



# Draft Genome Sequence of Megaplasmid-Bearing *Staphylococcus sciuri* Strain B9-58B, Isolated from Retail Pork

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**ABSTRACT** Here, we report the genome sequence of the megaplasmid-bearing *Staphylococcus sciuri* strain B9-58B, isolated from retail pork. This strain contains a 2,761,440-bp chromosome and a 162,858-bp megaplasmid. The genome contains putative genes involved in virulence, the stress response, and antimicrobial agent and heavy metal resistance.

*Staphylococcus* spp. are major foodborne pathogens that cause food poisoning through the production of enterotoxins (1). Numerous studies in the United States have revealed a high prevalence of multidrug-resistant *Staphylococcus* strains in food, indicating the potential acquisition and colonization of strains by food industry workers and consumers (2–4). We previously reported a high prevalence of *Staphylococcus aureus* in retail pork products, and isolates exhibited multidrug resistance and encoded toxins (5).

*Staphylococcus sciuri* is a coagulase-negative, oxidase-positive, and novobiocin-resistant species. While animals are the main reservoir for *S. sciuri* (6), this organism has also been isolated from humans, food, and the environment (4, 7, 8). Although coagulase-negative species are generally known as harmless members of the microbiota of human skin and mucous membranes, they are increasingly being identified as causative agents of health care- and community-associated infections. Recent research has proposed that *S. sciuri* functions as a reservoir of virulence and antibiotic resistance genes that facilitate *S. aureus* colonization and survival in different environments (9). Existing reports on foodborne *Staphylococcus* spp. have focused primarily on coagulase-positive *S. aureus* strains.

Here, we announce the draft genome sequence of *S. sciuri* strain B9-58B, which was previously isolated from retail pork and was initially identified as *S. aureus* by PCR using the *S. aureus*-specific primers Sa442-1 and Sa442-2 (5, 10). This isolate contains a 162,858-bp megaplasmid, which was confirmed by S1 nuclease pulsed-field gel electrophoresis, as described previously (11). This isolate was grown for 16 to 24 h in tryptic soy agar at 37°C under aerobic conditions, and genomic DNA was isolated and purified using a DNeasy blood and tissue kit (Qiagen, Valencia, CA). The library used for sequencing was prepared using the Nextera XT library preparation kit for small genomes (Illumina, Inc., San Diego, CA), and a MiSeq desktop sequencer was used to sequence the whole genome with an Illumina v2 reagent kit (2 × 250 cycles). The 500 cycles yielded approximately 229× coverage. The sequence reads that passed the initial MiSeq quality standards were assessed using CLC Genomics Workbench v12.0 (Qiagen) and were trimmed for adapters, low-quality reads, and short reads. A total of 3,202,194 paired-end reads, with a mean sequence length of 201.1 bp, were submitted to the comprehensive genome analysis service at PATRIC v3.5.39 (12) for *de novo* assembly using the default settings. The resulting contigs, with an  $N_{50}$  value of 249,127 bp and a GC content of 32.45%, were scaffolded by aligning them against the closest reference using Genome Finishing Module v1.9 of CLC Genomics Workbench v12.0 (Qiagen).

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The genome of *S. sciuri* B9-58B contained a single chromosome and a megaplasmid, which were 2,761,440 and 162,858 bp, respectively, with GC contents of 32.66% and 31.13%. While it was possible to close the chromosome, the plasmid remained unclosed, with 39 contigs. The B9-58B genome was annotated using the RAST tool kit. The chromosome contained 2,712 protein-coding sequences, 62 tRNA genes, and 16 rRNA genes, while the megaplasmid had 218 coding sequences. Multiple stress response genes, including osmotic, oxidative, heat/cold shock, and periplasmic stress-related genes, in addition to detoxification and carbon starvation genes, were chromosomally located. This strain contained genes conferring resistance to a variety of antibiotics and heavy metals, such as bacitracin, fluoroquinolones, copper, cobalt-zinc-cadmium, and mercury. Interestingly, the chromosome contained a *Mycobacterium* virulence operon that is involved in protein synthesis and facilitates invasion and intracellular resistance. The megaplasmid contained genes conferring resistance to cobalt, zinc, copper, cadmium, fosfomycin, and chromium.

**Data availability.** The raw sequence reads have been deposited in the Sequence Read Archive (SRA) under accession number [PRJNA555628](https://www.ncbi.nlm.nih.gov/sra/PRJNA555628). The whole-genome shotgun sequences for *Staphylococcus sciuri* strain B9-58B have been deposited in GenBank under accession number [CP041879](https://www.ncbi.nlm.nih.gov/genbank/CP041879) for the chromosome and accession numbers [CP041880](https://www.ncbi.nlm.nih.gov/genbank/CP041880), [CP041881](https://www.ncbi.nlm.nih.gov/genbank/CP041881), [CP041882](https://www.ncbi.nlm.nih.gov/genbank/CP041882), [CP041883](https://www.ncbi.nlm.nih.gov/genbank/CP041883), [CP041884](https://www.ncbi.nlm.nih.gov/genbank/CP041884), [CP041885](https://www.ncbi.nlm.nih.gov/genbank/CP041885), [CP041886](https://www.ncbi.nlm.nih.gov/genbank/CP041886), [CP041887](https://www.ncbi.nlm.nih.gov/genbank/CP041887), [CP041888](https://www.ncbi.nlm.nih.gov/genbank/CP041888), [CP041889](https://www.ncbi.nlm.nih.gov/genbank/CP041889), [CP041890](https://www.ncbi.nlm.nih.gov/genbank/CP041890), [CP041891](https://www.ncbi.nlm.nih.gov/genbank/CP041891), [CP041892](https://www.ncbi.nlm.nih.gov/genbank/CP041892), [CP041893](https://www.ncbi.nlm.nih.gov/genbank/CP041893), [CP041894](https://www.ncbi.nlm.nih.gov/genbank/CP041894), [CP041895](https://www.ncbi.nlm.nih.gov/genbank/CP041895), [CP041896](https://www.ncbi.nlm.nih.gov/genbank/CP041896), [CP041897](https://www.ncbi.nlm.nih.gov/genbank/CP041897), [CP041898](https://www.ncbi.nlm.nih.gov/genbank/CP041898), [CP041899](https://www.ncbi.nlm.nih.gov/genbank/CP041899), [CP041900](https://www.ncbi.nlm.nih.gov/genbank/CP041900), [CP041901](https://www.ncbi.nlm.nih.gov/genbank/CP041901), [CP041902](https://www.ncbi.nlm.nih.gov/genbank/CP041902), [CP041903](https://www.ncbi.nlm.nih.gov/genbank/CP041903), [CP041904](https://www.ncbi.nlm.nih.gov/genbank/CP041904), [CP041905](https://www.ncbi.nlm.nih.gov/genbank/CP041905), [CP041906](https://www.ncbi.nlm.nih.gov/genbank/CP041906), [CP041907](https://www.ncbi.nlm.nih.gov/genbank/CP041907), [CP041908](https://www.ncbi.nlm.nih.gov/genbank/CP041908), [CP041909](https://www.ncbi.nlm.nih.gov/genbank/CP041909), [CP041910](https://www.ncbi.nlm.nih.gov/genbank/CP041910), [CP041911](https://www.ncbi.nlm.nih.gov/genbank/CP041911), [CP041912](https://www.ncbi.nlm.nih.gov/genbank/CP041912), [CP041913](https://www.ncbi.nlm.nih.gov/genbank/CP041913), [CP041914](https://www.ncbi.nlm.nih.gov/genbank/CP041914), [CP041915](https://www.ncbi.nlm.nih.gov/genbank/CP041915), [CP041916](https://www.ncbi.nlm.nih.gov/genbank/CP041916), [CP041917](https://www.ncbi.nlm.nih.gov/genbank/CP041917), and [CP041918](https://www.ncbi.nlm.nih.gov/genbank/CP041918) for the plasmid.

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## REFERENCES

- Kadariya J, Smith TC, Thapaliya D. 2014. *Staphylococcus aureus* and staphylococcal food-borne disease: an ongoing challenge in public health. *Biomed Res Int* 2014:827965–827969. <https://doi.org/10.1155/2014/827965>.
- Ge B, Mukherjee S, Hsu CH, Davis JA, Tran TTT, Yang Q, Abbott JW, Ayers SL, Young SR, Crarey ET, Womack NA, Zhao S, McDermott PF. 2017. MRSA and multidrug-resistant *Staphylococcus aureus* in U.S. retail meats, 2010–2011. *Food Microbiol* 62:289–297. <https://doi.org/10.1016/j.fm.2016.10.029>.
- Jackson CR, Davis JA, Barrett JB. 2013. Prevalence and characterization of methicillin-resistant *Staphylococcus aureus* isolates from retail meat and humans in Georgia. *J Clin Microbiol* 51:1199–1207. <https://doi.org/10.1128/JCM.03166-12>.
- Marino M, Frigo F, Bartolomeoli I, Maifreni M. 2011. Safety-related properties of staphylococci isolated from food and food environments. *J Appl Microbiol* 110:550–561. <https://doi.org/10.1111/j.1365-2672.2010.04909.x>.
- Abdalrahman L, Wells H, Fakhr M. 2015. *Staphylococcus aureus* is more prevalent in retail beef livers than in pork and other beef cuts. *Pathogens* 4:182–198. <https://doi.org/10.3390/pathogens4020182>.
- Nemeghaire S, Vanderhaeghen W, Angeles Argudin M, Haesebrouck F, Butaye P. 2014. Characterization of methicillin-resistant *Staphylococcus sciuri* isolates from industrially raised pigs, cattle and broiler chickens. *J Antimicrob Chemother* 69:2928–2934. <https://doi.org/10.1093/jac/dku268>.
- Couto I, Wu SW, Tomasz A, de Lencastre H. 2003. Development of methicillin resistance in clinical isolates of *Staphylococcus sciuri* by transcriptional activation of the *mecA* homologue native to the species. *J Bacteriol* 185:645–653. <https://doi.org/10.1128/jb.185.2.645-653.2003>.
- Piessens V, Van Coillie E, Verbist B, Supré K, Braem G, Van Nuffel A, De Vuyst L, Heyndrickx M, De Vlieghe S. 2011. Distribution of coagulase-negative *Staphylococcus* species from milk and environment of dairy cows differs between herds. *J Dairy Sci* 94:2933–2944. <https://doi.org/10.3168/jds.2010-3956>.
- Nemeghaire S, Argudin MA, Feßler AT, Hauschild T, Schwarz S, Butaye P. 2014. The ecological importance of the *Staphylococcus sciuri* species group as a reservoir for resistance and virulence genes. *Vet Microbiol* 171:342–356. <https://doi.org/10.1016/j.vetmic.2014.02.005>.
- Martineau F, Picard FJ, Roy PH, Ouellette M, Bergeron MG. 1998. Species-specific and ubiquitous-DNA-based assays for rapid identification of *Staphylococcus aureus*. *J Clin Microbiol* 36:618–623.
- Marasini D, Fakhr M. 2014. Exploring PFGE for detecting large plasmids in *Campylobacter jejuni* and *Campylobacter coli* isolated from various retail meats. *Pathogens* 3:833–844. <https://doi.org/10.3390/pathogens3040833>.
- Wattam AR, Davis JJ, Assaf R, Boisvert S, Brettin T, Bun C, Conrad N, Dietrich EM, Disz T, Gabbard JL, Gerdes S, Henry CS, Kenyon RW, Machi D, Mao C, Nordberg EK, Olsen GJ, Murphy-Olson DE, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Vonstein V, Warren A, Xia F, Yoo H, Stevens RL. 2017. Improvements to PATRIC, the all-bacterial bioinformatics database and analysis resource center. *Nucleic Acids Res* 45:D535–D542. <https://doi.org/10.1093/nar/gkw1017>.