

Review

Basophils Orchestrating Eosinophils' Chemotaxis and Function in Allergic Inflammation

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Abstract: Eosinophils are well known to contribute significantly to Th2 immunity, such as allergic inflammations. Although basophils have often not been considered in the pathogenicity of allergic dermatitis and asthma, their role in Th2 immunity has become apparent in recent years. Eosinophils and basophils are present at sites of allergic inflammations. It is therefore reasonable to speculate that these two types of granulocytes interact in vivo. In various experimental allergy models, basophils and eosinophils appear to be closely linked by directly or indirectly influencing each other since they are responsive to similar cytokines and chemokines. Indeed, basophils are shown to be the gatekeepers that are capable of regulating eosinophil entry into inflammatory tissue sites through activation-induced interactions with endothelium. However, the direct evidence that eosinophils and basophils interact is still rarely described. Nevertheless, new findings on the regulation and function of eosinophils and basophils biology reported in the last 25 years have shed some light on their potential interaction. This review will focus on the current knowledge that basophils may regulate the biology of eosinophil in atopic dermatitis and allergic asthma.

Keywords: basophils; eosinophils; chemotaxis; eosinophil-basophil interaction; allergic inflammation; eosinophil infiltration; atopic dermatitis; allergic asthma



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1. Eosinophils and Basophils Are Pivotal Players in the Progression of Allergic Inflammation

Type 2 immunity in the lung and skin is complex involving different types of cells, cells of adaptive and innate immunity, and a multi-layered network of cytokines, chemokines, and effector mediators [1]. Several studies demonstrate that basophils and eosinophils are essential contributors in the initiation, progression, and maintenance of allergic inflammations.

Basophils are the least abundant circulating granulocytes representing 0.5–1% of all leukocytes. They resemble mast cells in several characteristic features, especially in the expression of the high-affinity immunoglobulin E (IgE) receptor FcεRI on their cell surface, which directly binds to the IgE. The cross-linking of FcεRI and IgE complexes upon allergen challenge leads to the release of immunoregulatory and effector mediators, including Th2 cytokines like interleukin-4 (IL-4) and IL-13, histamine, and leukotriene (LTC₄) [2]. Due to their resemblance to mast cells and scarcity, basophils have often been considered insignificant and analyzed only as a surrogate of the tissue-dwelling mast cells. However, recent studies revealed the unique functions of basophils in chronic allergic diseases such as allergic rhinitis and asthma [3]. Infiltration of basophils and not mast cells during late-phase allergic reaction in the lungs is pivotal for the severity of the inflammatory response as evidenced by the observation that most histamine positive cells are basophils and the simultaneous absence of mediators like tryptase exclusively secreted by mast cells [4]. Histamine and pro-inflammatory lipid mediators (like LTC₄) released by activated basophils increase vascular permeability and mucus production and promote contraction of smooth muscle cells surrounding the bronchi thereby promoting

bronchoconstriction [2]. A non-redundant function of basophils in the context of allergic inflammation is the rapid secretion of IL-4 and IL-13 at levels greater than any other cells capable of secreting these cytokines, including Th2 cells, mast cells, and eosinophils [5]. The release of IL-13 stimulates goblet cells to produce more mucus and to proliferate [6]. In synergy with IL-13, IL-4 provides additional signals for isotype switching to IgE in B cells [5]. Moreover, it has been shown that IL-4 and IL-13 trigger CCL11 release and vascular cell adhesion molecule-1 (VCAM-1) expression from endothelial cells resulting in infiltration of eosinophils [2]. By producing IL-4 and IL-13, basophils regulate infiltration of eosinophils [7], which is confirmed in a recent study using basophil-deficient mice [8]. Furthermore, basophils also secrete various chemoattractants, including IL-8 (own observation [9]) and CCL5 (RANTES) [7], which promote the influx of potent inflammatory cells, neutrophils, eosinophils, and macrophages. Several lines of evidence from murine models suggest basophils' direct role in antigen presentation for Th2 responses through MHC-II expression [10–12], initiation, and maintenance of IgE-mediated Th2 responses possibly independent of T cells and mast cells (reviewed in [13]). Despite this evidence from mouse models of the multifaceted roles of basophils in Th2 allergic inflammation, the crucial function of basophils in human allergic diseases remains undefined because of inadequate supporting evidence.

In addition to the infiltration of basophils, migration of eosinophils to inflammation sites is another essential characteristic of allergic inflammation. Eosinophils are also relatively rare (about 1–5% of all leukocytes) circulating granulocytes. The content of eosinophil granules differs from that of basophils. It mainly comprises cationic proteins such as major basic protein (MPB), eosinophil cationic protein (ECP), eosinophil peroxidase, and eosinophil-derived neurotoxin [2]. However, some of the compounds secreted by eosinophils are also found in basophils, such as IL-4 and IL-13 [2]. In the context of allergic inflammation, the release of cationic mediators has been shown to cause injury to epithelial tissue damage and thereby contribute to airway hyperresponsiveness [14]. MBP released by eosinophils may cause a positive feedback loop since it provokes histamine release and LTC₄ from basophils [15].

Basophils and eosinophils are present at sites of allergic inflammation in the airways and skin [4,16–21]. Namely, increased numbers of basophils are reported in the lung tissues of fatal asthmatics [16], in sputum and bronchoalveolar lavage (BAL) fluid from asthmatics after allergen challenge [4,20]. Interestingly, upon allergen challenge, the increase in basophil percentages in BAL fluids negatively correlate with basophil percentages in the blood, indicating the recruitment of basophils from the circulation into the lung [20]. The presence of basophil infiltrates in atopic allergic inflammation in human skin was established using basophil granule-specific monoclonal antibody (mAb) [18]. In an extension of this observation, Ying and co-workers demonstrated, besides basophils, eosinophils infiltrate 6 h after allergen challenge in late-phase reaction [22]. Consistently, concomitant recruitment of basophils and eosinophils is reported in the skin lesions of eosinophilic pustular folliculitis and atopic dermatitis [21,23]. In asthma, elevated levels of eosinophil count and their mediator, MBP in BAL fluid from asthmatics are also reported [24]. A comparative study between basophils and eosinophils in atopic asthmatics identifies their presence in bronchial biopsies, and their numbers are further enhanced after allergen challenge in the late-phase of asthmatic and skin reactions [17]. Similarly, an increase in numbers of both airway basophils and eosinophils in atopic asthmatics' sputum after allergen inhalation challenge is observed [19]. Interestingly, it has been demonstrated that eosinophil-derived MBP directly activates the release of histamine and LTC₄ from human basophils [15,25]. Consistent with this observation, in several experimental allergy models, basophils and eosinophils appear to be closely linked by directly influencing each other or indirectly due to being responsive to similar cytokines and chemokines [7,8,26]. Therefore, it is reasonable that basophils and eosinophils interact with each other at sites of allergic inflammation. In 1995, Thomas et al. reviewed the potential functional network between basophils and eosinophils [27]. New findings in recent years give some more insights into

the mechanisms of interaction between these two types of granulocytes. Eosinophils are increased in allergic inflammation and are often considered the main actors of the late phase of allergic rashes. However, although present at low numbers, basophils seem to be pivotal for allergic inflammation progression and maintenance. We will summarize whether and how basophils can regulate the function of eosinophils in allergic dermatitis and asthma.

2. Chemotaxis

There are several potential mechanisms on how cells of different types may regulate each other. One of them is undoubtedly the attraction of a particular cell type from circulation to a specific tissue guided by an increasing gradient of chemoattractants. This type of cell migration is known as chemotaxis. It is well established that tissue-residing mast cells play an essential role in triggering infiltration of eosinophils and basophils to sites of inflammation (reviewed in [28]). Moreover, IL-5, produced by allergen-specific T-cells, is considered a key molecule of eosinophil chemotaxis [29,30], whereby systemically delivered IL-5 seems to be superior to IL-5 expressed locally in the airways on impacting airway eosinophilia [31]. Whether basophils and eosinophils attract each other to sites of inflammation such as irradiated skin lesions or inflamed airways is less well documented. Nevertheless, studies using basophil and eosinophil-depleted mice shed some light on this topic and will be summarized in the next section (Tables 1 and 2).

Table 1. Mouse models of allergic skin diseases.

| Model | Sensitization | Challenge | Way of Basophil Assessment | Diminished Eosinophil Infiltration | Reference |
|---------|--------------------------|----------------------------|------------------------------------|------------------------------------|-----------|
| IgE-CAI | i.v. TNP-specific IgE | s.q. TNP-OVA | CD49b ^{pos} cell fraction | Yes | [32] |
| | | | Anti-CD200R3 (B103) | Yes | [33] |
| | | | <i>Mcpt8Cre</i> BAC | Yes | [34] |
| | | | BasoDTR mice | Yes | [35] |
| ICD | i.p NAC | NAC and croton oil on skin | Anti-FcεRIα (MAR-1) | Yes | [36] |
| | | | BasTRECK | Yes | [36] |

IgE-CAI, IgE-chronic allergic inflammation; OVA, ovalbumin; ICD, irritant contact dermatitis; NAC, N-Acetylcystein.

Table 2. Mouse models of allergic airway inflammation.

| Model | Sensitization | Challenge | Way of Basophil Assessment | Diminished Eosinophil Infiltration | Reference |
|--------------------|---------------|-----------|----------------------------|------------------------------------|-----------|
| OVA-induced asthma | i.p. OVA-alum | i.t. OVA | Anti-FcεRIα (MAR-1) | No | [37] |
| | | | <i>Mcpt8Cre</i> BAC | No | [34] |
| | i.p. OVA-alum | i.n. OVA | Anti-FcεRIα (MAR-1) | Yes | [38] |
| | | | Anti-CD200R3 (B103) | Yes | [38] |
| HDM-induced asthma | i.n. HDM | i.n. HDM | Anti-FcεRIα (MAR-1) | Yes | [39] |
| | i.n. HDM | i.n. HDM | Anti-CD200R3 (B103) | Yes | [39] |

OVA, Ovalbumin; HDM, house dust mite.

2.1. Contribution of Basophils to Regulate Eosinophil Infiltration to Inflamed Skin

By subcutaneous injection of TNP-conjugated ovalbumin into anti-TNP IgE transgenic mice [40], Mukai and co-workers were able to induce an immediate as well as a late and chronic phase of IgE-mediated chronic allergic dermatitis (IgE-CAI) as evident by infiltration of granulocytes and ear swelling 2–4 days after injection. Interestingly, the induction and progression of early, late, and chronic allergic inflammation was not dependent on T cells, NK-T cells, and NK cells. In contrast, mast cells impacted granulocyte infiltration during the immediate and late but not the chronic phase of allergic skin inflammation [32].

Basophil-depleted mouse models did not exist at that time point. However, some indirect evidence that basophils are regulators of eosinophil infiltration during the chronic phase of allergic inflammation was given by the fact that transfer of the DX5 (CD49b) positive cell fraction, consisting 15–25% of basophils, into Fc γ R $^{-/-}$ mice restored IgE-CAI [32].

More direct evidence that basophils regulate eosinophil infiltration and disease progression in IgE-CAI is given in a murine model in which basophils were depleted by the usage of the basophil specific anti-CD200R3 antibody (Ba103). Depletion of basophils inhibited the infiltration of eosinophils and the development and maintenance of IgE-CAI. In contrast, in passive cutaneous anaphylaxis, mediated mainly by mast cells, and contact hypersensitivity, a model for type IV hypersensitivity, the clinical symptoms and eosinophil infiltration were not affected by basophil depletion [33]. These findings are in line with studies performed in constitutively basophil-depleted *Mcpt8Cre* BAC transgenic mice [34].

Expressing the human diphtheria toxin (DT) receptor under *CD203* or the eosinophil peroxidase promoter, basophil, and eosinophil depletion, respectively, was obtained upon administration of DT [35]. IgE-CIA was abolished in basophil-depleted BasoDTR mice, confirming the essential role of basophils to control eosinophil infiltration. Besides, Th2 cytokines (IL-4, IL-13, IL-6) and Th1 cytokines (IFN γ and IL-12) were reduced. Notably, CCL24 (eotaxin-2) expression, known to regulate eosinophils' infiltration, was significantly decreased in BasoDTR mice. In eosinophil-ablated EoDTR mice, IgE-CIA was diminished as well. Unfortunately, whether the attraction of basophils to inflamed skin was affected in the EoDTR mice was not addressed. Nevertheless, although the cytokines as mentioned above were expressed to a lesser degree in EoDTR mice compared to wild-type mice, EoDTR mice showed higher amounts of these cytokines than BasoDTR mice suggesting that basophils initiate and eosinophils ameliorate IgE-CIA [35].

In line with the IgE-CAI mouse models are reported data of croton oil-induced irritant contact dermatitis (ICD). Basophils infiltrated lesion skin within 3 h, followed by eosinophil migration at 24 h upon induction of ICD, suggesting that basophils have the capacity to imprint attraction of eosinophils by the simple fact that basophil infiltration occurs earlier than the one of eosinophil. Further analysis in anti-Fc ϵ RI α antibody (MAR-1)-treated and basophil-depleted Bas TRECK mice confirmed a significant reduction of eosinophils in basophil-depleted mice [7].

Therefore, these mouse studies point to basophils' vital role in regulating eosinophil infiltration to irradiated skin lesions during chronic allergic dermatitis. However, in contrast to these mouse models, in atopic asthma patients in whom late-phase skin reaction was induced, eosinophil infiltration to inflamed skin occurred prior to basophil infiltration, whereby maximal numbers of eosinophils were already observed at 6 h followed by basophils infiltration at 24 h [17,22]. In skin lesions of atopic dermatitis, the accumulation of both basophils and eosinophils are reported [21,36]. It has been widely reported that eosinophil recruitment is associated with increased expression of adhesion molecules, including VCAM-1 [41,42] and specific chemotactic factors such as CCL11 [43]. Interestingly, in basophil-depleted mice, eosinophil infiltration was found abrogated, and furthermore, the basophil-specific deletion of IL-4 showed diminished VCAM-1 up-regulation on endothelial cells [8]. Consistently, the accumulation of basophils that express IL-4 in skin lesions of atopic dermatitis patients was observed [36]. Taken together, it is plausible that infiltrated basophil represent a source of IL-4 that aids in eosinophil recruitment in atopic dermatitis. However, further investigations are necessary to better understand the functional significance of skin-infiltrating basophils in human atopic dermatitis.

2.2. The Controversial Role of Basophils in Affecting Eosinophil Infiltration to Sites of Inflammation in the Airways

Findings in mouse models of allergic airway inflammation are less consistent than the ones of IgE-CAI and chronic contact dermatitis. For instance, a mouse study of OVA-induced late asthmatic response revealed that basophil infiltration to sites of allergic airway inflammation and the release of IL-4, IL-5, and IL-13 into BAL fluids occurs before eosinophil, T cell, and monocyte infiltration [37]. However, although depletion of basophils

and mast cells by anti-Fc ϵ RI α MAR-1 antibody reduced the early phase of specific airway resistance (sRAW), the late phase of sRAW and late eosinophil infiltration to lung tissue was not affected [44]. Consistently, using the basophil-depleted *Mcpt8Cre* BAC transgenic mice, Ohnmacht and their research team demonstrated that the infiltration of eosinophils was not abrogated upon OVA challenge [34]. In contrast, MAR-1 and Ba103 treatment resulted in a significant reduction of eosinophil number in BAL fluid and airways, and reduced IL-4 expression in lung tissue in the study conducted by Zhong et al. [38]. Besides, whether basophils have the capacity to contribute to eosinophil infiltration seems to not only depend on the approach of basophil depletion, but also on the allergen used to induce airway inflammation. For instance, Hammad and co-workers compared the effects of the MAR-1 and Ba103 antibody, respectively, on the infiltration of eosinophils and Th2 cytokine secretion in BAL fluids in an allergic airway inflammation models using house dust mite (HDM) [39]. They found that MAR-1 antibody administration, which recognizes Fc ϵ RI α expressed on basophils, dendritic cells, and mast cells, resulted in a significant reduction of eosinophil infiltration and Th2 cytokine production. In contrast, treatment with the basophil specific anti-CD200R3 antibody Ba103 did not significantly affect Th2 cytokine release in BAL fluids. Furthermore, it resulted in a less pronounced but still significant reduction of eosinophil infiltration compared to MAR-1 treated mice [39]. Whether eosinophil depletion affects infiltration of basophils has not been studied in detail. However, interestingly enough, in an HDM-asthma model, it has been shown that eosinophil depletion by the usage of anti-CCR3 antibodies resulted in an 80 to 85% reduction of eosinophils in BAL and lung but did not affect numbers of Th2 cells and airway remodeling. These findings were confirmed in Δ dblGATA eosinophil-ablated mice [45].

In general, basophils' role in regulating eosinophils' trafficking seems different in allergic skin and airway inflammation. The reasons for this are manifold and certainly include the fact that allergic asthma and atopic dermatitis are two separate diseases with different underlying mechanisms. In addition, conflicting results can be explained by the undeniable fact that mouse models do not always represent human diseases. We refer the reader to the review of Graham et al. summarizing allergic mouse models and their contribution to understand the human diseases [46].

2.3. Overview of the General Mechanisms of Eosinophil and Basophil Chemotaxis

The regulation of eosinophils' chemotaxis has been investigated in many studies. Before we describe the potential mechanism of how basophils may guide the infiltration of eosinophils to inflamed skin, we will shortly outline the most general means of chemotactic regulation of basophils and eosinophils (also summarized in Figure 1).

2.4. The CCR3/CCL11/CCL24 Axis

CCL11 (eotaxin-1), CCL24 (eotaxin-2), CCL26 (eotaxin-3), CCL5 (RANTES), CCL3 (MIP-1 α), and CCL7, CCL8, CCL13 (MCP-3, -2 and 4), all recognized by CCR3, are prominent eosinophil-active chemokines. CCR3 is constitutively expressed on eosinophils from mice [47] and humans [48] and on basophils from humans [49] but not from mice. Murine models of allergic airway inflammation demonstrated the prominent role of CCR3 in regulating infiltration of eosinophils to inflamed lung tissue. Depending on the immunization protocol that has been used to induce airway inflammation, numbers of eosinophils in BAL fluid and lung tissue were modestly [50] or strongly reduced in CCR3-deficient mice [51,52]. It is less well known whether CCR3 regulates infiltration of basophils to inflamed airways to a similar extent as eosinophils since basophils from mice do not express CCR3. Nevertheless, Sénéchal and co-workers demonstrated in a mouse model of human skin xenografts and autologous transfer of PBMC from allergic donors that basophil infiltration to inflamed skin was not affected by anti-CCR3 antibody administration prior to allergen challenge [53].

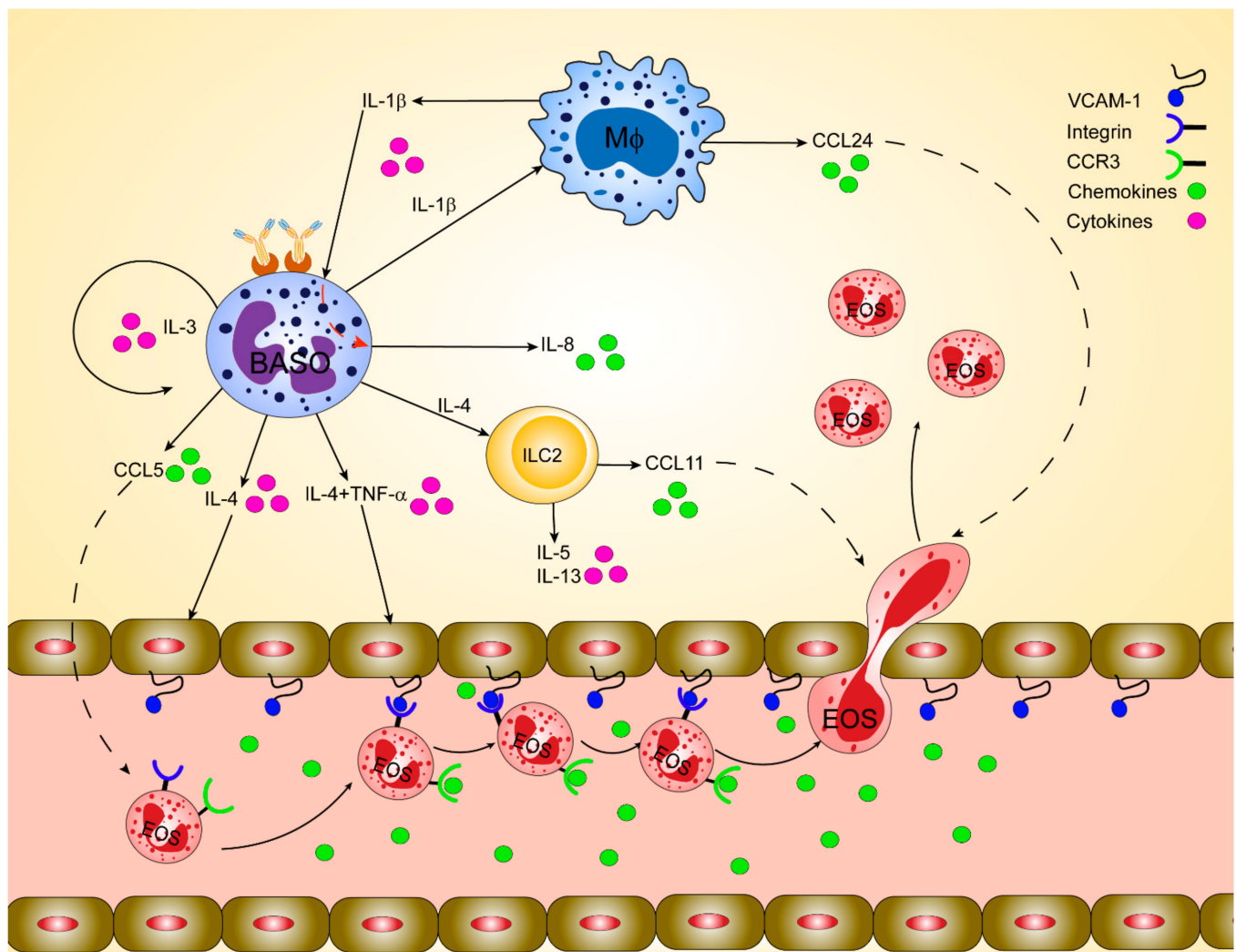


Figure 1. Orchestration of basophil-mediated eosinophil recruitment during allergic inflammation in humans. Basophils play a crucial immunoregulatory role in releasing cytokines such as interleukin (IL)-4 and IL-13 at the inflammatory tissue foci. Basophils-derived IL-4 induces the upregulation of the adhesion molecules like vascular cell adhesion molecule-1 (VCAM-1) on endothelial cells, which facilitates eosinophil infiltration. Basophil-derived IL-4 in synergy with tumor necrosis factor-alpha (TNF- α) stimulates fibroblasts and endothelial cells to produce eotaxins, including CCL11 and CCL24, which can, in turn, attract CCR3positive eosinophils. IL-4 activates lung-resident group 2 innate lymphoid cells (ILC2) to release IL-5, IL-13, and CCL11, stimulating eosinophils' infiltration. IL-13 stimulates airway macrophages to release CCL24, which regulates eosinophil migration to the airways. Infiltrated inflammatory cells like macrophages and monocytes in the airways activate and secrete IL-1 β , which in turn induce basophils to release the potent neutrophil chemoattractant, IL-8. IL-3 secreted by T cells, mast cells, or even basophils themselves induces the expression of CCL5 in basophils, which in turn regulates the trafficking of eosinophils.

CCL11, CCL24, and CCL26 are high-affinity ligands of CCR3. The levels of plasma CCL11 are significantly increased in atopic dermatitis patients [54] and during asthma exacerbation compared to the stable phase of asthma [55]. Moreover, elevated levels of CCL11 in plasma seem to have functional consequences since they are inversely associated with predicted FEV1 [56]. In line with this, is the observation that in the airways of asthma patients compared with healthy controls mRNA of CCL11 and CCL24 are increased at steady state. However, in contrast to CCL26 mRNA, CCL11 and CCL24 mRNAs are not further upregulated in the airways upon allergen challenge [57]. The same accounts for atopic dermatitis patients in whom serum levels of CCL26 correlate with disease severity while CCL24 does not [58].

Interestingly, CCL11 and CCL24 might have different importance in health and disease. On the one hand, CCL11, which, in humans, is constitutively expressed in the small intestine and colon and to a lesser extent in heart, kidney, and pancreas [59–61], seems to be more prominent than CCL24 in regulating eosinophil infiltration to the gastrointestinal tract at baseline in healthy state [62,63]. CCL24, on the other hand, seems to dominate the regulation of eosinophil infiltration under pathophysiological conditions. For instance, CCL24 and CCL11/CCL24 double-deficient mice show fewer eosinophils in BAL fluids 20 h upon OVA-induced experimental airway inflammation, while numbers of eosinophils in CCL11 knockouts were comparable to wild-type control mice [40]. This might be explained by the fact that in contrast to CCL11, the expression of CCL24 is prolonged up to 120 h upon allergen challenge and thus has the capacity to regulate eosinophil infiltration at later time points. Consistently, targeted gene disruption of CCL11 results in reduced numbers of eosinophils in BAL fluid after 18 h but not after 48 h OVA-challenge [64]. In terms of basophil chemotaxis, *in vitro* tests demonstrated that CCL11 and CCL24 are chemoattractants of human basophils, whereby CCL11 seems to be more potent than CCL24 in attracting basophils [49,65,66]. Eosinophil- and basophil-active chemotaxis of CCL11 and CCL24 and their temporal expression was confirmed in humans upon cutaneous allergen challenge. However, expression of CCR3, CCL11, and CCL24 did not correlate with numbers of basophils upon cutaneous allergen challenge questioning whether the eotaxin-CCR3 axis is essential in regulating infiltration of basophils to inflamed skin in humans [22].

Besides the eotaxin-family, the other known agonists of CCR3 have been demonstrated to impact eosinophil and basophil attraction. For instance, increased concentrations of CCL5 and CCL3 have been detected in BAL fluid of asthma patients compared to healthy non-smokers [67]. The chemotactic capacity of these ligands has been demonstrated by *in vitro* tests using trans-wells in which BAL fluid was added to one side and purified eosinophils from asthma patients to the other side. Importantly, the usage of anti-CCL5 and anti-CCL7 antibody demonstrated inhibition of eosinophil chemotaxis [67].

There are several recent reviews summarizing findings on the regulation of eosinophil and basophil chemotaxis to inflamed skin and lung and the further effects of the corresponding chemokines such as induction of granulocyte degranulation [68,69].

2.5. The GM-CSF Cytokine Family Axis

Members of the GM-CSF cytokine family (IL-3, IL-5, and GM-CSF) tremendously affect the maturation, development, and biology of eosinophils and basophils. Several studies investigated the contribution of IL-3, IL-5, and GM-CSF on chemotaxis (reviewed [70]). For instance, IL-5 and GM-CSF are found elevated in the BAL fluids from allergic subjects after the antigen challenge [71–73]. Interestingly, it has also been observed that eosinophils from BAL fluid of antigen challenged-allergic subjects showed enhanced transendothelial migration, suggesting eosinophils are activated *in vivo* by IL-5 and GM-CSF [74].

Receptors for all three GM-CSF cytokine family consist of a cytokine specific α -chain (IL-3R α , IL-5R α , or GM-CSFR α) and a common β -chain [75]. Both blood basophils and eosinophils express receptors for all members of the GM-CSF cytokine family. The sensitivity and magnitude of cell's responses elicited by different GM-CSF family members are different and dependent on the density of surface expression of the corresponding receptor α -chains on basophils and eosinophils [76]. For instance, IL-3R α mRNA is abundantly expressed in basophils, whereas in eosinophils, IL-5R α mRNA is dominant among all the α -chains of three GM-CSF cytokine receptors [77]. Consistently, among the three GM-CSF cytokines, IL-3 potently activates various basophil functions, whereas IL-5 efficiently activates eosinophils [76].

IL-3, IL-5, and GM-CSF play an important role in adhesion and migration of basophils and eosinophils across endothelial cells via the upregulation of β 2 integrins (CD11/CD18) expression [74,77–81]. IL-3 was observed to be more potent in upregulating CD11b expression in basophils than in eosinophils, while IL-5 and GM-CSF were almost equipotent in

both cell types [79]. In studies using IL-3, IL-5, and GM-CSF cytokines at low concentrations (150 pg/mL), IL-3 seems to be less efficient than IL-5 or GM-CSF on eosinophils' migration across endothelial cells [74]. On the other hand, at higher concentrations of all the three GM-CSF cytokine members (5 ng/mL), eosinophil migration was equally enhanced [80]. Interestingly, by inhibiting the re-consumption of IL-3 by basophils using a neutralizing antibody, Schroeder et al. demonstrated that basophils release IL-3 upon cross-linking the high-affinity IgE receptor [82]. Hence one may speculate that basophils that infiltrated inflamed tissue attract eosinophils by releasing IL-3 upon allergen binding.

IL-3, IL-5, and GM-CSF effectively prime eosinophils for enhanced chemotactic responses to various stimuli such as platelet-activating factor, leukotriene B₄, and complement C5a [83]. The most characterized synergism to promote eosinophilia is the interaction between IL-5 and eotaxin. Studies conducted on eosinophil trafficking in animal models indicate that IL-5 and eotaxin synergistically regulate eosinophils' migration during allergy [84–86].

2.6. Mechanisms of How Basophil Imprint Eosinophil Migration to Inflamed Skin and Lung

Basophils constitute only a minor population infiltrating the tissue compared to eosinophils. For instance, in IgE-CAI basophil numbers, which were identified as FcεRIpos, c-Kitneg, accounted for only 2% of infiltrating cells during the chronic phase, in contrast to eosinophils that accounted for the majority of infiltrating cells, namely 40% [32]. Therefore, it is justified to ask how such a minority can have such a significant influence. The study of Cheng et al. gives some more mechanistic insights into basophil and eosinophil infiltration dependency. In three different-IgE-dependent models of atopic dermatitis, basophil-derived IL-4 was shown to induce VCAM-1 expression on endothelial cells of the skin, which resulted in infiltration of eosinophils [8]. Consistently, by using transgenic basophil-depleted mice, eosinophil infiltration was abrogated. Moreover, by eliminating IL-4 from basophils, the study demonstrates the non-redundant role of IL-4-producing basophils in upregulating VCAM-1 on endothelial cells and regulating infiltration of eosinophils. These findings are consistent with the fact that the interaction between α₄β₁ integrin, expressed on eosinophils and VCAM-1, displayed by endothelial cells, plays a significant role in the adhesion of eosinophils to tissue (reviewed in [87]). Thus, basophil-derived IL-4 might be a possible mechanism for how eosinophil infiltration to inflamed skin is regulated. Moreover, intranasal injection of soluble IL-4R before OVA-challenge reduced VCAM-1 expression on endothelial cells of blood vessels in the airways and blocked eosinophil infiltration [88]. Consistently, in IL-4 deficient mice, fewer eosinophils were observed in BAL fluid than in wild-type mice [89]. Besides, it is well established that in synergy with TNF-α, IL-4 promotes the release of CCL11 [90,91], CCL24 [92], and CCL26 [93] from endothelial cells and fibroblasts. Unfortunately, basophils have not been addressed in these studies. Notably, however, an increased level of IL-4 concomitantly with basophil infiltration upon parasite infection was observed, and basophils are known to be the major source of IL-4 [94–97]. Interestingly, induction of eotaxin-family members is STAT6 dependent [98,99]. Voehniger et al. observed in lung tissue a non-T non-B cell population that produce IL-4, which was responsible for eosinophil infiltration and dependent on STAT6 [96]. One may therefore assume that basophil-derived IL-4 induces eotaxin-family members in a STAT6-dependent manner, which in turn results in eosinophil infiltration. A papain-induced airway inflammation model further gives evidence that basophils indirectly trigger eosinophil infiltration. Namely, basophil-derived IL-4 activated lung-resident group 2 innate lymphoid cells (ILC2s) to release IL-5, IL-13, and CCL11, which stimulate infiltration of eosinophils [26].

Besides IL-4, basophils are known to produce IL-13 upon FcεRI cross-linking [100,101]. Pope and their research team demonstrate that intratracheal injection of IL-13 into CCL24-deficient mice resulted in reduced numbers of eosinophils in BAL fluid. Interestingly, macrophages in the airway lumen seemed to be the major source of CCL24 [63]. It is

therefore quite reasonable to speculate that basophil-derived IL-13 stimulates airway macrophages to release CCL24, which regulates eosinophil recruitment to the airways.

Further, *in vitro* studies revealed that basophils regulate eosinophils' infiltration in co-operation with fibroblast, whereby basophil-derived TNF- α and IL-4 seem to stimulate CCL11 expression in fibroblast, which in turn attract eosinophils through CCR3. Furthermore, IL-3 induces high expression of CCL5 in basophils, which is further enhanced by co-culturing of basophils with fibroblast and basophil-derived TNF- α [7]. It is, therefore, likely that fibroblast-derived CCL11 and IL-3-mediated induction of CCL5 regulate the trafficking of eosinophils [102]. The importance of IL-3 in regulating attraction of eosinophils to airways is further given by the fact that IL-3 is a very potent inducer of IL-4 and IL-13 from human basophils [76,103,104]. Interestingly, basophil-derived IL-4 and IL-13 can be inhibited by a blocking anti-IL-3R α antibody [76] and INF- α treatment [104]. Hence, it would be interesting to investigate whether, under such treatments, eosinophil infiltration can be blocked. Moreover, IL-1 β induces in synergy with IL-3 the release of IL-8 (own observation [9]), which was shown to cause eosinophil chemotaxis [105] and thus provides another mechanism basophil regulate recruitment of eosinophils.

Besides cytokines and chemokines, basophil specific proteases contribute to the recruitment of eosinophils to inflamed tissues. Namely, mouse mast cell proteases (mMCP)-8, which is expressed explicitly in mouse basophils, triggers leukocyte infiltration by upregulating the expression of CCL24 (reviewed in [106]). Interestingly, mMCP-8 is highly homologous to human Granzyme B, which is released upon activating IL-3-primed basophils with various agonists (anti-Fc ϵ RI α , C5a, fMLP) [107]. Whether Granzyme B contributes to eosinophils chemotaxis in humans has, however, not yet been addressed. In addition, mMCP-11, a protease preferentially expressed in basophils, seems to attract eosinophils indirectly by proteolytic cleavage of a yet unidentified serum protein, whose product triggers recruitment of eosinophils. Consistently, mMCP-11-deficient mice show a diminished IgE-CIA. Curiously, however, mMCP-11-deficient basophils produce IL-4 in a normal range upon MCP-2 stimulation (reviewed in [106]), indicating that other mechanisms than IL-4-dependent induction of VCAM-1 contribute to eosinophil infiltration.

3. Cell-to-Cell Interaction at Sites of Inflammation

Another obvious mechanism, besides chemotaxis, in which cells of different origins may influence each other is the cell-to-cell contact. A recent book chapter by Landolina et al. summarizes the interaction of mast cells and eosinophils. The authors describe the formation of an allergic effector unit (AEU), enabling an enhanced function of both cell types [108]. In addition, studies have demonstrated that basophils and fibroblasts/endothelial cells directly interact. For instance, direct intercellular contact between basophils and fibroblasts was needed to mediate basophil-derived IL-8, CCL2, and CCL5 secretion upon NOD2/TLR2 stimulation [109]. Moreover, Fc ϵ RI and integrin signaling resulted in clustering of basophils at endothelial junctions upon allergen challenge in IgE-DNP sensitized mice. This arrest allowed the local increase of basophil-derived IL-4, which activates IL-4 receptor-expressing endothelial cells to upregulate VCAM-1 [8]. Consistently, the work of Schroeder and colleagues suggests that cell-to-cell interaction between basophil and lung epithelial cells triggers histamine, IL-4, and IL-13 release from basophils [110]. Moreover, studies on whether basophils may act [10,12,111,112] or not [113] as antigen-presenting cells (APC) stimulated the interest in investigating whether and if so, how basophils interact with T cells and dendritic cells via cell-to-cell contact. For instance, by using YFP fluorescence expressing basophils and OVA-specific T cells that expressed the red fluorescence protein DsRed, Sullivan and co-workers demonstrated that basophils interact with T cells in lung tissue but not in lymph nodes upon an *N. brasiliensis* infection [114]. These findings challenged, on one side, the role of basophils as APC in lymph nodes, but underscore that basophils interact with T cells by direct cell-to-cell contact resulting in worsening of Th2 inflammation [115]. Several studies published a few years later further

investigated the interaction between basophils and different subpopulations of T cells in variable inflammatory conditions in more detail (reviewed in [116]).

Unfortunately, however, whether basophils and eosinophils have direct cell-to-cell interaction during allergic inflammation is poorly defined. Nevertheless, the simultaneous presence of basophils and eosinophils at sites of inflamed skin was observed in chronic allergic dermatitis (ICD) patients giving evidence of basophil and eosinophil couple formation *in vivo*. Additionally, the same study confirmed the vicinity of basophils and eosinophils at inflammatory foci in an IgE-CAI mouse model [33]. Consistently, basophils promote activation of eosinophils by direct cell interaction, which induces activation markers (CD69, CD86, and intercellular adhesion molecule-1) on eosinophils [7]. Recently, basophils, eosinophils, and T cells have been found to co-localize in lung tissue of COPD patients [117]. These few studies imply that eosinophils and basophils co-localize in inflamed tissue and may affect each other's function by cell-to-cell contact. However, currently, the volume of research on that topic is small, not allowing sound conclusions. Moreover, direct evidence that basophil and eosinophils form a network of soluble mediators and the corresponding receptors are mainly missing, too. It is reasonable to speculate that a soluble mediator-receptor network exists between these two types of granulocytes simply because both eosinophils and basophils produce and release a plethora of cytokines, chemokines, and growth factors [13,118], as stated elsewhere [27].

4. Common Molecular Targets on Basophils and Eosinophils for Novel Therapies in Allergic Inflammation

The simultaneous presence of both basophils and eosinophils has been identified at the inflammation sites in chronic allergic disorders such as dermatitis, and asthma. Although they are considerably different in their ability to respond to stimuli, they share similar characteristics in terms of surface receptors and the inflammatory mediators they secrete. For instance, both basophils and eosinophils respond to inflammatory stimuli such as IL-3, IL-5, IL-33, and generate inflammatory cytokines such as IL-4 and IL-13 during allergic inflammation. Therefore, targeting both basophils and eosinophils together would be an effective anti-allergic therapeutic strategy. In this section, we summarize the possible common molecular targets for basophils and eosinophils.

IL-5-IL-5R α axis: Three monoclonal antibodies targeting the IL-5-IL-5R α axis have been developed for clinical purpose. Among them, mepolizumab and reslizumab are humanized mAb that target and neutralize circulating IL-5. Both of them have been approved for the treatment of adults with severe eosinophilic asthma [119]. Though basophil progenitors are unlikely to depend on IL-5 for development, blood basophil counts measured in routine clinical laboratories suggest they decrease following mepolizumab treatment [120]. However, a recent study by Wright et al. has demonstrated that mepolizumab does not alter the blood basophil count, and therefore, clinical benefit is likely to be independent of basophils [121]. Like mepolizumab, reslizumab treatment profoundly decreased circulating eosinophil counts in patients with asthma [122], but its effects on basophils are not investigated. Benralizumab is directed against the membrane-expressed IL-5R α -chain and induces antibody-mediated cellular cytotoxicity (ADCC) in eosinophils, depleting eosinophils both in blood and tissues [123]. Expression of the IL-5R α -chain by basophils makes it a potential target of benralizumab. Laviolette et al. showed that blood basophils counts were profoundly reduced with benralizumab treatment, but in this study, basophils have been studied less extensively, and the clinical relevance is unknown [124].

IL-3-IL-3R α axis: We have previously shown that blocking of the IL-3R α -chain with the CSL360, a humanized anti-IL-3R α mAb, significantly ameliorates the basophil phenotype and function [76]. Yet, there are no clinical trials conducted with this mAb. CSL362 is another humanized anti-IL-3R α mAb derived from clone 7G3 that targets and neutralizes IL-3. As predicted from *in vitro* and *ex vivo* studies, CSL362 produced a marked, dose-dependent and sustained depletion of peripheral blood basophils and plasmacytoid dendritic cells (pDCs) [125]. However, there has been no investigation conducted whether CSL362 affect eosinophil blood counts.

Anti-alarmins: Airway epithelium is in constant contact with various external stimuli, such as infectious agents, environmental allergens, and atmospheric pollutants. In response to such stimuli, airway epithelium releases epithelial-derived cytokines, such as IL-25, IL-33 collectively known as “alarmins”. Among them, IL-33 has been shown to activate both basophils and eosinophil phenotype, making them attractive targets for biologics [126]. Monoclonal antibodies targeting IL-33 or its receptor, ST2, are in clinical development, with several in phase 2 trials [127]. Nevertheless, anti-IL-33 and ST2 treatment reduced airway inflammation in a murine model of asthma and found that both treatments reduced the total cell counts and eosinophil counts in BAL fluid and significantly reduced the Th2 cytokine [128]. Recently, Jeannne Allinne et al. conducted a murine study with IL-33 neutralizing antibody REGN3500, which is currently in clinical development to treat asthma and COPD. Anti-IL-33 neutralizing antibody REGN3500 actively reduced the established HDM-induced lung infiltration by eosinophils, ST2-positive CD4+ T-cell numbers, and Th2 cytokine levels in lungs, and blocks the further increase in lung neutrophilia and levels of pro-inflammatory cytokines [129]. However, whether anti-IL-33 or anti-ST2 treatment affects basophils has not been investigated in the above-mentioned studies.

Anti-CCR3: The potential of CCR3 as a therapeutic target was established through the observations that CCR3-null mice and eotaxin-1 and eotaxin-2 double-knockout mice displayed significantly reduced allergen-induced airway eosinophil recruitment [130]. A short-term study was conducted in patients with allergic asthma and control subjects using a CCR3 antagonist, AXP1275. No significant effect on the number of circulating eosinophils and sputum eosinophils and metachromatic cells (mast cells and basophils) was observed [131]. R321 is a novel, biased nanoparticle CCR3 antagonist that inhibits G-protein signaling but not β -arrestin-mediated CCR3 internalization and degradation. Intravenously administered R321 significantly reduces eosinophil recruitment into the blood, lungs, and airways and prevents airway hyperresponsiveness in a mouse eosinophilic asthma model [132]. Nevertheless, the effect of R321 on basophils is still unknown.

CRTH2 and PGD2 axis: Chemoattractant receptor-homologous molecule (CRTH2) is highly expressed on human peripheral blood basophils and eosinophils and has been shown to mediate chemotaxis of eosinophils and basophils. Prostaglandin D2 (PGD2), one of the key mediators released by degranulating eosinophils and basophils upon allergen exposure, is the principal ligand of CRTH2. Royer et al. showed that AZ11665362, a novel CRTH2 antagonist, is more potent than Ramatroban (BAYu3405, the only clinically available CRTH2 antagonist) in inhibiting the migration of both human eosinophils and basophils. Besides, AZ11665362 abrogates the PGD2-induced mobilization of eosinophils from the bone marrow of the guinea pig [133]. However, more extensive clinical studies are necessary to understand the clinical relevance of AZ11665362.

Anti-inflammatory cytokines IL-4 and IL-13: Among the recently developed anti-IL-4 and anti-IL-13 biologic drugs, dupilumab is very promising given its ability to inhibit the biological effects of both IL-4 and IL-13. Dupilumab is a human mAb that is directed against the shared α -subunit of the IL-4 receptors. It blocked the signaling from IL-4 and IL-13 and showed efficacy in patients with moderate-to-severe asthma and airway and peripheral eosinophilia [134]. Dupilumab is the only biologic for asthma treatment that simultaneously reduces the severe exacerbations but on the other hand transiently increased blood eosinophilia [135–137]. The increase in the blood eosinophils is consistent with the hypothesis that dupilumab blocks the migration of eosinophils into tissue by inhibiting IL-4 and IL-13-mediated production of eotaxin and VCAM-1 expression. Dupilumab was also approved for treatments in adults with moderate-severe atopic dermatitis [138]. However, the result on eosinophil counts from different clinical studies with dupilumab treatment in atopic dermatitis is controversial. For instance, no changes in eosinophil count was observed for dupilumab treatment up to 16 weeks [139]. In contrast, Yamauchi et al. showed ameliorated blood eosinophilia for up to 32 weeks following dupilumab treatment in patients with atopic dermatitis [140]. However, the effects of dupilumab on basophils has not been investigated.

Small-molecule inhibitors targeting pro-survival proteins: The therapeutic potential of directly inhibiting pro-survival proteins was unveiled with the development of small-molecule inhibitors targeting B cell lymphoma 2 (BCL-2) homology domain 3 (BH3) mimetics. We have shown that ABT-199, an inhibitor for BCL-2, potently induces apoptosis in human basophils but not in human eosinophils [141,142]. More importantly, our previously published studies demonstrated that ABT-263, a selective inhibitor for both BCL-2 and BCL-2-like 1 (BCL-xL), induced cell death of human eosinophils and basophils [141]. However, pre-clinical data predicted a rapid and concentration dependent thrombocytopenia as a side effect of ABT-263 [143].

5. Concluding Remarks

We summarized findings on how basophils regulate the infiltration of eosinophils, and there are a few indications that these two types of granulocytes form cell-to-cell contact in tissue. The most recent findings demonstrating that basophils mediate eosinophil infiltration were achieved by mouse and in vitro models [7,8,109] and were published up to seven years ago. Apparently, there is still a considerable gap of knowledge on this research topic. Furthermore, studies on possible targets to attenuate and treat allergic inflammation rarely consider both eosinophils and basophils. This could be due to the technical difficulties in isolating and phenotyping these minority cells at the inflammatory sites. With the advent of new approaches like imaging mass cytometry-based studies could help gain more insights and direct evidence on basophil–eosinophil interactions at inflammatory sites. Considering that identifying which Th2-endotypes is involved in the pathogenicity of allergy seems to be a determinant of whether a specific treatment is successful or not [110], we think it will be essential to study the interaction of basophils and eosinophils in more details in the future. The simultaneous study of basophils and eosinophils could result in better profiling of Th2-endotypes, allowing individualized treatment.

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References

1. Akdis, C.A.; Arkwright, P.D.; Brügggen, M.-C.; Busse, W.; Gadina, M.; Guttman-Yassky, E.; Kabashima, K.; Mitamura, Y.; Vian, L.; Wu, J.; et al. Type 2 immunity in the skin and lungs. *Allergy* **2020**, *75*, 1582–1605. [[CrossRef](#)]
2. Stone, K.D.; Prussin, C.; Metcalfe, D.D. IgE, mast cells, basophils, and eosinophils. *J. Allergy Clin. Immunol.* **2010**, *125*, S73–S80. [[CrossRef](#)]
3. Siracusa, M.C.; Kim, B.S.; Spergel, J.M.; Artis, D. Basophils and allergic inflammation. *J. Allergy Clin. Immunol.* **2013**, *132*, 789–801. [[CrossRef](#)] [[PubMed](#)]
4. Guo, C.B.; Liu, M.C.; Galli, S.J.; Bochner, B.S.; Kagey-Sobotka, A.; Lichtenstein, L.M. Identification of IgE-bearing cells in the late-phase response to antigen in the lung as basophils. *Am. J. Respir. Cell Mol. Biol.* **1994**, *10*, 384–390. [[CrossRef](#)]
5. Devouassoux, G.; Foster, B.; Scott, L.M.; Metcalfe, D.D.; Prussin, C. Frequency and characterization of antigen-specific IL-4- and IL-13- producing basophils and T cells in peripheral blood of healthy and asthmatic subjects. *J. Allergy Clin. Immunol.* **1999**, *104*, 811–819. [[CrossRef](#)]
6. Marone, G.; Granata, F.; Pucino, V.; Pecoraro, A.; Heffler, E.; Loffredo, S.; Scadding, G.W.; Varricchi, G. The Intriguing Role of Interleukin 13 in the Pathophysiology of Asthma. *Front. Pharmacol.* **2019**, *10*, 1387. [[CrossRef](#)] [[PubMed](#)]
7. Nakashima, C.; Otsuka, A.; Kitoh, A.; Honda, T.; Egawa, G.; Nakajima, S.; Nakamizo, S.; Arita, M.; Kubo, M.; Miyachi, Y.; et al. Basophils regulate the recruitment of eosinophils in a murine model of irritant contact dermatitis. *J. Allergy Clin. Immunol.* **2014**, *134*, 100–107.e12. [[CrossRef](#)]
8. Cheng, L.E.; Sullivan, B.M.; Retana, L.E.; Allen, C.D.; Liang, H.-E.; Locksley, R.M. IgE-activated basophils regulate eosinophil tissue entry by modulating endothelial function. *J. Exp. Med.* **2015**, *212*, 513–524. [[CrossRef](#)]
9. Iype, J.; Odermatt, A.; Bachmann, S.; Coeudevez, M.; Fux, M. IL-1b Promotes Immunoregulatory Responses in Human Blood Basophils. *Allergy* **2021**, *1*. [[CrossRef](#)]

10. Perrigoue, J.G.; Saenz, S.A.; Siracusa, M.C.; Allenspach, E.J.; Taylor, B.C.; Giacomini, P.R.; Nair, M.G.; Du, Y.; Zaph, C.; Van Rooijen, N.; et al. MHC class II-dependent basophil-CD4+ T cell interactions promote TH2 cytokine-dependent immunity. *Nat. Immunol.* **2009**, *10*, 697–705. [[CrossRef](#)] [[PubMed](#)]
11. Yoshimoto, T.; Yasuda, K.; Tanaka, H.; Nakahira, M.; Imai, Y.; Fujimori, Y.; Nakanishi, K. Basophils contribute to TH2-IgE responses in vivo via IL-4 production and presentation of peptide-MHC class II complexes to CD4+ T cells. *Nat. Immunol.* **2009**, *10*, 706–712. [[CrossRef](#)] [[PubMed](#)]
12. Sokol, C.L.; Chu, N.-Q.; Yu, S.; Nish, S.A.; Laufer, T.M.; Medzhitov, R. Basophils function as antigen-presenting cells for an allergen-induced T helper type 2 response. *Nat. Immunol.* **2009**, *10*, 713–720. [[CrossRef](#)] [[PubMed](#)]
13. Van Beek, A.A.; Knol, E.F.; De Vos, P.; Smelt, M.J.; Savelkoul, H.F.J.; Van Neerven, R.J.J. Recent Developments in Basophil Research: Do Basophils Initiate and Perpetuate Type 2 T-Helper Cell Responses? *Int. Arch. Allergy Immunol.* **2013**, *160*, 7–17. [[CrossRef](#)]
14. Hogan, S.P.; Rosenberg, H.F.; Moqbel, R.; Phipps, S.; Foster, P.S.; Lacy, P.; Kay, A.B.; Rothenberg, M.E. Eosinophils: Biological Properties and Role in Health and Disease. *Clin. Exp. Allergy* **2008**, *38*, 709–750. [[CrossRef](#)]
15. O'Donnell, M.C.; Ackerman, S.J.; Gleich, G.J.; Thomas, L.L. Activation of basophil and mast cell histamine release by eosinophil granule major basic protein. *J. Exp. Med.* **1983**, *157*, 1981–1991. [[CrossRef](#)]
16. Koshino, T.; Teshima, S.; Fukushima, N.; Takaishi, T.; Hirai, K.; Miyamoto, Y.; Arai, Y.; Sano, Y.; Ito, K.; Morita, Y. Identification of basophils by immunohistochemistry in the airways of post-mortem cases of fatal asthma. *Clin. Exp. Allergy* **1993**, *23*, 919–925. [[CrossRef](#)]
17. Macfarlane, A.J.; Kon, O.M.; Smith, S.J.; Zeibecoglou, K.; Khan, L.; Barata, L.T.; McEuen, A.R.; Buckley, M.G.; Walls, A.F.; Meng, Q.; et al. Basophils, eosinophils, and mast cells in atopic and nonatopic asthma and in late-phase allergic reactions in the lung and skin. *J. Allergy Clin. Immunol.* **2000**, *105*, 99–107. [[CrossRef](#)]
18. Irani, A.-M.A.; Huang, C.; Xia, H.-Z.; Kepley, C.; Nafie, A.; Fouda, E.D.; Craig, S.; Zweiman, B.; Schwartz, L.B. Immunohistochemical detection of human basophils in late-phase skin reactions. *J. Allergy Clin. Immunol.* **1998**, *101*, 354–362. [[CrossRef](#)]
19. Gauvreau, G.M.; Lee, J.M.; Watson, R.M.; Irani, A.-M.A.; Schwartz, L.B.; O'Byrne, P.M. Increased Numbers of Both Airway Basophils and Mast Cells in Sputum after Allergen Inhalation Challenge of Atopic Asthmatics. *Am. J. Respir. Crit. Care Med.* **2000**, *161*, 1473–1478. [[CrossRef](#)] [[PubMed](#)]
20. Dijkstra, D.; Hennig, C.; Hansen, G.; Biller, H.; Krug, N.; Hohlfeld, J.M. Identification and quantification of basophils in the airways of asthmatics following segmental allergen challenge. *Cytom. Part. A* **2014**, *85*, 580–587. [[CrossRef](#)] [[PubMed](#)]
21. Ito, Y.; Satoh, T.; Takayama, K.; Miyagishi, C.; Walls, A.F.; Yokozeki, H. Basophil recruitment and activation in inflammatory skin diseases. *Allergy* **2011**, *66*, 1107–1113. [[CrossRef](#)] [[PubMed](#)]
22. Ying, S.; Robinson, D.S.; Meng, Q.; Barata, L.T.; McEuen, A.R.; Buckley, M.G.; Walls, A.F.; Askenase, P.W.; Kay, A.B. C-C Chemo-kines in Allergen-Induced Late-Phase Cutaneous Responses in Atopic Subjects: Association of Eotaxin with Early 6-Hour Eosinophils, and of Eotaxin-2 and Monocyte Chemoattractant Protein-4 with the Later 24-Hour Tissue Eosinophilia, and Relationship to Basophils and Other C-C Chemokines (Monocyte Chemoattractant Protein-3 and RANTES). *J. Immunol.* **1999**, *163*, 3976–3984.
23. Satoh, T.; Ito, Y.; Miyagishi, C.; Yokozeki, H. Basophils Infiltrate Skin Lesions of Eosinophilic Pustular Folliculitis (Ofuji's Disease). *ACTA Derm. Venereol.* **2011**, *91*, 371–372. [[CrossRef](#)] [[PubMed](#)]
24. Wardlaw, A.J.; Dunnette, S.; Gleich, G.J.; Collins, J.V.; Kay, A.B. Eosinophils and Mast Cells in Bronchoalveolar Lavage in Subjects with Mild Asthma: Relationship to Bronchial Hyperreactivity. *Am. Rev. Respir. Dis.* **1988**, *137*, 62–69. [[CrossRef](#)]
25. Metcalfe, D.D.; Pawankar, R.; Ackerman, S.J.; Akin, C.; Clayton, F.; Falcone, F.H.; Gleich, G.J.; Irani, A.-M.; Johansson, M.W.; Klion, A.D.; et al. Biomarkers of the involvement of mast cells, basophils and eosinophils in asthma and allergic diseases. *World Allergy Organ. J.* **2016**, *9*, 7. [[CrossRef](#)] [[PubMed](#)]
26. Motomura, Y.; Morita, H.; Moro, K.; Nakae, S.; Artis, D.; Endo, T.A.; Kuroki, Y.; Ohara, O.; Koyasu, S.; Kubo, M. Basophil-Derived Interleukin-4 Controls the Function of Natural Helper Cells, a Member of ILC2s, in Lung Inflammation. *Immunology* **2014**, *40*, 758–771. [[CrossRef](#)]
27. Thomas, L.L. Basophil and Eosinophil Interactions in Health and Disease. *Chem. Immunol.* **1995**, *61*, 186–207.
28. Shakoory, B.; Fitzgerald, S.M.; Lee, S.A.; Chi, D.S.; Krishnaswamy, G. The Role of Human Mast Cell-Derived Cytokines in Eosinophil Biology. *J. Interf. Cytokine Res.* **2004**, *24*, 271–281. [[CrossRef](#)]
29. Wardlaw, A.J. Eosinophil trafficking in asthma. *Clin. Med.* **2001**, *1*, 214–218. [[CrossRef](#)]
30. Van Oosterhout, A.J.; Fattah, D.; Van Ark, I.; Hofman, G.; Buckley, T.L.; Nijkamp, F.P. Eosinophil infiltration precedes development of airway hyperreactivity and mucosal exudation after intranasal administration of interleukin-5 to mice. *J. Allergy Clin. Immunol.* **1995**, *96*, 104–112. [[CrossRef](#)]
31. Wang, J.; Palmer, K.; Lotvall, J.; Milan, S.; Lei, X.F.; Matthaei, I.K.; Gauldie, J.; Inman, M.D.; Jordana, M.; Xing, Z. Circulating, but not local lung, IL-5 is required for the development of antigen-induced airways eosinophilia. *J. Clin. Investig.* **1998**, *102*, 1132–1141. [[CrossRef](#)] [[PubMed](#)]
32. Mukai, K.; Matsuoka, K.; Taya, C.; Suzuki, H.; Yokozeki, H.; Nishioka, K.; Hirokawa, K.; Etori, M.; Yamashita, M.; Kubota, T.; et al. Basophils Play a Critical Role in the Development of IgE-Mediated Chronic Allergic Inflammation Independently of T Cells and Mast Cells. *Immunology* **2005**, *23*, 191–202. [[CrossRef](#)] [[PubMed](#)]
33. Obata, K.; Mukai, K.; Tsujimura, Y.; Ishiwata, K.; Kawano, Y.; Minegishi, Y.; Watanabe, N.; Karasuyama, H. Basophils are essential initiators of a novel type of chronic allergic inflammation. *Blood* **2007**, *110*, 913–920. [[CrossRef](#)] [[PubMed](#)]

34. Ohnmacht, C.; Schwartz, C.; Panzer, M.; Schiedewitz, I.; Naumann, R.; Voehringer, D. Basophils Orchestrate Chronic Allergic Dermatitis and Protective Immunity against Helminths. *Immunology* **2010**, *33*, 364–374. [[CrossRef](#)]
35. Matsuoka, K.; Shitara, H.; Taya, C.; Kohno, K.; Kikkawa, Y.; Yonekawa, H. Novel Basophil- or Eosinophil-Depleted Mouse Models for Functional Analyses of Allergic Inflammation. *PLoS ONE* **2013**, *8*, e60958. [[CrossRef](#)] [[PubMed](#)]
36. Mashiko, S.; Mehta, H.; Bissonnette, R.; Sarfati, M. Increased frequencies of basophils, type 2 innate lymphoid cells and Th2 cells in skin of patients with atopic dermatitis but not psoriasis. *J. Dermatol. Sci.* **2017**, *88*, 167–174. [[CrossRef](#)]
37. Nabe, T.; Zindl, C.L.; Jung, Y.W.; Stephens, R.; Sakamoto, A.; Kohno, S.; Atkinson, T.P.; Chaplin, D.D. Induction of a late asthmatic response associated with airway inflammation in mice. *Eur. J. Pharmacol.* **2005**, *521*, 144–155. [[CrossRef](#)]
38. Zhong, W.; Su, W.; Zhang, Y.; Liu, Q.; Wu, J.; Di, C.; Zhang, Z.; Xia, Z. Basophils as a primary inducer of the T helper type 2 immunity in ovalbumin-induced allergic airway inflammation. *Immunology* **2014**, *142*, 202–215. [[CrossRef](#)]
39. Hammad, H.; Plantinga, M.; Deswarte, K.; Pouliot, P.; Willart, M.A.; Kool, M.; Muskens, F.; Lambrecht, B.N. Inflammatory dendritic cells—Not basophils—Are necessary and sufficient for induction of Th2 immunity to inhaled house dust mite allergen. *J. Exp. Med.* **2010**, *207*, 2097–2111. [[CrossRef](#)]
40. Matsuoka, K.; Taya, C.; Kubo, S.; Toyama-Sorimachi, N.; Kitamura, F.; Ra, C.; Yonekawa, H.; Karasuyama, H. Establishment of antigen-specific IgE transgenic mice to study pathological and immunobiological roles of IgE in vivo. *Int. Immunol.* **1999**, *11*, 987–994. [[CrossRef](#)]
41. Simon, D.; Braathen, L.R.; Simon, H.-U. Eosinophils and atopic dermatitis. *Allergy* **2004**, *59*, 561–570. [[CrossRef](#)] [[PubMed](#)]
42. Wakita, H.; Sakamoto, T.; Tokura, Y.; Takigawa, M. E-selectin and vascular cell adhesion molecule-1 as critical adhesion molecules for infiltration of T lymphocytes and eosinophils in atopic dermatitis. *J. Cutan. Pathol.* **1994**, *21*, 33–39. [[CrossRef](#)]
43. Yawalkar, N.; Ugucioni, M.; Schäfer, J.; Braunwalder, J.; Karlen, S.; Dewald, B.; Braathen, L.R.; Baggiolini, M. Enhanced Expression of Eotaxin and CCR3 in Atopic Dermatitis. *J. Investig. Dermatol.* **1999**, *113*, 43–48. [[CrossRef](#)]
44. Nabe, T.; Matsuya, K.; Akamizu, K.; Fujita, M.; Nakagawa, T.; Shioe, M.; Kida, H.; Takiguchi, A.; Wakamori, H.; Fujii, M.; et al. Roles of basophils and mast cells infiltrating the lung by multiple antigen challenges in asthmatic responses of mice. *Br. J. Pharmacol.* **2013**, *169*, 462–476. [[CrossRef](#)]
45. Fattouh, R.; Al-Garawi, A.; Fattouh, M.; Arias, K.; Walker, T.D.; Goncharova, S.; Coyle, A.J.; Humbles, A.A.; Jordana, M. Eosinophils Are Dispensable for Allergic Remodeling and Immunity in a Model of House Dust Mite-induced Airway Disease. *Am. J. Respir. Crit. Care Med.* **2011**, *183*, 179–188. [[CrossRef](#)] [[PubMed](#)]
46. Graham, M.T.; Nadeau, K.C. Lessons learned from mice and man: Mimicking human allergy through mouse models. *Clin. Immunol.* **2014**, *155*, 1–16. [[CrossRef](#)] [[PubMed](#)]
47. Gao, J.-L.; Sen, A.I.; Kitaura, M.; Yoshie, O.; Rothenberg, M.E.; Murphy, P.M.; Luster, A.D. Identification of a Mouse Eosinophil Receptor for the CC Chemokine Eotaxin. *Biochem. Biophys. Res. Commun.* **1996**, *223*, 679–684. [[CrossRef](#)] [[PubMed](#)]
48. Daugherty, B.L.; Siciliano, S.J.; DeMartino, A.J.; Malkowitz, L.; Sirotina, A.; Springer, M.S. Cloning, expression, and characterization of the human eosinophil eotaxin receptor. *J. Exp. Med.* **1996**, *183*, 2349–2354. [[CrossRef](#)] [[PubMed](#)]
49. Ugucioni, M.; Mackay, C.R.; Ochensberger, B.; Loetscher, P.; Rhis, S.; LaRosa, G.J.; Rao, P.; Ponath, P.D.; Baggiolini, M.; Dahinden, A.C. High expression of the chemokine receptor CCR3 in human blood basophils. Role in activation by eotaxin, MCP-4, and other chemokines. *J. Clin. Investig.* **1997**, *100*, 1137–1143. [[CrossRef](#)]
50. Ma, W.; Bryce, P.J.; Humbles, A.A.; Laouini, D.; Yalcindag, A.; Alenius, H.; Friend, D.S.; Oettgen, H.C.; Gerard, C.; Geha, R.S. CCR3 is essential for skin eosinophilia and airway hyperresponsiveness in a murine model of allergic skin inflammation. *J. Clin. Investig.* **2002**, *109*, 621–628. [[CrossRef](#)]
51. Humbles, A.A.; Lu, B.; Friend, D.S.; Okinaga, S.; Lora, J.; Al-Garawi, A.; Martin, T.R.; Gerard, N.P.; Gerard, C. The murine CCR3 receptor regulates both the role of eosinophils and mast cells in allergen-induced airway inflammation and hyperresponsiveness. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 1479–1484. [[CrossRef](#)] [[PubMed](#)]
52. Pope, S.M.; Zimmermann, N.; Stringer, K.F.; Karow, M.L.; Rothenberg, M.E. The Eotaxin Chemokines and CCR3 Are Fundamental Regulators of Allergen-Induced Pulmonary Eosinophilia. *J. Immunol.* **2005**, *175*, 5341–5350. [[CrossRef](#)] [[PubMed](#)]
53. Sénéchal, S.; Fahy, O.; Gentina, T.; Vorng, H.; Capron, M.; Walls, A.F.; McEuen, A.R.; Buckley, M.G.; Hamid, Q.; Wallaert, B.; et al. CCR3-Blocking Antibody Inhibits Allergen-Induced Eosinophil Recruitment in Human Skin Xenografts from Allergic Patients. *Lab. Investig.* **2002**, *82*, 929–939. [[CrossRef](#)] [[PubMed](#)]
54. Kaburagi, Y.; Shimada, Y.; Nagaoka, T.; Hasegawa, M.; Takehara, K.; Sato, S. Enhanced production of CC-chemokines (RANTES, MCP-1, MIP-1 α , MIP-1 β , and eotaxin) in patients with atopic dermatitis. *Arch. Dermatol. Res.* **2001**, *293*, 350–355. [[CrossRef](#)]
55. Lilly, C.M.; Woodruff, P.G.; Camargo, C.A.; Nakamura, H.; Drazen, J.M.; Nadel, E.S.; Hanrahan, J.P. Elevated plasma eotaxin levels in patients with acute asthma. *J. Allergy Clin. Immunol.* **1999**, *104*, 786–790. [[CrossRef](#)]
56. Nakamura, H.; Weiss, S.T.; Israel, E.; Luster, A.D.; Drazen, J.M.; Lilly, C.M. Eotaxin and Impaired Lung Function in Asthma. *Am. J. Respir. Crit. Care Med.* **1999**, *160*, 1952–1956. [[CrossRef](#)]
57. Berkman, N.; Ohnona, S.; Chung, F.K.; Breuer, R. Eotaxin-3 but Not Eotaxin Gene Expression Is Upregulated in Asthmatics 24 Hours after Allergen Challenge. *Am. J. Respir. Cell Mol. Biol.* **2001**, *24*, 682–687. [[CrossRef](#)] [[PubMed](#)]
58. Kagami, S.; Kakinuma, T.; Saeki, H.; Tsunemi, Y.; Fujita, H.; Nakamura, K.; Takekoshi, T.; Kishimoto, M.; Mitsui, H.; Torii, H.; et al. Significant elevation of serum levels of eotaxin-3/CCL26, but not of eotaxin-2/CCL24, in patients with atopic dermatitis: Serum eotaxin-3/CCL26 levels reflect the disease activity of atopic dermatitis. *Clin. Exp. Immunol.* **2003**, *134*, 309–313. [[CrossRef](#)]

59. Kitaura, M.; Nakajima, T.; Imai, T.; Harada, S.; Combadiere, C.; Tiffany, H.L.; Murphy, P.M.; Yoshie, O. Molecular Cloning of Human Eotaxin, an Eosinophil-selective CC Chemokine, and Identification of a Specific Eosinophil Eotaxin Receptor, CC Chemokine Receptor 3. *J. Biol. Chem.* **1996**, *271*, 7725–7730. [[CrossRef](#)] [[PubMed](#)]
60. Ponath, P.D.; Qin, S.; Ringler, D.J.; Clark-Lewis, I.; Wang, J.; Kassam, N.; Smith, H.; Shi, X.; Gonzalo, A.J.; Newman, W.; et al. Cloning of the human eosinophil chemoattractant, eotaxin. Expression, receptor binding, and functional properties suggest a mechanism for the selective recruitment of eosinophils. *J. Clin. Invest.* **1996**, *97*, 604–612. [[CrossRef](#)] [[PubMed](#)]
61. Garcia-Zepeda, E.A.; Rothenberg, M.E.; Ownbey, R.T.; Celestin, J.; Leder, P.; Luster, A.D. Human eotaxin is a specific chemoattractant for eosinophil cells and provides a new mechanism to explain tissue eosinophilia. *Nat. Med.* **1996**, *2*, 449–456. [[CrossRef](#)] [[PubMed](#)]
62. Matthews, A.N.; Friend, D.S.; Zimmermann, N.; Sarafi, M.N.; Luster, A.D.; Pearlman, E.; Wert, S.E.; Rothenberg, M.E. Eotaxin is required for the baseline level of tissue eosinophils. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 6273–6278. [[CrossRef](#)] [[PubMed](#)]
63. Pope, S.M.; Fulkeron, P.C.; Blanchard, C.; Akei, H.S.; Nikolaidis, N.M.; Zimmermann, N.; Molkentin, J.D.; Rothenberg, M.E. Identification of a Cooperative Mechanism Involving Interleukin-13 and Eotaxin-2 in Experimental Allergic Lung Inflammation. *J. Biol. Chem.* **2005**, *280*, 13952–13961. [[CrossRef](#)] [[PubMed](#)]
64. Rothenberg, M.E.; MacLean, J.A.; Pearlman, E.; Luster, A.D.; Leder, P. Targeted Disruption of the Chemokine Eotaxin Partially Reduces Antigen-induced Tissue Eosinophilia. *J. Exp. Med.* **1997**, *185*, 785–790. [[CrossRef](#)] [[PubMed](#)]
65. Yamada, H.; Hirai, K.; Miyamasu, M.; Iikura, M.; Misaki, Y.; Shoji, S.; Takaishi, T.; Kasahara, T.; Morita, Y.; Ito, K. Eotaxin Is a Potent Chemotaxin for Human Basophils. *Biochem. Biophys. Res. Commun.* **1997**, *231*, 365–368. [[CrossRef](#)]
66. Forssmann, U.; Ugucioni, M.; Loetscher, P.; Dahinden, C.A.; Langen, H.; Thelen, M.; Baggiolini, M. Eotaxin-2, a Novel CC Chemokine that Is Selective for the Chemokine Receptor CCR3, and Acts Like Eotaxin on Human Eosinophil and Basophil Leukocytes. *J. Exp. Med.* **1997**, *185*, 2171–2176. [[CrossRef](#)]
67. Alam, R.; York, J.; Boyars, M.; Stafford, S.; Grant, A.J.; Lee, J.; Forsythe, P.; Šim, T.; Ida, N. Increased MCP-1, RANTES, and MIP-1alpha in bronchoalveolar lavage fluid of allergic asthmatic patients. *Am. J. Respir. Crit. Care Med.* **1996**, *153*, 1398–1404. [[CrossRef](#)]
68. Ahmadi, Z.; Hassanshahi, G.; Khorramdelazad, H.; Zainodini, N.; Koochakzadeh, L. An Overlook to the Characteristics and Roles Played by Eotaxin Network in the Pathophysiology of Food Allergies: Allergic Asthma and Atopic Dermatitis. *Inflammation* **2016**, *39*, 1–15. [[CrossRef](#)]
69. Foster, P.S.; Maltby, S.; Rosenberg, H.F.; Tay, H.L.; Hogan, S.P.; Collison, A.M.; Yang, M.; Kaiko, G.E.; Hansbro, P.M.; Kumar, R.K.; et al. Modeling TH2 responses and airway inflammation to understand fundamental mechanisms regulating the pathogenesis of asthma. *Immunol. Rev.* **2017**, *278*, 20–40. [[CrossRef](#)]
70. Esnault, S.; Kelly, E.A. Essential mechanisms of differential activation of eosinophils by IL-3 compared to GM-CSF and IL-5. *Crit. Rev. Immunol.* **2017**, *36*, 429–444. [[CrossRef](#)]
71. Broide, D.H.; Paine, M.M.; Firestein, G.S. Eosinophils express interleukin 5 and granulocyte macrophage-colony-stimulating factor mRNA at sites of allergic inflammation in asthmatics. *J. Clin. Invest.* **1992**, *90*, 1414–1424. [[CrossRef](#)] [[PubMed](#)]
72. Johansson, M.W.; Kelly, E.A.B.; Busse, W.W.; Jarjour, N.N.; Mosher, D.F. Up-Regulation and Activation of Eosinophil Integrins in Blood and Airway after Segmental Lung Antigen Challenge. *J. Immunol.* **2008**, *180*, 7622–7635. [[CrossRef](#)] [[PubMed](#)]
73. Kato, M.; Liu, M.C.; Stealey, A.B.; Friedman, B.; Lichtenstein, L.M.; Permutt, S.; Schleimer, R.P. Production of granulocyte/macrophage colony-stimulating factor in human airways during allergen-induced late-phase reactions in atopic subjects. *Lymphokine Cytokine Res.* **1992**, *11*, 287–292. [[PubMed](#)]
74. Moser, R.; Fehr, J.; Olgiati, L.; Bruijnzeel, P.L. Migration of primed human eosinophils across cytokine-activated endothelial cell monolayers. *Blood* **1992**, *79*, 2937–2945. [[CrossRef](#)]
75. Dougan, M.; Dranoff, G.; Dougan, S.K. GM-CSF, IL-3, and IL-5 Family of Cytokines: Regulators of Inflammation. *Immunology* **2019**, *50*, 796–811. [[CrossRef](#)] [[PubMed](#)]
76. Kämpfer, S.S.; Odermatt, A.; Dahinden, C.A.; Fux, M. Late IL-3-induced phenotypic and functional alterations in human basophils require continuous IL-3 receptor signaling. *J. Leukoc. Biol.* **2016**, *101*, 227–238. [[CrossRef](#)] [[PubMed](#)]
77. Yamaguchi, M.; Koketsu, R.; Suzukawa, M.; Kawakami, A.; Iikura, M. Human Basophils and Cytokines/Chemokines. *Allergol. Int.* **2009**, *58*, 1–10. [[CrossRef](#)]
78. Bochner, B.S.; McKelvey, A.A.; Sterbinsky, A.S.; Hildreth, E.J.; Derse, C.P.; Klunk, A.D.; Lichtenstein, L.M.; Schleimer, R.P. IL-3 augments adhesiveness for endothelium and CD11b expression in human basophils but not neutrophils. *J. Immunol.* **1990**, *145*, 1832–1837.
79. Yoshimura-Uchiyama, C.; Yamaguchi, M.; Nagase, H.; Fujisawa, T.; Ra, C.; Matsushima, K.; Iwata, T.; Igarashi, T.; Yamamoto, K.; Hirai, K. Comparative effects of basophil-directed growth factors. *Biochem. Biophys. Res. Commun.* **2003**, *302*, 201–206. [[CrossRef](#)]
80. Ebisawa, M.; Yamada, T.; Bickel, C.; Klunk, D.; Schleimer, R.P. Eosinophil transendothelial migration induced by cytokines. III. Effect of the chemokine RANTES. *J. Immunol.* **1994**, *153*, 2153–2160.
81. Tanimoto, Y.; Takahashi, K.; Kimura, I. Effects of cytokines on human basophil chemotaxis. *Clin. Exp. Allergy* **1992**, *22*, 1020–1025. [[CrossRef](#)]
82. Schroeder, J.T.; Chichester, K.L.; Bieneman, A.P. Human Basophils Secrete IL-3: Evidence of Autocrine Priming for Phenotypic and Functional Responses in Allergic Disease. *J. Immunol.* **2009**, *182*, 2432–2438. [[CrossRef](#)]

83. Simson, L.; Foster, P.S. Chemokine and cytokine cooperativity: Eosinophil migration in the asthmatic response. *Immunol. Cell Biol.* **2000**, *78*, 415–422. [[CrossRef](#)]
84. Mould, A.W.; Matthaei, I.K.; Young, I.G.; Foster, P.S. Relationship between interleukin-5 and eotaxin in regulating blood and tissue eosinophilia in mice. *J. Clin. Investig.* **1997**, *99*, 1064–1071. [[CrossRef](#)] [[PubMed](#)]
85. Mould, A.W.; Ramsay, A.J.; Matthaei, K.I.; Young, I.G.; Rothenberg, M.E.; Foster, P.S. The Effect of IL-5 and Eotaxin Expression in the Lung on Eosinophil Trafficking and Degranulation and the Induction of Bronchial Hyperreactivity. *J. Immunol.* **2000**, *164*, 2142–2150. [[CrossRef](#)] [[PubMed](#)]
86. Collins, P.D.; Marleau, S.; Griffiths-Johnson, A.D.; Jose, P.J.; Williams, T.J. Cooperation between interleukin-5 and the chemokine eotaxin to induce eosinophil accumulation in vivo. *J. Exp. Med.* **1995**, *182*, 1169–1174. [[CrossRef](#)] [[PubMed](#)]
87. Johansson, M.W. Activation states of blood eosinophils in asthma. *Clin. Exp. Allergy* **2014**, *44*, 482–498. [[CrossRef](#)] [[PubMed](#)]
88. Henderson, W.R.; Chi, E.Y.; Maliszewski, C.R. Soluble IL-4 Receptor Inhibits Airway Inflammation Following Allergen Challenge in a Mouse Model of Asthma. *J. Immunol.* **2000**, *164*, 1086–1095. [[CrossRef](#)] [[PubMed](#)]
89. Brusselle, G.; Kips, J.; Joos, G.; Bluethmann, H.; Pauwels, R. Allergen-induced airway inflammation and bronchial responsiveness in wild-type and interleukin-4-deficient mice. *Am. J. Respir. Cell Mol. Biol.* **1995**, *12*, 254–259. [[CrossRef](#)]
90. Mochizuki, M.; Schröder, J.-M.; Christophers, E.; Yamamoto, S. IL-4 Induces Eotaxin in Human Dermal Fibroblasts. *Int. Arch. Allergy Immunol.* **1999**, *120*, 19–23. [[CrossRef](#)]
91. Mochizuki, M.; Bartels, J.; Mallet, I.A.; Christophers, E.; Schröder, J.M. IL-4 induces eotaxin: A possible mechanism of selective eosinophil recruitment in helminth infection and atopy. *J. Immunol.* **1998**, *160*, 60–68.
92. Zimmermann, N.; Hogan, S.P.; Mishra, A.; Brandt, E.B.; Bodette, T.R.; Pope, S.M.; Finkelman, F.D.; Rothenberg, M.E. Murine Eotaxin-2: A Constitutive Eosinophil Chemokine Induced by Allergen Challenge and IL-4 Overexpression. *J. Immunol.* **2000**, *165*, 5839–5846. [[CrossRef](#)]
93. Shinkai, A.; Yoshisue, H.; Koike, M.; Shoji, E.; Nakagawa, S.; Saito, A.; Takeda, T.; Imabeppu, S.; Kato, Y.; Hanai, N.; et al. A novel human CC chemokine, eotaxin-3, which is expressed in IL-4-stimulated vascular endothelial cells, exhibits potent activity toward eosinophils. *J. Immunol.* **1999**, *163*, 1602–1610. [[PubMed](#)]
94. Van Panhuys, N.; Prout, M.; Forbes, E.; Min, B.; Paul, W.E.; Le Gros, G. Basophils Are the Major Producers of IL-4 during Primary Helminth Infection. *J. Immunol.* **2011**, *186*, 2719–2728. [[CrossRef](#)]
95. Min, B.; Prout, M.; Hu-Li, J.; Zhu, J.; Jankovic, D.; Morgan, E.S.; Urban, J.F.; Dvorak, A.M.; Finkelman, F.D.; Legros, G.; et al. Basophils Produce IL-4 and Accumulate in Tissues after Infection with a Th2-inducing Parasite. *J. Exp. Med.* **2004**, *200*, 507–517. [[CrossRef](#)] [[PubMed](#)]
96. Voehringer, D.; Shinkai, K.; Locksley, R.M. Type 2 Immunity Reflects Orchestrated Recruitment of Cells Committed to IL-4 Production. *Immunology* **2004**, *20*, 267–277. [[CrossRef](#)]
97. Nouri-Aria, K.T.; Irani, A.-M.A.; Jacobson, M.R.; O'Brien, F.; Varga, E.M.; Till, S.J.; Durham, S.R.; Schwartz, L.B. Basophil recruitment and IL-4 production during human allergen-induced late asthma. *J. Allergy Clin. Immunol.* **2001**, *108*, 205–211. [[CrossRef](#)] [[PubMed](#)]
98. Hoeck, J.; Woisetschläger, M. Activation of Eotaxin-3/CCL26 Gene Expression in Human Dermal Fibroblasts Is Mediated by STAT6. *J. Immunol.* **2001**, *167*, 3216–3222. [[CrossRef](#)] [[PubMed](#)]
99. Hoeck, J.; Woisetschläger, M. STAT6 Mediates Eotaxin-1 Expression in IL-4 or TNF- α -Induced Fibroblasts. *J. Immunol.* **2001**, *166*, 4507–4515. [[CrossRef](#)]
100. Ochensberger, B.; Daepf, G.C.; Rihs, S. Dahinden Human Blood Basophils Produce Interleukin-13 in Response to IgE-Receptor-Dependent and -Independent Activation. *Blood* **1996**, *88*, 3028–3037. [[CrossRef](#)]
101. Gibbs, B.F.; Haas, H.; Falcone, F.H.; Albrecht, C.; Vollrath, I.B.; Noll, T.; Wolff, H.H.; Amon, U. Purified human peripheral blood basophils release interleukin-13 and preformed interleukin-4 following immunological activation. *Eur. J. Immunol.* **1996**, *26*, 2493–2498. [[CrossRef](#)]
102. Rot, A.; Krieger, M.; Brunner, T.; Bischoff, S.C.; Schall, T.J.; Dahinden, A.C. RANTES and macrophage inflammatory protein 1 α induce the migration and activation of normal human eosinophil granulocytes. *J. Exp. Med.* **1992**, *176*, 1489–1495. [[CrossRef](#)] [[PubMed](#)]
103. Brunner, T.; Heusser, C.H. Dahinden Human Peripheral Blood Basophils Primed by Interleukin 3 (IL-3) Produce IL-4 in Response to Immunoglobulin E Receptor Stimulation. *J. Exp. Med.* **1993**, *177*, 605–611. [[CrossRef](#)]
104. Hagmann, B.R.; Odermatt, A.; Kaufmann, T.; Dahinden, C.A.; Fux, M.; Hagmann, B.R. Balance between IL-3 and type I interferons and their interrelationship with FasL dictates lifespan and effector functions of human basophils. *Clin. Exp. Allergy* **2017**, *47*, 71–84. [[CrossRef](#)] [[PubMed](#)]
105. Sehmi, R.; Cromwell, O.; Wardlaw, A.J.; Moqbel, R.; Kay, A.B. Interleukin-8 is a chemo-attractant for eosinophils purified from subjects with a blood eosinophilia but not from normal healthy subjects. *Clin. Exp. Allergy* **1993**, *23*, 1027–1036. [[CrossRef](#)] [[PubMed](#)]
106. Karasuyama, H.; Miyake, K.; Yoshikawa, S.; Kawano, Y.; Yamanishi, Y. How do basophils contribute to Th2 cell differentiation and allergic responses? *Int. Immunol.* **2018**, *30*, 391–396. [[CrossRef](#)] [[PubMed](#)]
107. Tschopp, C.M.; Spiegl, N.; Didichenko, S.; Lutmann, W.; Julius, P.; Virchow, J.C.; Hack, C.E.; Dahinden, C.A. Granzyme B, a novel mediator of allergic inflammation: Its induction and release in blood basophils and human asthma. *Blood* **2006**, *108*, 2290–2299. [[CrossRef](#)] [[PubMed](#)]

108. Landolina, N.; Gangwar, R.S.; Levi-Schaffer, F. Mast Cells' Integrated Actions with Eosinophils and Fibroblasts in Allergic Inflammation. *Adv. Immunol.* **2015**, *125*, 41–85. [[CrossRef](#)]
109. Jiao, D.; Wong, C.-K.; Qiu, H.-N.; Dong, J.; Cai, Z.; Chu, M.; Hon, K.-L.; Tsang, M.S.-M.; Lam, C.W.-K. NOD2 and TLR2 ligands trigger the activation of basophils and eosinophils by interacting with dermal fibroblasts in atopic dermatitis-like skin inflammation. *Cell. Mol. Immunol.* **2015**, *13*, 535–550. [[CrossRef](#)]
110. Schroeder, J.T.; Bieneman, A.P. Activation of Human Basophils by A549 Lung Epithelial Cells Reveals a Novel IgE-Dependent Response Independent of Allergen. *J. Immunol.* **2017**, *199*, 855–865. [[CrossRef](#)]
111. Yoshimoto, T. Basophils as Th2-inducing antigen-presenting cells. *Int. Immunol.* **2010**, *22*, 543–550. [[CrossRef](#)]
112. Kim, S.; Shen, T.; Min, B. Basophils Can Directly Present or Cross-Present Antigen to CD8 Lymphocytes and Alter CD8 T Cell Differentiation into IL-10-Producing Phenotypes. *J. Immunol.* **2009**, *183*, 3033–3039. [[CrossRef](#)] [[PubMed](#)]
113. Voskamp, A.L.; Prickett, S.R.; Mackay, F.; Rolland, J.M.; O'Hehir, R.E. MHC Class II Expression in Human Basophils: Induction and Lack of Functional Significance. *PLoS ONE* **2013**, *8*, e81777. [[CrossRef](#)] [[PubMed](#)]
114. Sullivan, B.M.; Liang, H.-E.; Bando, J.K.; Wu, D.; Cheng, L.E.; McKerrow, J.K.; Allen, C.D.C.; Locksley, R.M. Genetic analysis of basophil function in vivo. *Nat. Immunol.* **2011**, *12*, 527–535. [[CrossRef](#)] [[PubMed](#)]
115. Wakahara, K.; Van, V.Q.; Baba, N.; Bégin, P.; Rubio, M.; Delespesse, G.; Sarfati, M. Basophils are recruited to inflamed lungs and exacerbate memory Th2 responses in mice and humans. *Allergy* **2012**, *68*, 180–189. [[CrossRef](#)]
116. Sarfati, M.; Wakahara, K.; Chapuy, L.; Delespesse, G. Mutual Interaction of Basophils and T Cells in Chronic Inflammatory Diseases. *Front. Immunol.* **2015**, *6*, 399. [[CrossRef](#)] [[PubMed](#)]
117. Jogdand, P.; Siddhuraj, P.; Mori, M.; Sanden, C.; Jönsson, J.; Walls, A.F.; Kearley, J.; Humbles, A.A.; Kolbeck, R.; Bjermer, L.; et al. Eosinophils, basophils and type 2 immune microenvironments in COPD-affected lung tissue. *Eur. Respir. J.* **2020**, *55*, 1900110. [[CrossRef](#)]
118. Davoine, F.; Lacy, P. Eosinophil Cytokines, Chemokines, and Growth Factors: Emerging Roles in Immunity. *Front. Immunol.* **2014**, *5*, 570. [[CrossRef](#)]
119. Simon, H.-U.; Yousefi, S.; Germic, N.; Arnold, I.C.; Haczku, A.; Karaulov, A.V.; Simon, D.; Rosenberg, H.F. The Cellular Functions of Eosinophils: Collegium Internationale Allergologicum (CIA) Update 2020. *Int. Arch. Allergy Immunol.* **2019**, *181*, 11–23. [[CrossRef](#)]
120. Büttner, C.; Lun, A.; Splettstoesser, T.; Kunkel, G.; Renz, H. Monoclonal anti-interleukin-5 treatment suppresses eosinophil but not T-cell functions. *Eur. Respir. J.* **2003**, *21*, 799–803. [[CrossRef](#)]
121. Wright, A.K.A.; Diver, S.; McCarthy, J.; Marvin, A.; Soares, M.; Thornton, T.; Bourne, M.; Craner, M.; Evans, H.; Edwards, S.; et al. Mepolizumab does not alter the blood basophil count in severe asthma. *Allergy* **2019**, *74*, 2488–2490. [[CrossRef](#)]
122. Kips, J.C.; O'Connor, B.J.; Langley, S.J.; Woodcock, A.; Kerstjens, H.A.M.; Postma, D.S.; Danzig, M.; Cuss, F.; Pauwels, R.A. Effect of SCH55700, a Humanized Anti-Human Interleukin-5 Antibody, in Severe Persistent Asthma. *Am. J. Respir. Crit. Care Med.* **2003**, *167*, 1655–1659. [[CrossRef](#)]
123. Roufosse, F. Targeting the Interleukin-5 Pathway for Treatment of Eosinophilic Conditions Other than Asthma. *Front. Med.* **2018**, *5*, 49. [[CrossRef](#)]
124. Laviolette, M.; Gossage, D.L.; Gauvreau, G.; Leigh, R.; Olivenstein, R.; Katial, R.; Busse, W.W.; Wenzel, S.; Wu, Y.; Datta, V.; et al. Effects of benralizumab on airway eosinophils in asthmatic patients with sputum eosinophilia. *J. Allergy Clin. Immunol.* **2013**, *132*, 1086–1096.e5. [[CrossRef](#)] [[PubMed](#)]
125. Herzog, E.; Busfield, S.; Biondo, M.; Vairo, G.; Dewitte, M.; Pragst, I.; Dickneite, G.; Nash, A.; Zollner, S. Pharmacodynamic Activity and Preclinical Safety of CSL362, a Novel Humanised, Affinity Matured Monoclonal Antibody Against Human Interleukin 3 Receptor. *Blood* **2012**, *120*, 1524. [[CrossRef](#)]
126. Pecaric-Petkovic, T.; Didichenko, S.A.; Kaempfer, S.; Spiegl, N.; Dahinden, C.A. Human basophils and eosinophils are the direct target leukocytes of the novel IL-1 family member IL-33. *Blood* **2009**, *113*, 1526–1534. [[CrossRef](#)]
127. Porsbjerg, C.M.; Sverrild, A.; Lloyd, C.M.; Menzies-Gow, A.N.; Bel, E.H. Anti-alarmins in asthma: Targeting the airway epithelium with next-generation biologics. *Eur. Respir. J.* **2020**, *56*, 2000260. [[CrossRef](#)]
128. Lee, H.Y.; Rhee, C.K.; Kang, J.Y.; Byun, J.H.; Choi, J.Y.; Kim, S.J.; Kim, Y.K.; Kwon, S.S.; Lee, S.Y. Blockade of IL-33/ST2 ameliorates airway inflammation in a murine model of allergic asthma. *Exp. Lung Res.* **2014**, *40*, 66–76. [[CrossRef](#)] [[PubMed](#)]
129. Allinne, J.; Scott, G.; Lim, W.K.; Birchard, D.; Erjefält, J.S.; Sandén, C.; Ben, L.-H.; Agrawal, A.; Kaur, N.; Kim, J.H.; et al. IL-33 blockade affects mediators of persistence and exacerbation in a model of chronic airway inflammation. *J. Allergy Clin. Immunol.* **2019**, *144*, 1624–1637.e10. [[CrossRef](#)]
130. Fulkerson, P.C.; Fischetti, C.A.; McBride, M.L.; Hassman, L.M.; Hogan, S.P.; Rothenberg, M.E. A central regulatory role for eosinophils and the eotaxin/CCR3 axis in chronic experimental allergic airway inflammation. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 16418–16423. [[CrossRef](#)] [[PubMed](#)]
131. Gauvreau, G.M.; Fitzgerald, J.M.; Boulet, L.P.; Watson, R.M.; Hui, L.; Villeneuve, H.; Scime, T.X.; Schlatman, A.R.; Obminski, C.; Kum, J.; et al. The effects of a CCR3 inhibitor, AXP1275, on allergen-induced airway responses in adults with mild-to-moderate atopic asthma. *Clin. Exp. Allergy* **2018**, *48*, 445–451. [[CrossRef](#)] [[PubMed](#)]
132. Grozdanovic, M.; Laffey, K.G.; Abdelkarim, H.; Hitchinson, B.; Harijith, A.; Moon, H.-G.; Park, G.Y.; Rousslang, L.K.; Masterson, J.C.; Furuta, G.T.; et al. Novel peptide nanoparticle-biased antagonist of CCR3 blocks eosinophil recruitment and airway hyperresponsiveness. *J. Allergy Clin. Immunol.* **2019**, *143*, 669–680.e12. [[CrossRef](#)] [[PubMed](#)]

133. Royer, J.F.; Schratl, P.; Carrillo, J.J.; Jupp, R.; Barker, J.; Weyman-Jones, C.; Beri, R.; Sargent, C.; Schmidt, J.A.; Lang-Loidolt, D.; et al. A novel antagonist of prostaglandin D2 blocks the locomotion of eosinophils and basophils. *Eur. J. Clin. Investig.* **2008**, *38*, 663–671. [[CrossRef](#)] [[PubMed](#)]
134. Wenzel, S.; Ford, L.; Pearlman, D.; Spector, S.; Sher, L.; Skobieranda, F.; Wang, L.; Kirkesseli, S.; Rocklin, R.; Bock, B.; et al. Dupilumab in Persistent Asthma with Elevated Eosinophil Levels. *N. Engl. J. Med.* **2013**, *368*, 2455–2466. [[CrossRef](#)] [[PubMed](#)]
135. Menzella, F.; Galeone, C.; Lusuardi, M.; Simonazzi, A.; Castagnetti, C.; Ruggiero, P.; Facciolongo, N. Near-fatal asthma responsive to mepolizumab after failure of omalizumab and bronchial thermoplasty. *Ther. Clin. Risk Manag.* **2017**, *ume 13*, 1489–1493. [[CrossRef](#)]
136. Fulkerson, P.C.; Rothenberg, M.E. Targeting eosinophils in allergy, inflammation and beyond. *Nat. Rev. Drug Discov.* **2013**, *12*, 117–129. [[CrossRef](#)] [[PubMed](#)]
137. Castro, M.; Corren, J.; Pavord, I.D.; Maspero, J.; Wenzel, S.; Rabe, K.F.; Busse, W.W.; Ford, L.; Sher, L.; Fitzgerald, J.M.; et al. Dupilumab Efficacy and Safety in Moderate-to-Severe Uncontrolled Asthma. *N. Engl. J. Med.* **2018**, *378*, 2486–2496. [[CrossRef](#)]
138. Beck, L.A.; Thaçi, D.; Hamilton, J.D.; Graham, N.M.; Bieber, T.; Rocklin, R.; Ming, J.E.; Ren, H.; Kao, R.; Simpson, E.; et al. Dupilumab Treatment in Adults with Moderate-to-Severe Atopic Dermatitis. *N. Engl. J. Med.* **2014**, *371*, 130–139. [[CrossRef](#)]
139. De Wijs, L.; Bosma, A.; Erler, N.; Hollestein, L.; Gerbens, L.; Middelkamp-Hup, M.; Kunkeler, A.; Nijsten, T.; Spuls, P.; Hijnen, D. Effectiveness of dupilumab treatment in 95 patients with atopic dermatitis: Daily practice data. *Br. J. Dermatol.* **2020**, *182*, 418–426. [[CrossRef](#)]
140. Yamauchi, T.; Sasaki, S.; Lee, E.S.; Tamura, T.; Seki, M.; Miwa, T.; Kobayashi, K.; Saruta, Y.; Kitami, Y.; Sueki, H.; et al. Dupilumab treatment ameliorates clinical and hematological symptoms, including blood eosinophilia, in patients with atopic dermatitis. *Int. J. Dermatol.* **2020**, *60*, 190–195. [[CrossRef](#)]
141. Rohner, L.; Reinhart, R.; Hagmann, B.; Odermatt, A.; Babirye, A.; Kaufmann, T.; Fux, M. FcεRI cross-linking and IL-3 protect human basophils from intrinsic apoptotic stress. *J. Allergy Clin. Immunol.* **2018**, *142*, 1647–1650.e3. [[CrossRef](#)] [[PubMed](#)]
142. Rohner, L.; Reinhart, R.; Iype, J.; Bachmann, S.; Kaufmann, T.; Fux, M. Impact of BH3-mimetics on Human and Mouse Blood Leukocytes: A Comparative Study. *Sci. Rep.* **2020**, *10*, 1–8. [[CrossRef](#)] [[PubMed](#)]
143. Souers, A.J.; Levenson, J.D.; Boghaert, E.R.; Ackler, S.L.; Catron, N.D.; Chen, J.; Dayton, B.D.; Ding, H.; Enschede, S.H.; Fairbrother, W.J.; et al. ABT-199, a potent and selective BCL-2 inhibitor, achieves antitumor activity while sparing platelets. *Nat. Med.* **2013**, *19*, 202–208. [[CrossRef](#)] [[PubMed](#)]