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RESEARCH ARTICLE

The effect of interleukin 10 polymorphisms on breast cancer susceptibility in Han women in Shaanxi Province

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Abstract

Background

Previous studies have reported on several genetic variants related to breast cancer, but a substantial proportion of mutation loci have not yet been identified. In the current study, we aimed to evaluate the association between single nucleotide polymorphisms (SNPs) of interleukin-10 (*IL-10*) and susceptibility to breast cancer in Shaanxi Han women in China.

Methods

Six SNPs were genotyped in 530 breast cancer patients and 628 healthy women from the First Affiliated Hospital of Xi'an Jiaotong University Hospital. Odds ratios and 95% confidence intervals were calculated by unconditional logistic regression analysis to assess the association between breast cancer risk and polymorphisms of six loci.

Results

Two SNPs, rs3024490 and rs1800871, were found to be significantly different between breast cancer patients and healthy women. These SNPs also increased the risk of breast cancer in co-dominant and dominant models. Moreover, another SNP, rs1554286, was significantly associated with an increased risk of breast cancer in the co-dominant model. Functional assessments indicated that these three variants may influence the expression and transcription factor binding of *IL-10*.

Competing interests: The authors have declared that no competing interests exist.

Abbreviations: Cls, Confidence intervals; eQTL, Expression quantitative trait loci; ER, estrogen receptor; GTEx, Genotype-tissue expression; GWAS, Genome-wide association study; HER2, human epidermal growth factor receptor 2; HWE, Hardy-Weinberg Equilibrium; IL-10, Interleukin-10; LD, Linkage disequilibrium; MAF, minor allele frequency; ORs, Odds ratios; PR, progesterone receptor; SD, standard deviation; SNPs, Single nucleotide polymorphisms.

Conclusions

Our findings suggest that variants of *IL-10* may be likelihood risk factors for the development and progression of breast cancer. Future studies should replicate this study and evaluate functional assessments in Chinese Han women and women from other regions.

Background

Breast cancer is a life-threatening cancer among women and is the sixth most common cause of death among women in China [1]. The American Cancer Society estimated that there were 252,710 new breast cancer cases in the United States in 2017, and approximately 40,610 deaths in women occurred due to breast cancer [2]. Although numerous environmental factors are well known to cause breast cancer, genetic factors have been confirmed to play a crucial role in the progression of the disease [3, 4].

A growing number of genome-wide association studies have found that variants in genes can significantly affect the susceptibility to breast cancer [5, 6]. However, currently identified single nucleotide polymorphisms (SNPs) can only shed partial insight into the inherited causes of breast cancer, and the precise mechanisms underlying breast cancer remain unclear. Therefore, it is valuable to explore novel SNP loci for assessing the risk of breast cancer. The human interleukin-10 gene (IL-10) is located on chromosome 1q32.1 and contains four introns and five exons [7]. IL-10 is a highly pleiotropic cytokine that is produced by various cells. Studies have shown that IL-10 plays an important role in the initiation and development of breast cancer [8]. Expression of IL-10 is higher in breast cancer patients than in healthy persons [9, 10]. Ahmad et al. [11] demonstrated that IL-10 has an inhibitory effect on the migration of breast cancer cell lines, and this study also found that breast cancer patients with higher IL-10 levels show better disease-free survival and breast cancer-specific survival in the early stages of invasive breast cancer. Thus, IL-10 has been suggested as a potential prognostic marker for breast cancer [12], and there may be a genetic correlation between *IL-10* and breast cancer susceptibility. Several studies [13, 14] have investigated the relationships between IL-10 polymorphisms and breast cancer risk, but have not completely clarified.

Here, we genotyped six SNPs (rs1554286, rs1518111, rs3021094, rs3790622, rs3024490, rs1800871) of IL-10 genes in a case-control study to explore the association between polymorphisms of *IL-10* and the risk of breast cancer in Northwest Chinese Han women. Our data shed new light on the association between IL-10 SNPs and breast cancer susceptibility in Shaanxi Han women in China.

Methods

Ethics approval and consent to participate

All participants were informed of the procedures and purpose of this study. Signed informed consent documents were obtained from both patients and healthy individuals. Study protocols were approved by the Ethical Committee of the First Affiliated Hospital of Xi'an Jiaotong University, and complied with the ethical standards of the Ethical Committee and World Medical Association Declaration of Helsinki. All subsequent research analyses were carried out in accordance with approved guidelines and regulations.

Study participants

The case and control study included 530 breast cancer patients and 628 healthy controls, who were recruited from September 2016 to June 2019 at the First Affiliated Hospital of Xi'an Jiaotong University. The case groups confirmed breast cancer through histological examination, and all cases were diagnosed with a new breast cancer event. Patients diagnosed with other types of cancer and / or receiving hormone therapy, radiotherapy or chemotherapy were excluded. The control groups were healthy women who had undergone an annual health assessment and were recruited from a health checkup center affiliated to our institution. These women did not have any positive signs of cancer and had no history of malignant family history. The case and control groups were all Chinese Han women living in Shaanxi Province, and there was no blood relationship between them.

SNP selection and genotyping

Total genomic DNA was isolated using a GoldMag-Mini Whole Blood Genomic DNA Purification Kit (GlodMag Co. Ltd., Xi'an, China) from peripheral blood, according to the manufacturer's protocol. The Agena Bioscience Assay Design Suite V2.0 software (https://agenacx. com/online-tools/) was used to design the extended primer. SNPs were genotyped using the MassARRAY Nanodispenser (Agena Bioscience, San Diego, CA, USA) and MassARRAY iPLEX platform (Agena Bioscience, San Diego, CA, USA), according to the standard protocol recommended by the manufacturer.

According to the standard protocol recommended by the manufacturer, two laboratory personnel used the MassARRAY Nanodispenser (Agena Bioscience, San Diego, CA, USA) and MassARRAY iPLEX platform (Agena Bioscience, San Diego, CA, USA) to genotype SNPs in a double-blind manner. About 10% of the samples were randomly selected for repeated geno-typing to confirm the results, and the reproducibility was 100%. The Agena Bioscience TYPER software (version 4.0) was used for data management and analyses.

In this study, six SNPs (rs1554286, rs1518111, rs3021094, rs3790622, rs3024490, rs1800871) of *IL-10* were selected from the DbSNP (http://www.hapmap.org/index.html.en) and SNP Consortium (http://snp.cshl.org/) databases based on the following criteria. We took allele frequency into consideration during genotyping. The lower frequency alleles were coded as minor alleles. All SNPs were selected at a minor allele frequency >5% in the 1,000 genome project (http://www.internationalgenome.org/). Chinese Han Beijing population, and the genotype distribution of SNPs in the control group was in accordance with Hardy-weinberg equilibrium (HWE) (p> 0.05). The genotyping of Agena MassARRAY RS1000 for these SNPs exceeded 95.0%.

Statistical analysis

Microsoft Excel and SPSS 18.0 statistical package (SPSS, Chicago, IL, USA) were used for statistical analyses. All *p*-values were based on two-sided tests, and $p \le 0.05$ was considered as the threshold of statistical significance. Welch's t-test was used to compare ages between cases and controls. The validation of each SNP frequency in control subjects was tested for departure from the Hardy-Weinberg Equilibrium (HWE) using an exact test, and the difference in allele frequency distribution among cases and controls was assessed using Pearson's chi-square test. Subsequently, unconditional logistic regression analysis was used to evaluate breast cancer susceptibility under four genotype models (co-dominant, dominant, recessive, and additive model). The SNPStats software platform (https://www.snpstats.net/start.htm) and Haploview software package (version 4.2) [15] were used to perform the linkage disequilibrium (LD) and analyze the association between haplotypes and the risk of breast cancer. The effects of *IL-10*

polymorphisms on the risk of breast cancer were expressed as odds ratios (ORs) and 95% confidence intervals (CIs) [16].

Functional annotation

In silico analysis of breast cancer-associated SNPs on gene expression was assessed using the genotype-tissue expression (GTEx) database of quantitative trait loci (eQTL) variants. This is used to determine the biological effects of genetic variants (http://www.gtexportal.org). Mapping of eQTLs provides a powerful approach to uncover the genetic factors underlying altered gene expression [17].

Regulome DB (http://www.regulomedb.org/) was used to determine the effect of *IL-10* SNPs on allele-specific transcription factor binding. Regulome DB utilizes chromatin immunoprecipitation-sequencing data and chromatin state information across many cell types, as well as eQTL information, for the functional annotation of variants. The variants are scored based on predicted potential effects caused by the variant residing in a functionally important region of the genome. The lower the score the higher impact on protein binding and expression of the target gene.

In addition, HaploReg (http://pubs.broadinstitute.org/mammals/haploreg/haploreg.php) has been used to explore annotations of the noncoding genome at variants on haplotype blocks [18]. We used HaploReg (version 4.1) to assess the rs3024490- and rs1800871-tagged SNPs using LD information from the 1000 Genomes Project with $r^2 \ge 0.8$.

Results

Participant characteristics

A total of 530 breast cancer patients and 628 healthy individuals were enrolled. Demographics and clinical characteristics are shown in Table 1. The mean ages of cases and controls were 50.69 and 51.04 years, respectively. Welch's t-test revealed significant differences in age between cases and controls (p = 0.001).

Variables		Cases (N = 530)	Controls (N = 628)	<i>p</i> -value
Age, years (mean±SD)		50.69 ± 11.74	51.04 ± 9.64	0.001 ^a
Age, years	> 50	276 (52.08%)	336 (53.50%)	
	≤ 50	254 (47.92%)	292 (46.50%)	
Tumor classification	< 2mm	232 (43.77%)		
	$\geq 2mm$	298 (56.23%)		
ER status	Negative	220 (41.51%)		
	Positive	310 (58.49%)		
PR status	Negative	278 (52.45%)		
	Positive	252 (47.55%)		
HER2 status	Negative	208 (39.25%)		
	Positive	322 (60.75%)		
Menopause Status	Pre-menopause	216 (40.75%)		
	Post-menopause	314 (59.25%)		

Table 1. Characteristics of cases and controls.

SD, standard deviation; ER: estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor 2.

^a*p*-value was calculated using Welch's t test.

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SNP ID	Band	Position	Role	Alleles A/B	MAF		<i>p</i> -HWE	OR(95%CI)	^a p-value
					Case	Control			
rs1554286	1q32.1	206944233	Intron (boundary)	G/A	0.358	0.344	0.168	1.07(0.84-1.36)	0.605
rs1518111	1q32.1	206944645	Intron (boundary)	C/T	0.358	0.338	0.043#	1.1(0.86-1.4)	0.456
rs3021094	1q32.1	206944952	Intron	G/T	0.436	0.447	0.022#	0.95(0.76-1.2)	0.692
rs3790622	1q32.1	206945163	Intron	A/G	0.081	0.08	0.429	1.02(0.67-1.57)	0.910
rs3024490	1q32.1	206945311	Intron	C/A	0.355	0.334	0.099	1.09(0.86-1.4)	0.468
rs1800871	1q32.1	206946634	Promoter	G/A	0.355	0.334	0.099	1.09(0.86-1.4)	0.468

Table 2. Allele frequencies in cases and controls and odds ratio estimates for breast cancer.

SNP, single nucleotide polymorphism; MAF, minor allele frequency; HWE, Hardy-Weinberg Equilibrium; ORs, odds ratios; CI, confidence interval; A, minor allele; B, major allele.

[#]Sites with HWE, p < 0.05, are excluded.

^ap-values were calculated using Pearson's Chi-square test.

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Association between IL-10 polymorphisms and breast cancer risk

Detailed SNP data, including the position, allele, and minor allele frequency, are presented in Table 2. Two SNPs, rs1518111 and rs3021094, were not found to be in HWE in the control participants (p < 0.05). Therefore, these two SNPs were excluded from subsequent statistical analyses. We assumed the minor allele of SNPs as the risk allele to analyze the correlation with breast cancer susceptibility. Our results show that polymorphisms of all candidate SNPs were not associated with breast cancer risk (p > 0.05).

Results from the genotype models are shown in Table 3. Rs1554286 was associated with an increased risk of breast cancer in the co-dominant model (OR = 1.52, 95% CI = 1.07–2.16, p = 0.018). After adjusting for age, the OR was 1.51 (95% CI = 1.07–2.15, p = 0.018). Rs3024490 was also associated with an increased risk of breast cancer in the co-dominant model (OR = 1.65, 95% CI = 1.16–2.33, p = 0.004) and dominant model (OR = 1.43, 95% CI = 1.03–2.00, p = 0.033). After adjusting for age, significant associations were found in the co-dominant model (OR = 1.64, 95% CI = 1.16–2.33, p = 0.004) and dominant model (OR = 1.64, 95% CI = 1.16–2.33, p = 0.004) and dominant model (OR = 1.64, 95% CI = 1.16–2.33, p = 0.004) and dominant model (OR = 1.64, 95% CI = 1.16–2.33, p = 0.004) and dominant model (OR = 1.64, 95% CI = 1.16–2.33, p = 0.035). In addition, the co-dominant model (OR = 1.64, 95% CI = 1.16–2.33, p = 0.035) showed associations between rs1800871 and an increased risk of breast cancer in multiple comparisons adjusting for age. No significant associations were found for rs3790622.

Association between haplotypes and breast cancer risk

Finally, LD analysis was used to assess the association between haplotypes and breast cancer susceptibility. Four SNPs (rs1554286, rs3790622, rs3024490 and rs1800871) had a strong LD, and the D' value was 1. This indicated that these four SNPs tended to be co-inherited (Fig 1). Furthermore, the haplotypes were not significantly associated with breast cancer susceptibility (Table 4).

Functional assessment of breast cancer-associated SNPs on IL-10 expression

After the association study, the effects of rs1554286, rs3024490, and rs1800871 on *IL-10* expression were evaluated using the GTEx database (Table 5). We found that these three SNPs

SNP_ID	Model	Genotype	Genotype	Frequencies	Without adjus	tment	With adjustment		
			Case	Control	OR(95%CI)	pa	OR(95%CI)	<i>p</i> b	
rs1554286	Co-dominant	A/A	200 (37.7%)	282 (44.9%)	1	0.018*	1	0.018*	
		A/G	280 (52.8%)	260 (41.4%)	1.52 (1.07-2.16)		1.51 (1.07-2.15)		
		G/G	50 (9.4%)	86 (13.7%)	0.82 (0.47-1.43)		0.82 (0.47-1.42)		
	Dominant	A/A	200 (37.7%)	282 (44.9%)	1	0.081	1	0.084	
		A/G-G/G	330 (62.3%)	346 (55.1%)	1.34 (0.96–1.88)		1.34 (0.96–1.87)		
	Recessive	A/A-A/G	480 (90.6%)	542 (86.3%)	1	0.11	1	0.11	
		G/G	50 (9.4%)	86 (13.7%)	0.66 (0.39–1.11)		0.65 (0.39-1.10)		
	Log-additive				1.07 (0.84–1.36)	0.6	1.07 (0.83-1.36)	0.62	
s3790622	Co-dominant	G/G	444 (84.1%)	267 (85%)	1	0.62	1	0.62	
		A/G	82 (15.5%)	88 (14%)	1.12 (0.71–1.78)		1.12 (0.71–1.78)		
		A/A	2 (0.4%)	6 (1%)	0.40 (0.04-3.88)		0.40 (0.04-3.90)		
	Dominant	G/G	444 (84.1%)	534 (85%)	1	0.76	1	0.75	
		A/G-A/A	84 (15.9%)	94 (15%)	1.07 (0.68–1.69)		1.08 (0.68-1.69)		
	Recessive	G/G-A/G	526 (99.6%)	622 (99%)	1	0.39	1	0.39	
		A/A	2 (0.4%)	6 (1%)	0.39 (0.04-3.81)		0.40 (0.04-3.83)		
	Log-additive				1.02 (0.67–1.57)	0.91	1.03 (0.67-1.57)	0.91	
s3024490	Co-dominant	A/A	200 (37.7%)	292 (46.5%)	1	0.004*	1	0.004*	
		A/C	284 (53.6%)	252 (40.1%)	1.65 (1.16-2.33)		1.64 (1.16-2.33)		
		C/C	46 (8.7%)	84 (13.4%)	0.80 (0.45-1.41)		0.80 (0.45-1.41)		
	Dominant	A/A	200 (37.7%)	292 (46.5%)	1	0.033*	1	0.035*	
		A/C-C/C	330 (62.3%)	336 (53.5%)	1.43 (1.03-2.00)		1.43 (1.02-2.00)		
	Recessive	A/A-A/C	484 (91.3%)	544 (86.6%)	1	0.072	1	0.071	
		C/C	46 (8.7%)	84 (13.4%)	0.62 (0.36-1.05)		0.61 (0.36-1.05)		
	Log-additive				1.10 (0.86-1.40)	0.46	1.09 (0.86-1.40)	0.47	
s1800871	Co-dominant	A/A	200 (37.7%)	292 (46.5%)	1	0.004*	1	0.004*	
		A/G	284 (53.6%)	252 (40.1%)	1.65 (1.16-2.33)		1.64 (1.16-2.33)		
		G/G	46 (8.7%)	84 (13.4%)	0.80 (0.45-1.41)		0.80 (0.45-1.41)		
	Dominant	A/A	200 (37.7%)	292 (46.5%)	1	0.033*	1	0.035*	
		A/G-G/G	330 (62.3%)	336 (53.5%)	1.43 (1.03-2.00)		1.43 (1.02-2.00)		
	Recessive	A/A-A/G	484 (91.3%)	544 (86.6%)	1	0.072	1	0.071	
		G/G	46 (8.7%)	84 (13.4%)	0.62 (0.36-1.05)		0.61 (0.36-1.05)		
	Log-additive				1.10 (0.86-1.40)	0.46	1.09 (0.86-1.40)	0.47	

Table 3. Association between dominant single nucleotide	polymorphisms (SNPs) and the risk of breast cance	r in multiple inheritance models (adjusted by age).

SNP, single nucleotide polymorphism; OR, odds ratio; 95% CI, 95% confidence interval.

^a*p*-values were calculated using unconditional logistic regression analysis.

^b*p*-values were calculated using unconditional logistic regression analysis with adjustments for age.

 $^*p \leq 0.05$ indicates statistical significance.

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variants significantly affected the expression of *IL-10* in whole blood. Furthermore, Regulome DB was used to annotate the possible functional effect. Rs1554286 likely affected eQTL + TF binding / DNase peak and had a Regulome DB score of 1f, which is classified as "Likely to affect binding and linked to expression of a gene target". Rs3024490 had a Regulome DB score of 6, which is classified as "minimal binding evidence". In addition, rs1800871 was likely to affect TF binding + any motif + DNase peak, and it had a Regulome DB score of 3a, which is classified as "less likely to affect binding".

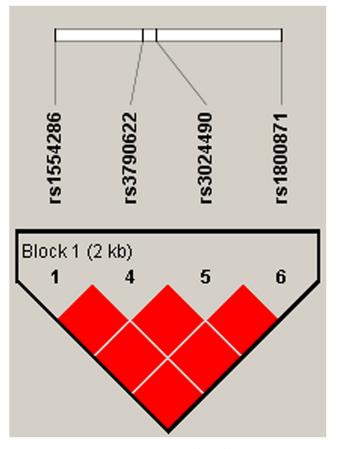


Fig 1. Haplotype block map for part of the (SNPs) in the IL-10 gene. Four SNPs in the haplotype map (rs1554285, rs3790622, rs3024490, and rs1800871) had linkage disequilibrium (LD). A standard color frame is used to show the LD pattern. The D value was 1. These four SNPs tended to be co-inherited.

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Moreover, we used the HaploReg database to get three genetic variants tagged by the rs3024490 and rs1800871 with $r^2 \ge 0.8$. These three polymorphisms were located in *IL-10* or upstream of *IL-10*. Detailed data, including LD information and predicted function, on these variants are shown in Table 6. These results indicated the functional importance of rs1554286, rs3024490, and rs1800871, which were associated with breast cancer.

Haplotype	rs1554286	rs3790622	rs3024490	rs1800871	Freq-case	Freq-control	Without adjustment		With adjustment	
							OR(95%CI)	p ^a	OR(95%CI)	p ^b
1	A	G	A	А	0.560	0.576	1	-	1	-
2	G	G	С	G	0.355	0.334	1.10 (0.85-1.42)	0.48	1.09 (0.85-1.41)	0.49
3	А	А	А	A	0.082	0.080	1.06 (0.68–1.64)	0.80	1.06 (0.68–1.64)	0.80
rare	*	*	*	*	0.003	0.010	0.48 (0.11-2.15)	0.34	0.47 (0.11-2.13)	0.33

Freq, frequency; ORs, odds ratios; CI, confidence interval.

 ${}^{\mathrm{a}}p$ -values were calculated using unconditional logistic regression analysis without adjustment.

^b*p*-values were calculated using unconditional logistic regression analysis with an adjustment for age.

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SNP	NES	<i>p</i> -value	Tissue	Regulome DB Score	Function
rs1554286	0.30	2.2×10 ⁻⁹	Whole Blood	1f	Likely to affect binding and linked to expression of a gene target
rs3024490	0.25	6.7×10^{-5}	Nerve-Tibial	6	Minimal binding evidence
rs3024490	0.28	4.5×10^{-9}	Whole Blood		
rs1800871	0.25	7.3×10 ⁻⁵	Nerve-Tibial	3a	Less likely to affect binding
rs1800871	0.28	4.5×10^{-9}	Whole Blood		

Table 5. Functional annotation results.

NES, Normalized effect size; 1f, eQTL + TF binding / DNase peak; 3a, TF binding + any motif + DNase peak; 6, other.

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Discussion and conclusions

It is known that the polymorphisms of *IL-10* may contribute to the risk of breast cancer. Therefore, we selected six polymorphic sites that were located in the intron or promoter of *IL-10* and investigated their association with breast cancer susceptibility in Shaanxi Han women in China. The results show that the rs1554286 variant was associated with an increased risk of breast cancer in the co-dominant model. Additionally, rs3024490 and rs1800871 were associated with an increased breast cancer risk in co-dominant and dominant models. *In silico* analysis of the SNPs revealed that rs1554286, rs3024490, and rs1800871 may play important roles in the occurrence and development of breast cancer via regulating the expression of the target gene, *IL-10*. Our findings suggest that polymorphisms of *IL-10* may influence the risk of breast cancer among a group of Northwest Chinese Han women.

IL-10 is an important cytokine in immunity and cancer [19]. It is generally speculated that IL-10 plays a role in inhibiting the initiation and development of tumors by activating natural killer cells and cytotoxic T lymphocytes. Surviving tumor cells express IL-10, thus allowing them to escape immune recognition by reducing the expression of MHC class II [20–23]. A large number of association studies have found that polymorphisms of *IL-10* are involved in the susceptibility to many diseases, such as Bechcet's disease [24], systemic lupus erythematosus [25], and ulcerative colitis [26]. Meanwhile, some studies have indicated that IL-10 plays a vital role in the progression, invasion, migration, and growth of breast cancer [27, 28]. These results indicated that polymorphisms of *IL-10* have an effect on the development of breast

Table 6. Breast cancer-associated single nucleotide polymorphisms (SNPs) with variants with $r^2 > 0.8$.

SNP	Chr: Position ^a	LD (r ²)	LD (D')	Allele ^b	Gene	HaploReg
rs1518111	1: 206771300	0.91	1	C/T	IL-10 (intron)	Promoter histone marks, Enhancer histone marks, DNAse, Proteins bound, Motifs changed, NHGRI/EBI GWAS hits, GRASP QTL hits, Selected eQTL hits
rs1518110	1: 206771516	0.89	0.98	C/A	IL-10 (intron)	Promoter histone marks, Enhancer histone marks, DNAse, Motifs changed, Selected eQTL hits
rs3024490	1: 206771966	1	1	C/A	IL-10 (intron)	Promoter histone marks, Enhancer histone marks, Motifs changed, GRASP QTL hits, Selected eQTL hits
rs1800872	1: 206773062	0.99	1	G/T	<i>IL-10</i> (upstream)	Promoter histone marks, Enhancer histone marks, Proteins bound, Motifs changed, GRASP QTL hits, Selected eQTL hits
rs1800871	1: 206773289	1	1	G/A	<i>IL-10</i> (upstream)	Promoter histone marks, Enhancer histone marks, DNAse, Motifs changed, NHGRI/EBI GWAS hits, Selected eQTL hits

SNP, single nucleotide polymorphism; LD, linkage disequilibrium; eQTL, quantitative trait loci; GWAS, Genome-wide association study.

^aBased on NCBI build 37 of the human genome.

^bReference allele/effect allele.

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cancer. Further research is needed to elucidate the relationships among IL-10, tumorigenesis, and progression.

In our study, we found a correlation between the rs1554286, rs3024490, or rs1800871 and the risk of breast cancer in the Northwest Chinese Han women. Several studies have reported a relationship between *IL-10* polymorphisms and breast cancer susceptibility. Slattery et al. reported that allele "G" of rs1554286 and rs1800871 are associated with the risk of breast cancer among women with Native American ancestry [8]. However, a meta-analysis showed that the rs1800871 polymorphism had no relationship with breast cancer risk in Caucasians [29]. Our results were inconsistent with these findings. The rs1800871, which is located in the promoter, may alter the transcriptional regulation of miRNA on *IL-10*. Our functional prediction also suggested that these variants may influence the expression and transcription factor binding of *IL-10*.

No significant results were found for rs1518111, rs3021094, and rs3790622. A variant of rs1518111 has been suggested to correlate with the development of lymphedema following breast cancer treatment [30]. According to SNPInfo analysis, rs3021094 is likely to alter a putative transcription factor binding site for the transcription of *IL-10* [31], and this SNP was significantly associated with poorer overall survival in patients with colorectal cancer [32]. The haplotype "GACC" formed by rs1518111, rs3021094, rs3790622, and rs1800871 was significantly related to the risk of Bechet's disease [33]. However, to date, there is limited information on the association of these three SNPs and the risk of breast cancer. Future studies are needed to confirm our findings in a larger sample size.

Although our study was powered adequately, there were several limitations. First, our study focused on Chinese Han women who lived in Shaanxi province. Second, although we identified significant associations between six SNPs in *IL-10* and breast cancer susceptibility, the mechanisms responsible for these associations remain unclear. In the future, we will want to explore the specific factors which connect the SNPs and the occurrence and development of breast cancer, containing transcriptional regulator and signal pathway.

Overall, this study provided a valuable argument that *IL-10* may be a risk factor for the development and progression of breast cancer. Future studies should replicate these findings and involve women from other regions.

Supporting information

S1 Data. (XLSX)

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Author Contributions

Conceptualization: Miao Li, Xinhan Zhao. Data curation: Miao Li, Chenli Yue, Guoquan Jin. Formal analysis: Chenli Yue. Investigation: Xiaoxiao Zuo. Methodology: Miao Li, Xiaoxiao Zuo, Hulin Guo. Project administration: Xinhan Zhao. Resources: Guoquan Jin, Guanying Wang.

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References

- Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, et al. Cancer statistics in China, 2015. CA: a cancer journal for clinicians. 2016; 66(2):115–32. Epub 2016/01/26. <u>https://doi.org/10.3322/caac.21338</u> PMID: 26808342.
- Siegel RL, Miller KD, Jemal A. Cancer Statistics, 2017. CA: a cancer journal for clinicians. 2017; 67 (1):7–30. Epub 2017/01/06. https://doi.org/10.3322/caac.21387 PMID: 28055103.
- Stephens PJ, Tarpey PS, Davies H, Van Loo P, Greenman C, Wedge DC, et al. The landscape of cancer genes and mutational processes in breast cancer. Nature. 2012; 486(7403):400–4. Epub 2012/06/ 23. https://doi.org/10.1038/nature11017 PMID: 22722201
- Gray J, Druker B. Genomics: the breast cancer landscape. Nature. 2012; 486(7403):328–9. Epub 2012/06/23. https://doi.org/10.1038/486328a PMID: 22722187.
- Zheng W, Zhang B, Cai Q, Sung H, Michailidou K, Shi J, et al. Common genetic determinants of breastcancer risk in East Asian women: a collaborative study of 23 637 breast cancer cases and 25 579 controls. Human molecular genetics. 2013; 22(12):2539–50. Epub 2013/03/29. <u>https://doi.org/10.1093/</u> hmg/ddt089 PMID: 23535825
- Michailidou K, Beesley J, Lindstrom S, Canisius S, Dennis J, Lush MJ, et al. Genome-wide association analysis of more than 120,000 individuals identifies 15 new susceptibility loci for breast cancer. Nature genetics. 2015; 47(4):373–80. Epub 2015/03/10. https://doi.org/10.1038/ng.3242 PMID: 25751625
- Eskdale J, Kube D, Tesch H, Gallagher G. Mapping of the human IL10 gene and further characterization of the 5' flanking sequence. Immunogenetics. 1997; 46(2):120–8. Epub 1997/01/01. <u>https://doi.org/ 10.1007/s002510050250 PMID: 9162098.</u>
- Slattery ML, Herrick JS, Torres-Mejia G, John EM, Giuliano AR, Hines LM, et al. Genetic variants in interleukin genes are associated with breast cancer risk and survival in a genetically admixed population: the Breast Cancer Health Disparities Study. Carcinogenesis. 2014; 35(8):1750–9. Epub 2014/03/ 29. https://doi.org/10.1093/carcin/bgu078 PMID: 24670917
- Llanes-Fernandez L, Arango-Prado Mdel C, Alcocer-Gonzalez JM, Guerra-Yi ME, Franco-Odio S, Camacho-Rodriguez R, et al. Association between the expression of IL-10 and T cell activation proteins loss in early breast cancer patients. Journal of cancer research and clinical oncology. 2009; 135 (2):255–64. Epub 2008/07/25. https://doi.org/10.1007/s00432-008-0446-7 PMID: 18651178.
- Heckel MC, Wolfson A, Slachta CA, Schwarting R, Salgame P, Katsetos CD, et al. Human breast tumor cells express IL-10 and IL-12p40 transcripts and proteins, but do not produce IL-12p70. Cellular immunology. 2011; 266(2):143–53. Epub 2010/11/09. https://doi.org/10.1016/j.cellimm.2010.09.010 PMID: 21055733.
- Ahmad N, Ammar A, Storr SJ, Green AR, Rakha E, Ellis IO, et al. IL-6 and IL-10 are associated with good prognosis in early stage invasive breast cancer patients. Cancer Immunology Immunotherapy. 2018; 67(4):1–13. https://doi.org/10.1007/s00262-017-2106-8 PMID: 29256156
- Gerger A, Renner W, Langsenlehner T, Hofmann G, Knechtel G, Szkandera J, et al. Association of interleukin-10 gene variation with breast cancer prognosis. Breast cancer research and treatment. 2010; 119(3):701–5. Epub 2009/05/14. https://doi.org/10.1007/s10549-009-0417-y PMID: 19437115.
- Yu KD, Chen AX, Yang C, Fan L, Huang AJ, Shao ZM. The associations between two polymorphisms in the interleukin-10 gene promoter and breast cancer risk. Breast Cancer Res Treat. 2012; 131(1):27– 31. Epub 2010/08/26. https://doi.org/10.1007/s10549-010-1133-3 PMID: 20737205.
- Pooja S, Chaudhary P, Nayak LV, Rajender S, Saini KS, Deol D, et al. Polymorphic variations in IL-1β, IL-6 and IL-10 genes, their circulating serum levels and breast cancer risk in Indian women. Cytokine. 2012; 60(1):122–8. Epub 2012/07/24. https://doi.org/10.1016/j.cyto.2012.06.241 PMID: 22818022.
- Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics (Oxford, England). 2005; 21(2):263–5. Epub 2004/08/07. <u>https://doi.org/10.1093/bioinformatics/bth457</u> PMID: 15297300.

- Bland JM, Altman DG. Statistics notes. The odds ratio. BMJ (Clinical research ed). 2000; 320 (7247):1468. Epub 2000/05/29. https://doi.org/10.1136/bmj.320.7247.1468 PMID: 10827061
- Consortium. G. The Genotype-Tissue Expression (GTEx) project. Nature genetics. 2013; 45(6):580–5. Epub 2013/05/30. https://doi.org/10.1038/ng.2653 PMID: 23715323
- Ward LD, Kellis M. HaploReg v4: systematic mining of putative causal variants, cell types, regulators and target genes for human complex traits and disease. Nucleic acids research. 2016; 44(D1):D877– 81. Epub 2015/12/15. https://doi.org/10.1093/nar/gkv1340 PMID: 26657631
- Saraiva M, O'Garra A. The regulation of IL-10 production by immune cells. Nature reviews Immunology. 2010; 10(3):170–81. Epub 2010/02/16. https://doi.org/10.1038/nri2711 PMID: 20154735.
- Sato T, Terai M, Tamura Y, Alexeev V, Mastrangelo MJ, Selvan SR. Interleukin 10 in the tumor microenvironment: a target for anticancer immunotherapy. Immunologic research. 2011; 51(2–3):170–82. Epub 2011/12/06. https://doi.org/10.1007/s12026-011-8262-6 PMID: 22139852.
- 21. Dennis KL, Blatner NR, Gounari F, Khazaie K. Current status of interleukin-10 and regulatory T-cells in cancer. Current opinion in oncology. 2013; 25(6):637–45. Epub 2013/10/01. <u>https://doi.org/10.1097/</u>CCO.0000000000000 PMID: 24076584
- Ostapchuk YO, Cetin EA, Perfilyeva YV, Yilmaz A, Skiba YA, Chirkin AP, et al. Peripheral blood NK cells expressing HLA-G, IL-10 and TGF-beta in healthy donors and breast cancer patients. Cellular immunology. 2015; 298(1–2):37–46. Epub 2015/09/13. <u>https://doi.org/10.1016/j.cellimm.2015.09.002</u> PMID: 26362675.
- Alotaibi MR, Hassan ZK, Al-Rejaie SS, Alshammari MA, Almutairi MM, Alhoshani AR, et al. Characterization of Apoptosis in a Breast Cancer Cell Line after IL-10 Silencing. Asian Pacific journal of cancer prevention: APJCP. 2018; 19(3):777–83. Epub 2018/03/28. https://doi.org/10.22034/APJCP.2018.19. 3.777 PMID: 29582634
- Mizuki N, Meguro A, Ota M, Ohno S, Shiota T, Kawagoe T, et al. Genome-wide association studies identify IL23R-IL12RB2 and IL10 as Behcet's disease susceptibility loci. Nature genetics. 2010; 42 (8):703–6. Epub 2010/07/14. https://doi.org/10.1038/ng.624 PMID: 20622879.
- Gateva V, Sandling JK, Hom G, Taylor KE, Chung SA, Sun X, et al. A large-scale replication study identifies TNIP1, PRDM1, JAZF1, UHRF1BP1 and IL10 as risk loci for systemic lupus erythematosus. Nature genetics. 2009; 41(11):1228–33. Epub 2009/10/20. <u>https://doi.org/10.1038/ng.468</u> PMID: 19838195
- Franke A, Balschun T, Karlsen TH, Sventoraityte J, Nikolaus S, Mayr G, et al. Sequence variants in IL10, ARPC2 and multiple other loci contribute to ulcerative colitis susceptibility. Nature genetics. 2008; 40(11):1319–23. Epub 2008/10/07. https://doi.org/10.1038/ng.221 PMID: 18836448.
- Valle Oseguera CA, Spencer JV. Human cytomegalovirus interleukin-10 enhances matrigel invasion of MDA-MB-231 breast cancer cells. Cancer cell international. 2017; 17:24. Epub 2017/02/24. <u>https://doi.org/10.1186/s12935-017-0399-5</u> PMID: 28228690
- Liu C, Sun B, Xu B, Meng X, Li L, Cong Y, et al. A panel containing PD-1, IL-2Ralpha, IL-10, and CA15-3 as a biomarker to discriminate breast cancer from benign breast disease. Cancer management and research. 2018; 10:1749–61. Epub 2018/07/10. https://doi.org/10.2147/CMAR.S160452 PMID: 29983594
- Dai ZJ, Wang XJ, Zhao Y, Ma XB, Kang HF, Min WL, et al. Effects of interleukin-10 polymorphisms (rs1800896, rs1800871, and rs1800872) on breast cancer risk: evidence from an updated meta-analysis. Genetic testing and molecular biomarkers. 2014; 18(6):439–45. Epub 2014/04/12. https://doi.org/ 10.1089/gtmb.2014.0012 PMID: 24720854.
- Leung G, Baggott C, West C, Elboim C, Paul SM, Cooper BA, et al. Cytokine candidate genes predict the development of secondary lymphedema following breast cancer surgery. Lymphatic research and biology. 2014; 12(1):10–22. Epub 2014/02/08. https://doi.org/10.1089/lrb.2013.0024 PMID: 24502445
- Xu Z, Taylor JA. SNPinfo: integrating GWAS and candidate gene information into functional SNP selection for genetic association studies. Nucleic acids research. 2009; 37(suppl_2):W600–W5. Epub 2009/ 05/07. https://doi.org/10.1093/nar/gkp290 PMID: 19417063
- Ting WC, Chen LM, Huang LC, Hour MJ, Lan YH, Lee HZ, et al. Impact of interleukin-10 gene polymorphisms on survival in patients with colorectal cancer. Journal of Korean medical science. 2013; 28 (9):1302–6. Epub 2013/09/10. https://doi.org/10.3346/jkms.2013.28.9.1302 PMID: 24015034
- Wu Z, Zheng W, Xu J, Sun F, Chen H, Li P, et al. IL10 polymorphisms associated with Behcet's disease in Chinese Han. Human immunology. 2014; 75(3):271–6. Epub 2013/11/26. <u>https://doi.org/10.1016/j.</u> humimm.2013.11.009 PMID: 24269690.