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# Highly monodisperse Pd-Ni nanoparticles supported on rGO as a rapid, sensitive, reusable and selective enzyme-free glucose sensor

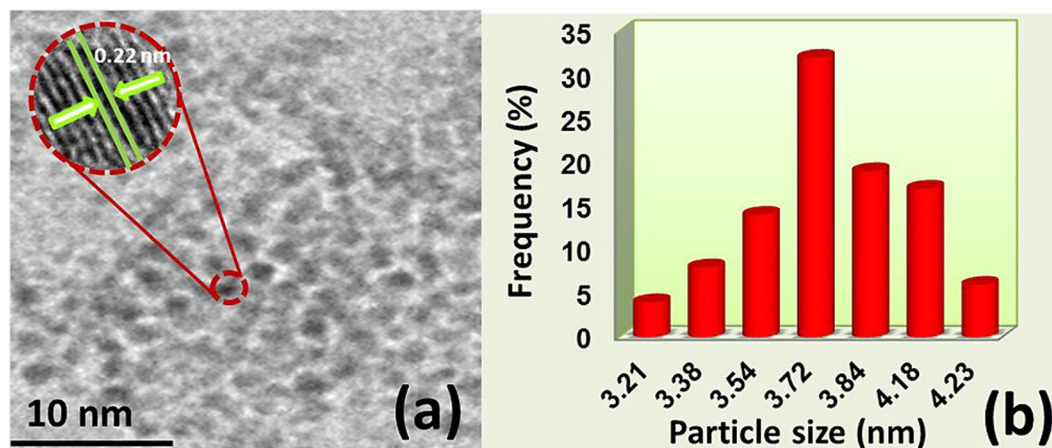
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In this work, highly monodispersed palladium-nickel (Pd-Ni) nanoparticles supported on reduced graphene oxide (rGO) were synthesized by the microwave-assisted methodology. The synthesized nanoparticles were used for modification of a glassy carbon electrode (GCE) to produce our final product as PdNi@rGO/GCE, which were utilized for non-enzymatic detecting of glucose. In the present study, electrochemical impedance spectroscopy (EIS), chronoamperometry (CA) and, cyclic voltammetry (CV) methods were implemented to investigate the sensing performance of the developed glucose electrode. The modified electrode, PdNi@rGO/GCE, exhibited very noticeable results with a linear working range of 0.05–1.1 mM. Moreover, an ultralow detection limit of 0.15  $\mu$ M was achieved. According to the results of amperometric signals of the electrodes, no significant change was observed, even after 250 h of operation period. In addition, the highly monodisperse PdNi@rGO/GCE was utilized to electrochemical detection of glucose in real serum samples. In light of the results, PdNi@rGO/GCE has shown an excellent sensing performance and can be used successfully in serum samples for glucose detection and it is suitable for practical and clinical applications.

In today's world, there are many chronic diseases that people struggling, and diabetes is one of them. High levels of blood sugar can be a sign of diabetes which considered one of the most important health problems worldwide. Since 1960, very important developments have been progressed in the determination of biomolecules through electrochemical systems<sup>1-3</sup>. Those electrochemical systems were applied in a wide range of applications, for instance, glucose oxidation, therapeutic approaches, water treatment, biofuel cell development and other analytical approaches in the food sector<sup>4-8</sup>. Glucose oxidation is considered a major step in the oxidation of organic materials and has been studied by many scientists<sup>9-11</sup>.

The development of novel materials that can be used in electrocatalytic processes still has great importance<sup>12,13</sup>. In order to achieve electrochemical detection of glucose, enzymatic glucose sensors are commonly used<sup>14-17</sup>. However, enzymatic glucose sensors have some disadvantages, for instance, being expensive, discontinuity, and immobilization problem. For example, glucose oxidase, one of the most commonly used enzyme for enzymatic glucose detection, it rapidly loses its activity and can be permanently damaged at higher operation temperatures (> 40 °C) if pH is less than 2 or higher than 8. Moreover, humidity can affect these enzymatic sensors. Incorrectly measured levels of glucose might cause prescribing overdose of insulin which leads hypoglycemia<sup>18</sup>. The stability problems in enzymatic systems hinder the applicability and development of these systems<sup>19</sup>. Non-enzymatic glucose sensors are better alternatives due to their superior properties, such as cost-effectiveness, high sensitivity, better performance in glucose oxidation, and stability. In literature, there are various nanotechnology applications

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**Figure 1.** (a) TEM and HR-TEM image (inlay); (b) particle size histogram for PdNi@rGO.

developed for the detection of glucose; metals (Au, Ni, Co, Cu, In, or Ru), metal oxides (cobalt (II) oxide, nickel oxide hydroxide, or ruthenium (IV) oxide), and metallic composites (Au-Pt, Cr-Ni, Cu-Pd)<sup>12,20–23</sup> are among them. In addition, several carbon-based compounds, such as carbon nanotubes (CNTs) and graphene, have been proposed to utilize as electrocatalysts for oxidation of glucose and increase the reaction yield on the surface of the sensor<sup>3,24,25</sup>. Among these carbon-based support materials, reduced graphene oxide (rGO) has unique properties such as good electrical conductivity, wide surface area, mechanical and chemical stability. These properties enable the rGO or rGO-containing composites, favorable for sensor applications. rGO is a very suitable supporting material for biosensors and electrochemical sensors<sup>26–28</sup>. Recently, metal nanoparticles and rGO composites have been presented, and they exhibited high sensitivity in sensing small molecules, such as H<sub>2</sub>O<sub>2</sub>, glucose, and methanol. A very few studies reported the using of rGO as a catalyst with very high sensitivity, selectivity, and durability. However, there is no study on the using of PdNi-rGO combination as a glucose sensor<sup>29–31</sup>. We also tested the stability of synthesized PdNi@rGO due to Ostwald ripening effect which may cause small metal particles can be detached from carbonous support and clustered to form larger particles<sup>32</sup>.

This study aims to the utilization of reduced graphene oxide with PdNi nanoparticles (PdNi@rGO) to enhance the corresponding activity of glucose detection. For this purpose, highly monodisperse PdNi@rGO was used to modify a glassy carbon electrode (GCE). PdNi@rGO/GCE was developed for the detection of glucose in biologic samples as a non-enzymatic sensor. Fabrication and characterization of the developed sensor were demonstrated by high resolution-transmission electroscop microscope (HR-TEM), Raman spectroscopy, X-ray diffraction (XRD), and X-ray photoelectron spectroscopy (XPS). Prepared nanoparticle enhanced electrode was examined by electrochemical techniques to uncover its selectivity, stability, sensitivity and detection limit as a non-enzymatic biosensor for glucose detection.

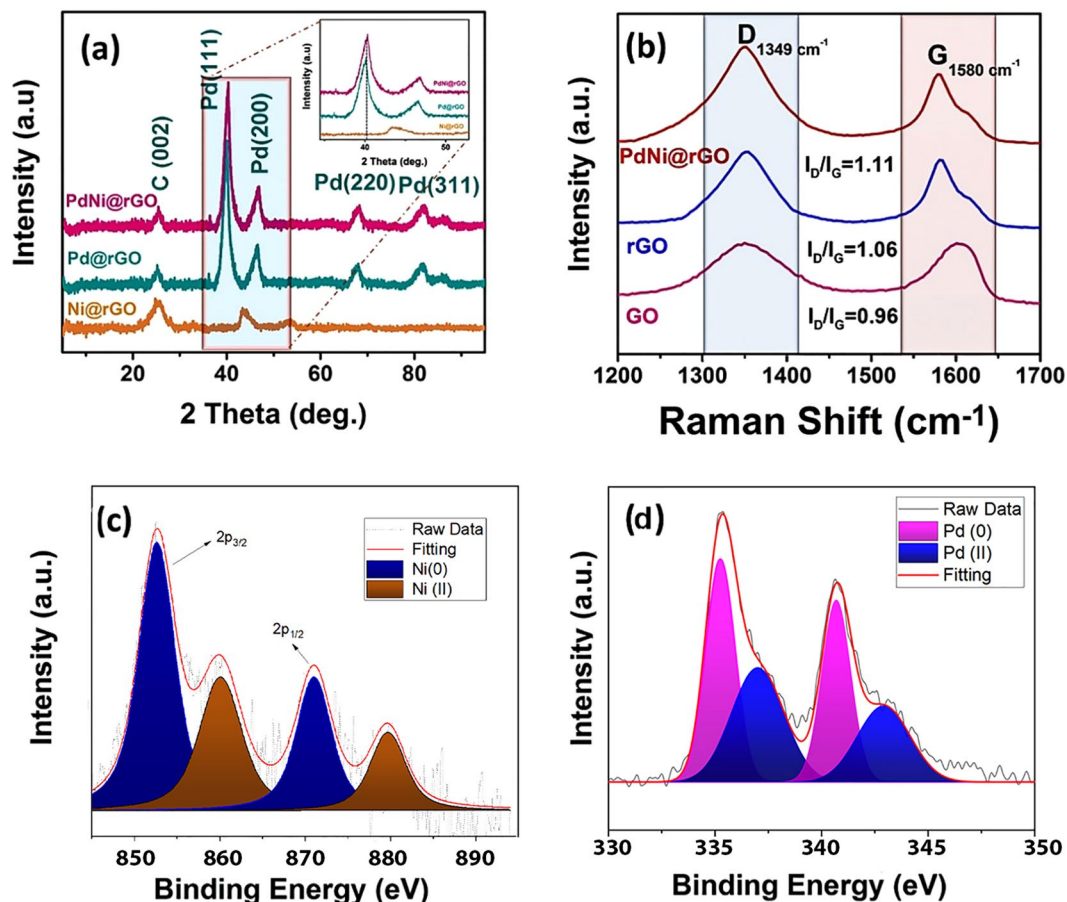
## Experimental

**Synthesis of highly monodisperse PdNi@rGO nanoparticles.** A modified microwave synthesis method adapted from previous studies<sup>33–36</sup> in literature was conducted for a consistent producing of monodisperse nanomaterials. For this aim, equal amounts (0.25 mmol of each) of PdCl<sub>2</sub> and Ni(Ac)<sub>2</sub> were dissolved in 20 mL ethylene glycol with vigorous mixing, in the presence of oleylamine (OA) which is used for stabilization of metal nanoparticles. The pH adjustment was achieved using a NaOH-ethylene glycol solution. In this work, ethylene glycol was added for reducing of PdCl<sub>2</sub> and Ni(Ac)<sub>2</sub>. The mixture was located in a microwave oven for 60 s (5 times) at 1200 W. Finally, the resultant was filtered and washed using deionized water and acetone. Synthesis of GO and rGO is given in the Supporting Information section. The produced PdNi bimetallic nanoparticles were mixed with (0.50 mmol) rGO using a sonicator to produce a uniform dispersion. The solution was then subjected to microwave irradiation (1200 W) for another 60 s (5 times). Obtained PdNi@rGO nanoparticles were dried under vacuum at room temperature.

**Preparation of PdNi@rGO/GCE modified electrode.** The GC electrode surface was polished to mirror-like using 0.3- $\mu$ m and 0.05- $\mu$ m Al<sub>2</sub>O<sub>3</sub> powder and washed with distilled H<sub>2</sub>O and ethanol for 2 min. 1 mg PdNi@rGO were dispersed in 10 mL N,N-dimethylformamide (DMF) in an ultrasonic bath, and a black solution obtained. 20  $\mu$ L of the solution was cast at the surface of the glassy carbon electrode and the solvent was evaporated at 50 °C.

## Results and Discussion

**Characterization of nanoparticles.** HR-TEM, Raman, XRD, and XPS analyses were utilized to characterize PdNi@rGO nanocomposites, the detailed examination of characterization methods was given in Supporting Information section (see Appendix). TEM and HR-TEM images of PdNi@rGO was given in Fig. 1(a). As can be seen, PdNi nanoparticles were monodispersely distributed on the surface of rGO and no agglomeration was observed. The synthesized nanoparticles were mostly spherical shaped and have an average particle size of



**Figure 2.** (a) The XRD and (b) Raman spectra of PdNi@rGO NPs; XPS spectra of (c) Ni 2p and (d) Pd 3d.

3.72 ± 0.51 nm (Fig. 1(b)). In addition, HR-TEM analysis presented the atomic lattice fringe of 0.22 nm and this is totally in agreement with the nominal Pd (111) range<sup>37,38</sup>.

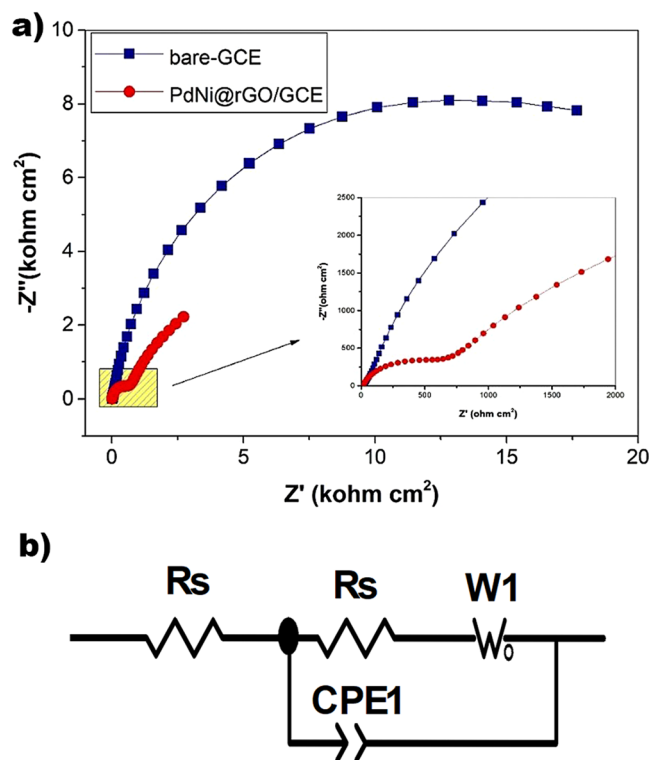
XRD analyses were implemented to identify the structure of crystal formations and the average crystal size of PdNi@rGO. In Fig. 2(a), typical crystal structures observed in our samples can be seen. The crystal planes of Pd (111), (200), (220) and (311) are attributed to 2θ peaks of 40.3°, 46.4°, 68.4°, and 82.5°, respectively, which reveals a face-centered cubic (fcc) crystal structure. The diffraction peaks in Fig. 2(a) exhibited higher 2θ values (slightly shifted when compared with the pure Pd), this indicates the existence of the monodisperse PdNi@rGO alloy. Additionally, the peak observed at 25.5° implying that the reduction of GO to reduced graphene oxide was achieved successfully. The XRD results of PdNi revealed that there was no significant nickel diffraction peak, due to the fact that synthesized nanoparticle has stronger signals for the palladium, which is associated with the amorphous structure. Using the Eq. 1, the average size of crystal particles was calculated as 3.21 ± 0.51 nm. The results are compatible with the result obtained from TEM analysis and also previous studies in the literature<sup>39–43</sup>.

$$d(\text{\AA}) = \frac{k\lambda}{\beta \cos \theta} \quad (1)$$

Where  $k$  is the coefficient (0.9),  $\lambda$  is the wavelength of the X-ray (= 1.54056 Å),  $\beta$  is the full width of the respective diffraction peak at the half-maximum (rad), and  $\theta$  is the angle at the position of peak maximum (rad). With the aim of finding the lattice parameter values ( $\alpha$ PdNi), the diffraction peak of PdNi (220) was used. By the using of Eq. 2, the lattice parameter value of the PdNi@rGO was found to be 3.88 Å, which is a little lower than 3.89 Å (for pure Pd)<sup>44,45</sup>.

$$\sin \theta = \frac{\lambda \sqrt{h^2 + k^2 + l^2}}{2a} \quad (\text{for cubic structure}) \quad (2)$$

Raman analysis conducted for distinguishing irregular and regular carbon structures. Raman spectra of GO, rGO, and PdNi@rGO were displayed in Fig. 2(b). Two distinct peaks observed at 1349 cm<sup>-1</sup> and 1580 cm<sup>-1</sup> corresponding to D ( $E_{2g}$  phonon) and G ( $A_{1g}$  symmetry) bands. The D band is related to the disordered carbon atoms, while the G band is associated to sp<sup>2</sup>-hybridized graphitic carbon atoms. The I<sub>D</sub>/I<sub>G</sub> ratios (the intensity ratio of the D band to the G band which is used for seeing modifications and defects in nanomaterials) were calculated as



**Figure 3.** (a) Nyquist plots of bare-GCE and Pd-Ni@rGO modified electrodes, (b) equivalent circuit diagram.

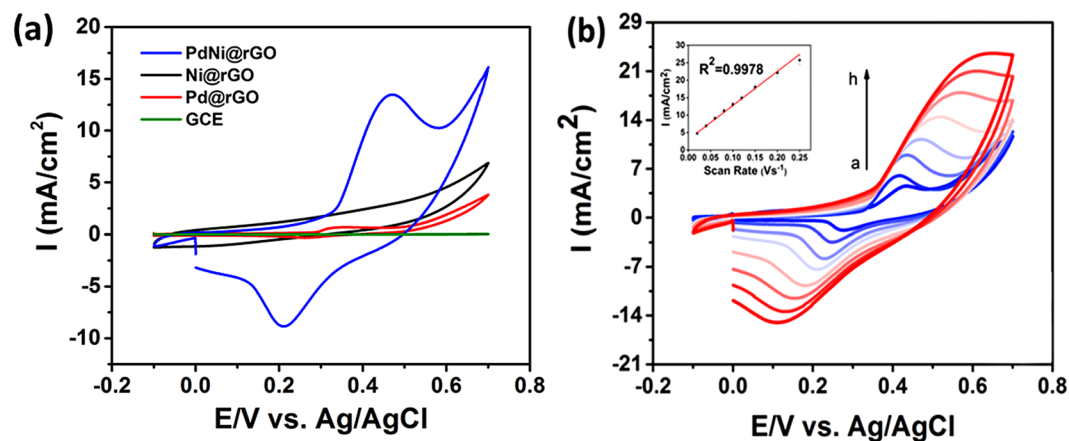
0.96, 1.06 and 1.1 for GO, rGO and PdNi@rGO, respectively. The increasing  $I_D/I_G$  ratio indicates that GO deoxygenated, and rGO became functionalized with PdNi nanoparticles.

XPS analysis was utilized to investigate the surface composition and the oxidation states of Pd and Ni atoms in PdNi@rGO. The XPS spectra for Pd 3d and Ni 2p were displayed in Fig. 2(c,d). The Gaussian-Lorentzian method was employed to examine the Pd 3d and Ni 2p regions of the spectrum. The relative density of the species was assessed by counting the integral of peaks. The binding energies ( $\pm 0.3$  eV) were measured by reference to the C 1s peak at 283.6 eV. The peaks observed at 852.6 eV/871.0 eV and 860.0 eV/879.7 eV corresponded to metallic Ni and Ni (II), respectively (Fig. 2(c)). Similarly, in Fig. 2(d), peaks observed at 335.3 eV/340.7 eV and 337.0 eV/342.9 eV corresponded to metallic Pd and Pd (II), respectively. The intensity of the peaks showed that Pd and Ni exist predominantly in metallic form. The peaks of Pd(II) and Ni(II) observed in XPS spectra are the probable results of chemical sorption of oxygen or surface oxidation while nanomaterials were producing. A low energy shift of Ni 2p<sub>3/2</sub> peak indicates an alloying process of PdNi.

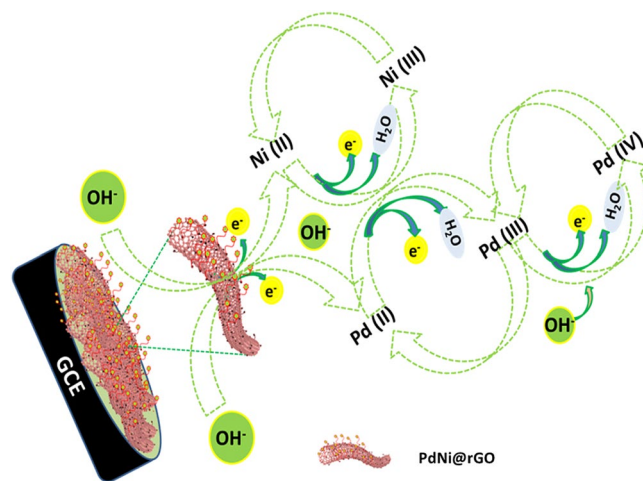
**Electrochemical properties of the PdNi@rGO/GCE.** *Electrochemical impedance characterization of electrodes.* The bare-GCE and Pd-Ni@rGO modified electrodes were tested with EIS. Nyquist plots of the bare and modified electrodes were shown in Fig. 3(a), and related electrical equivalent circuit diagram was shown in Fig. 3(b). The EIS data were fitted by Zview software. According to fitted EIS results, Pd-Ni@rGO modified electrodes exhibited a nearly straight line with significantly lower charge-transfer resistance ( $R_{ct} = 600 \Omega \text{ cm}^2$ ), while the bare-GCE exhibited a semi-circle in the high-frequency range ( $R_{ct} = 19844 \Omega \text{ cm}^2$ ). This indicates the prepared Pd-Ni@rGO enhanced electrochemical process on the electrode/electrolyte interface in contrast to a relatively slow electrochemical performance of bare GCE. Our findings support that electrical conductivity and electron transfer rates are increased as a result of decreasing charge transfer resistance.

*Cyclic voltammetry measurement results.* In order to analyze the glucose oxidation in bare and modified glassy carbon electrodes(GCE), cyclic voltammetry (CV) measurements were conducted. Figure 4(a) represents the comparative results of CV experiments. Electrochemical activities of GCE, monometallic electrodes (Ni@rGO/GCE and Pd@rGO/GCE), and PdNi@rGO/GCE electrode were measured at the coexistence of 0.1 mM glucose in 0.1 M NaOH medium. All the CV tests were conducted in a potential range of  $-0.2$  V to  $+0.8$  V and at  $50 \text{ mV s}^{-1}$  scan rate. For Ni@rGO/GCE and Pd@rGO/GCE modified electrodes, intensities of the anodic and cathodic peaks were found to be close. However, for the PdNi@rGO/GCE modified electrode, the intensity of the cathodic and anodic peaks exhibited a significant increase. This can be explained by the larger surface area of PdNi@rGO/GCE and enhanced electron transfer rate. Moreover, the anodic peak current of the modified PdNi@rGO/GCE electrode was higher than the PdNi/GCE electrode (Fig. S2). The results demonstrated that the PdNi@rGO/GCE showed enhanced electrochemical activity for glucose electrooxidation to gluconolactone compared to GCE and modified with monometallic nanoparticles.





**Figure 4.** (a) Cyclic voltammograms of GCE, Pd@rGO, Ni@rGO and PdNi@rGO (scan rate:  $50 \text{ mV s}^{-1}$ , in  $0.1 \text{ M NaOH}$  and  $0.1 \text{ mM}$  glucose). (b) Cyclic voltammograms of PdNi@rGO/GCE at different scanning rates. ( $1 \text{ mM}$  glucose, and scan rates from  $20$  to  $180 \text{ mV s}^{-1}$ ).

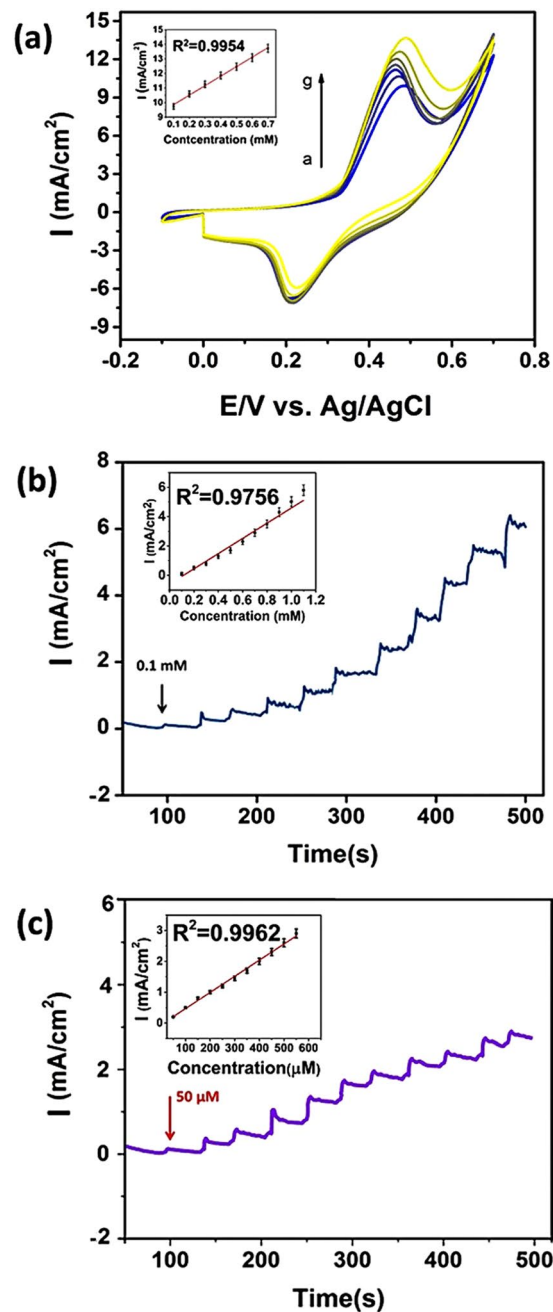


**Figure 5.** The mechanism for the activation of Ni-Pd@rGO/GCE in the alkali medium.

The electrochemical properties of PdNi@rGO/GCE were also examined at various scan rates using  $0.3 \text{ mM}$  glucose in  $0.1 \text{ M NaOH}$  (Fig. 4(b)). An increased redox current was achieved by increasing scan rate from  $20$  to  $180 \text{ mV s}^{-1}$ . This is explained by a diffusion-controlled redox reaction that occurred on PdNi@rGO/GCE. Here, it should be noted that  $0.1 \text{ M NaOH}$  is used for all electrochemical experiments and it can be expected a possible shift to positive potential values.

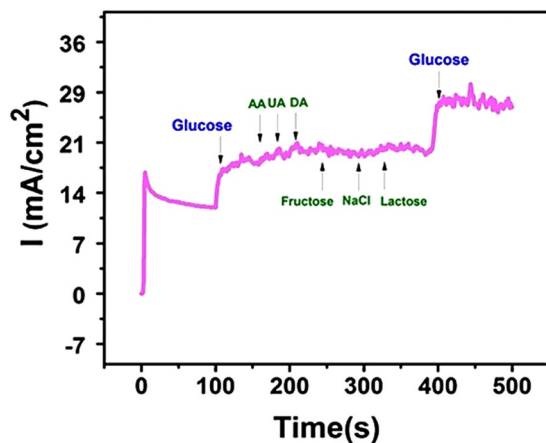
The mechanism for the activation of PdNi@rGO/GCE was presented in Fig. 5: (i) anodic scan (under the alkaline conditions), Pd (III) and Ni (III) were formed from the Pd (II) and Ni (II); (ii) at higher potentials, Pd (IV) was formed from Pd (III); and (iii) in the cathodic scan, Pd (II) and Ni (II) formation was observed.

**Amperometric measurements and optimization studies.** CV experiments were also conducted with the different amounts of glucose (from  $0.1 \text{ mM}$  to  $0.7 \text{ mM}$ ) to investigate the concentration effect on the performance of PdNi@rGO/GCE electrode. The relationship between the glucose concentration and the peak intensity of oxidation was given in Fig. 6(a). A proportional relationship was observed between them up to  $0.7 \text{ mM}$ . The amperometric measurements of PdNi@rGO/GCE electrode were displayed in Fig. 6(b,c). The results were obtained at  $0.50 \text{ V}$  potential, in  $0.1 \text{ M NaOH}$  medium, with an increased concentration of glucose ( $0.1$ – $1.0 \text{ mM}$ , with  $0.1 \text{ mM}$  successive addition steps and,  $50$  to  $500 \mu\text{M}$ , with  $50 \mu\text{M}$  successive injections). The modified electrode exhibited a rapid response when glucose was introduced and the current became steady-state within  $3$ – $5$  seconds. The linear correlation between glucose concentration and the current was displayed as inset images. The respective correlation coefficients ( $R^2$ ) were found to be very high at  $0.9756$  and  $0.9962$ . In addition, the detection limit (LOD) of PdNi@rGO/GCE electrode was  $0.15 \mu\text{M}$ ,  $S/N = 3$  (Table S1) that supports the proposed sensor have excellent sensitivity and very low detection limit.



**Figure 6.** (a) CVs for the PdNi@rGO/GCE at various concentrations of glucose (0.1–0.7 mM). The linear range plot was shown in the inset. (b) Amperometric i-t curve responses (0.1 to 1.1 mM), inset shows the corresponding calibration plots (c) Amperometric i-t curve responses at different glucose concentrations (50 to 500  $\mu$ M), inset shows the corresponding calibration plots.

*Electrochemical interference analysis of PdNi@rGO/GCE.* In real serum samples, there may be some interferences with some carbohydrates (fructose and lactose) and some biological molecules (e.g. uric acid (UA), dopamine (DA), and ascorbic acid (AA)) which naturally exist in the human blood. Thus, the selectivity of the sensor is a crucial parameter in terms of clinical applicability. Therefore, the interference effect was also examined. For this aim, successive addition of 0.1 mM of AA, DA, fructose, lactose, sodium chloride, and UA was done into 1 mM of glucose. The interference experiment was performed in 0.1 M of NaOH solution at 0.50 V to test the selectivity of PdNi@rGO/GCE. The amperometric i-t curve responses were given in Fig. 7. A significant increase in the amperometric current response was observed with the addition of 1.0 mM glucose. However, the current response did not change slightly with the injection of these potential interferences into the same solution. The responsive signal for additional molecules is very low compared to the glucose signal and these signals can be ignored. The results supported that PdNi@rGO/GCE is highly selective towards the detection of glucose in the coexistence of these interferences.



**Figure 7.** The effect of interferents (with the addition of AA, UA, DA, fructose, NaCl and lactose (0.1 mM for each) respectively) on the amperometric response of PdNi@rGO/GCE. Experiments were conducted at +0.50 V, in alkali medium, and successive addition of 1.0 mM glucose.

Samples	Glucose concentration (mM) (PdNi@rGO/GCE)	Relative standard deviation (RSD) (N = 5)	Glucose concentration (mM) (commercial device)
1	5.50	3.8%	5.30
2	3.50	5.4%	3.32
3	6.00	4.5%	5.74
4	3.10	1.6%	3.05
5	6.75	3.05%	6.55

**Table 1.** Glucose concentrations in serum samples obtained by PdNi@rGO/GCE electrode and a commercial glucose meter.

The applicability of PdNi@rGO/GCE electrode in daily use was investigated with real serum samples which contain many interference molecules. For these investigations, 10 mL of NaOH (0.1 M) was mixed with 20  $\mu$ L of human blood serum. PdNi@rGO/GCE electrode was used and amperometric measurements were conducted. The glucose concentrations of each sample were also measured with a commercial glucose monitoring meter designed for using in daily life (GluNEO Lite, IGM-1003A with GDH-FAD test strip enzyme by Infopia Co., Korea) and results were compared in Table 1. Five different samples were used for comparison and it was observed that the proposed sensor showed an excellent agreement with the commercial glucose meter. However, PdNi@rGO/GCE gave slightly higher signal responses than the commercial glucose sensor and the relative standard deviation (N = 5) was found between 1.6 and 5.4%.

## Conclusions

In summary, PdNi@rGO nanomaterial was synthesized successfully by a microwave-assisted method for producing a modified GCE that will be utilized in electrochemical glucose detection. A non-enzymatic electrochemical biosensor, PdNi@rGO/GCE, was developed and used for catalyzed conversion of glucose to gluconolactone, successfully. This novel glucose sensor also showed a low detection limit (0.15  $\mu$ M), wide linear range (0.05–1.1 mM), a high sensitivity (37.5 mA mM<sup>-1</sup> cm<sup>-2</sup> at +0.5 V), and a very good reproducibility (Table S1). Remarkably, modifying the GCE surface with PdNi@rGO significantly improved the accuracy and selectivity of the glucose measurement. Moreover, glucose detection with developed electrode did not affect by interferences, and highly satisfied results were obtained in serum samples. Based on the excellent electrochemical performance, it can be suggested that PdNi@rGO/GCE is a promising electrode as a non-enzymatic glucose sensor. The long-term stability tests showed that, after eight weeks keeping in dry and cold conditions, PdNi@rGO/GCE modified electrode indicated excellent stability. In the light of these results, PdNi@rGO/GCE is an excellent candidate for fabricating glucose biosensors in clinical and biotechnology fields.

**Ethical Statement.** Authors confirm that all experiments were performed in accordance with relevant guidelines and regulations. Authors confirm that all methods were carried out in accordance with relevant guidelines and regulations. Authors confirm that all experimental protocols were approved by Dumlupınar University Scientific Research and Publication Ethics Committee. Authors confirm that informed consent was obtained from all subjects or, if subjects are under 18, from a parent and/or legal guardian.

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## References

- Crouch, E., Cowell, D. C., Hoskins, S., Pittson, R. W. & Hart, J. P. A novel, disposable, screen-printed amperometric biosensor for glucose in serum fabricated using a water-based carbon ink. *Biosens. Bioelectron.* **21**, 712–718 (2005).
- Clark, L. C. & Lyons, C. Electrode Systems for Continuous Monitoring in Cardiovascular Surgery. *Ann. N. Y. Acad. Sci.* **102**, 29–45 (1962).
- Erken, E. *et al.* New Pt(0) Nanoparticles as Highly Active and Reusable Catalysts in the C1–C3 Alcohol Oxidation and the Room Temperature Dehydrocoupling of Dimethylamine-Borane (DMAB). *J. Clust. Sci.* **27**, 9–23 (2016).
- Yildiz, Y. *et al.* Different ligand based monodispersed Pt nanoparticles decorated with rGO as highly active and reusable catalysts for the methanol oxidation. *Int. J. Hydrogen Energy* **42**, 13061–13069 (2017).
- Başkaya, G. *et al.* Rapid, sensitive, and reusable detection of glucose by highly monodisperse nickel nanoparticles decorated functionalized multi-walled carbon nanotubes. *Biosens. Bioelectron.* **91**, 728–733 (2017).
- Yildiz, Y. *et al.* Highly Monodisperse Pt/Rh Nanoparticles Confined in the Graphene Oxide for Highly Efficient and Reusable Sorbents for Methylene Blue Removal from Aqueous Solutions. *ChemistrySelect* **2**, 697–701 (2017).
- Ertan, S., Şen, F., Şen, S. & Gökagaç, G. Platinum nanocatalysts prepared with different surfactants for C1–C3 alcohol oxidations and their surface morphologies by AFM. *J. Nanoparticle Res.* **14**, 922–934 (2012).
- Li, C. *et al.* Electrochemical Deposition: An Advanced Approach for Templated Synthesis of Nanoporous Metal Architectures. *Acc. Chem. Res.* **51**, 1764–1773 (2018).
- Heller, A. & Feldman, B. Electrochemical glucose sensors and their applications in diabetes management. *Chemical Reviews* **108**, 2482–2505 (2008).
- Jiang, L. C. & Zhang, W. D. A highly sensitive nonenzymatic glucose sensor based on CuO nanoparticles-modified carbon nanotube electrode. *Biosens. Bioelectron.* **25**, 1402–1407 (2010).
- Park, S., Boo, H. & Chung, T. D. Electrochemical non-enzymatic glucose sensors. *Analytica Chimica Acta* **556**, 46–57 (2006).
- Ataee-Esfahani, H., Wang, L. & Yamauchi, Y. Block copolymer assisted synthesis of bimetallic colloids with Au core and nanodendritic Pt shell. *Chem. Commun.* **46**, 3684 (2010).
- Ataee-Esfahani, H. *et al.* Mesoporous Metallic Cells: Design of Uniformly Sized Hollow Mesoporous Pt-Ru Particles with Tunable Shell Thicknesses. *Small* **9**, 1047–1051 (2013).
- Park, S., Chung, T. D. & Kim, H. C. Nonenzymatic glucose detection using mesoporous platinum. *Anal. Chem.* **75**, 3046–3049 (2003).
- Li, Y., Song, Y. Y., Yang, C. & Xia, X. H. Hydrogen bubble dynamic template synthesis of porous gold for nonenzymatic electrochemical detection of glucose. *Electrochem. commun.* **9**, 981–988 (2007).
- Shen, Z. *et al.* Highly sensitive nonenzymatic glucose sensor based on nickel nanoparticle-attapulgite-reduced graphene oxide-modified glassy carbon electrode. *Talanta* **159**, 194–199 (2016).
- Li, L. H. & Zhang, W. D. Preparation of carbon nanotubes supported platinum nanoparticles by an organic colloidal process for nonenzymatic glucose sensing. In *Microchimica Acta* vol. 163, 305–311 Springer Vienna, (2008).
- Hwang, D.-W., Lee, S., Seo, M. & Chung, T. D. Recent advances in electrochemical non-enzymatic glucose sensors – A review. *Anal. Chim. Acta* **1033**, 1–34 (2018).
- Toghill, K. E. & Compton, R. G. Electrochemical non-enzymatic glucose sensors: A perspective and an evaluation. *Int. J. Electrochem. Sci.* **5**, 1246–1301 (2010).
- Jena, B. K. & Raj, C. R. Enzyme-free amperometric sensing of glucose by using gold nanoparticles. *Chem. - A Eur. J.* **12**, 2702–2708 (2006).
- Cui, H. F., Ye, J. S., Liu, X., Zhang, W. D. & Sheu, F. S. Pt-Pb alloy nanoparticle/carbon nanotube nanocomposite: A strong electrocatalyst for glucose oxidation. *Nanotechnology* **17**, 2334–2339 (2006).
- Kang, X., Mai, Z., Zou, X., Cai, P. & Mo, J. A sensitive nonenzymatic glucose sensor in alkaline media with a copper nanocluster/multiwall carbon nanotube-modified glassy carbon electrode. *Anal. Biochem.* **363**, 143–150 (2007).
- Rong, L. Q., Yang, C., Qian, Q. Y. & Xia, X. H. Study of the nonenzymatic glucose sensor based on highly dispersed Pt nanoparticles supported on carbon nanotubes. *Talanta* **72**, 819–824 (2007).
- Çelik, B. *et al.* Nearly monodisperse carbon nanotube furnished nanocatalysts as highly efficient and reusable catalyst for dehydrocoupling of DMAB and C1 to C3 alcohol oxidation. *Int. J. Hydrogen Energy* **41**, 3093–3101 (2016).
- Karatepe, Ö. *et al.* Enhanced electrocatalytic activity and durability of highly monodisperse Pt@PPy-PANI nanocomposites as a novel catalyst for the electro-oxidation of methanol. *RSC Adv.* **6**, 50851–50857 (2016).
- Liu, Y. *et al.* Horseradish peroxidase supported on porous graphene as a novel sensing platform for detection of hydrogen peroxide in living cells sensitively. *Biosens. Bioelectron.* **87**, 101–107 (2017).
- Zhang, C. *et al.* Facile fabrication of Pt-Ag bimetallic nanoparticles decorated reduced graphene oxide for highly sensitive non-enzymatic hydrogen peroxide sensing. *Talanta* **159**, 280–286 (2016).
- Demirkan, B. *et al.* Composites of Bimetallic Platinum-Cobalt Alloy Nanoparticles and Reduced Graphene Oxide for Electrochemical Determination of Ascorbic Acid, Dopamine, and Uric Acid. *Sci. Rep.* **9**, 12258 (2019).
- Pei, S. & Cheng, H. M. The reduction of graphene oxide. *Carbon N. Y.* **50**, 3210–3228 (2012).
- Wu, B., Zhao, N., Hou, S. & Zhang, C. Electrochemical synthesis of polypyrrole, reduced graphene oxide, and gold nanoparticles composite and its application to hydrogen peroxide biosensor. *Nanomaterials* **6**, 220 (2016).
- Shahnavaz, Z., Woi, P. M. & Alias, Y. Electrochemical sensing of glucose by reduced graphene oxide-zinc ferrosinels. *Appl. Surf. Sci.* **379**, 156–162 (2016).
- Li, C. *et al.* Emerging Pt-based electrocatalysts with highly open nanoarchitectures for boosting oxygen reduction reaction. *Nano Today* **21**, 91–105 (2018).
- Sreejesh, M., Dhanush, S., Rossignol, F. & Nagaraja, H. S. Microwave assisted synthesis of rGO/ZnO composites for non-enzymatic glucose sensing and supercapacitor applications. *Ceram. Int.* **43**, 4895–4903 (2017).
- Xu, X., Shen, J., Li, N. & Ye, M. Microwave-assisted synthesis of graphene/CoMoO<sub>4</sub> nanocomposites with enhanced supercapacitor performance. *J. Alloys Compd.* **616**, 58–65 (2014).
- Liu, X. *et al.* Microwave-assisted synthesis of CdS-reduced graphene oxide composites for photocatalytic reduction of Cr(vi). *Chem. Commun.* **47**, 11984 (2011).
- She, X. *et al.* Microwave-assisted synthesis of Mn<sub>3</sub>O<sub>4</sub> nanoparticles@reduced graphene oxide nanocomposites for high performance supercapacitors. *Mater. Res. Bull.* **70**, 945–950 (2015).
- Yao, C. F., Zhuang, L., Cao, Y. L., Ai, X. P. & Yang, H. X. Hydrogen release from hydrolysis of borazane on Pt- and Ni-based alloy catalysts. *Int. J. Hydrogen Energy* **33**, 2462–2467 (2008).
- Çelik, B. *et al.* Monodispersed palladium-cobalt alloy nanoparticles assembled on poly(N-vinyl-pyrrolidone) (PVP) as a highly effective catalyst for dimethylamine borane (DMAB) dehydrocoupling. *RSC Adv.* **6**, 24097–24102 (2016).
- Çelik, B. *et al.* Highly Monodisperse Pt(0)@AC NPs as Highly Efficient and Reusable Catalysts: The Effect of The Surfactant on Their Catalytic Activities in Room Temperature Dehydrocoupling of DMAB. *Catal. Sci. Technol.* **6**, 1685–1692 (2016).
- Clark, T. J., Whittell, G. R. & Manners, I. Highly efficient colloidal cobalt- and rhodium-catalyzed hydrolysis of H<sub>3</sub>N-BH<sub>3</sub> in air. *Inorg. Chem.* **46**, 7522–7527 (2007).
- Ozturk, Z., Sen, F., Sen, S. & Gokagac, G. The Preparation and Characterization of Nano-sized Pt-Pd/C Catalysts and Comparison of Their Superior Catalytic Activities for Methanol and Ethanol Oxidation. *J. Mater. Sci.* **47**, 8134–8144 (2012).



42. Şen, F., Gökağaç, G. & Şen, S. High Performance Pt Nanoparticles Prepared by New Surfactants for C1 to C3 Alcohol Oxidation Reactions. *J. Nanoparticle Res.* **15**, 1979–88 (2013).
43. Şen, F. & Gökağaç, G. Pt nanoparticles synthesized with new surfactants: Improvement in C 1-C3 alcohol oxidation catalytic activity. *J. Appl. Electrochem.* **44**, 199–207 (2014).
44. Şen, S., Şen, F. & Gökağaç, G. Preparation and characterization of nano-sized Pt–Ru/C catalysts and their superior catalytic activities for methanol and ethanol oxidation. *Phys. Chem. Chem. Phys.* **13**, 6784 (2011).
45. Erken, E., Esirden, I., Kaya, M. & Sen, F. A rapid and novel method for the synthesis of 5-substituted 1H-tetrazole catalyzed by exceptional reusable monodisperse Pt NPs@AC under the microwave irradiation. *RSC Adv.* **5**, 68558–68564 (2015).

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## Author contributions

K.C., S.K.G., S.K., E.S.I., and F.S. participated in the study design and coordination, conducted molecular studies and participated in the data analysis, discussion and manuscript preparation; K.C. and F.T. performed the characterization analysis; A.S. and K.A. carried out experimental studies and participated in the coordination. All authors have read and approved the article.

## Competing interests

The authors declare no competing interests.

## Additional information

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