

## **Optical coherence tomography angiography in retinitis pigmentosa**

### A narrative review

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#### Abstract

Retinitis pigmentosa (RP) is a group of inherited retinal disorders characterized by progressive rod and cone photoreceptor degeneration. Changes in retinal vasculature have long been associated with RP. Optical coherence tomography angiography (OCTA) is a novel imaging technology that enables noninvasive visualization of the retinal and choroidal microvasculature. OCTA enables quantification of microvascular changes in the retinal capillary plexus and choriocapillaris, in addition to qualitative feature description. Therefore, OCTA has the potential to become an important tool for better understanding, early detection, progression, and treatment of RP. In this review, we focus on the applications of OCTA in clinical research on RP. We also discuss future improvements in the OCTA technology for RP management. We believe that the advancement of the OCTA technique will ultimately lead to a better understanding of RP and aid in the prevention of visual impairment.

**Abbreviations:** CCP = choriocapillaris plexus, CD = cone density, CDFI = color Doppler flow imaging, CRMBs = computational retinal microvascular biomarkers, CSI = choroidal stromal index, DCP = deep capillary plexus, FA = fluorescein angiography, FAF = fundus autofluorescence, FAZ = foveal avascular zone, FVs = flow voids, ICP = intermediate capillary plexus, IS/OS = inner/outer segment, MRI = magnetic resonance imaging, OCT = optical coherence tomography, OCTA = optical coherence tomography angiography, OCTARA = OCTA ratio analysis, OMAG = OCT-based microangiography, OMAGC = optical micro angiography complex, PD = perfusion density, PR-OCTA = projection-resolved OCTA, RGCs = retinal ganglion cells, RNFL = retinal nerve fiber layer, RP = retinitis pigmentosa, RPC = radial peripapillary capillary, RPE = retinal pigment epithelium, SCP = superficial capillary plexus, SSADA = split-spectrum amplitude decorrelation angiography, SVC = superficial vascular complex, UWF-CP = ultra-wide-field color fundus photography, UWF-FA = ultra-wide field fluorescein angiography, VDI = vessel diameter index, VDisp = vessel dispersion, VLD = vessel length density, VR = vessel rarefaction, VT = vessel tortuosity, WF-OCTA = wide-field OCTA

**Keywords:** macular microvasculature, optical coherence tomography angiography, retinal biomarkers, retinal function, retinitis pigmentosa, vessel density

### 1. Introduction

Retinitis pigmentosa (RP) is a group of genetically diverse inherited retinal dystrophies characterized by progressive degeneration of rod and cone photoreceptors.<sup>[1]</sup> RP patients usually suffer from impaired dark adaption, night blindness, visual field constriction, and central vision deterioration around 40 years of age.<sup>[2,3]</sup> The relationship between ocular hemodynamics and RP has not been fully understood. Previous studies have confirmed the reductions in blood flow in RP patients,<sup>[4]</sup> questions have yet to be answered regarding the role of vascular dysfunction, as well as vascular abnormalities in the foveal and parafoveal regions in the degeneration of photoreceptors. Owing to recent advances in imaging technology, optical coherence tomography angiography (OCTA) enables non-invasive visualization and quantitative assessment of the retinal and choroidal microvasculature, which shows great potential in providing diagnostic, prognostic, and perhaps therapeutic biomarkers of ocular hemodynamics in RP patients.<sup>[5]</sup>

Our purpose is to conduct a database search of all published studies that focus on the use of OCTA in clinical research on RP to better understand this retinal disorder. Further, we envision that advancements in OCTA technology could help clinicians in RP diagnosis, follow-up, and treatment in the near future.

### 2. Methods

A literature review was performed using PubMed, including all original studies registered until February 2021. Using the

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The datasets generated during and/or analyzed during the current study are not publicly available but are available from the corresponding author on reasonable request.

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keyword "RP" and "OCTA", 38 relevant publications published from 2016 to 2021 were retrieved.

Ethics committee approval was not requested because it is not needed for narrative reviews of the literature.

## 3. Current methods for retinal vasculature evaluation in RP

Evaluation of retinal vasculature was historically dependent on fluorescein angiography (FA), which could demonstrate abnormal retinal and choroidal vasculature in patients with RP, including prolonged transit time, narrowed vessels, and lower dye concentration.<sup>[6]</sup> However, FA is invasive and has been reported to cause hepatic, renal, or allergic complications due to the need for intravenous injection and the possibility of leakage of dye.<sup>[7]</sup> Fundus autofluorescence (FAF) is also useful in evaluating the retinal status in RP patients. Photoreceptor loss corresponds to hypo-autofluorescence in the peripheral fundus, whereas areas of hyper-autofluorescence correspond to increased lipofuscin in the retinal pigment epithelium (RPE).<sup>[8]</sup> However, FAF produces a lower signal strength than FA and is monochromatic which does not produce multicolored images.<sup>[9]</sup> Optical coherence tomography (OCT) has been used to evaluate RP to reveal decreased foveal thickness, interruption of the photoreceptor inner/outer segment (IS/OS) junction, and significantly reduced choroidal thickness.<sup>[4,10]</sup> OCT structural changes correlate with the deterioration of sight.<sup>[11]</sup> While these measurements are useful, OCT does not possess the ability to visualize microvascular changes during RP progression. The above modalities provide information about the structures of interest but do not provide details of vascular structure and blood flow.

OCTA is a recent technological advancement that allows for the acquisition of high-resolution, depth-resolved retinal images of both retinal and choroidal vascular layers in a rapid, non-invasive manner.<sup>[12]</sup> With the advantages of differentiating the superficial capillary plexus (SCP), deep capillary plexus (DCP), and choriocapillaris plexus (CCP), OCTA has been broadly applied in the early detection of vascular abnormalities and diagnosis of vascular pathology of many inherited retinal dystrophies, including RP.<sup>[13]</sup> It can also be used to provide a quantitative assessment of the microcirculation and microvasculature of the retina and choroid in various layers.<sup>[14]</sup> Since RP development and progression are both associated with retinal and choroidal vascular changes (as either a primary or a secondary effect), this technology has the potential to bring forward new information about the pathophysiology of RP, as well as to help clinicians with RP diagnosis and management.

#### 4. Applications of OCTA in RP

#### 4.1. Terminology in OCTA

Various OCTA algorithms have been used by different commercially available OCTA devices: the split-spectrum amplitude-decorrelation angiography (SSADA) algorithm, the optical microangiography complex (OMAGC) algorithm, the OCTbased microangiography (OMAG) algorithm, the OCTA ratio analysis (OCTARA) algorithm, and the amplitude decorrelation algorithm (Table 1).

Several areas of the retina were assessed in the selected studies (Fig. 1). Macular scans were centered on the fovea (Fig. 1A). The "whole image" macular was defined as the whole surface of the scan (generally  $3 \times 3$  or  $6 \times 6$  mm). The fovea was defined as the central 1-mm circle on the macular scan. The parafovea was defined as the central 3-mm circle on

#### Table 1

Algorithms used by various OCTA machines

		Commercia	alized OCTA Machines		
	AngioVue	AngioPlex	SSOCT Angio	Spectralis OCTA	AngioScan
Company Algorithm	OptoVue SSADA	Zeiss OMAGC	Topcon OCTARA	Heidelberg Amplitude decorrelation	Nidex Modified OMAG
A-scans/s Image dimensions	70,000 3 × 3 mm, 4.5 × 4.5 mm,	68,000 3×3mm,	100,000 3×3mm, 4.5×4.5mm,	85,000 10°×10,	53,000 3 × 3 mm, 6 × 6 mm,
-	6×6mm, 8×8mm, 6×10 mm	6×6 mm	6×6 mm	15°×15°, 20°×20°, 15°×30°	9×9mm, 12×9 mm



Figure 1. OCTA images of the macular area and the optic disc. (A) OCTA image (OptoVue) of the superficial vascular network centered on the fovea in a healthy subject. (B) OCTA image of the same subject depicting the FAZ area. (C) OCTA image (Zeiss) of the peripapillary area centered on the optic nerve head in a healthy subject. OCTA = optical coherence tomography angiography, FAZ = foal avascular zone.

the macular scan, except for the fovea, whereas the perifovea was defined as the central 6-mm circle on the macular scan, except for the fovea. The foveal avascular zone (FAZ) was defined as a round capillary-free zone within the macula on OCTA images of the superficial vascular network (Fig. 1B). Optic disc scans were centered on the optic disc (Fig. 1C). The neural canal opening, which terminates the RPE/Bruch membrane complex, was used to define the optic disc area. The peripapillary area was used to describe both the circumpapillary and whole-image peripapillary areas. The whole image peripapillary scan was defined as the entire area of the optic disc scan. Scans assessing the optic disc, the peripapillary area, and the whole image peripapillary area were generally  $4.5 \times 4.5$  mm wide.

#### 4.2. OCTA analyses in RP

Thirty-eight published papers evaluating the role of OCTA in RP were identified through a literature search using PubMed. Eighteen cross-sectional clinical studies were chosen after scrutiny (Dr. Lu and Dr. Chiu). For an overview of all these articles, see Table 2.

#### 4.3. Vessel density (VD) analysis in RP

4.3.1. Retinal blood flow. The pathogenesis of RP is quite complex and is mainly related to genetic alterations at the photoreceptor and RPE cell levels, ultimately leading to retinal degeneration.<sup>[6]</sup> However, the importance of the involvement of both the inner retina and vascular supply has been increasingly recognized in recent years.<sup>[15]</sup> Photoreceptor cell loss with the reduction of oxygen consumption has been suggested, and thus decreased the need for oxygen delivery from the retinal circulation in the pathology of eyes with RP. Such changes in oxygen diffusion are assumed to cause attenuation of the vessels.<sup>[16]</sup> Histopathological studies showed that the features of RP included vessel narrowing and sclerosis, followed by thickening of the blood vessel wall and lumen occlusion.<sup>[17]</sup> In agreement with these histopathological findings, abundant evidence has proved that vascular changes (e.g. perivascular cuffing, arteriolar attenuation, and reduced ocular blood flow) feature RP which was hypothesized to be part of the pathogenic process with advanced technologies.<sup>[4]</sup> Reduced retinal blood flow velocity and vascular diameter were demonstrated with the use of magnetic resonance imaging (MRI)<sup>[18]</sup> and color Doppler flow imaging (CDFI)<sup>[19]</sup> in RP patients. However, each of these techniques has its limitations, such as being qualitative or invasive.

OCTA parameters have been demonstrated to enable earlier detection of circulatory alterations compared to other conventional methods.<sup>[19]</sup> Moreover, OCTA has potential advantages over conventional techniques for assessing retinal terminal vessels. Previous studies showed that both SCP and DCP vessel densities are significantly decreased in early-[20,21] and middle- and late-stage RP<sup>[22]</sup> after comparison with healthy objects. Moreover, a more profound involvement of the deep layer with reduced retinal vasculature signal was found (parafoveal VD: DCP, P=.001; SCP, P=.009).<sup>[20]</sup> This finding was later proved by one study that enrolled 110 eyes of RP and 32 control eyes (Parafoveal VD: DCP, P<.001; SCP, P=.66).<sup>[23]</sup> Another study demonstrated that the most severe vascular impairment occurred in the parafoveal flow area (DCP, P=.004; SCP, P=.007).<sup>[24]</sup> Most recently, one study also showed that vascular alteration in RP might begin at the level of DCP, while the change in the SCP would occur later in the evolution of the disease.<sup>[25]</sup>

However, these conventional OCTA images suffer from projection artifacts, which limit the ability to accurately separate and quantify microvasculature into the three distinct macular vascular layers corresponding to histologic studies<sup>[26,27]</sup>: superficial vascular complex (SVC), intermediate capillary plexus (ICP), and DCP. Advanced projection-resolved OCTA (PR-OCTA) with improved vascular segmentation metrics is required for better visualization and illustration. One study group found that deeper retinal plexuses (both ICP and DCP) were primarily damaged by RP, compared to SVC using the self-developed PR-OCTA algorithm.<sup>[28]</sup>

4.3.2. Choroidal blood flow. Choroidal vascular features are important in RPE atrophic disorders because of their role in oxygen and nutrition supply as well as metabolic exchanges to the outer retina and RPE. Early histopathology studies have shown the absence of choriocapillaris in RP, which has lost photoreceptors and forms bone-spicule pigments.[17] However, when analyzing the blood flow at the CCP layer, there are controversies in the findings of different studies. Vessel densities of CCP in middle- and late-stage RP patients were reported to be remarkably lower than those in healthy subjects (P=.024),<sup>[22]</sup> while other studies reported no differences in CCP vessel densities between RP patients and normal controls.<sup>[20,24]</sup> These discrepancies could be explained by the limitations of conventional OCTA devices that suffer from projection artifacts and penetration depth. With advanced OCTA technology, accurate choroidal changes can be detected by SS-OCTA which has a light source of 1050nm. Guduru et al demonstrated that flow voids (FVs) in patients with RP were significantly reduced compared to healthy subjects (FV area: P < .01), indicating that the compromised choriocapillaris might be a result or cause of RP.<sup>[29]</sup> Manual segmentation lines for OCTA images were added to separate deep choroids at different levels (L1, L2, and L3) from Bruch's membrane to obtain more precise data on the whole choroid in one study,[30] and a more significant reduction in vascular density was found in L1, which could be explained by subsequent retinal degeneration. Recently, wide-angle OCTA (8×8mm or 12×12mm) has been applied to investigate choriocapillaris defects, for the fact that small angle OCTA  $(3 \times 3 \text{ mm or } 6 \times 6 \text{ mm})$  failed to detect differences in choriocapillaris in RP patients since the peripheral retina is more likely to be affected at the early stage.<sup>[1,31]</sup>

4.3.3. Blood flow in optic disc and peripapillary area. The radial peripapillary capillary (RPC) network is a unique vascular plexus around the optic disc within the retinal nerve fiber layer (RNFL). It originates from the central retinal artery and branches out and runs parallel to the axon direction of the retinal ganglion cells (RGCs). Therefore, it is believed that the blood flow of the RPC, as well as the thickness of the RNFL around the optic disc, is reduced in RP patients, although the specific mechanism has not been clarified. In particular, the biomicroscopic aspect of the "pale" optic disc, together with its typical vascular attenuation, suggests the presence of vascular damage in the optic disc and peripapillary area. With the newly developed OCTA technique, the RPC network can be visualized separately. In one cross-sectional study that included 11 RP patients (22 eyes) and 16 age-matched control subjects (16 eyes), RPC vessel densities in the optic disc and peripapillary areas were found to be significantly reduced in RP patients along with a reduction in RNFL thickness, indicating that vascular changes in these areas might be associated with RGC death and RNFL thinning, which are the two pathological processes accompanying inner retinal disorganization.<sup>[32]</sup>

#### 4.4. Macular microvasculature analysis in RP

Many studies have suggested that macular microvasculature changes caused by decreased blood flow might be indicated in the development of RP and the pathogenesis of RP in the macular area, particularly in the final loss of central vision. It has been confirmed that there is a reduction in macular vascular density in the eyes of patients with RP, as shown by different

Clinical cross-sectional	studies u	Ising OC	TA in RP patients						
	Groups	and # of				Summ	arv of the Besults		
Article	r   <b>6</b>	Control	OCTA Instrument	Scan Area	<b>OCTA</b> Parameter	RP	Control	P value	Functional Test
Vessel Density Analysis Toto et al (2016) <sup>[22]</sup>	28 eves		RTV/ne-XR Avanti	3 × 3 mm (fovea)	Vessel density (SCP)-parafovea	42 2 + 3 4%	51 4 + 2 3%	< 001	mfFRG
	2010				Vessel density (DCP)-parafovea	$42.7 \pm 6.2\%$	$56.6 \pm 2.2\%$	<.001	2
					Vessel density (CCP)-parafovea	$65.3 \pm 2.7\%$	$67.2 \pm 1.4\%$	=.024	
Battaglia Parodi et al. (2017) <sup>[20]</sup>	32 eyes	30 eyes	Swept-source OCT DRI Topcon Triton	$3 \times 3 \mathrm{mm}$ (fovea)	Vessel density (SCP)-parafovea	$29.5 \pm 6.8\%$	$34.1 \pm 4.3\%$	=.009	/
					Vessel density (DCP)-parafovea	$28.7 \pm 7.5\%$	$35.5 \pm 4.3\%$	=.001	
					Vessel density (CCP)-parafovea	$51.0 \pm 4.4\%$	$51.3 \pm 2.2\%$	=.716	
					FAZ area (SCP)	$0.277 \pm 0.133  \text{mm}^2$	$0.277 \pm 0.133  \text{mm}^2$	=.350	
					FAZ area (DCP)	$0.541 \pm 0.211  \text{mm}^2$	$0.243 \pm 0.157  \text{mm}^2$	<.001	
Sugahara et al. (2017) <sup>[23]</sup>	110 eyes	32 eyes	RTVue-XR Avanti	3 × 3 mm (fovea)	Vessel density (SCP)-parafovea	$47.0 \pm 4.9\%$	/	=.66	ERG
					Vessel density (DCP)-parafovea	$52.4 \pm 5.5\%$	/	<.001	
					Vessel density (CCP)-parafovea	$51.0 \pm 4.4\%$	/	=.46	
					FAZ area (SCP)	$0.342 \pm 0.198 \mathrm{mm^2}$	/	=.003	
					FAZ area (DCP)	$0.429 \pm 0.154 \text{mm}^2$	/	=.45	
Mastropasqua et al. (2017) <sup>[32]</sup>	19 eyes	16 eyes	RTVue-XR Avanti	$4.5 \times 4.5 \text{ mm}(\text{optic})$	RPC vessel density-disc	$46.5 \pm 7.1\%$	$45.4 \pm 10.6\%$	=.754	/
-					RPC vessel density-peripapillary	$52.5 \pm 5.0\%$	$57.2 \pm 5.1\%$	=.011	
Takagi et al. (2018) <sup>[24]</sup>	50 eyes	22 eyes	RTVue-XR Avanti	$3 \times 3 \mathrm{mm}$ (fovea)	Flow area (SCP)-parafovea	$3.99 \pm 0.38 \mathrm{mm^2}$	$4.32 \pm 0.27  \text{mm}^2$	=.007	Visual field
					Flow area (DCP)-parafovea	$4.06 \pm 0.71 \text{ mm}^2$	$4.44 \pm 0.37  \text{mm}^2$	=.004	
					Flow area (CCP)-parafovea	$5.43 \pm 0.17  \text{mm}^2$	$5.47 \pm 0.13  \text{mm}^2$	=.353	
					FAZ area (SCP)	$0.30 \pm 0.09  \text{mm}^2$	$0.36 \pm 0.07  \text{mm}^2$	=.006	
					FAZ area (DCP)	$0.41 \pm 0.13  \text{mm}^2$	$0.42 \pm 0.09  \text{mm}^2$	=.237	
Koyanagi et al. (2018) <sup>[21]</sup>	73 eyes	36 eyes	RTVue-XR Avanti	$3 \times 3 \mathrm{mm}$ (fovea)	Flow density (SCP)-fovea	27.1 (11.0–45.8)%	29.1 (22.2–40.1)%	=.309	Visual field
					Flow density (DCP)-fovea	24.5 (8.32-45.8)%	24.7 (17.7–34.1)%	=.757	
					Flow density (SCP)-parafovea	43.8 (34.6–54.6)%	54.7 (41.0–61.1)%	<.001	
					Flow density (DCP)-parafovea	50.1 (39.7–61.1)%	61.7 (55.0–65.5)%	<.001	
					FAZ area (SCP)	0.231 (0.08–1.048) mm <sup>2</sup>	0.225 (0.089-0.371) mm <sup>2</sup>	=0.309	
					FAZ area (DCP)	0.240 (0.085–1.102) mm <sup>2</sup>	0.249 (0.109–0.451) mm <sup>2</sup>	=.890	
Guduru et al. (2018) <sup>[29]</sup>	70 eyes	37 eyes	Swept-source OCT DRI Topcon Triton	6 × 6 mm (fovea)	Flow voids number	$55.5 \pm 20.1$	$30.7 \pm 16.3$	<.01	/
					Flow voids area	$0.33 \pm 0.12  \text{mm}^2$	$0.18 \pm 0.10  \text{mm}^2$	<.01	
Wang et al., 2019 <sup>[19]</sup>	40 eyes	26 eyes	Cirrus HD-OCT 5000	3 × 3 mm (fovea)	Vessel area density-fovea	$20.5 \pm 5.4\%$	$27.5 \pm 5.5\%$	<.01	/
					Vessel area density-temporal	$35.5 \pm 4.2\%$	$45.1 \pm 1.8\%$	<.01	
					Vessel area density-superior	$36.9 \pm 3.8\%$	$46.6 \pm 1.8\%$	<.01	
					Vessel area density-inferior	$36.7 \pm 4.2\%$	$45.9 \pm 1.9\%$	<.01	
					Vessel area densitv-nasal	$36.9 \pm 3.8\%$	$45.8 \pm 1.8\%$	<.01	

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(Continued)

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Table2 (Continued)									
	Groups Ey	and # of res				Summar	ry of the Results		
Article	RP	Control	<b>OCTA Instrument</b>	Scan Area	OCTA Parameter	RP	Control	<i>P</i> value	Functional Test
Hagag et al. (2019) <sup>[28]</sup>	44 eyes	34 eyes	RTVue-XR Avanti	6 × 6 mm (fovea)	Vessel density (SVC)-parafovea Without CME With CME	68.42±11.27% 71.19±5.42%	$65.76 \pm 7.4\%$ $65.76 \pm 7.4\%$	=.48 =.005	~
					Vessel density (SVC)-perifovea Without CME With CME	$65.86 \pm 4.7\%$ $66.07 \pm 5.7\%$	$65.74 \pm 5.24\%$ $65.74 \pm 5.24\%$	=.56 =.83	
					vessel density (Ler)-paratovea Without CME With CME Vessel density (ICP)-perifovea	47.65±9.71% 47.89±6.25	$50.46 \pm 5.65\%$ $50.46 \pm 5.65\%$	=.24 =.11	
					without Civie With CME Vessel density (DCP)-parafovea Without CME	43./4±/.26 39.87±4.73 21.12±6.84	40.73±7.1% 46.73±7.1% 21.16+4.04%	=.14 <.001 - 76	
					With CME With CME Vessel density (DCP)-perifovea	20.79±6.76	21.16±4.04%	=. 	
					Without CME With CME Outer retinal thickness-parafovea	$17.74 \pm 7.72$ 13.10 ± 4.90	25.87 ± 5.71% 25.87 ± 5.71%	<.001 <.001	
					Without CME With CME Outer retinal thickness-perifovea	119.15 ± 32.18 µm 79.56 ± 26.66 µm	147.91 ± 10.53 μm 147.91 ± 10.53 μm	<.001 <.001	
					Without CME With CME Under retinal thickness-parafovea	84.73±28.21 µm 43.79±13.95 µm	133.08 ± 8.05 µm 133.08 ± 8.05 µm	<.001 <.001	
					Without a kine way to be account with the ME With CME Inner retinal thickness-perifovea	192.24 ± 22.10 µm 227.87 ± 33.39 µm	179.36 ±10.86 μm 179.36 ±10.86 μm	<.08 <.001	
					Without CME With CME	175.58 ± 22.59 μm 188.3 ± 27.47 μm	150.59 ± 10.44 µm 150.59 ± 10.44 µm	<.001 <.001	
Miyata et al. (2019) <sup>[31]</sup>	43 eyes	12 eyes	PLEX Eite 9000	12×12mm (fovea)	Residual choroicapillaris area Concentric group	$44.7 \pm 20.2 \text{ mm}^2$	$144.0 \pm 0 \mathrm{mm^2}$	<.001	Visual field
Falfoul et al. (2020) <sup>(25)</sup>	70 eyes	34 eyes	Swept-source OCT DRI Topcon Triton	3 × 3 mm (fovea)	Vermicular youp Vessel density (SCP)-parafovea Vessel density (DCP)-parafovea FAZ area (SCP)	1.24. 1 ± 19.111111 <sup>-</sup> 25.99 ± 5.3% 25.04 ± 5.53% 369 69 + 142 11m <sup>2</sup>	$144.0 \pm 01000$ 29.74 ± 3% $34.47 \pm 2.37\%$ 312 + 119.002	=.002 =0.0026 <.001	mfERG
Corazza et al. (2020) <sup>(30)</sup>	40 eyes	24 eyes	Swept-source OCT DRI Topcon Triton	4.5×4.5 mm (fovea)	FAZ area (DCP) Vascular density (superficial) Vascular density (deep) Vascular density (choriocapillaris) Secondariation fine 1	575.96 ± 162.94 µm <sup>2</sup> 37.23 ± 3.99% 38.56 ± 6.3% 49.58 ± 3.43%	362±107 µm <sup>2</sup> 39.28±1.54% 42.48±3.66% 51.16±1.38% 51.60±2.01%	<ul> <li>&lt;.001</li> <li>=.0064</li> <li>=.198</li> <li>003</li> </ul>	MP1
					Segmentation line 2 Segmentation line 3	49.70±7.96% 51.13±5.62%	$45.14 \pm 3.30\%$ $47.54 \pm 4.82\%$	 =.15 =.848	
Shen et al. (2020) <sup>[1]</sup>	63 eyes	96 eyes	RTVue-XR Avanti;	3 × 3 mm (fovea)	Vessel density (SCP)-parafovea Vessel density (SCP)-perifovea	$39.63 \pm 6.01\%$ $39.63 \pm 6.01\%$	$50.68 \pm 3.83\%$ $46.65 \pm 4.21\%$	<.001 <.001	Visual field
			VG200 SVision Imaging	6 × 6 mm (fovea); 12 × 12 mm (fovea)	Vessel density (JCP)-paratovea Vessel density (DCP)-perifovea CVI-parafovea	48.10±5.17% 45.57±5.52% 0.25±0.04	$55.03 \pm 3.32\%$ $51.23 \pm 5.11\%$ $0.28 \pm 0.06$	<.001 <.001 =.113	
					CVI-perifovea	$0.18 \pm 0.07$	$0.25 \pm 0.05$	=.003	

(Continued)

5

	Groups a Eye	ind # of 's				Summa	rry of the Results		
Article	RP	Control	<b>OCTA Instrument</b>	Scan Area	OCTA Parameter	RP	Control	<i>P</i> value	Functional Test
Macular Microvasculature Analysis Inooka et al. (2018) <sup>[35]</sup>	53 eyes	46 eyes	Cirrus HD-OCT 5000	3 × 3 mm (fovea)	Perfusion density (whole) Perfusion density (superficial)	$0.3257 \pm 0.0462$ $0.3854 \pm 0.0166$	$0.3895 \pm 0.0204$ $0.4166 \pm 0.0080$	<.001 <.001	Visual field
					Perfusion density (deep) Vessel length density (whole)	$0.2929 \pm 0.0476$ $17.566 \pm 2.938  \text{mm}^{-1}$	$0.3475 \pm 0.0298$ 22.034 ± 1.371 mm <sup>-1</sup>	<ul><li>&lt; 001</li><li>&lt; 001<td></td></li></ul>	
					vesser lengtn density (superiiciai) Vessel length density (deep) Vessel dismotor index (whole)	20.203 ± 1.170 mm <sup>-1</sup> 14.766 ± 2.711 mm <sup>-1</sup> 0.0166 ± 0.0006 mm	22.040±0.700mm <sup>-1</sup> 18.448±1.769mm <sup>-1</sup>	<ul><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li><li>.001</li>&lt;</ul>	
					Vessel diameter index (whole) Vessel diameter index (superficial)	$0.0190 \pm 0.0004$ mm	0.0184±0.0003 mm	<.001	
					Vessel diameter index (deep) FAZ area	$0.0199 \pm 0.0005 \text{ mm}^2$ $0.3091 \pm 0.091 \text{ mm}^2$	$0.0188 \pm 0.0004 \text{ mm}$ $0.2310 \pm 0.065 \text{ mm}^2$	<.001 <.001	
Arrigo et al. (2019) <sup>[36]</sup> 3	32 eyes	32 eyes	Swept-source OCT DRI Topcon Triton	$3 \times 3 \text{ mm}$ (fovea)	Vessel density (SCP)-parafovea	$0.39 \pm 0.02$	$0.41 \pm 0.01$	<.01	Visual field
				$4.5 \times 4.5 \text{ mm}$ (optic)	vessel density (JUCP)-paratovea Vessel dispersion (SCP)-parafovea	0.30±0.03 24±15	0.45±0.01 11±4	01 01	
					Vessel dispersion (DCP) -parafovea	16±12	11 = 3	<.01	
					Vessel tortuosity (SCP)-parafovea	$4.80 \pm 0.29$	$7.2 \pm 0.31$	<.01	
					Vessel tortuosity (DCP)-parafovea	$4.42 \pm 0.49$	$7.84 \pm 0.34$	<.01	
					Vessel rarefaction (SCP)-parafovea	$0.66 \pm 0.04$	$1.80 \pm 0.32$	<.01	
l in et al. (2019) <sup>[33]</sup>	37 eves	54 eves	RTVne-XR Avanti	$3 \times 3 \mathrm{mm}$ (fovea)	Vessel raretaction (UCP)-paratovea Vessel density (SCP)-narafoveal	$0.62 \pm 0.03$	$1.09 \pm 0.2$	<.01	
					Moderate	$41.83 \pm 4.56\%$	$48.95 \pm 3.73\%$	<.001	
					Severe	$41.64 \pm 4.93\%$	$48.95 \pm 3.73\%$	<.001	
					Vessel density (DCP)-parafoveal				
					Moderate	$47.7 \pm 9.44\%$	$52.84 \pm 3.4\%$	=.026	
					Severe	$39.58 \pm 9.46\%$	$52.84 \pm 3.4\%$	<.001	
					Cone density-C1				
					Moderate	$22726 \pm 2648/mm^2$	$23691 \pm 2941/mm^2$	=.96	
					Severe	$16338 \pm 4139/mm^2$	23691 ± 2941/mm <sup>2</sup>	<.001	
					Modorato	10005 - 7177/2002	20070 , 0776/mm <sup>2</sup>	100 1	
					INIUUEI ALE Sevvere	13003 ± 242//IIIIIF 14751 + 3089/mm <sup>2</sup>	$23278 + 2776/mm^2$	- 001 - 001	
					Cone density-C3			-	
					Moderate	$18741 + 2317/mm^2$	$20974 + 2074/mm^2$	=.005	
					Severe	$14091 \pm 2863/mm^2$		<.001	
					Cone density-C4				
					Moderate	$18086 \pm 2086/mm^2$	$18734 \pm 1460/mm^2$	=.86	
					Severe	$13794 \pm 3030/mm^2$	$18734 \pm 1460/mm^2$	<.001	
					Cone density-Co				
					Moderate Severe	$1/4/3 \pm 2084/mm^{-1}$ $13428 \pm 3034/mm^{2}$	1 / 24/ ± 132 //mm <sup>4</sup> 1 7247 ± 1327/mm <sup>2</sup>	∪0.1= <.001	

Lu et al • Medicine (2022) 101:34

Table2 (Continued) (Continued)

	Groups Ey	and # of es				Summa	iry of the Results		
Article	RP	Control	<b>OCTA Instrument</b>	Scan Area	OCTA Parameter	RP	Control	P value	Functional Test
AttaAllah et al. (2020) <sup>[34]</sup>	30 eyes	24 eyes	RTVue-XR Avanti	6 × 6 mm (fovea)	Vessel density (SCP)-parafovea	45.2 ± 3%	$46.8 \pm 5.2\%$	=.191	_
					Vessel density (DCP)-parafovea	$44 \pm 5.8\%$	$55 \pm 5.2\%$	<.001	
					Vessel density (CCP)-parafovea	$63.8 \pm 2.4\%$	$65.8 \pm 1.8\%$	<.001	
					FAZ area (SCP)	$0.5 \pm 0.2 \mathrm{mm^2}$	$0.3 \pm 0.1  \text{mm}^2$	<.001	
					FAZ area (DCP)	$0.6 \pm 0.2 \mathrm{mm^2}$	$0.3 \pm 0.1  \text{mm}^2$	<.001	
Arrigo et al. (2020) <sup>[37]</sup>	45 eyes	45 eyes	Swept-source OCT DRI Topcon Triton	$3 \times 3 \mathrm{mm}$ (fovea)	Choroidal thickness				/
				$4.5 \times 4.5$ mm(optic)	Pattern 1 (early stage)	227 ±37 μm	305 ± 59 µm	=.685	
				-	Pattern 2 (advanced stage)	$218 \pm 44  \mu m$	$305 \pm 59  \mu m$	<.01	
					Pattern 3 (late stage)	$156 \pm 40 \ \mu m$	$305 \pm 59 \mu m$	<.01	
					Choroidal stromal index				
					Pattern 1 (early stage)	$0.03 \pm 0.01$	$0.03 \pm 0.01$	<.01	
					Pattern 2 (advanced stage)	$0.07 \pm 0.01$	$0.03 \pm 0.01$	<.01	
								<.01	

types of OCTA devices. Lin *et al.* evaluated the macular structural changes in the parafoveal regions in normal subjects and mild- or late-stage RP patients with objectively quantified cone density (CD) and microvascular density and showed significant cone loss in RP patients.<sup>[33]</sup> AttaAllah et al demonstrated a reduction in macular microvascular density in all studied layers on OCTA as well as macular structural changes such as EZ disruption and FAZ enlargement.<sup>[34]</sup> However, findings regarding the FAZ area have been controversial. Parodi et al found that the FAZ area was significantly enlarged only at the level of the DCP in RP eyes.<sup>[20]</sup> Conversely, Koyanagi et al found a significant enlargement of the superficial, but not deep, FAZ area, which was relatively preserved until the mild-to-late stages.<sup>[21]</sup>

Quantitative OCTA parameters help to identify retinal vascular abnormalities in patients with RP. Measurements of geometric vascular features using OCTA biomarkers may become a useful tool to monitor disease activity and the efficacy of new therapeutic modalities. Inooka et al showed that both qualitative and quantitative changes in microvascular density and morphology are useful for assessing the pathophysiology of RP.[35] With an automated program, indices of the microvascular density, perfusion density (PD), and vessel length density (VLD) were found to be significantly reduced, and the vessel diameter index (VDI) was significantly increased in RP patients. Arrigo et al analyzed the changes in vascular features at the level of both the macula and optic nerve between RP patients and healthy controls.<sup>[36]</sup> Their results revealed a statistically significant difference in all the calculated OCTA parameters, including vessel density, vessel tortuosity, vessel dispersion, and vessel rarefaction, with VT and VR being the most reliable biomarkers to describe the abnormalities of geometric vascular features in RP patients. In addition, quantitative measurements of choroidal features, including choroidal stromal index (CSI), VT, VDisp, and vessel density, revealed further detailed information regarding the changes in choroidal patterns in RP patients which were found to be associated with different RP clinical forms as well as with different progression after 1 year using SS-OCTA.<sup>[37]</sup>

# 5. Correlation of OCTA parameters with retinal function analysis in RP

Correlation studies between the quantitative vascular OCTA parameters and retinal function (measured either with objective methods such as multifocal ERG or subjective methods such as visual acuity, visual field, and microperimetry) could better clarify whether any functional impairment corresponds to vascular signal changes. Functional dysregulation of retinal and choroidal changes seems to occur in patients with late-stage RP. Toto et al demonstrated that both SCP and DCP vessel densities in the macular region are correlated with the macular function, as well as with the GCC thickness.[22] Liu et al analyzed the correlation of choroidal small/middle and large vessel density with retinal photoreceptor cells and visual function in patients with RP, which demonstrated that choroidal microcirculation was a prominent factor affecting the visual acuity, visual field, and ERG b-wave amplitude in patients with RP.<sup>[38]</sup> This may provide new insights into the mechanisms and treatment of RP. With the advantage of SS-OCTA, various types of choriocapillaris defects can be defined according to the choroidal vascular structure, and the degree of choriocapillaris defects was correlated with the BCVA, Humphrey indexes, and microperimetry index.<sup>[1]</sup> Falfoul et al also found a statistically significant correlation between macular function and parafoveal DCP density.<sup>[25]</sup>

# 6. Limitations and future visions for OCTA in RP management

The relationship between retinal and choroidal vascular changes and retinal function should be further confirmed in a larger population. Moreover, the nature of the cross-sectional design of these previous studies makes it insufficient to precisely assess the cause-effect relationship between retinal and choroidal vasculature changes and retinal function. Further prospective studies with longer follow-up periods combining retinal function studies and microvasculature changes will provide a better understanding of the pathophysiology of RP patients.

OCTA is a useful tool for monitoring RP disease progression and may be used to measure retinal vascular parameters as outcomes in clinical trials. It is important, however, to recognize its limitations, such as susceptibility to motion artifacts, projection artifacts, limited comparability among different OCTA devices, and restricted contribution of information regarding the grade of disease activity. With the continuing improvement of OCTA technology, PR-OCTA can enable the clean visualization of some retinal plexuses and vascular pathologies using post-processing algorithms to reduce projection artifacts.<sup>[39]</sup> The ability of the newly developed wide-field OCTA (WF-OCTA) was also compared to that of ultra-wide-field fluorescein angiography (UWF-FA) and ultra-wide-field color fundus photography (UWF-CP) for retinal disorder detection.<sup>[40]</sup> Retinal biomarkers from OCTA images can facilitate the clinical management of retinal disorders. However, current commercially available OCTA devices are not able to provide a wide field of quantifiable retinal biomarkers because of the limitations of their analysis software. An "all-in-one" metric that can generate comprehensive retinal biomarkers, including parameters depicting geometric vasculature features with accurate definition, as well as those quantifying blood flows with improved classification and segmentation, for retinal disorder analysis is of great importance and convenience for both clinicians and patients. Hence, we have recently invented a method for establishing computational retinal microvascular biomarkers (CRMBs) through a knowledge-driven computerized automatic analytical system based on fractal analysis using OCTA images (manuscript in preparation for submission). We anticipate that these CRMBs will directly lead to a new classification of RP patients, facilitating better understanding, early detection, timely treatment, and improved quality of life.

#### 7. Conclusions

The application of OCTA is beneficial for studying alterations in the retinal vasculature in the progression of RP. In this review, we summarised the current OCTA findings in clinical research on RP and envisioned future advancements in OCTA. We believe that the development of OCTA is a major contribution to advancing ophthalmic imaging. This will help us to better understand the etiology and pathology of RP. It will also facilitate the diagnosis, monitoring, and treatment of RP.

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#### **Author Contributions**

Bingwen Lu had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data interpretation.

Concept and design: Bingwen Lu.

Acquisition, analysis, or interpretation of data: Bingwen Lu.

Drafting of the manuscript: Bingwen Lu.

Critical revision of the manuscript for important intellectual content: All authors.

Supervision: Like Xie

#### References

- Shen C, Li Y, Wang Q, et al. Choroidal vascular changes in retinitis pigmentosa patients detected by optical coherence tomography angiography. BMC Ophthalmol. 2020;20:384.
- [2] Rezaei KA, Zhang Q, Chen CL, et al. Retinal and choroidal vascular features in patients with retinitis pigmentosa imaged by OCT based microangiography. Graefes Arch Clin Exp Ophthalmol. 2017;255:1287–95.
- [3] Fahim A. Retinitis pigmentosa: recent advances and future directions in diagnosis and management. Curr Opin Pediatr. 2018;30:725–33.
- [4] Lang M, Harris A, Ciulla TA, et al. Vascular dysfunction in retinitis pigmentosa. Acta Ophthalmol. 2019;97:660–4.
- [5] Spaide RF, Fujimoto JG, Waheed NK, et al. Optical coherence tomography angiography. Prog Retin Eye Res. 2018;64:1–55.
- [6] Merin S, Auerbach E. Retinitis pigmentosa. Surv Ophthalmol. 1976;20:303–46.
- [7] Takayama K, Ito Y, Kaneko H, et al. Comparison of indocyanine green angiography and optical coherence tomographic angiography in polypoidal choroidal vasculopathy. Eye (Lond). 2017;31:45–52.
- [8] Robson AG, Saihan Z, Jenkins SA, et al. Functional characterisation and serial imaging of abnormal fundus autofluorescence in patients with retinitis pigmentosa and normal visual acuity. Br J Ophthalmol. 2006;90:472–9.
- [9] Pichi F, Aggarwal K, Neri P, et al. Choroidal biomarkers. Indian J Ophthalmol. 2018;66:1716–26.
- [10] Nakazawa M, Hara A, Ishiguro SI. Optical coherence tomography of animal models of retinitis pigmentosa: from animal studies to clinical applications. Biomed Res Int. 2019;2019:8276140.
- [11] Funatsu J, Murakami Y, Nakatake S, et al. Direct comparison of retinal structure and function in retinitis pigmentosa by co-registering microperimetry and optical coherence tomography. PLoS One. 2019;14:e0226097.
- [12] Rocholz R, Corvi F, Weichsel J, et al. OCT angiography (OCTA) in retinal diagnostics. 2019. In: Bille JF, (eds). High Resolution Imaging in Microscopy and Ophthalmology: New Frontiers in Biomedical Optics [Internet]. Cham (CH): Springer; 2019. Chapter 6. PMID: 32091844.
- [13] Ong SS, Patel TP, Singh MS. Optical coherence tomography angiography imaging in inherited retinal diseases. J Clin Med. 2019;8:2078.
- [14] Chu Z, Lin J, Gao C, et al. Quantitative assessment of the retinal microvasculature using optical coherence tomography angiography. J Biomed Opt. 2016;21:66008.
- [15] Murakami Y, Ikeda Y, Akiyama M, et al. Correlation between macular blood flow and central visual sensitivity in retinitis pigmentosa. Acta Ophthalmol. 2015;93:e644–8.
- [16] Zong Y, Lin L, Yi C, et al. Retinal vessel oxygen saturation and vessel diameter in retinitis pigmentosa at various ages. Graefes Arch Clin Exp Ophthalmol. 2016;254:243–52.
- [17] Milam AH, Li ZY, Fariss RN. Histopathology of the human retina in retinitis pigmentosa. Prog Retin Eye Res. 1998;17:175–205.
- [18] Zhang Y, Harrison JM, Nateras OS, et al. Decreased retinal-choroidal blood flow in retinitis pigmentosa as measured by MRI. Doc Ophthalmol. 2013;126:187–97.
- [19] Wang XN, Zhao Q, Li DJ, et al. Quantitative evaluation of primary retinitis pigmentosa patients using colour Doppler flow imaging and optical coherence tomography angiography. Acta Ophthalmol. 2019;97:e993–7.
- [20] Battaglia Parodi M, Cicinelli MV, Rabiolo A, et al. Vessel density analysis in patients with retinitis pigmentosa by means of optical coherence tomography angiography. Br J Ophthalmol. 2017;101:428–32.
- [21] Koyanagi Y, Murakami Y, Funatsu J, et al. Optical coherence tomography angiography of the macular microvasculature changes in retinitis pigmentosa. Acta Ophthalmol. 2018;96:e59–67.
- [22] Toto L, Borrelli E, Mastropasqua R, et al. Macular features in retinitis pigmentosa: correlations among ganglion cell complex thickness, capillary density, and macular function. Invest Ophthalmol Vis Sci. 2016;57:6360–6.
- [23] Sugahara M, Miyata M, Ishihara K, et al. Optical coherence tomography angiography to estimate retinal blood flow in eyes with retinitis pigmentosa. Sci Rep. 2017;7:46396.
- [24] Takagi S, Hirami Y, Takahashi M, et al. Optical coherence tomography angiography in patients with retinitis pigmentosa who have normal visual acuity. Acta Ophthalmol. 2018;96:e636–42.
- [25] Falfoul Y, Elleuch I, El Matri K, et al. Multimodal imaging in retinitis pigmentosa: correlations among microvascular changes, macular function and retinal structure. J Curr Ophthalmol. 2020;32:170–7.
- [26] Campbell JP, Zhang M, Hwang TS, et al. Detailed vascular anatomy of the human retina by projection-resolved optical coherence tomography angiography. Sci Rep. 2017;7:42201.

- [27] Yu DY, Cringle SJ. Oxygen distribution and consumption within the retina in vascularised and avascular retinas and in animal models of retinal disease. Prog Retin Eye Res. 2001;20:175–208.
- [28] Hagag AM, Wang J, Lu K, et al. Projection-resolved optical coherence tomographic angiography of retinal plexuses in retinitis pigmentosa. Am J Ophthalmol. 2019;204:70–9.
- [29] Guduru A, Al-Sheikh M, Gupta A, et al. Quantitative assessment of the choriocapillaris in patients with retinitis pigmentosa and in healthy individuals using OCT angiography. Ophthalmic Surg Lasers Imaging Retina. 2018;49:e122–8.
- [30] Corazza P, Cirafici P, Testa V, et al. Vascular density and retinal function in patients with retinitis pigmentosa evaluated by swept-source OCT angiography and microperimetry. Ophthalmologica. 2020;244:27–33.
- [31] Miyata M, Oishi A, Hasegawa T, et al. Concentric choriocapillaris flow deficits in retinitis pigmentosa detected using wide-angle swept-source optical coherence tomography angiography. Invest Ophthalmol Vis Sci. 2019;60:1044–9.
- [32] Mastropasqua R, Borrelli E, Agnifili L, et al. Radial peripapillary capillary network in patients with retinitis pigmentosa: an optical coherence tomography angiography study. Front Neurol. 2017;8:572.
- [33] Lin R, Shen M, Pan D, et al. Relationship between cone loss and microvasculature change in retinitis pigmentosa. Invest Ophthalmol Vis Sci. 2019;60:4520–31.

- [34] AttaAllah HR, Mohamed AAM, Hamid MA. Quantification of macular microvascular changes in retinitis pigmentosa using optical coherence tomography angiography. Clin Ophthalmol. 2020;14:1705–13.
- [35] Inooka D, Ueno S, Kominami T, et al. Quantification of macular microvascular changes in patients with retinitis pigmentosa using optical coherence tomography angiography. Invest Ophthalmol Vis Sci. 2018;59:433–8.
- [36] Arrigo A, Romano F, Albertini G, et al. Vascular patterns in retinitis pigmentosa on swept-source optical coherence tomography angiography. J Clin Med. 2019;8:1425.
- [37] Arrigo A, Bordato A, Romano F, et al. Choroidal patterns in retinitis pigmentosa: correlation with visual acuity and disease progression. Transl Vis Sci Technol. 2020;9:17.
- [38] Liu R, Lu J, Liu Q, et al. Effect of choroidal vessel density on the ellipsoid zone and visual function in retinitis pigmentosa using optical coherence tomography angiography. Invest Ophthalmol Vis Sci. 2019;60:4328–35.
- [39] Hormel TT, Jia Y, Jian Y, et al. Plexus-specific retinal vascular anatomy and pathologies as seen by projection-resolved optical coherence tomographic angiography. Prog Retin Eye Res. 2021;80:100878.
- [40] Pichi F, Smith SD, Abboud EB, et al. Wide-field optical coherence tomography angiography for the detection of proliferative diabetic retinopathy. Graefes Arch Clin Exp Ophthalmol. 2020;258:1901–9.