

The Biorepository and Integrative Genomics resource for inclusive genomics: insights from a diverse pediatric and admixed cohort

Silvia Buonaiuto^{1*}, Franco Marsico^{1*}, Akram Mohammed², Lokesh K
Chinthala², Ernestine K Amos-Abanyie¹, Regeneron Genetics Center³, Pjotr
Prins¹, Kyobeni Mozhui^{1,4}, Robert J Rooney⁵, Robert W Williams⁴, Robert L
Davis², Terri H Finkel^{3,5}, Chester W Brown^{1,3}, and Vincenza Colonna^{1,5,6}

**Equal Contribution*

¹Dept of Genetics, Genomics and Informatics, UTHSC, USA

²Center for Biomedical Informatics, UTHSC, USA

³Regeneron Genetics Center, Tarrytown, NY, USA

⁴Department of Preventive Medicine, Division of Preventive Medicine, UTHSC, USA

³Dept of Pediatrics, Division of Genetics, UTHSC, USA

⁵Dept of Pediatrics, Division of Rheumatology, UTHSC, USA

⁶Institute of Genetics and Biophysics, National Research Council, Naples, 80111, Italy

Abstract

The Biorepository and Integrative Genomics (BIG) Initiative in Tennessee has developed a pioneering resource to address gaps in genomic research by linking genomic, phenotypic, and environmental data from a diverse Mid-South population, including underrepresented groups. We analyzed 13,152 genomes from BIG and found significant genetic diversity, with 50% of participants inferred to have non-European or several types of admixed ancestry. Ancestry within the BIG cohort is stratified, with distinct geographic and demographic patterns, as

23 African ancestry is more common in urban areas, while European ancestry is more common
24 in suburban regions. We observe ancestry-specific rates of novel genetic variants, which are
25 enriched for functional or clinical relevance. Disease prevalence analysis linked ancestry and
26 environmental factors, showing higher odds ratios for asthma and obesity in minority groups,
27 particularly in the urban area. Finally, we observe discrepancies between self-reported race and
28 genetic ancestry, with related individuals self-identifying in differing racial categories. These
29 findings underscore the limitations of race as a biomedical variable. BIG has proven to be an
30 effective model for community-centered precision medicine. We integrated genomics educa-
31 tion, and fostered great trust among the contributing communities. Future goals include cohort
32 expansion, and enhanced genomic analysis, to ensure equitable healthcare outcomes.

33 **1 Introduction**

34 The history of human genetic research, from foundational concepts of genetic mapping to the de-
35 velopment of genome-wide association studies, highlights how how a series of technical break-
36 throughs have progressively dismantled biases, and paved the way for more inclusive studies.
37 Comprehensive human linkage studies began in the 1980s with the discovery that restriction length
38 fragment polymorphisms could map Mendelian loci [1]. Several years before the human genome
39 was sequenced, Risch and Merikangas proposed that genotyping large numbers of '*diallelic poly-*
40 *morphisms*' could uncover the genetic basis of common diseases [2], laying the foundation for
41 genome-wide association studies in the early 2000s. However, limited genome data and infor-
42 mative markers led early genetic studies to focus on low-heterogeneity populations like Finnish
43 and Icelandic cohorts, in which power to detect linkage were well matched to informative Single
44 Nucleotide Polymorphisms (SNPs) [3, 4, 5, 6]. The initial SNP arrays had relative low coverage
45 — barely adequate for the most genetically tractable European populations —, and much more
46 limited utility for more diverse populations, especially highly heterogeneous African cohorts[7, 8].

47 Newer high density arrays has enabled the inclusion of non-European populations [9, 10, 11],
48 but they have now been overshadowed by affordable whole exome and genome sequencing and by
49 the welcome addition of deep sequencing of a much more representative swath of humanity [12, 13,
50 14, 15, 16]. However, even supposedly unbiased sequencing suffers from a strong structural bias —
51 namely the reliance on the procrustean expedient of using a single “reference” genome. These final

52 barriers to genetic equity were finally overcome last year with the publication of comprehensive
53 human pangenome assemblies that do not reify a single haploid genome as “The Human” [17, 18].
54 This advancement will allow a minimally biased exploration of genome-phenome-environment
55 relations of almost any human population.

56 Because of the slow progress in technology and data availability described above, along with
57 other important contributing factors, to date most genetic data available for human research has
58 predominantly originated from European populations, introducing a bias in medical research and
59 healthcare that fails to accurately represent the genetic diversity of the global human population
60 [19, 20, 21, 22, 23]. Genetic risk assessments based on European ancestry cohorts yield less accu-
61 rate outcomes for non-European populations, as seen with *CYP2C19* gene variants, which affect
62 drug metabolism and increase risks of misdiagnosis or delayed treatment [24, 25, 26]. While the
63 importance of including ethnically diverse populations in studies of quantitative trait evolution is
64 well known [27], the underrepresentation of diverse populations in genetic research exacerbates
65 health inequities and limits understanding of disease genetics across ancestries, further deepen-
66 ing existing treatment disparities. This underrepresentation underscores the urgent need for more
67 inclusive and diverse genetic studies to improve global health outcomes, leading to a surge of
68 initiatives aimed at addressing these disparities (e.g., [16, 28, 29, 30]).

69 The Biorepository and Integrative Genomics (BIG) Initiative of Tennessee (US), is a multi-
70 institute initiative that has developed a biorepository resource from a diverse Mid-South population
71 in the US, including African Americans, and rural populations in Appalachia, which are dispro-
72 portionately impacted by chronic diseases and the associated costs of healthcare [31, 32]. The BIG
73 biospecimens and their genomic data are linked to de-identified electronic health records, with the
74 purpose of creating a platform for genomics-based research that includes underrepresented popu-
75 lations and to support future personalized healthcare delivery platforms [33]. The initial focus of
76 BIG on building a large and diverse cohort for genetically informed treatment and prevention of
77 pediatric conditions, has now been expanded to a state-wide program that enrolls participants of
78 any age with the goal of building genome-phenome-environment data for 100,000 Tennesseans.

79 Here we report on the analysis of 13,152 genomes from the BIG collection. We demonstrate
80 that the BIG is a genetically diverse and ethnically rich study population, representing a unique
81 and valuable resource for inclusive genomics. Our findings highlight ancestry-specific diversity

82 and genetic burden, underscoring the critical need of inclusive sets of data. Finally, we show that
83 self-reported race does not accurately reflect genetic ancestry and should be cautiously applied as
84 a covariate in genetic analyses.

85 **2 Results**

86 **A robust foundation for inclusive genomics studies**

87 To date, the BIG initiative has consented over 42,000 participants with electronic health records
88 and collected more than 15,000 biosamples from five collection sites (**Fig 1A**). The BIG cohort
89 is predominantly pediatric, with 87% of participants under 18 years old. At the time of sample
90 collection, participant ages ranged from infancy to 90 years, with an average age of 8.4 years and a
91 median age of 6.2 years (Fig. **S1**). BIG stands out as one of the largest cohorts focused on diverse
92 ancestries, providing a substantial representation of different ethnic backgrounds [34, 35, 36, 37,
93 38, 39, 40] (**Table S1**). Notably, it is among the few cohorts specifically enriched for children
94 with diseases, unlike most pediatric cohorts that typically recruit healthy mother-child pairs during
95 pregnancy [41, 42, 43, 35, 36, 38, 40].

96 Since 2017, the BIG initiative has developed the Memphis Genomics Educational Network
97 (*MEMGEN*) to engage the Memphis Shelby County Public School District (MSCPSD) commu-
98 nity in genomics education. *MEMGEN* has reached students in seven public high schools (with
99 plans to expand to 25), providing hands-on genomic experiences and ethical discussions that in-
100 spire STEM careers and academic growth in underserved communities. Community engagement
101 is strengthened through advisory boards like the Le Bonheur Family Partners Council, supporting
102 the BIG initiative since 2015, and the UTHSC Community Advisory Board, representing seven-
103 teen grassroots organizations. These boards ensure research and educational efforts align with
104 community needs, fostering a community-centered approach to precision medicine and addressing
105 health disparities.

106 **Capturing broad diversity and several types of admixture**

107 Within the BIG cohort, we identified and phased 6.8 million high-confidence variable sites, evenly
108 distributed across the genome (**Fig S2**) through exome sequencing and genotype-by-sequencing
109 data from 13,152 individuals. We used this genetic information to understand the ancestry com-
110 position of BIG by performing supervised ancestry deconvolution [44], with 1000 Genomes and
111 HGDP as reference populations [12, 13]. While we observe a clear, uninterrupted continuum of
112 ancestry, we subdivided the data set into seven ancestry groups to account for admixture and fur-
113 ther characterize our cohort (**Fig 1B**). In practice, individuals were classified as not-admixed if
114 more than 85% of their global ancestry corresponded to a single group. The choice of an 85%
115 threshold reflects the understanding that genetic ancestry exists on a continuum, therefore defining
116 discrete categories implies setting thresholds and making arbitrary decisions ([29] see Methods
117 section). Furthermore, ancestral contributions over 10-15% are generally considered accurate and
118 significant, while lower proportions are often linked to shorter ancestral segments and higher error
119 rates [45].

120 According to this ancestry-based grouping, 50% of participants relate to individuals of non-
121 European origin in the reference data sets. In particular, 20% of the BIG individuals are similar
122 to Africans in the reference sets, and 30% present admixed origins, with two-way and multiple-
123 admixture patterns (**Fig 1B**). These figures, projected on all consented individuals, indicate that
124 over 20k consented samples are likely of non-European or admixed origin, placing BIG among the
125 largest pediatric cohorts with many admixed children (**Table S1**).

126 The distribution of inferred ancestry groups by zip code shows ancestry stratification, with
127 prevalence of European ancestry in the suburbs and areas surrounding Memphis (**Fig 1C, S3**).
128 Stratification appears even more marked when visualized by single ancestry (**Fig 1D**). A high dis-
129 similarity index [46] between EUR and AFR (0.67) is observed, highlighting relevant geographic
130 difference, while AFR and EUR-AFR (0.24) are the most evenly distributed pair, indicating much
131 closer spatial overlap (**Fig S3C**). This evidence indicates that BIG individuals with similar ancestry
132 often share a similar environment, implying that geography could act as a confounding factor if
133 not accounted for in association analyses.

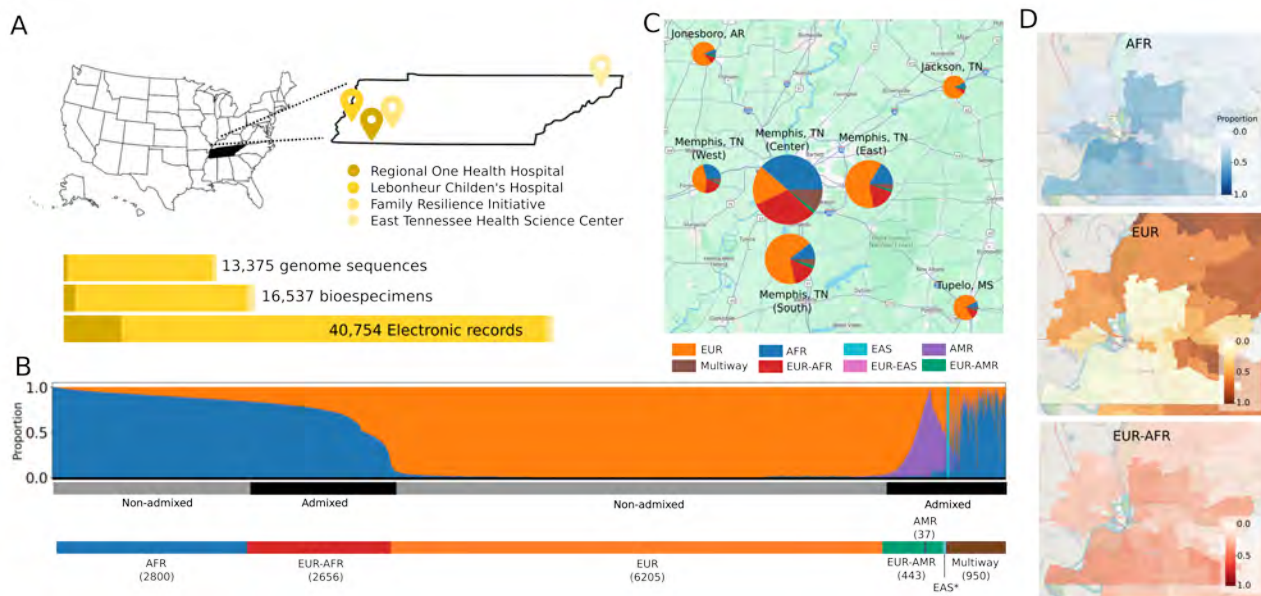


Figure 1: Geographic distribution and global ancestry deconvolution of individuals from the BIG initiative - (A) Overview of data collected across four sites in Tennessee, US. **(B)** Global ancestry deconvolution of 13,152 sequenced individuals, based on reference populations in the 1000 Genomes and HGDP data sets. Each vertical bar represents one individual, colors are proportional to inferred ancestry. For further analyses, individuals were grouped based on the ancestry proportions in seven categories (colored bar, number of individuals per category in parentheses), and classified as admixed or not (black and gray bar) **(C)** Proportion of individuals corresponding to each ancestry stratified by the zip code. **(D)** Prevalence of ancestries by zip code - EUR: European; AFR: African; EAS: East-Asian; AMR: Indigenous-American.

134 Integrating genetic, phenotypic, and environmental information

135 Electronic health records are an integral part of the BIG cohort, covering a range of Phecode
 136 categories [47], with gastrointestinal and respiratory medical conditions among the most repre-
 137 sented (**Fig S4**). We examined the prevalence of obesity, hypertension, diabetes and asthma, four
 138 health conditions commonly associated with minority groups and local environmental influences
 139 [48]. BIG children have a high incidence of diabetes and asthma (363 and 697 cases, respectively,
 140 **Fig 2A**), while adults have a more balanced incidence across these same four diseases (**Fig S5**).
 141 Ancestry categories such as AFR and EUR-AFR, are major contributors across conditions, and
 142 we observed higher odds ratios for obesity and asthma in minority groups (all individuals self-
 143 identified as belonging to non-White racial groups) compared to 200 randomly selected conditions
 144 (**Fig 2B**).

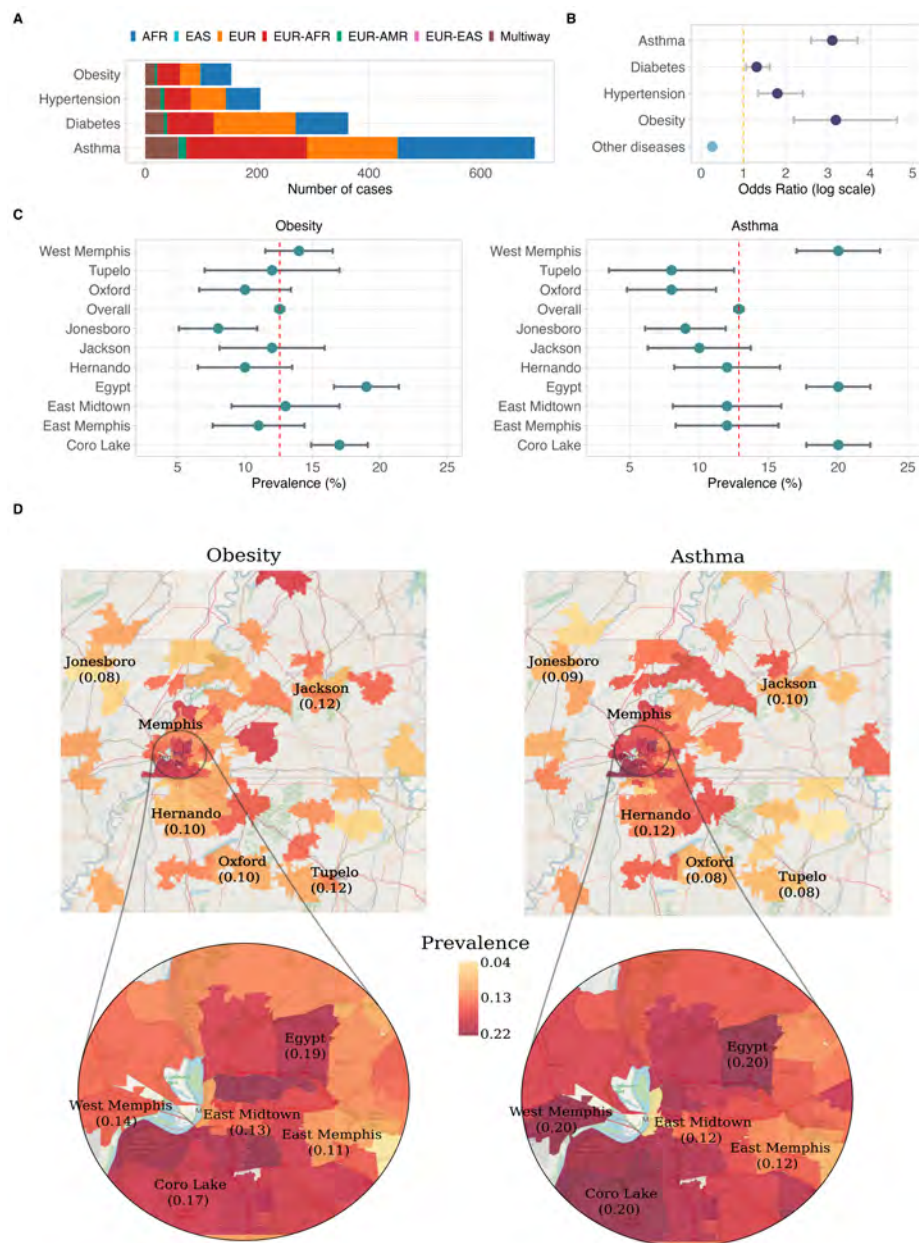


Figure 2: Prevalence of diseases common in health disparities populations. (A) Number of cases stratified by inferred ancestry categories. **(B)** Odds ratios for asthma, diabetes, hypertension, obesity compared to odds ratio of two hundred random diseases, observed among individuals self-identifying as belonging to non-White racial groups. **(C)** Prevalence of obesity and asthma by zone. This is defined as the proportion of cases in the total population. The 95% confidence intervals are calculated using the Wald method. **(D)** The map displays zones color-coded by prevalence levels in locations with more than 100 total individuals. The Memphis Metropolitan area, characterized by high population density, is zoomed in.

145 Analysis of disease prevalence by zip code suggests a notable environmental component for
146 obesity and asthma. In particular, three suburban areas around Memphis exhibit above-average
147 prevalence for both conditions, with asthma being 1.7 times more prevalent in these zones com-
148 pared to the overall prevalence in BIG ($\approx 20\%$ versus 12.8% CI95 [12.51-13.19] **Fig 2C**). While
149 these analyses are only preliminary, the resulting observations underscore the value of the BIG
150 dataset in linking genetic, phenotypic, and environmental information, enabling a multidimen-
151 sional understanding of health disparities.

152 **Ancestry-specific diversity and genetic burden**

153 Our joint principal component analysis (PCA) of the BIG and 1000 Genomes datasets (**Fig 3A**)
154 reveals significant genetic diversity in the BIG dataset, with mixed ancestry groups contributing
155 to the spread and overlap between clusters corresponding to African, American, East Asian, and
156 European individuals in the 1000 Genomes. In contrast, the 1000 Genomes dataset exhibits more
157 distinct clustering with minimal overlap, reflecting more clearly defined ancestral groups. These
158 results underscore the BIG dataset's value in capturing admixture and genetic diversity not repre-
159 sented in the 1000 Genomes, highlighting the importance of including diverse and admixed popu-
160 lations in genetic studies to better capture the full spectrum of human variation.

161 As expected, the average number of genetic differences from the reference human genome
162 varies by ancestry [12]. Individuals with African or admixed African ancestry typically have, on
163 average, $\sim 85k$ more variable sites compared to other ancestry groups (**Fig 3B**). This observation
164 underscores the risk of bias in using a single reference sequence and its associated genomic an-
165 notations. The genetic diversity represented within BIG would be more accurately modeled by a
166 pangenomic approach [17].

167 Our dataset includes 771,717 novel single nucleotide variants (11.2% of the total), which are
168 absent from major databases such as gnomAD, 1000 Genomes Project, or Human Genome Di-
169 versity Project [12, 13, 49]. Novel variants are mostly rare and private to ancestries, as expected
170 (**Fig S6**). The rough number of novel variants per individual is higher within inferred admixed
171 ancestries, Americans, and Asians (**Fig 3C**). Some novel variants have important functional con-
172 sequences on the gene product (**Fig S6**, VEP classification [50]: 2.8% high impact, including

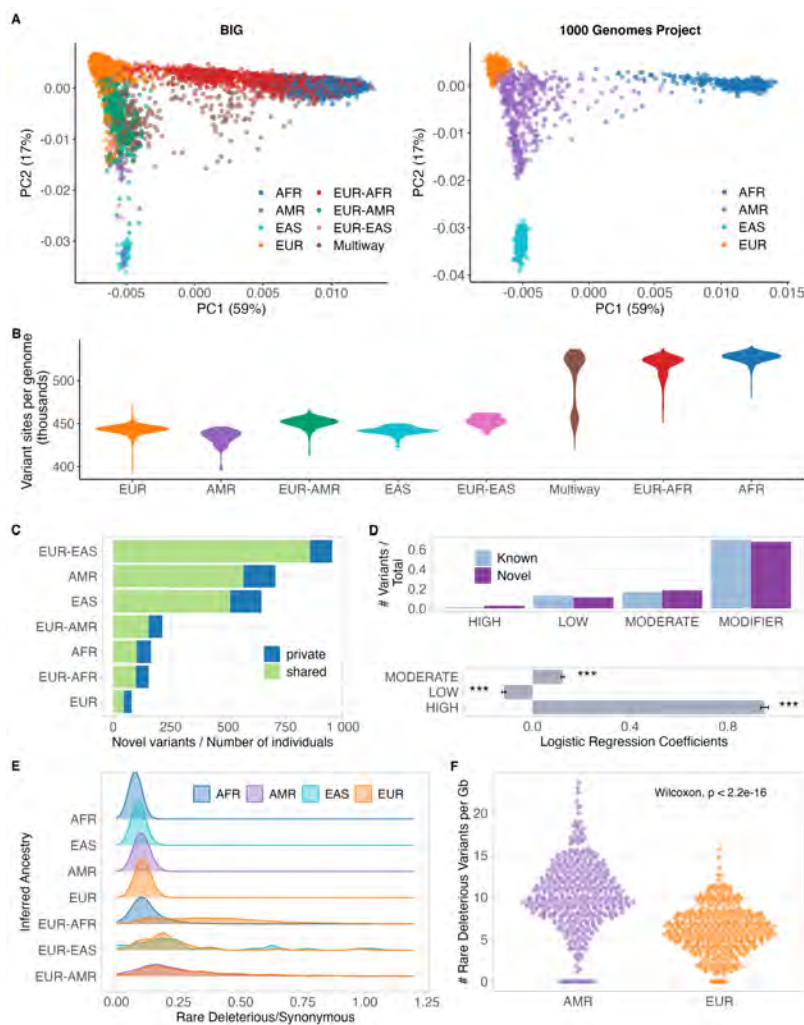


Figure 3: Genetic variability and genetic burden in the BIG cohort - (A) Joint principal component analysis of genetic data from individuals in the BIG and in the 1000 Genomes populations, represented separately for clarity. Colors represent inferred genetic ancestry. The first two principal components explain 76% of the variance captured by the first 20 PCs. **(B)** Number of variable sites per genome compared to the reference sequence as a function of inferred ancestry. **(C)** Estimate of the number of novel variants by individuals per ancestry with indication of variants private to the ancestry **(D)** Proportion of known and novel variants across different impact categories. **(E)** Rare deleterious-to-synonymous variant ratio across inferred ancestries. The peaks and spreads of these distributions highlight variation in the frequency of deleterious mutations across ancestries, reflecting potential differences in genetic diversity, mutation load, and evolutionary pressures. **(F)** Count of rare deleterious variants in EUR-AMR admixed individuals, which have the highest deleterious-to-synonymous ratio. Variant counts are assigned based on the inferred ancestry of the genomic regions where they are found. This means individuals are counted twice: once for their AMR ancestry regions and once for their EUR ancestry regions.

173 frameshift variants, stop/start gain/loss and splicing affecting variants; 19.7%: missense) and po-
174 tential implications for disease association (11.0% predicted to be deleterious by SIFT [51]; 7.9%
175 considered probably or possibly damaging by PolyPhen [52]). Notably, the rate of high impact
176 annotation in novel variants is double compared to known (logistic regression coefficient $\beta=0.95$,
177 p-value<0.001, **Table S3, Fig. 3D**)

178 Genetic burden by ancestry was evaluated as the distribution of rare deleterious (alternate al-
179 lele frequency <1% in the total BIG samples, predicted to have high impact or missense with
180 SIFT<0.05 and Polyphen>0.85) versus rare synonymous genetic variants across different an-
181 cestral groups. Among non-admixed groups, African individuals display the lowest deleteri-
182 ous/synonymous ratio, whereas European individuals exhibit the highest (**Fig 3C**). Admixed popu-
183 lations show broader distributions in deleterious/synonymous ratios, with the European-American
184 group demonstrating the highest ratios. In EUR-AMR group, the average number of rare deleteri-
185 ous variants per Gb is significantly higher in the AMR tracts compared to EUR ones (**Fig 3D, Fig**
186 **S7**) as shown in other studies [53], likely due to demography and founder effect [54, 55].

187 Overall, the remarkable breadth of genetic diversity observed underscores BIG's value as a
188 comprehensive resource for exploring genetic variation, enhancing disease association studies, and
189 promoting equitable genomic research in underrepresented populations.

190 **Discrepancies between self-reported race and inferred genetic ancestry**

191 We compared counts of individuals in self-reported racial categories with those in inferred genetic
192 ancestry categories, with some racial categories aggregated for simplicity (Table S2). The number
193 of self-reported White individuals aligns closely with those inferred as Europeans, while partici-
194 pants identifying as Black or African American appear distributed between two genetic ancestry
195 categories: Africans and admixed African-Europeans. For other racial groups, the patterns are
196 more diverse and complex (**Fig 4A**).

197 We evaluated the fraction of the genome shared identical by descent (IBD) among all possi-
198 ble pairs of individuals and compared with self-reported race. Predictably, IBD genome sharing
199 was higher among individuals within the same self-reported race. However, we also detected IBD
200 relationships greater than the 2nd degree (compatible with 1st cousin or uncle-nephew relation-

201 ship) between individuals of different self-reported races (**Fig 4B**). This observation suggests that
202 genetically related individuals may self-identify differently with respect to socially constructed
203 categories like race.

204 The relationship between self-reported race and inferred ancestry was further examined among
205 pairs of individuals who identified as belonging to the same race. In some instances, the self-
206 reported race of a pair differed from that of other pairs within the same ancestry category (**Fig**
207 **4C**). For example, one pair of first-degree relatives (sharing approximately 50% of their genome)
208 who both self-reported as White were found to have differing inferred ancestries: one individ-
209 ual was classified as having African ancestry, while the other showed a mixture of African and
210 European ancestries (represented by the orange triangle in the AFR; EUR-AFR category in **Fig**
211 **4C**). Similarly, among three pairs of individuals self-reporting as Black or African American, one
212 member of each pair was inferred to have European ancestry (represented by the purple triangle
213 in the EUR; EUR-AFR category in **Fig 4C**). These findings highlight the limitations of using
214 self-reported race as a category for analyzing genetic variation.

215 **3 Discussion**

216 The BIG cohort is a genetically diverse and ethnically inclusive pediatric resource, addressing
217 the historic underrepresentation of non-European populations in genomics research. With 87% of
218 participants under 18 and 50% of non-European ancestry—including 20% closely aligning with
219 African reference populations and 30% exhibiting complex admixture patterns—it offers broad
220 genetic variability and significant potential to represent human genomic diversity. Previous com-
221 parative studies have shown that admixed African populations from Tennessee rank among those
222 with the highest proportion of African ancestry in the United States [56]. Notably, individuals from
223 Memphis exhibit the greatest genetic diversity within their African ancestry component compared
224 to thirteen other similar populations [57]. Although our study is not explicitly comparative, these
225 findings position the African and admixed African individuals in the BIG cohort as being among
226 the most genetically diverse populations globally.

227 This diversity facilitated the discovery of new genetic variants, many with clinical relevance.
228 We have indications of ancestry-specific burden in admixed individuals. While this is an intrigu-

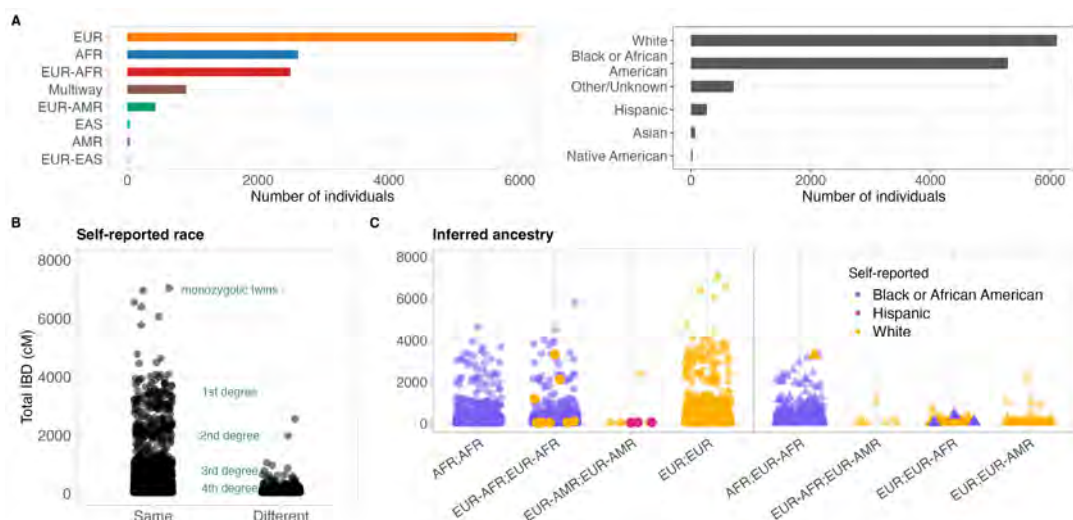


Figure 4: **Poor alignment between self-reported race and genetic ancestry.** (A) Counts of individuals per inferred ancestry (left) and self-reported race (right). (B) Genome segments shared Identical By Descent (IBD) in centimorgans (cM) between all individual pairs in BIG, categorized by whether individuals self-reported the same or different race. In some instances, individuals who self-report as belonging to different races are related at the third-degree level (e.g., first cousins) or even as close as second-degree relatives (e.g., half-siblings), as indicated by the IBD analysis. (C) IBD genome sharing and inferred ancestry among individuals self-reporting the same race (color-coded). In some cases the self-reported race of a pair deviates from the patterns observed in other pairs within the same ancestry category.

229 ing observation, it certainly deserves further investigation before any definitive conclusions can be
230 reached. We believe that several factors, including sample size, stratification effects, and demog-
231 raphy, must be carefully considered to achieve a more solid conclusion. This again underscores
232 the importance of ensuring that relevant populations are well represented, as failing to do so risks
233 leading to erroneous conclusions.

234 As a model for studying health disparities, the BIG cohort reveals higher odds ratios for obesity
235 and asthma among minority groups, driven by genetic and environmental factors, as reflected in
236 zip-code-specific disease patterns. We show that the BIG cohort has the potential to integrate ge-
237 nomic data, electronic health records, and environmental information to thoroughly analyze these
238 and other common diseases. [58] With relevance to disease mapping, our study highlights how self-
239 identified racial categories often fail to align with genetic ancestry, as seen in other studies [59].
240 The value of using race in biomedical research has been a longstanding topic of debate [60, 61].
241 Race is predominantly a socio-cultural construct, reflecting identity and social experiences rather

242 than genetic heritage [62]. Nevertheless, race can serve as a useful framework for describing health
243 disparities in societies where racial categories are deeply embedded in social structures [59], and
244 there have been increasing calls for greater inclusion of underrepresented individuals in genetic
245 and biomedical research to help clarify the relationship between race and ancestry [63, 64].

246 A peculiar feature of the BIG cohort is the inclusion of many admixed individuals, encom-
247 passing three distinct types of admixture. Admixed populations constitute a significant part of
248 global genetic diversity and present unique statistical challenges in the analysis of genetic varia-
249 tion, leading to their frequent exclusion from genomics and medical research. Admixture can be
250 used to map quantitative traits and to detect positive selection [65, 66], requiring smaller sample
251 sizes compared to other mapping techniques [67]. Admixture mapping leverages local ancestry
252 inference to associate traits with an unusually high proportion of ancestry from one of the parental
253 populations around the disease-causing locus [68, 69, 70] and it has been successfully used - as an
254 example - to map Alzheimer's disease [71].

255 All the findings from the BIG study hold significant implications primarily for the scientific
256 community, however, and most importantly, BIG pioneers a model for inclusive genomic studies,
257 emphasizing community engagement to align research efforts with the needs of the contributing
258 communities. Clinically, the insights gained from BIG can inform precision medicine initiatives for
259 historically underserved populations, particularly in regions of Tennessee, where African Amer-
260 icans and others face a disproportionate burden of chronic disease. Through *MEMGEN* local
261 students and families engage with hands-on genomics education and ethical aspects of genetic re-
262 search, which demystifies the science and inspires interest in STEM fields, promoting inclusivity
263 by respecting cultural contexts and building trust.

264 A future key priority for the BIG initiative is to expand its participant base to include adults,
265 allowing for a comprehensive study across all age groups and an even broader spectrum of genetic
266 diversity. Continued community education is also a priority to sustain engagement and participa-
267 tion in the BIG initiative. Another important priority is to adopt a pangenomic approach in genetic
268 data analysis to better represent the genetic diversity within the cohort. Moving toward an inclusive
269 genome model that integrates multiple ancestries and population-specific variants will enhance the
270 accuracy of variant identification and genetic association studies for individuals in the BIG cohort.

271 By embracing this pangenomic approach, the BIG initiative can establish a new benchmark

272 for inclusive genomics, ensuring that research benefits all participants by reflecting their unique
273 genetic backgrounds.

274 In conclusion, the BIG initiative can continue to lead in inclusive genomics, creating a resource
275 that supports equitable health outcomes and advances the field toward a truly representative model
276 of precision medicine.

277 **4 Methods**

278 **Ethics**

279 This study adhered to the ethical principles outlined in the Declaration of Helsinki for medical
280 research involving human subjects. This study was conducted in accordance with ethical standards
281 and is approved by the Institutional Review Board (IRB) of UTHSC (IRB number: 23-09204-
282 NHSR). Written informed consent was obtained from all participants; for pediatric subjects, con-
283 sent was provided by their legal guardians or next of kin. To ensure confidentiality, all data were
284 de-identified prior to analysis.

285 **Sample collection sites**

286 Le Bonheur Children's Hospital (LBCH, Memphis, TN) - LBCH is the primary pediatric care cen-
287 ter in Memphis, and serves a predominantly African American population in an area marked by
288 significant health disparities. Recruitment at this site was launched in October 2015 and spans
289 inpatient rooms, ICUs, outpatient clinics, and the emergency department. Information from ge-
290 nomic DNA extracted from leftover blood collected during routine care is linked to de-identified
291 electronic health record data. Leftover samples are not always available for collection, although
292 they can be collected on a subsequent visit. This explains the discrepancy between the number of
293 consented participants and collected biosamples.

294 Regional One Health (ROH, Memphis, TN) - ROH is a leading healthcare provider in Mem-
295 phis, providing comprehensive care to underserved and vulnerable communities in the same ge-
296 ographical area of LBCH. In May 2022, the BIG Initiative extended its reach to ROH, focusing
297 on adult genomic research. Participants are recruited across hospital settings, with DNA collected

298 from leftover blood during standard care and linked to de-identified EHR data. This expansion
299 complements BIG's pediatric focus at LBCH by including a diverse adult population.

300 East Tennessee State University (ETSU, Johnson City, TN) - The BIG Initiative expanded to
301 ETSU in May 2023 to include the Appalachian region, emphasizing adult participant recruitment.
302 DNA samples are collected through dedicated blood draws and linked to de-identified EHR data.
303 ETSU's inclusion aligns with BIG's commitment to engaging rural and underserved populations,
304 complementing efforts at LBCH and ROH to create a robust, diverse genomic database for advanc-
305 ing precision medicine across the Mid-South and Appalachia.

306 Family Resilience Initiative (FRI, Memphis, TN) - Launched in January 2019, the Family Re-
307 silience Initiative (FRI) examines the impact of adverse childhood experiences (ACEs) and social
308 determinants of health on long-term outcomes. The program enrolls mother-child dyads from the
309 Memphis region, collecting sputum and/or blood samples at four visits spaced six months apart.
310 Samples are processed through BIG's operational pipeline for DNA isolation, cortisol measure-
311 ments, and clinical assessments. By linking biological and environmental data, FRI aims to under-
312 stand ACEs' physiological and epigenetic effects, providing insights to guide tailored interventions
313 and improve family health in vulnerable communities.

314 **DNA sequencing**

315 The 13,152 samples were processed with NEB/Kapa reagents, captured with the Twist Comprehen-
316 sive Exome Capture design, enhanced by Regeneron-designed spikes targeting sequencing geno-
317 typing sites. Among the sequenced samples average coverage is 20X for 95.2%, with 99.3% above
318 90%, highlighting the overall quality of the data. The genotyping spike targets an additional $\approx 1.4M$
319 variants in the human genome. Genotyping call rate (percentage of SNP / indels targeted geno-
320 typing at which a call can be made) is 99.0%. CHIP targets mean coverage, crucial for detecting
321 low-frequency variations, averages at 100X. All samples were sequenced on an Illumina NovaSeq
322 6000 system on S4 flow cells sequencer using 2x75 paired-end sequencing.

323 **Variant identification**

324 Sequence reads were aligned by the Burrows-Wheeler Aligner (BWA) MEM [72] to the GRCh38
325 assembly of the human reference genome in an alt-aware manner. Duplicates were marked using
326 Picard, and mapped reads were sorted using sambamba [73]. DeepVariant v0.10.0 with a custom
327 exome model was used for variant calling [74], and the GLnexus v1.2.6 tool was used for joint
328 variant calling [75]. The variants were annotated using a Variant Effect Predictor (VEP 110) [50].
329 Phasing was performed using ShapeIT v5 [76].

330 **Global and Local Ancestry inference**

331 To characterize the genetic admixture within the BIG cohort, we performed a global and local an-
332 cestry inference (LAI) analysis using RFMix v.2.0; [https://github.com/slowkoni/](https://github.com/slowkoni/rfmix)
333 [rfmix](https://github.com/slowkoni/rfmix) [44]. Reference samples included those of the 1000 Genomes Project and the Human
334 Genome Diversity Project (HGDP), using the recently developed joint call [77]. The merged
335 genotyping dataset, which combined BIG participants with reference samples, consisted of au-
336 toosomal variants. To select the reference samples, we followed a quality control previously used in
337 other studies [78]. To exclude reference samples with extensive admixture, we performed an un-
338 supervised cluster analysis using ADMIXTURE [79]. We selected 4 groups ($k = 4$), and reference
339 samples with a major group proportion > 0.99 were considered for the analysis. Four-way LAI
340 was performed with the number of terminal nodes for the random forest classifier set to 5 ($-n 5$),
341 the average number of generations since the expected addition set to 12 ($-G 12$), and ten rounds
342 of the expectation maximization algorithm (EM) ($-e 10$). Reference superpopulations selected at
343 the continental level were African (AFR), American (AMR), European (EUR), and Asian (EAS).
344 Specifically, AFR is represented by YRI (101), LWK (30), MSL (16), Mbuti (10), GWD (48),
345 ESN (64), Bantu South Africa (3), Bantu Kenya (10) and Biaka (21) groups. EUR contains Tus-
346 can (6), Sardinian (12), Orcadian (13), IBS (117), GBR (103), French (24), Bergamo Italian (9),
347 Basque (17) and CEU (114). AMR by Surui (6), Pima (10), PEL (10), Maya (16), Karitiana (7),
348 and CLM (7). Finally, EAS is represented by CHS (106) and CHB (39). Local ancestry inference
349 with RFMix2 was used to classify rare alleles ($AF < 0.01$), both synonymous and deleterious, by
350 ancestry. A custom script was developed to process phased VCFs with local ancestry calls, assign-

351 ing each allele to an ancestral population and generating ancestry-specific haplotype counts. This
352 approach enables the precise tracking of allelic ancestry in samples.

353 Discrete ancestry categories (AMR, AFR, EUR, EAS, EUR-AMR, EUR-AFR, and Multiway)
354 were defined based on the following criteria: (i) individuals with more than 85% of a single an-
355 cestry were categorized into single-ancestry groups; (ii) individuals with at least 15% contribution
356 from two ancestries, and a combined total of over 85%, were classified as two-way admixed; (iii)
357 individuals with significant contributions (greater than 15%) from three or more ancestries were
358 classified as Multiway. The number of individuals per ancestry group by ZIP code (based on
359 ZCTA5 Code Tabulation Areas from the 2020 U.S. Census) was used to map the proportion of
360 each ancestry within each location. The dissimilarity index [46] was calculated for ancestry cate-
361 gories with populations exceeding 500 individuals. To ensure reliable calculations, ZIP codes with
362 fewer than 100 total individuals were excluded from the analysis.

363 **About inferred population labels**

364 In this study, we use self-reported race and ethnicity, which are socially constructed and categori-
365 cal, alongside genetic ancestry proxies derived from methods like RFMix [44]. Although race and
366 ethnicity are discrete categories that reflect social and historical contexts, genetic ancestry arises
367 from continuous biological processes that capture paths through the ancestral recombination graph
368 [80]. To facilitate our analysis, we categorize genetic ancestry into regional groupings such as
369 AMR (ancestries from the Americas) or EUR (ancestries from Europe), but it is important to clar-
370 ify that these labels are not fixed or essentialized categories [81]. This grouping is useful only
371 because it helps us explore the demographic and environmental histories that shape the variation
372 of complex genetic traits. This discretization is merely one arbitrary scale, and in several anal-
373 yses, we examine finer ancestral variation within these groupings using dimensionality reduction
374 techniques (PCA), unsupervised clustering (ADMIXTURE) and relatedness (e.g., IBD segment
375 analyses). We emphasize that such proxy cannot be equated with historical racial categories that
376 have been used to justify inequality [82]. In fact, a part of the results section is focused on showing
377 the discrepancies between both categories.

378 **About self-reported race**

379 Race is self-reported by enrolled patients at the time of admission to the hospital. The admission
380 staff select the race code from a drop-down list of possible race categories according to HL7 stan-
381 dards for race and ethnicity [https://hl7-definition.caristix.com/v2/HL7v2.5/](https://hl7-definition.caristix.com/v2/HL7v2.5/Tables/0005)
382 [Tables/0005](https://hl7-definition.caristix.com/v2/HL7v2.5/Tables/0005). It is possible to select multiple race codes from the drop-down list in case people
383 associate themselves with multiple races.

384 **Clinical Data**

385 The clinical data associated with BIG participants are extracted from the EHR (Electronic Health
386 Records) system in flat files and shared with UTHSC through a secure file transfer protocol. These
387 data include demographics, visits, diagnoses, procedures, prescribed and administered medica-
388 tions, labs, and vital signs. These data elements are converted to a limited data set (LDS) and
389 mapped to a common data model, the OMOP (Observational Medical Outcomes Partnership)
390 CDM. To support the analysis, the ICD9/10 diagnosis codes are assigned to PheCodes.

391 **Diversity and population structure analyses**

392 Joint PCA, considering BIG and 1000GP cohorts, was performed in order to compare genetic di-
393 versity. We used the bigsnpr R package protocol for PCA analysis (<https://privefl.github.io/bigsnpr>)
394 [83]. Briefly, this involved using King software [84] to estimate kinship coefficients and remove
395 first and second-degree relatives (cutoff < 0.0884). LD clumping ($r < 0.2$) and exclusion of
396 long-range LD regions were based on Mahalanobis distances. Outliers were identified with K-
397 nearest-neighbor. The first 20 PCs were computed using truncated SVD. After excluding outliers,
398 we projected related individuals in the PC space. Variants with $MAF < 0.01$ were excluded. For
399 ADMIXTURE analyses, we performed unsupervised clustering with $k = 3, 4, 5,$ and 6 . We applied
400 standard quality control filters, including LD pruning and removal of variants with $MAF < 0.01$.
401 Logistic regression was performed in R.

402 **Relatedness and Identical By Descent analysis**

403 To analyze relatedness and infer family relationships, we used KING software to calculate kinship
404 coefficients and determine the probability of sharing zero IBD (identity by descent) [84]. Quality
405 control for kinship inference included removing variants with high missingness, filtering by MAF
406 > 0.01 , and performing LD pruning.

407 To identify IBD segments, we used hap-ibd in the phased data set comprising 13,152 genomes,
408 focusing on autosomal loci [85]. Hap-ibd was executed with a minimum seed parameter of 2 cM
409 to detect IBD segments of at least this length. The inferred IBD segments were post-processed
410 using the protocol developed by Browning et al. [86], particularly the merge-ibd-segments tool,
411 with default parameters. Gaps with at most one discordant homozygote and less than 0.6 cM were
412 removed.

413 **Code availability**

414 The scripts used for QC, PCA, local and global ancestry deconvolution, and IBD analysis are
415 available on <https://github.com/SilviaBuonaiuto/BIG>

416 **5 Data availability**

417 The BIG data presented here is potentially identifiable human data, and therefore its availability
418 is somewhat restricted. However, we strongly support data availability in general. Data used for
419 this study can be shared after University of Tennessee Health Science Center institutional IRB and
420 BIG Research Oversight Committee review and approval <https://uthsc.edu/cbmi/big/>.
421 Please contact the authors for further information.

422 **6 Acknowledgments**

423 We extend our gratitude to all the individuals and their families who generously contributed to
424 the BIG initiative. We would like to thank Carol Hendrix and the consent teams in Memphis
425 and in Johnson City for oversight of recruitment and sample collection; Kito Lord, from ROH;

426 James Adkins, and Jonathan Patrick Moorman from ETSU; Jason Yaun, Sandra Arnold from FRI;
427 Marcella Vacca; Scott Strome; Jon McCullers; David Haines; Peter Buckley, G. Nicholas Verne,
428 and Pamela Beckley from UTHSC; Trey Eubanks from Le Bonheur Children’s Hospital; the BIG
429 Community Advisory Board.

430 The authors gratefully acknowledge support from the Center for Integrative and Translational
431 Genomics at UTHSC (SB, FM, RWW, PP, VC); NIH/NIGMS (R01GM123489 to PP); NSF (PPoSS
432 Award 2118709 to PP); the NIH/NHLBI (RO1 HL170151 to THF); The Rady Children’s Institute
433 for Genomic Medicine (THF); the Children’s Foundation of Memphis (THF); the Urban Child
434 Institute; the Children’s Foundation Research Institute, Children’s Foundation of Memphis; the
435 Assisi Foundation (CWB).

436 **7 Author contribution**

437 SB, FM, PP, KM, RWW, RLD, THF, CWB, VC; Data curation: SB, FM, AM, LKC; Formal
438 Analysis: SB, FM, AM, EKA, VC; Funding acquisition: PP, RJR, RWW, RLD, THF, CWB, VC;
439 Investigation: SB, FM, VC; Methodology: SB, FM, VC; Project administration: ; Resources: PP,
440 RWW, CWB; Software: PP; Supervision: PP, RJR, RWW, RLD, THF, CWB, VC; Validation: ;
441 Visualization: SB, FM, VC; Writing – original draft: SB, FM, EKA, RWW, RLD, THF, CWB,
442 VC; Writing – review & editing: SB, FM, AM, LKC, PP, KM, EKA, RJR, RWW, RLD, THF,
443 CWB, VC.

444 **Extended Affiliation [3]:**

445 3. Regeneron Genetics Center, Tarrytown, NY, USA.

446 RGC Management & Leadership Team Aris Baras, Goncalo Abecasis, Adolfo Ferrando, Gio-
447 vanni Coppola, Andrew Deubler, Aris Economides, Luca A Lotta, John D Overton, Jeffrey G
448 Reid, Alan Shuldiner, Katherine Siminovitch, Jason Portnoy, Marcus B Jones, Lyndon Mitnaul,
449 Alison Fenney, Jonathan Marchini, Manuel Allen Revez Ferreira, Maya Ghoussaini, Mona Nafde,
450 William Salerno.

451 Sequencing & Lab Operations John D Overton, Christina Beechert, Erin Fuller, Laura M Cre-
452 mona, Eugene Kalyuskin, Hang Du, Caitlin Forsythe, Zhenhua Gu, Kristy Guevara, Michael Lat-
453 tari, Alexander Lopez, Kia Manoochehri, Prathyusha Challa, Manasi Pradhan, Raymond Reynoso,
454 Ricardo Schiavo, Maria Sotiropoulos Padilla, Chenggu Wang, Sarah E Wolf, Hang Du, Kristy
455 Guevara.

456 Clinical Informatics Amelia Averitt, Nilanjana Banerjee, Dadong Li, Sameer Malhotra, Justin
457 Mower, Mudasar Sarwar, Deepika Sharma, Sean Yu, Aaron Zhang, Muhammad Aqeel.

458 Genome Informatics & Data Engineering Jeffrey G Reid, Mona Nafde, Manan Goyal, George
459 Mitra, Sanjay Sreeram, Rouel Lanche, Vrushali Mahajan, Sai Lakshmi Vasireddy, Gisu Eom, Kr-
460 ishna Pawan Punuru, Sujit Gokhale, Benjamin Sultan, Pooja Mule, Eliot Austin, Xiaodong Bai,
461 Lance Zhang, Sean O’Keeffe, Razvan Panea, Evan Edelstein, Ayesha Rasool, William Salerno,
462 Evan K Maxwell, Boris Boutkov, Alexander Gorovits, Ju Guan, Lukas Habegger, Alicia Hawes,
463 Olga Krasheninina, Samantha Zarate, Adam J Mansfield, Lukas Habegger.

464 Analytical Genetics & Data Science Goncalo Abecasis, Manuel Allen Revez Ferreira, Joshua
465 Backman, Kathy Burch, Adrian Campos, Liron Ganel, Sheila Gaynor, Benjamin Geraghty, Arko-
466 pravo Ghosh, Salvador Romero Martinez, Christopher Gillies, Lauren Gurski, Joseph Herman,
467 Eric Jorgenson, Tyler Joseph, Michael Kessler, Jack Kosmicki, Adam Locke, Priyanka Nakka,
468 Jonathan Marchini, Karl Landheer, Olivier Delaneau, Maya Ghoussaini, Anthony Marcketta, Joelle
469 Mbatchou, Arden Moscati, Aditeya Pandey, Anita Pandit, Jonathan Ross, Carlo Sidore, Eli Stahl,
470 Timothy Thornton, Sailaja Vedantam, Rujin Wang, Kuan-Han Wu, Bin Ye, Blair Zhang, Andrey
471 Ziyatdinov, Yuxin Zou, Jingning Zhang, Kyoko Watanabe, Mira Tang, Frank Wendt, Suganthi
472 Balasubramanian, Suying Bao, Kathie Sun, Chuanyi Zhang.

473 Therapeutic Area Genetics Adolfo Ferrando, Giovanni Coppola, Luca A Lotta, Alan Shuldiner,
474 Katherine Siminovitch, Brian Hobbs, Jon Silver, William Palmer, Rita Guerreiro, Amit Joshi,
475 Antoine Baldassari, Cristen Willer, Sarah Graham, Ernst Mayerhofer, Erola Pairo Castineira,
476 Mary Haas, Niek Verweij, George Hindy, Jonas Bovijn, Tanima De, Parsa Akbari, Luanluan
477 Sun, Olukayode Sosina, Arthur Gilly, Peter Dornbos, Juan Rodriguez-Flores, Moeen Riaz, Manav
478 Kapoor, Gannie Tzoneva, Momodou W Jallow, Anna Alkelai, Ariane Ayer, Veera Rajagopal, Sahar
479 Gelfman, Vijay Kumar, Jacqueline Otto, Neelroop Parikshak, Aysegul Guvenek, Jose Bras, Silvia
480 Alvarez, Jessie Brown, Jing He, Hossein Khiabani, Joana Revez, Kimberly Skead, Valentina
481 Zavala, Jae Soon Sul, Lei Chen, Sam Choi, Amy Damask, Nan Lin, Charles Paulding.

482 Research Program Management and Strategic Initiatives Marcus B Jones, Esteban Chen, Michelle
483 G LeBlanc, Jason Mighty, Jennifer Rico-Varela, Nirupama Nishtala, Nadia Rana, Jaimee Hernan-
484 dez.

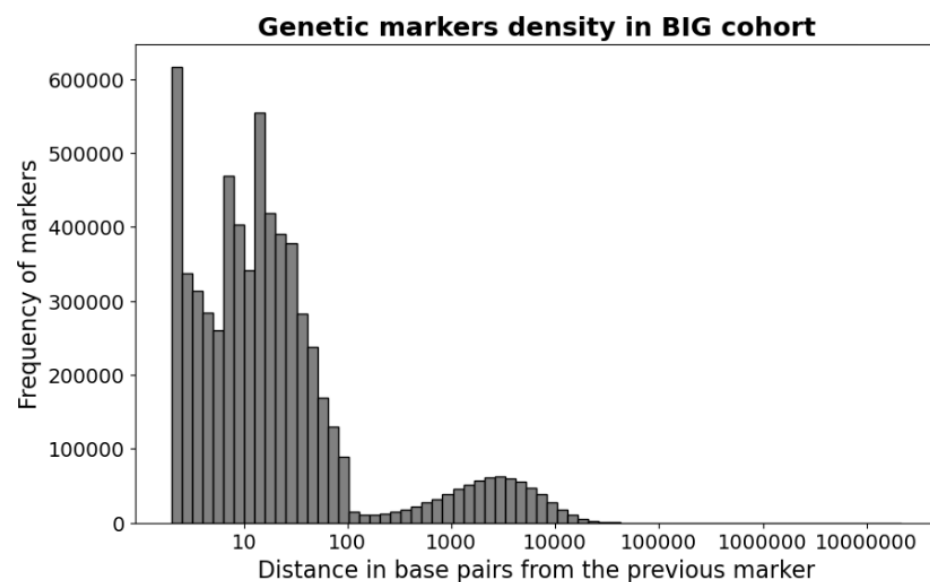
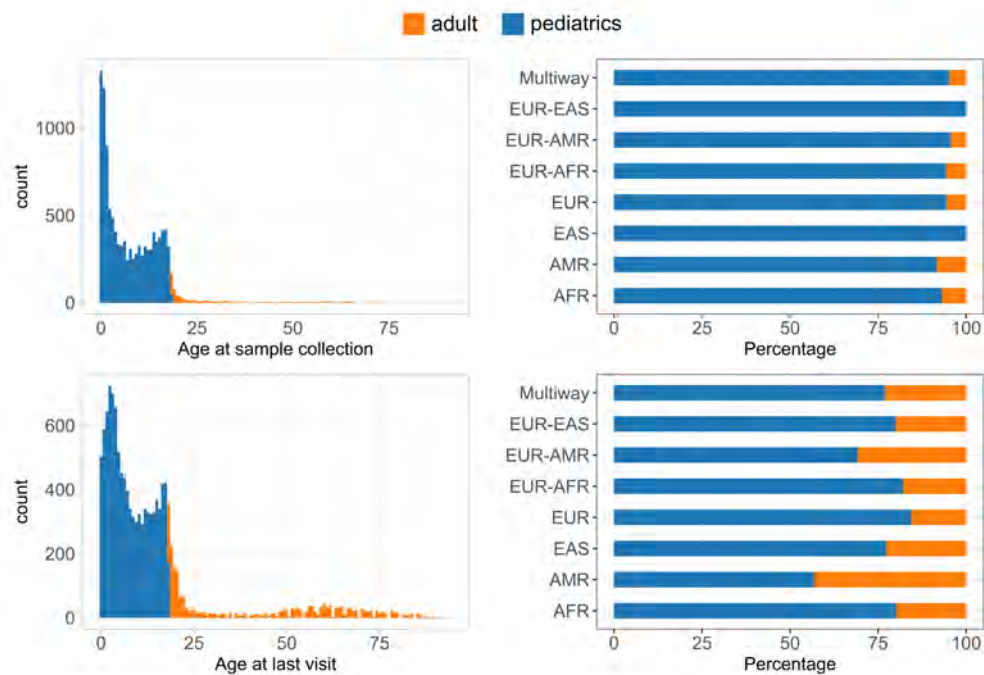
485 Senior Partnerships and Business Operations

486 Alison Fenney, Randi Schwartz, Jody Hankins, Anna Han, Samuel Hart.

487 Business Operations and Administrative Coordinators

488 Ann Perez-Beals, Gina Solari, Johannie Rivera-Picart, Michelle Pagan, Sunilbe Siceron.

489 **8 Supplementary Figures**



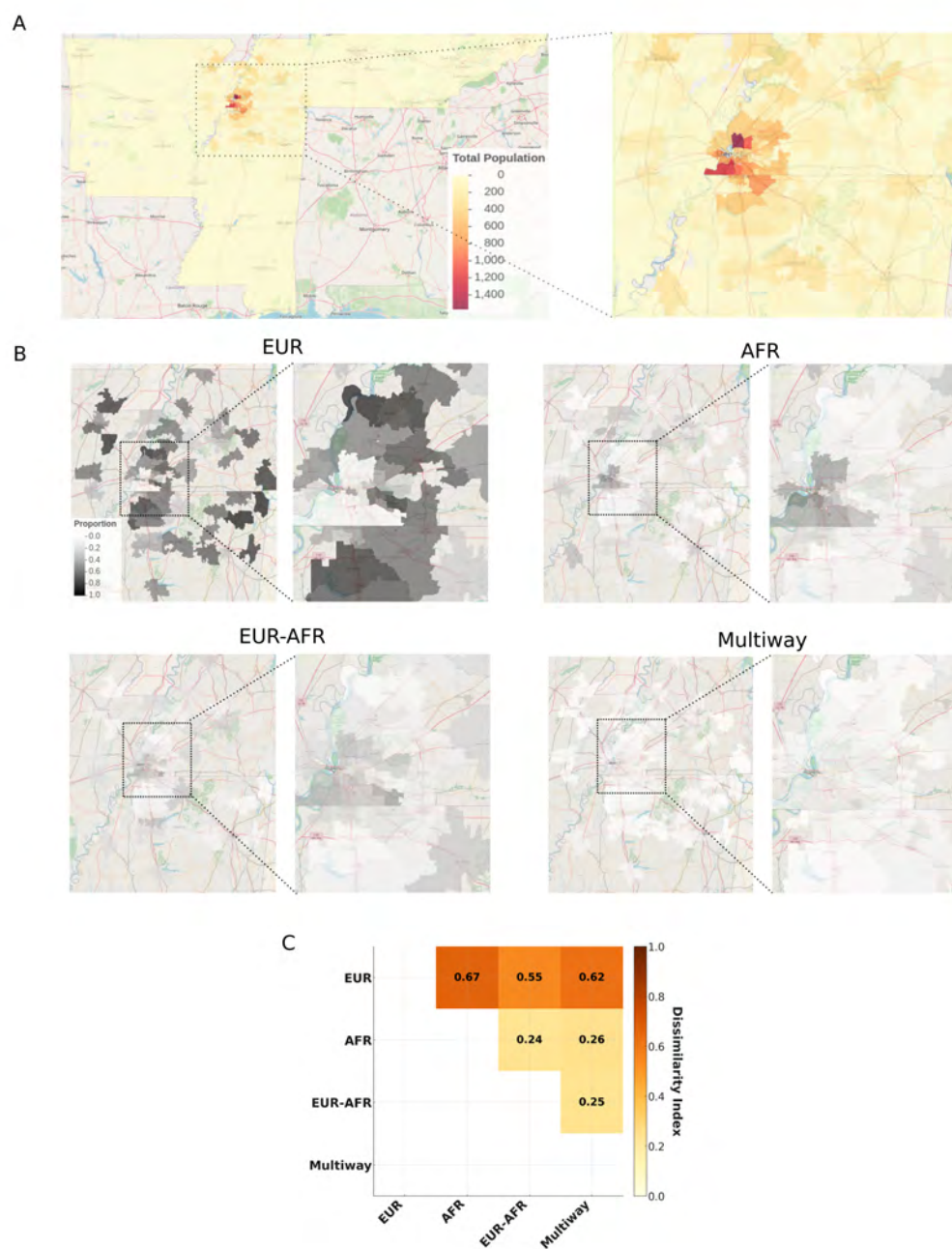


Figure S3: Demography of enrolled participants. (A) Number of enrolled participants by ZIP code. The region surrounding Memphis City is zoomed in. (B) Proportion of individuals by zip code for inferred ancestries with more than 500 individuals. (C) Pairwise Dissimilarity index between ancestries, considering the proportion in each zip code. The dissimilarity index measures the extent of segregation between two groups across geographic areas, indicating the proportion of one group that would need to relocate to achieve an even distribution relative to the other group, with values ranging from 0 (perfect integration) to 1 (complete segregation).

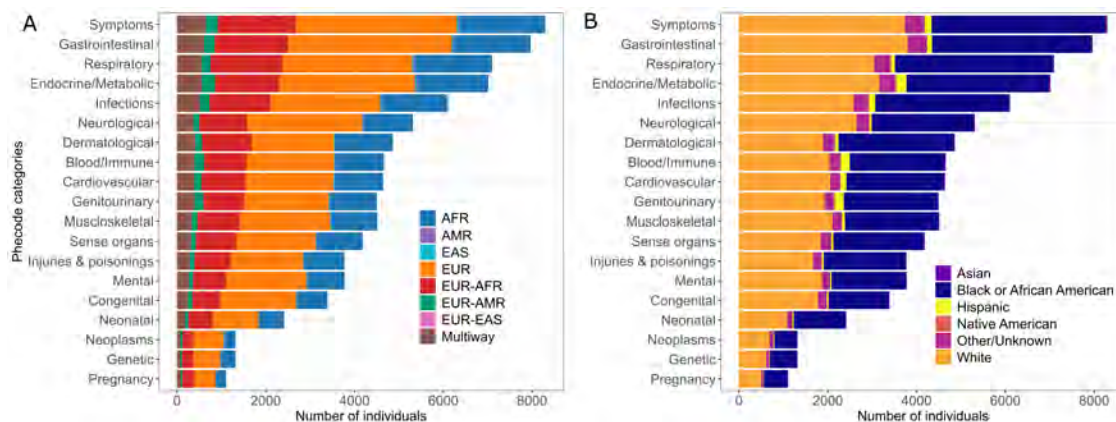


Figure S4: **Phenotypes prevalence in electronic health records in participants with sequence data** stratified by inferred ancestry (A) and self-reported race (B). Phenotypes are grouped into Phecode categories. Distribution of ancestries in Phecode categories reflect the global distribution of ancestry

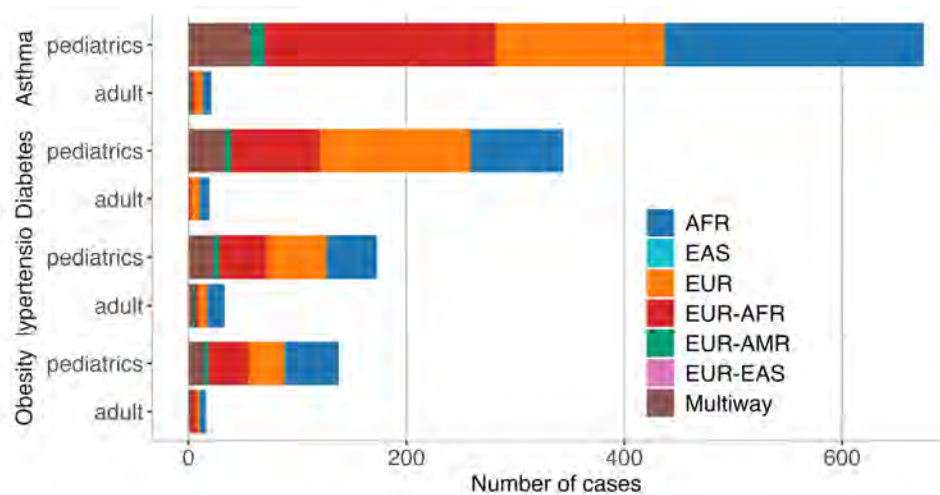


Figure S5: **Count of cases across Asthma, Diabetes, Hypertension, and Obesity.** Cases are categorized by pediatric and adult populations and color-coded by inferred ancestry groups: AFR (African), EAS (East Asian), EUR (European), EUR-AFR (European-African), EUR-AMR (European-American), EUR-EAS (European-East Asian), and Multiway.

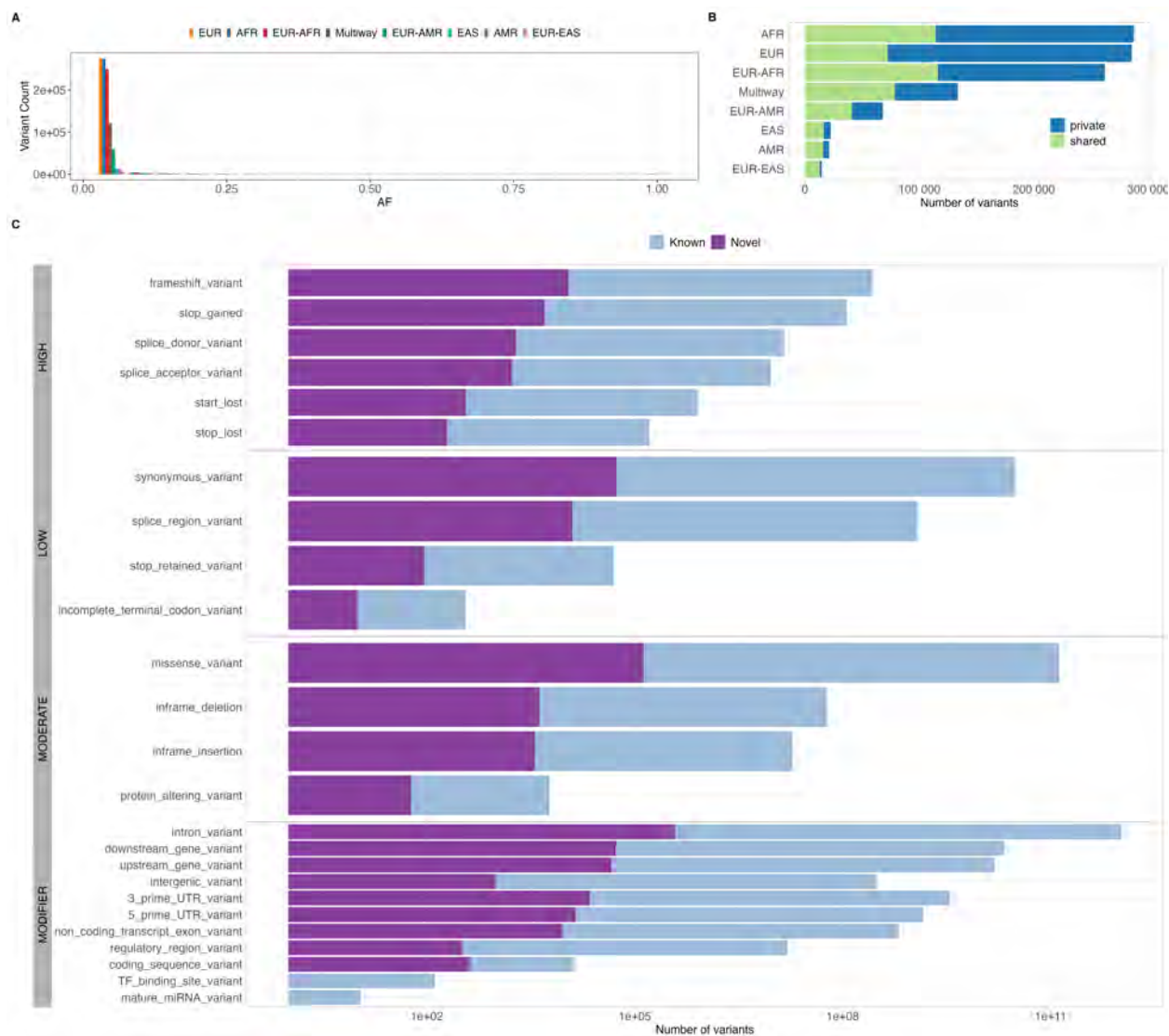


Figure S6: Features of novel variants. (A) Allele frequency spectrum showing the prevalence of rare variants. (B) Counts of variants by ancestry stratified by private and shared with another one or more ancestries. (C) Counts of variants (log scale) by annotated consequences for novel and known variants.

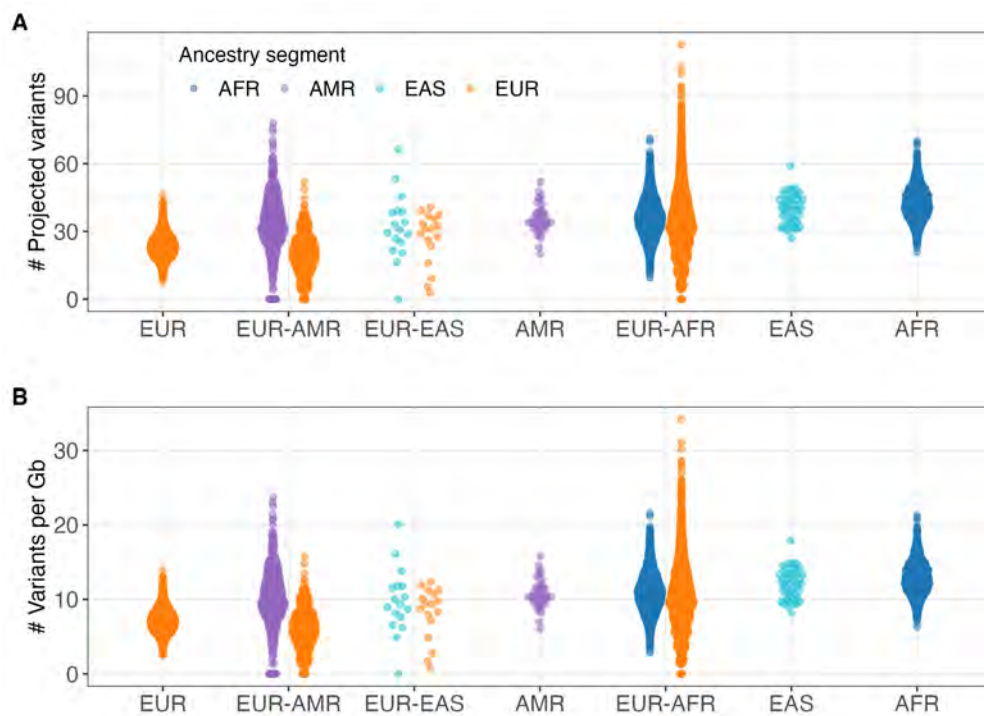


Figure S7: **Counts per individual of rare deleterious variant by ancestry.** Rare deleterious variants are defined as having alternate allele frequency $<1\%$ in the total BIG samples, and classified as high impact or missense with $SIFT < 0.05$ and $Polyphen > 0.85$. Variants counts take into account the inferred ancestry of the genomic tract in which they are located, therefore individuals in admixed groups individuals are represented twice. In panel (A) counts per ancestry tract are normalized by the proportion of ancestry and therefore the y-axis represent the projection as the ancestry tract was as long as the whole genome. In panel (B) we report counts per Gb.

490 **9 Supplementary Tables**

Table S1: Examples of large pediatric cohorts. Although the list is not exhaustive, it is intended to provide context for understanding BIG’s position in terms of size, diversity, and data availability.

Reported Ancestry Representation	Cohort Name	Size	Start Date	Reported Ancestry	Study Group Type	EHR Availability	Genetic Data
Predominantly one ancestry	Avon Longitudinal Study of Parents and Children (ALSPAC) [41]	14,000 children	1991	Predominantly European descent, reflecting the population of the Avon area in the UK	Mother-Child	Has EHR	Has genetic data
	Copenhagen Prospective Studies on Asthma in Childhood (COPSAC) [87]	700 children	1998	Primarily Danish, reflecting the population of Denmark	Children Only	No EHR	Has genetic data
	The Norwegian Mother and Child Cohort Study (MoBa) [42]	114,500 children	1999	Predominantly Norwegian, reflecting the population of Norway	Mother-Child	Has EHR	Has genetic data
	Longitudinal Study of Australian Children (LSAC) [88]	10,000 children	2004	Predominantly Australian, with representation from various ethnic backgrounds	Children Only	No EHR	No genetic data
	All Our Families (AOF) Cohort [43]	3,000 families	2008	Primarily of European descent, reflecting the population of Calgary, Canada	Mother-Child	Has EHR	No genetic data
Diverse ancestries	Children of Philadelphia (CHOP)	100,000 children	2006	Diverse, reflecting the population of Philadelphia	Mother-Child	Yes	Has genetic data
	Childhood Cancer Survivor Study (CCSS) [34]	24,000 survivors	1994	Diverse, reflecting the population of North America	Children Only	Has EHR	Has genetic data
	The Boston Birth Cohort [35]	8,000 births	1998	Predominantly African American and Hispanic participants	Mother-Child	Has EHR	Has genetic data
	Generation R Study [36]	10,000 children	2002	Multi-ethnic urban population, including Dutch, Surinamese, Turkish, Moroccan, and others	Mother-Child	Has EHR	Has genetic data
	Pediatric Imaging, Neurocognition, and Genetics (PING) Study [37]	1,400 children	2009	Diverse, including African American, Asian, Hispanic, and non-Hispanic White participants	Children Only	No EHR	Has genetic data
	NICHD Fetal Growth Studies [38]	2,400 pregnancies	2009	Diverse, including African American, Asian, Hispanic, and non-Hispanic White participants	Mother-Child	Has EHR	Has genetic data
	Biorepository for Integrative Genomics (BIG) [33]	42,000	-	Diverse, including African American, Asian, Hispanic, and non-Hispanic White participants	Children Only	Has EHR	Has genetic data
	Healthy Brain Network (HBN) [39]	10,000 children	2015	Diverse, with efforts to include underrepresented populations	Children Only	No EHR	Has genetic data
	Environmental Influences on Child Health Outcomes (ECHO) [40]	50,000 children	2016	Diverse, with efforts to include underrepresented populations	Mother-Child	Has EHR	Has genetic data

Table S2: **Simplification of self-reported race entries in electronic health records.** The purpose of grouping is to simplify the analyses and eliminate the use of inaccurate or inappropriate terminology [89].

Original Category	Grouped Category
White or Caucasian Caucasian White	White
Black or African American African American	Black or African American
Asian Asian or Pacific Islander	Asian
Other/Unknown Other Declined Patient Declined to answer Unavailable Multiple	Other/Unknown

Table S3: **Prevalence of high impact variants among novel variants.** Estimates from the logistic regression.

Term	Estimate	Std. error	Statistic	p.value	conf.low
Intercept	-2.09	0.001	-1430	< 2e - 16	-2.10
HIGH	0.95	0.008	116	< 2e - 16	0.93
LOW	-0.118	0.004	-30.3	< 2e - 16	-0.125
MODERATE	0.12	0.003	38.8	< 2e - 16	0.118

491 **References**

- 492 [1] David Botstein, Raymond L White, Mark Skolnick, and Ronald W Davis. Construction of
493 a genetic linkage map in man using restriction fragment length polymorphisms. *American*
494 *journal of human genetics*, 32(3):314, 1980.
- 495 [2] Neil Risch and Kathleen Merikangas. The future of genetic studies of complex human dis-
496 eases. *Science*, 273(5281):1516–1517, 1996.
- 497 [3] Leena Peltonen, Anu Jalanko, and Teppo Varilo. Molecular genetics the finnish disease her-
498 itage. *Human molecular genetics*, 8(10):1913–1923, 1999.
- 499 [4] Jeff Gulcher, Agnar Helgason, and Kári Stefánsson. Genetic homogeneity of icelanders.
500 *Nature Genetics*, 26(4):395–395, 2000.
- 501 [5] Yali Xue, Massimo Mezzavilla, Marc Haber, Shane McCarthy, Yuan Chen, Vagheesh
502 Narasimhan, Arthur Gilly, Qasim Ayub, Vincenza Colonna, Lorraine Southam, et al. Enrich-
503 ment of low-frequency functional variants revealed by whole-genome sequencing of multiple
504 isolated european populations. *Nature communications*, 8(1):15927, 2017.
- 505 [6] Kalliope Panoutsopoulou, Konstantinos Hatzikotoulas, Dionysia Kiara Xifara, Vincenza
506 Colonna, Aliko-Eleni Farmaki, Graham RS Ritchie, Lorraine Southam, Arthur Gilly, Ioanna
507 Tachmazidou, Segun Fatumo, et al. Genetic characterization of greek population isolates re-
508 veals strong genetic drift at missense and trait-associated variants. *Nature communications*,
509 5(1):5345, 2014.
- 510 [7] Dan L Nicolae, Xiaoquan Wen, Benjamin F Voight, and Nancy J Cox. Coverage and charac-
511 teristics of the affymetrix genechip human mapping 100k snp set. *PLoS Genetics*, 2(5):e67,
512 2006.
- 513 [8] Lon R Cardon and Goncalo R Abecasis. Using haplotype blocks to map human complex trait
514 loci. *TRENDS in Genetics*, 19(3):135–140, 2003.
- 515 [9] DM Altshuler, RA Gibbs, L Peltonen, DM Altshuler, RA Gibbs, L Peltonen, et al. Interna-

- 516 tional hapmap 3 consortium: Integrating common and rare genetic variation in diverse human
517 populations. *Nature*, 467:52, 2010.
- 518 [10] Guanjie Chen, Daniel Shriner, Jie Zhou, Ayo Doumatey, Hanxia Huang, Norman P Gerry,
519 Alan Herbert, Michael F Christman, Yuanxiu Chen, Georgia M Dunston, et al. Development
520 of admixture mapping panels for african americans from commercial high-density snp arrays.
521 *BMC genomics*, 11:1–12, 2010.
- 522 [11] Arti Tandon, Nick Patterson, and David Reich. Ancestry informative marker panels for
523 african americans based on subsets of commercially available snp arrays. *Genetic epidemi-*
524 *ology*, 35(1):80–83, 2011.
- 525 [12] Adam Auton, Gonçalo R Abecasis, David M Altshuler, Richard M Durbin, David R Bentley,
526 Aravinda Chakravarti, Andrew G Clark, Peter Donnelly, Evan E Eichler, Paul Flicek,
527 Stacey B Gabriel, Richard A Gibbs, Eric D Green, Matthew E Hurles, and Gil McVean. A
528 global reference for human genetic variation. *Nature*, 526:68–74, 2015.
- 529 [13] Anders Bergström, Shane A McCarthy, Ruoyun Hui, Mohamed A Almarri, Qasim Ayub,
530 Petr Danecek, Yuan Chen, Sabine Felkel, Pille Hallast, Jack Kamm, et al. Insights
531 into human genetic variation and population history from 929 diverse genomes. *Science*,
532 367(6484):eaay5012, 2020.
- 533 [14] H3Africa Consortium et al. Enabling the genomic revolution in africa: H3africa is de-
534 veloping capacity for health-related genomics research in africa. *Science (New York, NY)*,
535 344(6190):1346, 2014.
- 536 [15] Swapan Mallick, Heng Li, Mark Lipson, Iain Mathieson, Melissa Gymrek, Fernando Racimo,
537 Mengyao Zhao, Niru Chennagiri, Susanne Nordenfelt, Arti Tandon, et al. The simons genome
538 diversity project: 300 genomes from 142 diverse populations. *Nature*, 538(7624):201–206,
539 2016.
- 540 [16] Mayo Blegen Ashley L. 18 Wirkus Samantha J. 18 Wagner Victoria A. 18 Meyer Jeffrey G.
541 18 Cicek Mine S. 10 18 Biobank and All of Us Research Demonstration Project Teams Choi
542 Seung Hoan 14 <http://orcid.org/0000-0002-0322-8970> Wang Xin 14 <http://orcid.org/0000>

- 543 0001-6042-4487 Rosenthal Elisabeth A. 15. Genomic data in the all of us research program.
544 *Nature*, 627(8003):340–346, 2024.
- 545 [17] Wen-Wei Liao, Mobin Asri, Jana Ebler, Daniel Doerr, Marina Haukness, Glenn Hickey,
546 Shuangjia Lu, Julian K Lucas, Jean Monlong, Haley J Abel, et al. A draft human pangenome
547 reference. *Nature*, 617(7960):312–324, 2023.
- 548 [18] Erik Garrison, Andrea Guarracino, Simon Heumos, Flavia Villani, Zhigui Bao, Lorenzo
549 Tattini, Jörg Hagmann, Sebastian Vorbrugg, Santiago Marco-Sola, Christian Kubica, et al.
550 Building pangenome graphs. *Nature Methods*, pages 1–5, 2024.
- 551 [19] Christina V. Van Hout, Ioanna Tachmazidou, Joshua D. Backman, J. Scott Hoffman, Di Liu,
552 Ashutosh K. Pandey, Claudia Gonzaga-Jauregui, Shaista Khalid, Bingshan Ye, Niranjana
553 Banerjee, Matthew R. Nelson, and Gonçalo R. Abecasis. Whole exome sequencing and
554 characterization of coding variation in 49,960 individuals in the uk biobank. *Nature Commu-*
555 *nications*, 11(1):1–11, Jul 2020.
- 556 [20] Christina C. Kyriazis, Wei Wei, Jemma Danesh, Danish Saleheen, Roland D. Schmid, and
557 Emanuele Di Angelantonio. Human genetic diversity and disease: from outside africa to
558 within europe. *Communications Biology*, 6:353, 2023.
- 559 [21] Pardis C. Sabeti and David Reich. Genetic and archeological evidence for early human pop-
560 ulation structure. *Cell*, 179(6):1462–1474, 2019.
- 561 [22] Giorgio Sirugo, Scott M Williams, and Sarah A Tishkoff. The missing diversity in human
562 genetic studies. *Cell*, 177(1):26–31, 2019.
- 563 [23] Segun Fatumo, Tinashe Chikowore, Ananyo Choudhury, Muhammad Ayub, Alicia R Martin,
564 and Karoline Kuchenbaecker. A roadmap to increase diversity in genomic studies. *Nature*
565 *medicine*, 28(2):243–250, 2022.
- 566 [24] Sonia Moreno-Grau, Manvi Vernekar, Arturo Lopez-Pineda, Daniel Mas-Montserrat, Míriam
567 Barrabés, Consuelo D Quinto-Cortés, Babak Moatamed, Ming Ta Michael Lee, Zhenning
568 Yu, Kensuke Numakura, et al. Polygenic risk score portability for common diseases across
569 genetically diverse populations. *Human Genomics*, 18(1):93, 2024.

- 570 [25] Ola AlAzzeh and Youssef M Roman. The frequency of rs2231142 in *abcg2* among native
571 hawaiian and pacific islander subgroups: implications for personalized rosuvastatin dosing.
572 *Pharmacogenomics*, 24(3):173–182, 2023.
- 573 [26] David Twesigomwe, Britt I Drögemöller, Galen EB Wright, Clement Adebamowo, Godfred
574 Agongo, Palwendé R Boua, Mogomotsi Matshaba, Maria Paximadis, Michèle Ramsay, Gus-
575 tave Simo, et al. Characterization of *cyp2d6* pharmacogenetic variation in sub-saharan african
576 populations. *Clinical Pharmacology & Therapeutics*, 113(3):643–659, 2023.
- 577 [27] Michael A McQuillan, Chao Zhang, Sarah A Tishkoff, and Alexander Platt. The importance
578 of including ethnically diverse populations in studies of quantitative trait evolution. *Current*
579 *opinion in genetics & development*, 62:30–35, 2020.
- 580 [28] Edra K Ha, Daniel Shriner, Shawneequa L Callier, Lorinda Riley, Adebowale A Adeyemo,
581 Charles N Rotimi, and Amy R Bentley. Native hawaiian and pacific islander populations in
582 genomic research. *NPJ Genomic Medicine*, 9(1):45, 2024.
- 583 [29] Mashaal Sohail, María J Palma-Martínez, Amanda Y Chong, Consuelo D Quinto-
584 Cortés, Carmina Barberena-Jonas, Santiago G Medina-Muñoz, Aaron Ragsdale, Guadalupe
585 Delgado-Sánchez, Luis Pablo Cruz-Hervert, Leticia Ferreyra-Reyes, et al. Mexican biobank
586 advances population and medical genomics of diverse ancestries. *Nature*, 622(7984):775–
587 783, 2023.
- 588 [30] Luisa Pereira, Leon Mutesa, Paulina Tindana, and Michèle Ramsay. African genetic diversity
589 and adaptation inform a precision medicine agenda. *Nature Reviews Genetics*, 22(5):284–
590 306, 2021.
- 591 [31] Richard Crespo, Matthew Christiansen, Kim Tieman, and Richard Wittberg. An emerging
592 model for community health worker–based chronic care management for patients with high
593 health care costs in rural appalachia. *Preventing chronic disease*, 17:E13, 2020.
- 594 [32] Kate Beatty, Olivia Egen, John Dreyzehner, and Randy Wykoff. Poverty and health in ten-
595 nessee. *South Med J*, 113(1):1–7, 2020.

- 596 [33] Rony Jose, Robert Rooney, Naga Nagisetty, Robert Davis, and David Hains. Biorepository
597 and integrative genomics initiative: Designing and implementing a preliminary platform for
598 predictive, preventive and personalized medicine at a pediatric hospital in a historically dis-
599 advantaged community in the usa. *EPMA Journal*, 9:225–234, 2018.
- 600 [34] Leslie L Robison, Gregory T Armstrong, John D Boice, Eric J Chow, Stella M Davies,
601 Sarah S Donaldson, Daniel M Green, Sue Hammond, Anna T Meadows, Ann C Mertens,
602 et al. The childhood cancer survivor study: a national cancer institute–supported resource for
603 outcome and intervention research. *Journal of clinical oncology*, 27(14):2308–2318, 2009.
- 604 [35] Colleen Pearson, Tami Bartell, Guoying Wang, Xiumei Hong, Serena A Rusk, LingLing Fu,
605 Sandra Cerda, Blandine Bustamante-Helfrich, Wendy Kuohung, Christina Yarrington, et al.
606 Boston birth cohort profile: rationale and study design. *Precision nutrition*, 1(2):e00011,
607 2022.
- 608 [36] Marjolein N Kooijman, Claudia J Kruithof, Cornelia M van Duijn, Liesbeth Duijts, Oscar H
609 Franco, Marinus H van IJzendoorn, Johan C de Jongste, Caroline CW Klaver, Aad van der
610 Lugt, Johan P Mackenbach, et al. The generation r study: design and cohort update 2017.
611 *European journal of epidemiology*, 31:1243–1264, 2016.
- 612 [37] Terry L Jernigan, Timothy T Brown, Donald J Hagler Jr, Natacha Akshoomoff, Hauke
613 Bartsch, Erik Newman, Wesley K Thompson, Cinnamon S Bloss, Sarah S Murray, Nicholas
614 Schork, et al. The pediatric imaging, neurocognition, and genetics (ping) data repository.
615 *Neuroimage*, 124:1149–1154, 2016.
- 616 [38] Germaine M Buck Louis, Jagtshwar Grewal, Paul S Albert, Anthony Sciscione, Deborah A
617 Wing, William A Grobman, Roger B Newman, Ronald Wapner, Mary E D’Alton, Daniel
618 Skupski, et al. Racial/ethnic standards for fetal growth: the nichd fetal growth studies. *Amer-
619 ican journal of obstetrics and gynecology*, 213(4):449–e1, 2015.
- 620 [39] Lindsay M Alexander, Jasmine Escalera, Lei Ai, Charissa Andreotti, Karina Febre, Alexander
621 Mangone, Natan Vega-Potler, Nicolas Langer, Alexis Alexander, Meagan Kovacs, et al. An
622 open resource for transdiagnostic research in pediatric mental health and learning disorders.
623 *Scientific data*, 4(1):1–26, 2017.

- 624 [40] Christina H Park, Carol J Blaisdell, S Sonia Arteaga, Clay Mash, Susan Laessig, Manjit
625 Hanspal, Erin Luetkemeier, Leslie C Thompson, and Matthew W Gillman. How the environ-
626 mental influences on child health outcome (echo) cohort can spur discoveries in environmen-
627 tal epidemiology. *American Journal of Epidemiology*, page kwae073, 2024.
- 628 [41] Deborah A Lawlor, Melanie Lewcock, Louise Rena-Jones, Claire Rollings, Vikki Yip, Daniel
629 Smith, Rebecca M Pearson, Laura Johnson, Louise AC Millard, Nashita Patel, et al. The
630 second generation of the avon longitudinal study of parents and children (alspac-g2): a cohort
631 profile. *Wellcome Open Research*, 4, 2019.
- 632 [42] Per Magnus, Lorentz M Irgens, Kjell Haug, Wenche Nystad, Rolv Skjærven, and Camilla
633 Stoltenberg. Cohort profile: the norwegian mother and child cohort study (moba). *Internation-
634 al journal of epidemiology*, 35(5):1146–1150, 2006.
- 635 [43] Suzanne C Tough, Sheila W McDonald, Beverly Anne Collisson, Susan A Graham, Heather
636 Kehler, Dawn Kingston, and Karen Benzies. Cohort profile: the all our babies pregnancy
637 cohort (aob). *International Journal of Epidemiology*, 46(5):1389–1390k, 2017.
- 638 [44] Brian K Maples, Simon Gravel, Eimear E Kenny, and Carlos D Bustamante. Rfmix: a dis-
639 criminative modeling approach for rapid and robust local-ancestry inference. *The American
640 Journal of Human Genetics*, 93(2):278–288, 2013.
- 641 [45] Simon Gravel. Population genetics models of local ancestry. *Genetics*, 191(2):607–619,
642 2012.
- 643 [46] Otis Dudley Duncan and Beverly Duncan. A methodological analysis of segregation indexes.
644 *American sociological review*, 20(2):210–217, 1955.
- 645 [47] Lisa Bastarache. Using phecodes for research with the electronic health record: from phewas
646 to phers. *Annual review of biomedical data science*, 4(1):1–19, 2021.
- 647 [48] Christine Cole Johnson, Aruna Chandran, Suzanne Havstad, Xiuhong Li, Cynthia T McEvoy,
648 Dennis R Ownby, Augusto A Litonjua, Margaret R Karagas, Carlos A Camargo, James E
649 Gern, et al. Us childhood asthma incidence rate patterns from the echo consortium to identify
650 high-risk groups for primary prevention. *JAMA pediatrics*, 175(9):919–927, 2021.

- 651 [49] Konrad J Karczewski, Laurent C Francioli, Grace Tiao, Beryl B Cummings, Jessica Alföldi,
652 Qingbo Wang, Ryan L Collins, Kristen M Laricchia, Andrea Ganna, Daniel P Birnbaum, et al.
653 The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature*,
654 581(7809):434–443, 2020.
- 655 [50] William McLaren, Laurent Gil, Sarah E Hunt, Harpreet Singh Riat, Graham RS Ritchie,
656 Anja Thormann, Paul Flicek, and Fiona Cunningham. The ensembl variant effect predictor.
657 *Genome biology*, 17:1–14, 2016.
- 658 [51] Robert Vaser, Swarnaseetha Adusumalli, Sim Ngak Leng, Mile Sikic, and Pauline C Ng. Sift
659 missense predictions for genomes. *Nature protocols*, 11(1):1–9, 2016.
- 660 [52] Ivan Adzhubei, Daniel M Jordan, and Shamil R Sunyaev. Predicting functional effect of
661 human missense mutations using polyphen-2. *Current protocols in human genetics*, 76(1):7–
662 20, 2013.
- 663 [53] Zachary A Szpiech, Angel CY Mak, Marquitta J White, Donglei Hu, Celeste Eng, Esteban G
664 Burchard, and Ryan D Hernandez. Ancestry-dependent enrichment of deleterious homozy-
665 gotes in runs of homozygosity. *The American Journal of Human Genetics*, 105(4):747–762,
666 2019.
- 667 [54] Marcos Araújo Castro e Silva, Tiago Ferraz, Caina M Couto-Silva, Renan B Lemes, Kelly
668 Nunes, David Comas, and Tábita Hünemeier. Population histories and genomic diversity of
669 south american natives. *Molecular biology and evolution*, 39(1):msab339, 2022.
- 670 [55] Sara D Niedbalski and Jeffrey C Long. Novel alleles gained during the beringian isolation
671 period. *Scientific Reports*, 12(1):4289, 2022.
- 672 [56] Soheil Baharian, Maxime Barakatt, Christopher R Gignoux, Suyash Shringarpure, Jacob
673 Errington, William J Blot, Carlos D Bustamante, Eimear E Kenny, Scott M Williams,
674 Melinda C Aldrich, et al. The great migration and african-american genomic diversity. *PLoS*
675 *genetics*, 12(5):e1006059, 2016.
- 676 [57] Sharon R Browning, Brian L Browning, Martha L Daviglus, Ramon A Durazo-Arvizu, Neil

- 677 Schneiderman, Robert C Kaplan, and Cathy C Laurie. Ancestry-specific recent effective
678 population size in the americas. *PLoS genetics*, 14(5):e1007385, 2018.
- 679 [58] Kirsten Voorhies, Akram Mohammed, Lokesh Chinthala, Sek Won Kong, In-Hee Lee,
680 Alvin T Kho, Michael McGeachie, Kenneth D Mandl, Benjamin Raby, Melanie Hayes,
681 et al. Gsdmb/ormdl3 rare/common variants are associated with inhaled corticosteroid re-
682 sponse among children with asthma. *Genes*, 15(4):420, 2024.
- 683 [59] Tesfaye B Mersha and Tilahun Abebe. Self-reported race/ethnicity in the age of genomic
684 research: its potential impact on understanding health disparities. *Human genomics*, 9(1):1,
685 2015.
- 686 [60] Esteban González Burchard, Elad Ziv, Natasha Coyle, Scarlett Lin Gomez, Hua Tang, An-
687 drew J Karter, Joanna L Mountain, Eliseo J Pérez-Stable, Dean Sheppard, and Neil Risch.
688 The importance of race and ethnic background in biomedical research and clinical practice,
689 2003.
- 690 [61] Richard S Cooper. Race and genomics. *The New England journal of medicine*, 348(12):1166,
691 2003.
- 692 [62] Kellee White, Jourdyn A Lawrence, Nedelina Tchangalova, Shuo J Huang, and Jason L Cum-
693 mings. Socially-assigned race and health: a scoping review with global implications for
694 population health equity. *International journal for equity in health*, 19:1–14, 2020.
- 695 [63] Daphne O Martschenko, Hannah Wand, Jennifer L Young, and Genevieve L Wojcik. Includ-
696 ing multiracial individuals is crucial for race, ethnicity and ancestry frameworks in genetics
697 and genomics. *Nature genetics*, 55(6):895–900, 2023.
- 698 [64] Giorgio Sirugo, Sarah A Tishkoff, Scott M Williams, et al. The quagmire of race, genetic
699 ancestry, and health disparities. *The Journal of clinical investigation*, 131(11), 2021.
- 700 [65] Iman Hamid, Katharine L Korunes, Sandra Beleza, and Amy Goldberg. Rapid adaptation to
701 malaria facilitated by admixture in the human population of cabo verde. *Elife*, 10:e63177,
702 2021.

- 703 [66] Iman Hamid, Katharine L Korunes, Daniel R Schrider, and Amy Goldberg. Localizing post-
704 admixture adaptive variants with object detection on ancestry-painted chromosomes. *Molec-*
705 *ular Biology and Evolution*, 40(4):msad074, 2023.
- 706 [67] Meng Lin, Danny S Park, Noah A Zaitlen, Brenna M Henn, and Christopher R Gignoux.
707 Admixed populations improve power for variant discovery and portability in genome-wide
708 association studies. *Frontiers in genetics*, 12:673167, 2021.
- 709 [68] Nick Patterson, Neil Hattangadi, Barton Lane, Kirk E Lohmueller, David A Hafler, Jorge R
710 Oksenberg, Stephen L Hauser, Michael W Smith, Stephen J O’Brien, David Altshuler, et al.
711 Methods for high-density admixture mapping of disease genes. *The American Journal of*
712 *Human Genetics*, 74(5):979–1000, 2004.
- 713 [69] Eva Suarez-Pajes, Ana Díaz-de Usera, Itahisa Marcelino-Rodríguez, Beatriz Guillen-Guio,
714 and Carlos Flores. Genetic ancestry inference and its application for the genetic mapping of
715 human diseases. *International journal of molecular sciences*, 22(13):6962, 2021.
- 716 [70] Michael W Smith, Nick Patterson, James A Lautenberger, Ann L Truelove, Gavin J McDon-
717 ald, Alicja Waliszewska, Bailey D Kessing, Michael J Malasky, Charles Scafe, Ernest Le,
718 et al. A high-density admixture map for disease gene discovery in african americans. *The*
719 *American Journal of Human Genetics*, 74(5):1001–1013, 2004.
- 720 [71] Andréa RVR Horimoto, Diane Xue, Timothy A Thornton, and Elizabeth E Blue. Admixture
721 mapping reveals the association between native american ancestry at 3q13. 11 and reduced
722 risk of alzheimer’s disease in caribbean hispanics. *Alzheimer’s Research & Therapy*, 13:1–14,
723 2021.
- 724 [72] Heng Li and Richard Durbin. Fast and accurate short read alignment with burrows–wheeler
725 transform. *bioinformatics*, 25(14):1754–1760, 2009.
- 726 [73] Artem Tarasov, Albert J Vilella, Edwin Cuppen, Isaac J Nijman, and Pjotr Prins. Sambamba:
727 fast processing of ngs alignment formats. *Bioinformatics*, 31(12):2032–2034, 2015.
- 728 [74] Ryan Poplin, Pi-Chuan Chang, David Alexander, Scott Schwartz, Thomas Colthurst, Alexan-
729 der Ku, Dan Newburger, Jojo Dijamco, Nam Nguyen, Pegah T Afshar, et al. A universal snp

- 730 and small-indel variant caller using deep neural networks. *Nature biotechnology*, 36(10):983–
731 987, 2018.
- 732 [75] Michael F Lin, Ohad Rodeh, John Penn, Xiaodong Bai, Jeffrey G Reid, Olga Krasheninina,
733 and William J Salerno. Glnexus: joint variant calling for large cohort sequencing. *BioRxiv*,
734 page 343970, 2018.
- 735 [76] Olivier Delaneau, Bryan Howie, Anthony J Cox, Jean-François Zagury, and Jonathan Mar-
736 chini. Haplotype estimation using sequencing reads. *The American Journal of Human Ge-*
737 *netics*, 93(4):687–696, 2013.
- 738 [77] Zan Koenig, Mary T Yohannes, Lethukuthula L Nkambule, Xuefang Zhao, Julia K Goodrich,
739 Heesu Ally Kim, Michael W Wilson, Grace Tiao, Stephanie P Hao, Nareh Sahakian, et al. A
740 harmonized public resource of deeply sequenced diverse human genomes. *Genome Research*,
741 2024.
- 742 [78] Alex Mas-Sandoval, Sara Mathieson, and Matteo Fumagalli. The genomic footprint of social
743 stratification in admixing american populations. *Elife*, 12:e84429, 2023.
- 744 [79] David H Alexander and Kenneth Lange. Enhancements to the admixture algorithm for indi-
745 vidual ancestry estimation. *BMC bioinformatics*, 12:1–6, 2011.
- 746 [80] Anna CF Lewis, Santiago J Molina, Paul S Appelbaum, Bege Dauda, Anna Di Rienzo,
747 Agustin Fuentes, Stephanie M Fullerton, Nanibaa’A Garrison, Nayanika Ghosh, Eve-
748 lynn M Hammonds, et al. Getting genetic ancestry right for science and society. *Science*,
749 376(6590):250–252, 2022.
- 750 [81] Ethnicity Committee on the Use of Race, Engineering National Academies of Sciences, and
751 Medicine. Using population descriptors in genetics and genomics research: A new framework
752 for an evolving field. *NATIONAL ACADEMIES PRESS*, 2023.
- 753 [82] Helena Machado and Rafaela Granja. Emerging dna technologies and stigmatization. *Foren-*
754 *sic genetics in the governance of crime*, pages 85–104, 2020.

- 755 [83] Florian Privé, Hugues Aschard, Andrey Ziyatdinov, and Michael G.B. Blum. Efficient anal-
756 ysis of large-scale genome-wide data with two R packages: bigstatsr and bigsnpr. *Bioinfor-*
757 *matics*, 34(16):2781–2787, 2018.
- 758 [84] Ani Manichaikul, Josyf C Mychaleckyj, Stephen S Rich, Kathy Daly, Michèle Sale, and Wei-
759 Min Chen. Robust relationship inference in genome-wide association studies. *Bioinformatics*,
760 26(22):2867–2873, 2010.
- 761 [85] Ying Zhou, Sharon R Browning, and Brian L Browning. A fast and simple method for
762 detecting identity-by-descent segments in large-scale data. *The American Journal of Human*
763 *Genetics*, 106(4):426–437, 2020.
- 764 [86] Brian L Browning and Sharon R Browning. Improving the accuracy and efficiency of
765 identity-by-descent detection in population data. *Genetics*, 194(2):459–471, 2013.
- 766 [87] Hans Bisgaard. The copenhagen prospective study on asthma in childhood (copsac): design,
767 rationale, and baseline data from a longitudinal birth cohort study. *Annals of Allergy, Asthma*
768 *& Immunology*, 93(4):381–389, 2004.
- 769 [88] Matthew Gray and Diana Smart. Growing up in australia: the longitudinal study of aus-
770 tralian children: a valuable new data source for economists. *Australian Economic Review*,
771 42(3):367–376, 2009.
- 772 [89] Alice B Popejoy. Too many scientists still say caucasian. *Nature*, 596(7873):463–463, 2021.