an altered methylation pattern of genes responsible for vascular processes, which could explain the vascular dysfunction found in these mice [44].

Together, these studies provide evidence that ART can alter epigenetic reprogramming, which affects fetal growth and metabolism and could result in the development of metabolic alterations during adulthood (Figure 3). Considering the high stability of epigenetic modifications, it is possible that the alterations in the methylation patterns at early developmental stages persist later in life. However, there is also evidence that epigenetic changes are reversible and can be modulated by the environment during critical windows of susceptibility. In the case of ART, one study showed that epigenetic alterations found in blood during the neonatal period were attenuated in adulthood [101], suggesting that the adverse health outcomes associated with ART might not be permanent.

Transgenerational inheritance of disease

In the last decade, transgenerational epigenetics has been considered a critical underlying mechanism of transgenerational inheritance of environmentally influenced disease [102–104]. This phenomenon refers to the transmission of a specific phenotype to the next generation of offspring via epigenetic modifications in the germline [105]. Transgenerational epigenetics has been reported to be involved in the transmission of metabolic disturbances, such as insulin resistance, obesity, and metabolic disease, among others, to second and third generations, in studies on maternal/paternal nutritional effects [103, 106, 107]. Furthermore, it was reported that methylation alterations in the promoter region of *H19* were present in two subsequent generations in an IUGR rat model [75].

This mechanism of inheritance is an emerging topic in ART since a great number of ART-conceived children have now reached marriageable age, and the possibility of transgenerational inheritance of the adverse outcomes associated with ART should be taken into serious consideration. A few studies have addressed this subject in animal models. Calle et al. observed the transmission of metabolic abnormalities in two generations of mouse offspring generated by suboptimal embryo culture [52]. Li et al. found that differentially methylated regions of the brain in IVF mouse offspring persisted to a second generation [108]. Moreover, Xu et al. reported that alterations in the methylation pattern of several genes in the testis of adult mice were conserved in a second generation [109]. These studies reveal the need for more detailed analyses of ART-induced epigenetic alterations that could lead to the development of pathologies in adult life, not only for the health of the individual in question but also for that of future generations.

Phenotypical plasticity

The extent and severity of adverse metabolic outcomes associated with ART are still controversial, as not all IVF/ICSI children present the same degree of metabolic alterations. It is evident that ART procedures affect metabolic health and disease in a sex-specific manner. Female ART-conceived offspring are more susceptible to cardiovascular alterations and obesity at a young age than their counterpart male offspring [42, 46]. However, in male offspring, the alterations become more evident later in life, suggesting the possible risk of

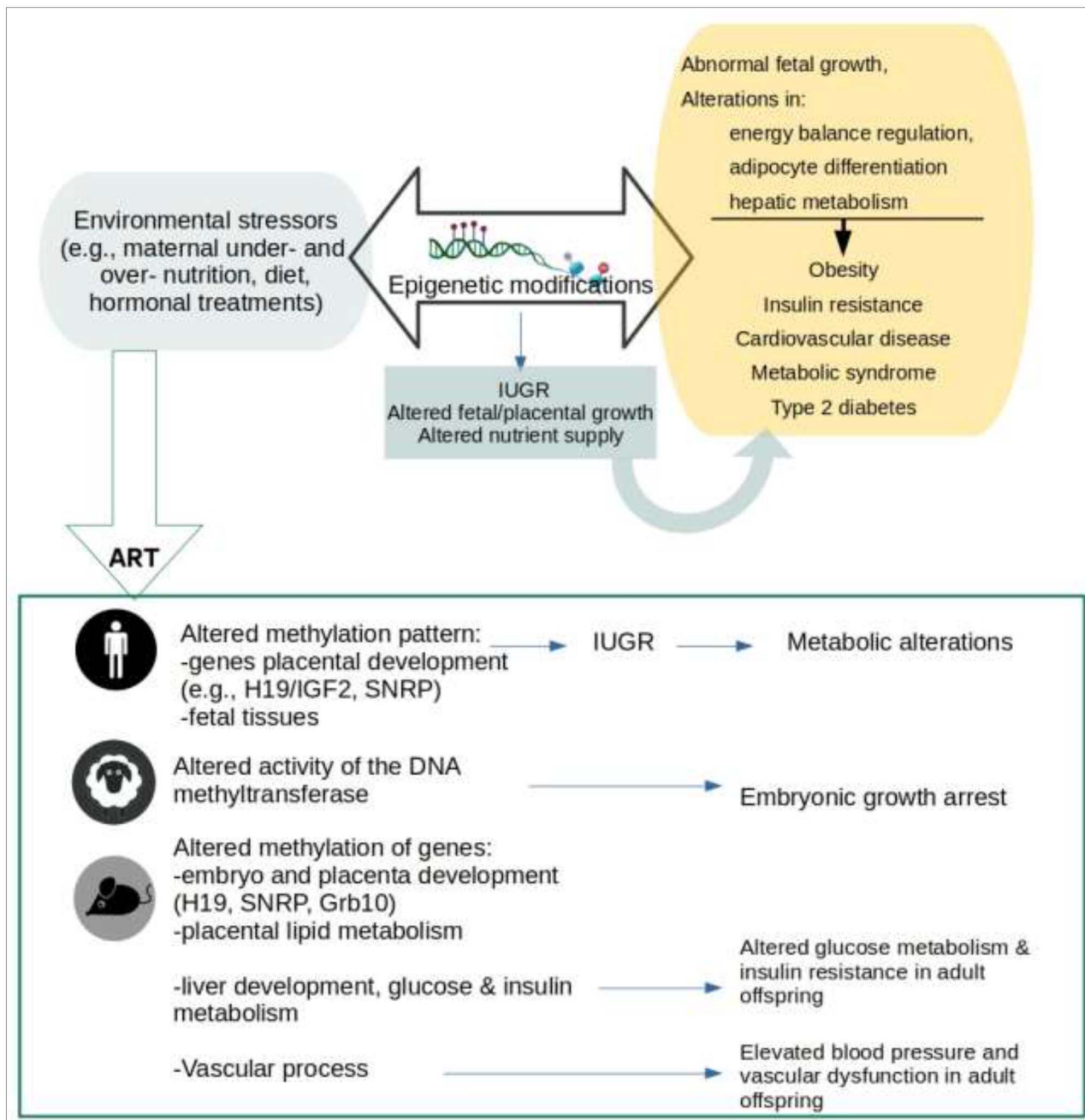


Figure 3. Epigenetics and ART. Adverse environmental factors can influence development through epigenetic modifications. Epigenetic modifications are associated with altered placental development and nutrient transport, as well as intrauterine growth restriction (IUGR), which can all lead to the onset of adult metabolic diseases. There is evidence from human and animal studies that ART can induce stable epigenetic modifications that result in adverse outcomes later in life.

further disease development [36, 42]. Sexual dimorphism is common in several metabolic alterations during adulthood (e.g., glucose metabolism and adipose tissue function), which can influence disease susceptibility [110]. Due to transcriptional sexual dimorphism, male and female genomes react differently to environmental stress, leading to sex-specific, long-term effects, and epigenetic alterations [111]. However, the difference between a mild or more severe phenotype could be explained by developmental plasticity. This phenomenon of the same environmental stressor leading to different phenotypes was observed in a rat model of in utero androgen excess, where

female offspring presented two distinctive phenotypes, both with alterations in reproductive and metabolic function. However, one phenotype presented more severe alterations and was less receptive to pharmacological treatment [112, 113]. Until recently, the long-term outcomes of developmental programming were considered to be irreversible; however, there is evidence of the reversibility of the postnatal programmed phenotype. In particular, some authors have observed improvements in some components of metabolic syndrome and cardiometabolic alterations with nutritional and exercise therapy [114, 115] during critical windows of susceptibility [116].

Thus, some of the milder adverse metabolic alterations found in ART-conceived children might be reversible with proper therapeutic interventions.

Conclusions and closing remarks

Research into the potential side effects of ART on metabolic health still has a long way to go. There is a serious need for long-term and transgenerational studies in ART-conceived children. In the paragraphs above, we have described evidence of signs of cardiometabolic alterations that are detectable at a young age, but do not appear to constitute a high risk of disease and morbidity. However, this possibility should be taken into consideration since CVD, T2D, and metabolic syndrome are chronic, adult-onset diseases, and the presence of at least one minor metabolic alteration can result in a high risk of disease later in life. Animal models have corroborated this theory, as several metabolic alterations and risk of adult disease have been reported. Studies have highlighted that at an advanced age, the alterations seem to be more severe and that a stressful environment has the ability to worsen them. We have also gathered evidence that ART is able to induce epigenetic changes that could be responsible for adverse health outcomes, and these epigenetic alterations can be transmitted across generations, culminating in the transmission of metabolic disease. However, there is evidence that the risk factors for metabolic syndrome could be reversed by changes in lifestyle (e.g., nutritional therapy, diet restrictions, physical activity), and whereas lifestyle interventions may be able to improve long-term health, the adverse outcomes may be difficult to overcome if developmental plasticity is no longer present. Thus, detection of metabolic and epigenetic alterations during the early years of ART-conceived children, as well as the proper planning of and counseling regarding therapeutic interventions, could help to prevent the development of adult-onset disease.

Conflict of interest: The authors have declared that no conflict of interest exists.

References

- Alberti KGMM, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, Fruchart JC, James WP, Loria CM, Smith SC Jr, International Diabetes Federation Task Force on Epidemiology and Prevention, National Heart, Lung, and Blood Institute et al. Harmonizing the metabolic syndrome: A joint interim statement of the international diabetes federation task force on epidemiology and prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. *Circulation* 2009; 120:1640–1645.
- Samson SL, Garber AJ. Metabolic syndrome. *Endocrinol Metab Clin North Am* 2014; 43:1–23.
- Nolan PB, Carrick-Ranson G, Stinear JW, Reading SA, Dalleck LC. Prevalence of metabolic syndrome and metabolic syndrome components in young adults: A pooled analysis. *Prev Med Rep* 2017; 7:211–215.
- Padmanabhan V, Cardoso RC, Puttabyatappa M. Developmental programming, a pathway to disease. *Endocrinology* 2016; 157:1328–1340.
- Li J, Tsuprykov O, Yang X, Hochar B. Paternal programming of offspring cardiometabolic diseases in later life. *J Hypertens* 2016; 34:2111–2126.
- Watkins AJ, Dias I, Tsuru H, Allen D, Emes RD, Moreton J, Wilson R, Ingram RJM, Sinclair KD. Paternal diet programs offspring health through sperm- and seminal plasma-specific pathways in mice. *Proc Natl Acad Sci USA* 2018; 115:10064–10069.
- Barker DJ. Fetal origins of coronary heart disease. *BMJ (Clinical Research Ed)* 1995; 311:171–174.
- Fleming TP, Velazquez MA, Eckert JJ. Embryos, DOHaD and David Barker. *J Dev Orig Health Dis* 2015; 6:377–383.
- Lucas A. Programming by early nutrition in man. *Ciba Found Symp* 1991; 156:38–50 discussion 50–55.
- Godfrey KM, Barker DJ. Fetal programming and adult health. *Public Health Nutr* 2001; 4:611–624.
- Hanson MA, Gluckman PD. Developmental origins of health and disease—global public health implications. *Best Pract Res Clin Obstet Gynaecol* 2015; 29:24–31.
- Kamel RM. Assisted reproductive technology after the birth of Louise Brown. *J Reprod Infertil* 2013; 14:96–109.
- Feuer SK, Camarano L, Rinaudo PF. ART and health: clinical outcomes and insights on molecular mechanisms from rodent studies. *Mol Hum Reprod* 2013; 19:189–204.
- Romundstad LB, Romundstad PR, Sunde A, von Düring V, Skjaerven R, Vatten LJ. Increased risk of placenta previa in pregnancies following IVF/ICSI; a comparison of ART and non-ART pregnancies in the same mother. *Hum Reprod* 2006; 21:2353–2358.
- Chen S, Sun F, Huang X, Wang X, Tang N, Zhu B, Li B. Assisted reproduction causes placental maldevelopment and dysfunction linked to reduced fetal weight in mice. *Sci Rep* 2015; 5. doi: 10.1038/srep10596.
- de Waal E, Vrooman LA, Fischer E, Ord T, Mainigi MA, Coutifaris C, Schultz RM, Bartolomei MS. The cumulative effect of assisted reproduction procedures on placental development and epigenetic perturbations in a mouse model. *Hum Mol Genet* 2015; 24:6975–6985.
- Zhu L, Zhang Y, Liu Y, Zhang R, Wu Y, Huang Y, Liu F, Li M, Sun S, Xing L, Zhu Y, Chen Y et al. Maternal and live-birth outcomes of pregnancies following assisted reproductive technology: A retrospective cohort study. *Sci Rep* 2016; 6:35141.
- Fidanza A, Toschi P, Zacchini F, Czernik M, Palmieri C, Scapolo P, Modlinski JA, Loi P, Ptak GE. Impaired placental vasculogenesis compromises the growth of sheep embryos developed in vitro. *Biol Reprod* 2014; 91:21.
- Toschi P, Czernik M, Zacchini F, Fidanza A, Loi P, Ptak GE. Evidence of placental autophagy during early pregnancy after transfer of in vitro produced (IVP) sheep embryos. *PLoS One* 2016; 11:e0157594.
- Belva F, Henriët S, Liebaers I, Van Steirteghem A, Celestin-Westreich S, Bonduelle M. Medical outcome of 8-year-old singleton ICSI children (born ≥ 32 weeks' gestation) and a spontaneously conceived comparison group. *Hum Reprod* 2007; 22:506–515.
- Ceelen M, Weissenbruch MM, Vermeiden JPW, Leeuwen FE, Delemarre-van de Waal HA. Cardiometabolic differences in children born after in vitro fertilization: follow-up study. *J Clin Endocrinol Metab* 2008; 93:1682–1688.
- Mohammadi M, Khedmati Morasae E, Maroufizadeh S, Almasi-Hashiani A, Navid B, Amini P, Omani-Samani R, Alizadeh A. Assisted reproductive technology and the risk of gestational diabetes mellitus: a systematic review and meta-analysis. *Middle East Fertil Soc J* 2020; 25:6.
- Bloise E, Feuer SK, Rinaudo PF. Comparative intrauterine development and placental function of ART concepti: implications for human reproductive medicine and animal breeding. *Hum Reprod Update* 2014; 20:822–839.
- Chen M, Heilbronn LK. The health outcomes of human offspring conceived by assisted reproductive technologies (ART). *J Dev Orig Health Dis* 2017; 8:388–402.
- Vrooman LA, Bartolomei MS. Can assisted reproductive technologies cause adult-onset disease? Evidence from human and mouse. *Reprod Toxicol* 2017; 68:72–84.
- Zhao J, Yan Y, Huang X, Li Y. Do the children born after assisted reproductive technology have an increased risk of birth defects? A systematic review and meta-analysis. *J Matern Fetal Neonatal Med* 2020; 33:322–333.
- Qin J, Liu X, Sheng X, Wang H, Gao S. Assisted reproductive technology and the risk of pregnancy-related complications and adverse pregnancy

- outcomes in singleton pregnancies: a meta-analysis of cohort studies. *Fertil Steril* 2016; 105:73–85.e1-6.
28. Schieve LA, Meikle SF, Ferre C, Peterson HB, Jeng G, Wilcox LS. Low and very low birth weight in infants conceived with use of assisted reproductive technology. *N Engl J Med* 2002; 346:731–737.
 29. Sjöblom C, Roberts CT, Wikland M, Robertson SA. Granulocyte-macrophage Colony-stimulating factor alleviates adverse consequences of embryo culture on fetal growth trajectory and placental morphogenesis. *Endocrinology* 2005; 146:2142–2153.
 30. Wei Y, Chen S, Huang X, Lam SM, Shui G, Sun F. Assisted reproduction causes intrauterus growth restriction by disrupting placental lipid metabolism. *BioRxiv* 2015; 030965. doi: 10.1101/030965.
 31. Nelissen ECM, van Montfoort APA, Smits LJM, Menheere PPCA, Evers JLH, Coonen E, Derhaag JG, Peeters LL, Coumans AB, Dumoulin JCM. IVF culture medium affects human intrauterine growth as early as the second trimester of pregnancy. *Hum Reprod* 2013; 28:2067–2074.
 32. Bay B, Lyngsø J, Hohwü L, Kesmodel US. Childhood growth of singletons conceived following in vitro fertilisation or intracytoplasmic sperm injection: a systematic review and meta-analysis. *BJOG* 2019; 126:158–166.
 33. Nobili V, Alisi A, Panera N, Agostoni C. Low birth weight and catch-up-growth associated with metabolic syndrome: a ten year systematic review. *Pediatric Endocrinology Reviews: PER* 2008; 6:241–247.
 34. Berends LM, Fernandez-Twinn DS, Martin-Gronert MS, Cripps RL, Ozanne SE. Catch-up growth following intra-uterine growth-restriction programmes an insulin-resistant phenotype in adipose tissue. *Int J Obes (Lond)* 2013; 37:1051–1057.
 35. Meister TA, Rimoldi SF, Soria R, von Arx R, Messerli FH, Sartori C, Scherrer U, Rexhaj E. Association of Assisted Reproductive Technologies with arterial hypertension during adolescence. *J Am Coll Cardiol* 2018; 72:1267–1274.
 36. Chen M, Wu L, Zhao J, Wu F, Davies MJ, Wittert GA, Norman RJ, Robker RL, Heilbronn LK. Altered glucose metabolism in mouse and humans conceived by IVF. *Diabetes* 2014; 63:3189–3198.
 37. Sakka SD, Loutradis D, Kanaka-Gantenbein C, Margeli A, Papastamatiki M, Papassotiriou I, Chrousos GP. Absence of insulin resistance and low-grade inflammation despite early metabolic syndrome manifestations in children born after in vitro fertilization. *Fertil Steril* 2010; 94:1693–1699.
 38. Pontesilli M, Painter RC, Grooten IJ, van der Post JA, Mol BW, Vrijkotte TGM, Repping S, Roseboom TJ. Subfertility and assisted reproduction techniques are associated with poorer cardiometabolic profiles in childhood. *Reprod Biomed Online* 2015; 30:258–267.
 39. Scherrer U, Rimoldi SF, Rexhaj E, Stuber T, Duplain H, Garcin S, de Marchi SF, Nicod P, Germond M, Allemann Y, Sartori C. Systemic and pulmonary vascular dysfunction in children conceived by assisted reproductive technologies. *Circulation* 2012; 125:1890–1896.
 40. Green MP, Mouat F, Miles HL, Hopkins SA, Derraik JGB, Hofman PL, Peek JC, Cutfield WS. Phenotypic differences in children conceived from fresh and thawed embryos in in vitro fertilization compared with naturally conceived children. *Fertil Steril* 2013; 99:1898–1904.
 41. Ceelen M, Weissenbruch MM, Roos JC, Vermeiden JPW, Leeuwen FE, Delemarre-van de Waal HA. Body composition in children and adolescents born after in vitro fertilization or spontaneous conception. *J Clin Endocrinol Metab* 2007; 92:3417–3423.
 42. Belva, F, Painter, R., Bonduelle, M., Roelants, M., Devroey, P., De Schepper, J., 2012. Are ICSI adolescents at risk for increased adiposity? *Hum Reprod* 27: 257–64
 43. Watkins AJ, Platt D, Papenbrock T, Wilkins A, Eckert JJ, Kwong WY, Osmond C, Hanson M, Fleming TP. Mouse embryo culture induces changes in postnatal phenotype including raised systolic blood pressure. *Proc Natl Acad Sci* 2007; 104:5449–5454.
 44. Rexhaj E, Paoloni-Giacobino A, Rimoldi SF, Fuster DG, Anderegg M, Somm E, Bouillet E, Allemann Y, Sartori C, Scherrer U. Mice generated by in vitro fertilization exhibit vascular dysfunction and shortened life span. *J Clin Invest* 2013; 123:5052–5060.
 45. Ramirez-Perez FI, Schenewerk AL, Coffman KL, Foote C, Ji T, Rivera RM, Martinez-Lemus LA. Effects of the use of assisted reproductive technologies and an obesogenic environment on resistance artery function and diabetes biomarkers in mice offspring. *PLoS One* 2014; 9:e112651. doi: 10.1371/journal.pone.0112651.
 46. Wang Q, Zhang Y, Le F, Wang N, Zhang F, Luo Y, Lou Y, Hu M, Wang L, Thurston LM, Xu X, Jin F. Alteration in the expression of the renin-angiotensin system in the myocardium of mice conceived by in vitro fertilization. *Biol Reprod* 2018; 99:1276–1288.
 47. Scott KA, Yamazaki Y, Yamamoto M, Lin Y, Melhorn SJ, Krause EG, Woods SC, Yanagimachi R, Sakai RR, Tamashiro KLK. Glucose parameters are altered in mouse offspring produced by assisted reproductive technologies and somatic cell nuclear transfer. *Biol Reprod* 2010; 83:220–227.
 48. Donjacour A, Liu X, Lin W, Simbulan R, Rinaudo PF. Vitro fertilization affects growth and glucose metabolism in a sex-specific manner in an outbred mouse model. *Biol Reprod* 2014; 90. doi: 10.1095/biolreprod.113.113134.
 49. Zheng MM, Cao HR, Zhang WY, Yan PP, Xu JY, Zhao HL, Zhu F, Zhang JJ, Li Y, Zhu H. Abnormal gene methylation during embryonic development after preimplantation genetic testing increases risk of liver-derived insulin resistance. *Ann N Y Acad Sci* 2018; 1425:70–81.
 50. Feuer SK, Liu X, Donjacour A, Lin W, Simbulan RK, Giritharan G, Piane LD, Kolahi K, Ameri K, Maltepe E, Rinaudo PF. Use of a mouse in vitro fertilization model to understand the developmental origins of health and disease hypothesis. *Endocrinology* 2014; 155:1956–1969.
 51. Sampino S, Zacchini F, Swiergiel AH, Modlinski AJ, Loi P, Ptak GE. Effects of blastomere biopsy on post-natal growth and behavior in mice. *Hum Reprod* 2014; 29:1875–1883.
 52. Calle A, Miranda A, Fernandez-Gonzalez R, Pericuesta E, Laguna R, Gutierrez-Adan A. Male mice produced by in vitro culture have reduced fertility and transmit organomegaly and glucose intolerance to their male offspring. *Biol Reprod* 2012; 87:34.
 53. Rinaudo P, Liu X, Lin W, Simbulan R, Feuer S, Donjacour A. Metabolic differences in serum and liver can explain the impaired glucose tolerance found in adult mice conceived by IVF. *Biol Reprod* 2012; 87:114–114.
 54. Gu L, Zhang J, Zheng M, Dong G, Xu J, Zhang W, Wu Y, Yang Y, Zhu H. A potential high risk for fatty liver disease was found in mice generated after assisted reproductive techniques. *J Cell Biochem* 2018; 119:1899–1910.
 55. Rinaudo P, Wang E. Fetal programming and metabolic syndrome. *Annu Rev Physiol* 2012; 74:107–130.
 56. Zhu Z, Cao F, Li X. Epigenetic programming and fetal metabolic programming. *Front Endocrinol* 2019; 10. doi: 10.3389/fendo.2019.00764.
 57. Holliday R. Epigenetics: A historical overview. *Epigenetics* 2006; 1:76–80.
 58. Handy DE, Castro R, Loscalzo J. Epigenetic modifications: basic mechanisms and role in cardiovascular disease. *Circulation* 2011; 123:2145–2156.
 59. Kim AY, Park YJ, Pan X, Shin KC, Kwak S-H, Bassas AF, Sallam RM, Park KS, Alfadda AA, Xu A, Kim JB. Obesity-induced DNA hypermethylation of the adiponectin gene mediates insulin resistance. *Nat Commun* 2015; 6:7585.
 60. Xie X, Lin T, Zhang M, Liao L, Yuan G, Gao H, Ning Q, Luo X. IUGR with infantile overnutrition programs an insulin-resistant phenotype through DNA methylation of peroxisome proliferator-activated receptor- γ coactivator-1 α in rats. *Pediatr Res* 2015; 77: 625–632.
 61. Masuyama H, Mitsui T, Eguchi T, Tamada S, Hiramatsu Y. The effects of paternal high-fat diet exposure on offspring metabolism with epigenetic changes in the mouse adiponectin and leptin gene promoters. *Am J Physiol Endocrinol Metab* 2016; 311:E236–E245.
 62. Barlow DP, Bartolomei MS. Genomic imprinting in mammals. *Cold Spring Harb Perspect Biol* 2014; 6:a018382.
 63. Bartolomei MS, Ferguson-Smith AC. Mammalian genomic imprinting. *Cold Spring Harb Perspect Biol* 2011; 3. doi: 10.1101/cshperspect.a002592.

64. Constância M, Hemberger M, Hughes J, Dean W, Ferguson-Smith A, Fundele R, Stewart F, Kelsey G, Fowden A, Sibley C, Reik W. Placental-specific IGF-II is a major modulator of placental and fetal growth. *Nature* 2002; 417:945–948.
65. Angiolini E, Fowden A, Coan P, Sandovici I, Smith P, Dean W, Burton G, Tycko B, Reik W, Sibley C. Regulation of placental efficiency for nutrient transport by imprinted genes. *Placenta* 2006; 27:98–102.
66. Ptak GE, D'Agostino A, Toschi P, Fidanza A, Zacchini F, Czernik M, Monaco F, Loi P. Post-implantation mortality of in vitro produced embryos is associated with DNA methyltransferase 1 dysfunction in sheep placenta. *Hum Reprod* 2013; 28:298–305.
67. Koukoura O, Sifakis S, Zaravinos A, Apostolidou S, Jones A, Hajjiounou J, Widschwendter M, Spandidos DA. Hypomethylation along with increased H19 expression in placentas from pregnancies complicated with fetal growth restriction. *Placenta* 2011; 32:51–57.
68. Apicella C, Ruano CSM, Méhats C, Miralles F, Vaiman D. The role of epigenetics in placental development and the Etiology of preeclampsia. *Int J Mol Sci* 2019; 20. doi: 10.3390/ijms20112837.
69. Maccani MA, Marsit CJ. Epigenetics in the placenta. *Am J Reprod Immunol* 2009; 62:78.
70. Masuyama H, Hiramatsu Y. Effects of a high-fat diet exposure in utero on the metabolic syndrome-like phenomenon in mouse offspring through epigenetic changes in Adipocytokine gene expression. *Endocrinology* 2012; 153:2823–2830.
71. Huang R-C, Galati JC, Burrows S, Beilin LJ, Li X, Pennell CE, van Eekelen J, Mori TA, Adams LA, Craig JM. DNA methylation of the IGF2/H19 imprinting control region and adiposity distribution in young adults. *Clin Epigenetics* 2012; 4:21.
72. Williams-Wyss O, Zhang S, MacLaughlin SM, Kleemann D, Walker SK, Suter CM, Cropley JE, Morrison JL, Roberts CT, Caroline McMillen I. Embryo number and periconceptual undernutrition in the sheep have differential effects on adrenal epigenotype, growth, and development. *Am J Physiol Endocrinol Metab* 2014; 307:E141–E150.
73. Hernández-Valero MA, Rother J, Gorlov I, Frazier M, Gorlova O. Interplay between polymorphisms and methylation in the H19/IGF2 gene region may contribute to obesity in Mexican-American children. *J Dev Orig Health Dis* 2013; 4:499–506.
74. Nelissen ECM, van Montfoort APA, Dumoulin JCM, Evers JLH. Epigenetics and the placenta. *Hum Reprod Update* 2011; 17:397–417.
75. Gonzalez-Rodriguez P, Cantu J, O'Neil D, Seferovic MD, Goodspeed DM, Suter MA, Aagaard KM. Alterations in expression of imprinted genes from the H19/IGF2 loci in a multigenerational model of intrauterine growth restriction (IUGR). *Am J Obstet Gynecol* 2016; 214: 625.e1–625.e11.
76. Reik W. Stability and flexibility of epigenetic gene regulation in mammalian development. *Nature* 2007; 447:425–432.
77. Surani MA, Hayashi K, Hajkova P. Genetic and epigenetic regulators of pluripotency. *Cell* 2007; 128:747–762.
78. Saitou M, Kagiwada S, Kurimoto K. Epigenetic reprogramming in mouse pre-implantation development and primordial germ cells. *Development* 2012; 139:15–31.
79. Kierszenbaum AL. Epigenetic reprogramming in germ cells and preimplantation embryos. In: Jorde LB, Little PFR, Dunn MJ, Subramaniam S (eds.), *Encyclopedia of Genetics, Genomics, Proteomics and Bioinformatics*. Chichester: John Wiley & Sons, Ltd; 2005: g103320.
80. Choux C, Binquet C, Carmignac V, Bruno C, Chapusot C, Barberet J, Lamotte M, Sagot P, Bourc'his D, Fauque P. The epigenetic control of transposable elements and imprinted genes in newborns is affected by the mode of conception: ART versus spontaneous conception without underlying infertility. *Hum Reprod* 2018; 33:331–340.
81. Rancourt RC, Harris HR, Michels KB. Methylation levels at imprinting control regions are not altered with ovulation induction or in vitro fertilization in a birth cohort. *Hum Reprod* 2012; 27:2208–2216.
82. Katari S, Turan N, Bibikova M, Erinle O, Chalian R, Foster M, Gaughan JP, Coutifaris C, Sapienza C. DNA methylation and gene expression differences in children conceived in vitro or in vivo. *Hum Mol Genet* 2009; 18:3769–3778.
83. Turan N, Katari S, Gerson LF, Chalian R, Foster MW, Gaughan JP, Coutifaris C, Sapienza C. Inter- and intra-individual variation in allele-specific DNA methylation and gene expression in children conceived using assisted reproductive technology. *PLoS Genet* 2010; 6:e1001033.
84. Nelissen ECM, Dumoulin JCM, Daunay A, Evers JLH, Tost J, van Montfoort APA. Placentas from pregnancies conceived by IVF/ICSI have a reduced DNA methylation level at the H19 and MEST differentially methylated regions. *Hum Reprod* 2013; 28:1117–1126.
85. Lou H, le F, Hu M, Yang X, Li L, Wang L, Wang N, Gao H, Jin F. Aberrant DNA methylation of IGF2-H19 locus in human Fetus and in spermatozoa from assisted reproductive technologies. *Reprod Sci* 2019; 26:997–1004.
86. Camprubí C, Iglesias-Platas I, Martin-Trujillo A, Salvador-Alarcon C, Rodriguez MA, Barredo DR, Court F, Monk D. Stability of genomic imprinting and gestational-age dynamic methylation in complicated pregnancies conceived following assisted reproductive technologies. *Biol Reprod* 2013; 89:50.
87. Melamed N, Choufani S, Wilkins-Haug LE, Koren G, Weksberg R. Comparison of genome-wide and gene-specific DNA methylation between ART and naturally conceived pregnancies. *Epigenetics* 2015; 10:474–483.
88. Mani S, Ghosh J, Coutifaris C, Sapienza C, Mainigi M. Epigenetic changes and assisted reproductive technologies. *Epigenetics* 2020; 15:12–25.
89. Lazaraviciute G, Kausar M, Bhattacharya S, Haggarty P, Bhattacharya S. A systematic review and meta-analysis of DNA methylation levels and imprinting disorders in children conceived by IVF/ICSI compared with children conceived spontaneously. *Hum Reprod Update* 2014; 20:840–852.
90. Young LE, Fernandes K, McEvoy TG, Butterwith SC, Gutierrez CG, Carolan C, Broadbent PJ, Robinson JJ, Wilmot I, Sinclair KD. Epigenetic change in IGF2R is associated with fetal overgrowth after sheep embryo culture. *Nat Genet* 2001; 27:153–154.
91. Farin PW, Piedrahita JA, Farin CE. Errors in development of fetuses and placentas from in vitro-produced bovine embryos. *Theriogenology* 2006; 65:178–191.
92. Hiendler S, Wirtz M, Mund C, Klempt M, Reichenbach H-D, Stojkovic M, Weppert M, Wenigerkind H, Elmlinger M, Lyko F, Schmitz OJ, Wolf E. Tissue-specific effects of in vitro fertilization procedures on genomic cytosine methylation levels in overgrown and normal sized bovine fetuses. *Biol Reprod* 2006; 75:17–23.
93. Bonk AJ, Li R, Lai L, Hao Y, Liu Z, Samuel M, Ferguson EA, Whitworth KM, Murphy CN, Antoniou E, Prather RS. Aberrant DNA methylation in porcine in vitro-, parthenogenetic-, and somatic cell nuclear transfer-produced blastocysts. *Mol Reprod Dev* 2008; 75: 250–264.
94. Deshmukh RS, Østrup O, Østrup E, Vejlsted M, Niemann H, Lucas-Hahn A, Petersen B, Li J, Callesen H, Hyttel P. DNA methylation in porcine preimplantation embryos developed in vivo and produced by in vitro fertilization, parthenogenetic activation and somatic cell nuclear transfer. *Epigenetics* 2011; 6:177–187.
95. de Waal E, Mak W, Calhoun S, Stein P, Ord T, Krapp C, Coutifaris C, Schultz RM, Bartolomei MS. In vitro culture increases the frequency of stochastic epigenetic errors at imprinted genes in placental tissues from mouse concepti produced through assisted reproductive technologies. *Biol Reprod* 2014; 90:22.
96. Doherty AS, Mann MR, Tremblay KD, Bartolomei MS, Schultz RM. Differential effects of culture on imprinted H19 expression in the preimplantation mouse embryo. *Biol Reprod* 2000; 62:1526–1535.
97. Fortier AL, Lopes FL, Darricarrère N, Martel J, Trasler JM. Superovulation alters the expression of imprinted genes in the midgestation mouse placenta. *Hum Mol Genet* 2008; 17:1653–1665.
98. Khosla S, Dean W, Brown D, Reik W, Feil R. Culture of preimplantation mouse embryos affects fetal development and the expression of imprinted genes. *Biol Reprod* 2001; 64:918–926.
99. Market-Velker BA, Zhang L, Magri LS, Bonvissuto AC, Mann MRW. Dual effects of superovulation: loss of maternal and paternal imprinted

- methylation in a dose-dependent manner. *Hum Mol Genet* 2010; 19:36–51.
100. Li B, Chen S, Tang N, Xiao X, Huang J, Jiang F, Huang X, Sun F, Wang X. Assisted reproduction causes reduced fetal growth associated with downregulation of paternally expressed imprinted genes that enhance fetal growth in mice. *Biol Reprod* 2016; 94. doi: [10.1095/biol-reprod.115.136051](https://doi.org/10.1095/biol-reprod.115.136051).
 101. Novakovic B, Lewis S, Halliday J, Kennedy J, Burgner DP, Czajko A, Kim B, Sexton-Oates A, Juonala M, Hammarberg K, Amor DJ, Doyle LW et al. Assisted reproductive technologies are associated with limited epigenetic variation at birth that largely resolves by adulthood. *Nat Commun* 2019; 10:1–12.
 102. Gluckman PD, Hanson MA, Beedle AS. Non-genomic transgenerational inheritance of disease risk. *BioEssays: news and reviews in molecular, cellular. Dev Biol* 2007; 29:145–154.
 103. Stegeman R, Buchner DA. Transgenerational inheritance of metabolic disease. *Semin Cell Dev Biol* 2015; 43:131–140.
 104. Szyf M. Nongenetic inheritance and transgenerational epigenetics. *Trends Mol Med* 2015; 21:134–144.
 105. Daxinger L, Whitelaw E. Understanding transgenerational epigenetic inheritance via the gametes in mammals. *Nat Rev Genet* 2012; 13:153–162.
 106. Martínez D, Pentinat T, Ribó S, Daviaud C, Bloks VW, Cebrià J, Villalmanzo N, Kalko SG, Ramón-Krauel M, Díaz R, Plösch T, Tost J et al. In utero undernutrition in male mice programs liver lipid metabolism in the second-generation offspring involving altered Lxra DNA methylation. *Cell Metab* 2014; 19:941–951.
 107. Jimenez-Chillaron JC, Ramon-Krauel M, Ribo S, Diaz R. Transgenerational epigenetic inheritance of diabetes risk as a consequence of early nutritional imbalances. *Proc Nutr Soc* 2016; 75: 78–89.
 108. Li L, Wang L, Xu X, Lou H, le F, Li L, Sheng J, Huang H, Jin F. Genome-wide DNA methylation patterns in IVF-conceived mice and their progeny: a putative model for ART-conceived humans. *Reprod Toxicol* 2011; 32:98–105.
 109. Xu X-R, Fu R-G, Wang L-Y, Wang N, Zhang F, le F, Li L, Li LJ, Liu XZ, Zheng YM, Lou HY, Jiang SW et al. Epigenetic inheritance of paternally expressed imprinted genes in the testes of ICSI mice. *Curr Pharm Des* 2014; 20:1764–1771.
 110. Geer EB, Shen W. Gender differences in insulin resistance, body composition. *and energy balance Gender Medicine* 2009; 6:60–75.
 111. Gilbert JS, Banek CT. Sex differences in the developmental programming of adult disease. *Sex Hormones* 2012. doi: [10.5772/26019](https://doi.org/10.5772/26019).
 112. Abruzzese GA, Heber MF, Ferreira SR, Velez LM, Reynoso R, Pignataro OP, Motta AB. Prenatal hyperandrogenism induces alterations that affect liver lipid metabolism. *J Endocrinol* 2016; 230:67–79.
 113. Heber MF, Ferreira SR, Abruzzese GA, Trinidad R, Pignataro OP, Vega M, Motta AB. Metformin improves ovarian insulin signaling alterations caused by fetal programming. *J Endocrinol* 2019. doi: [10.1530/JOE-18-0520](https://doi.org/10.1530/JOE-18-0520).
 114. Monzavi R, Dreimane D, Geffner ME, Braun S, Conrad B, Klier M, Kaufman FR. Improvement in risk factors for metabolic syndrome and insulin resistance in overweight youth who are treated with lifestyle intervention. *Pediatrics* 2006; 117:e1111–e1118.
 115. Li N, Guenancia C, Rigal E, Hachet O, Chollet P, Desmoulines L, Leloup C, Rochette L, Vergely C. Short-term moderate diet restriction in adulthood can reverse oxidative, cardiovascular and metabolic alterations induced by postnatal overfeeding in mice. *Sci Rep* 2016; 6. doi: [10.1038/srep30817](https://doi.org/10.1038/srep30817).
 116. Vickers MH, Sloboda DM. Strategies for reversing the effects of metabolic disorders induced as a consequence of developmental programming. *Front Physiol* 2012; 3. doi: [10.3389/fphys.2012.00242](https://doi.org/10.3389/fphys.2012.00242).