




Draft Genome Assemblies of Ionic Liquid-Resistant *Yarrowia lipolytica* PO1f and Its Superior Evolved Strain, YICW001

Caleb Walker,^a Seunghyun Ryu,^{a*} Sajeet Haridas,^b Hyunsoo Na,^b Matthew Zane,^b Kurt LaButti,^b Kerrie Barry,^b Igor V. Grigoriev,^b  Cong T. Trinh^a

^aDepartment of Chemical and Biomolecular Engineering, The University of Tennessee, Knoxville, Tennessee, USA

^bU.S. Department of Energy Joint Genome Institute, Walnut Creek, California, USA

ABSTRACT Adaptive laboratory evolution of *Yarrowia lipolytica* PO1f in the benchmark ionic liquid (IL; 1-ethyl-3-methylimidazolium acetate) produced a superior IL-tolerant microorganism, strain YICW001. Here, we report the genome sequences of PO1f and YICW001 to study the robustness of *Y. lipolytica* and its potential use as a microbial platform for producing fuels and chemicals.

Yarrowia lipolytica PO1f (ATCC MYA-2613), one of the most widely characterized strains of the *Yarrowia* clade (subphylum *Saccharomycotina*), is derivative of the French haploid strain W29 (ATCC 20460), engineered to remove extracellular protease production (1). This generally recognized as safe (GRAS) yeast (2) is regarded as an emerging bioenergy microbe due to its ability to consume complex sugars (3, 4) and produce specialty lipids (5–7), organic acids (8–10), and proteins (1, 11). Additionally, *Y. lipolytica* exhibits exceptional tolerance to high salinity (12), broad pH ranges (13), and ionic liquids (ILs) (14, 15). Microbial biocatalysis in IL is an attractive strategy for the production of high-value chemicals (16) and biofuels (17) because ILs effectively process recalcitrant biomass for fermentation (18–20). While ILs are toxic to most microbes at a concentration of 0.5 to 1.0% (vol/vol) (21, 22), PO1f exhibits robust growth and organic acid production in up to 10% (vol/vol) 1-ethyl-3-methyl imidazolium acetate (EMIM OAc) (14). Aiming to illuminate the underlying mechanisms of IL resistance, we report the genome sequences of the naturally IL-tolerant strain PO1f and its superior evolved mutant, strain YICW001, generated from 200 generations of adaptive laboratory evolution (15). To our knowledge, YICW001 is the most IL-tolerant microorganism, capable of growing in a variety of ILs at high levels, up to 18% IL.

PO1f and YICW001 were cultured in rich medium overnight, and their genomic DNA samples were collected using a Zymo Research (Irvine, CA) fungal/bacterial DNA miniprep kit (catalog no. D6005). Sequencing was performed at the U.S. Department of Energy Joint Genome Institute (DOE JGI) using the Illumina HiSeq 2500 platform as previously described (23). Briefly, 100 ng of DNA was sheared to 500-bp inserts with Covaris LE220 (Woburn, MA) with size selection solid-phase reversible immobilization (SPRI) beads (Beckman Coulter, Brea, CA). The KAPA-Illumina library creation kit (Kapa Biosystems, Boston, MA) was used to treat the resulting fragments with end repair, A tailing, and ligation of Illumina-compatible adapters (IDT, Inc., Skokie, IL). A clustered flow cell was generated using a TruSeq paired-end cluster kit v4 from the multiplexed libraries, and sequencing was performed following a 2 × 100-bp indexed run recipe using the HiSeq TruSeq sequencing by synthesis (SBS) sequencing kits v4. The raw sequence data were filtered using the JGI quality control (QC) pipeline, which integrates BBDuk (v36.94) (<http://bbtools.jgi.doe.gov>) and BBDuk. This pipeline removes contaminants, adapter sequences, right quality reads where quality drops to 0, reads containing 1 or more N bases, reads having an average quality score less than 13, and reads

Citation Walker C, Ryu S, Haridas S, Na H, Zane M, LaButti K, Barry K, Grigoriev IV, Trinh CT. 2020. Draft genome assemblies of ionic liquid-resistant *Yarrowia lipolytica* PO1f and its superior evolved strain, YICW001. Microbiol Resour Announc 9:e01356-19. <https://doi.org/10.1128/MRA.01356-19>.

Editor Jason E. Stajich, University of California, Riverside

This is a work of the U.S. Government and is not subject to copyright protection in the United States. Foreign copyrights may apply. Address correspondence to Cong T. Trinh, ctrinh@utk.edu.

* Present address: Seunghyun Ryu, CJ Research Center America, Woburn, Massachusetts, USA.

Received 7 November 2019

Accepted 2 February 2020

Published 27 February 2020

TABLE 1 Genome information and accession numbers

Strain	GenBank accession no.	SRA accession no.	BioProject accession no.	Genome size (Mbp)	Coverage (×)	No. of contigs	No. of gene models
PO1f	WHUU000000000	SRP115772	PRJNA347220	20.21	155.42	175	6,798
YICW001	WEZX000000000	SRP115773	PRJNA347221	20.22	166.48	150	6,800

containing a minimum length less than or equal to 41 bp or 33% of the full read length. Further, reads that were mapped at 95% identity to masked human, cat, dog, mouse, or common microbial contaminants were removed. Genome assembly on JGI QC-filtered genomic reads was carried out using SPAdes v3.11.1 (24) with the following parameters: `--phred-offset 33 --cov-cutoff auto -t 16 -m 115 --careful --12`. The genomes of PO1f and YICW001 were annotated with transcripts of FKP355 (25) using the JGI annotation pipeline (<https://mycocosm.jgi.doe.gov/programs/fungi/FungalGenomeAnnotationSOP.pdf>) (26, 27). The resultant genome assemblies are just over 20.2 Mbp, containing 49% median GC content and 6,798 to 6,800 gene models (Table 1).

Data availability. The whole-genome assemblies and annotation were deposited at DDBJ/EMBL/GenBank under the accession numbers listed in Table 1. The versions provided in this paper are the first versions.

ACKNOWLEDGMENTS

We acknowledge financial support from the DOE BER Genomic Science Program (DE-SC0019412) and the National Science Foundation (NSF 1511881). The sequencing project was supported by the BioEnergy Science Center (BESC) and the Center for Bioenergy Innovation (CBI), U.S. Department of Energy Bioenergy Research Centers, supported by the Office of Biological and Environmental Research in the DOE Office of Science. The work conducted by the U.S. Department of Energy Joint Genome Institute, a DOE Office of Science User Facility, is supported by the Office of Science of the U.S. Department of Energy under contract no. DE-AC02-05CH11231.

REFERENCES

- Madzak C, Tréton B, Blanchin-Roland S. 2000. Strong hybrid promoters and integrative expression/secretion vectors for quasi-constitutive expression of heterologous proteins in the yeast *Yarrowia lipolytica*. *J Mol Microbiol Biotechnol* 2:207–216.
- Groenewald M, Boekhout T, Neuvéglise C, Gaillardin C, van Dijck PWM, Wyss M. 2014. *Yarrowia lipolytica*: safety assessment of an oleaginous yeast with a great industrial potential. *Crit Rev Microbiol* 40:187–206. <https://doi.org/10.3109/1040841X.2013.770386>.
- Ryu S, Hipp J, Trinh CT. 2016. Activating and elucidating metabolism of complex sugars in *Yarrowia lipolytica*. *Appl Environ Microbiol* 82:1334–1345. <https://doi.org/10.1128/AEM.03582-15>.
- Ryu S, Trinh CT. 2018. Understanding functional roles of native pentose-specific transporters for activating dormant pentose metabolism in *Yarrowia lipolytica*. *Appl Environ Microbiol* 84:e02146-17. <https://doi.org/10.1128/AEM.02146-17>.
- Beopoulos A, Cescut J, Haddouche R, Uribealraea J-L, Molina-Jouve C, Nicaud J-M. 2009. *Yarrowia lipolytica* as a model for bio-oil production. *Prog Lipid Res* 48:375–387. <https://doi.org/10.1016/j.plipres.2009.08.005>.
- Blazek J, Hill A, Liu L, Knight R, Miller J, Pan A, Otoupal P, Alper HS. 2014. Harnessing *Yarrowia lipolytica* lipogenesis to create a platform for lipid and biofuel production. *Nat Commun* 5:3131. <https://doi.org/10.1038/ncomms4131>.
- Qiao K, Wasylenko TM, Zhou K, Xu P, Stephanopoulos G. 2017. Lipid production in *Yarrowia lipolytica* is maximized by engineering cytosolic redox metabolism. *Nat Biotechnol* 35:173–177. <https://doi.org/10.1038/nbt.3763>.
- Gao C, Yang X, Wang H, Rivero CP, Li C, Cui Z, Qi Q, Lin CSK. 2016. Robust succinic acid production from crude glycerol using engineered *Yarrowia lipolytica*. *Biotechnol Biofuels* 9:179. <https://doi.org/10.1186/s13068-016-0597-8>.
- Tan M-J, Chen X, Wang Y-K, Liu G-L, Chi Z-M. 2016. Enhanced citric acid production by a yeast *Yarrowia lipolytica* over-expressing a pyruvate carboxylase gene. *Bioprocess Biosyst Eng* 39:1289–1296. <https://doi.org/10.1007/s00449-016-1607-8>.
- Yovkova V, Otto C, Aurich A, Mauersberger S, Barth G. 2014. Engineering the alpha-ketoglutarate overproduction from raw glycerol by overexpression of the genes encoding NADP⁺-dependent isocitrate dehydrogenase and pyruvate carboxylase in *Yarrowia lipolytica*. *Appl Microbiol Biotechnol* 98:2003–2013. <https://doi.org/10.1007/s00253-013-5369-9>.
- Nicaud J-M, Madzak C, van den Broek P, Gysler C, Duboc P, Niederberger P, Gaillardin C. 2002. Protein expression and secretion in the yeast *Yarrowia lipolytica*. *FEMS Yeast Res* 2:371–379. [https://doi.org/10.1016/S1567-1356\(02\)00082-X](https://doi.org/10.1016/S1567-1356(02)00082-X).
- Andreishcheva EN, Isakova EP, Sidorov NN, Abramova NB, Ushakova NA, Shaposhnikov GL, Soares MI, Zvyagilskaya RA. 1999. Adaptation to salt stress in a salt-tolerant strain of the yeast *Yarrowia lipolytica*. *Biochemistry (Mosc)* 64:1061–1067.
- Epova E, Guseva M, Kovalyov L, Isakova E, Deryabina Y, Belyakova A, Zylkova M, Shevelev A. 2012. Identification of proteins involved in pH adaptation in extremophile yeast *Yarrowia lipolytica*. In Heazlewood JL, Petzold CJ (ed), *Proteomic applications in biology*. IntechOpen, London, United Kingdom. <https://doi.org/10.5772/28791>.
- Ryu S, Labbe N, Trinh CT. 2015. Simultaneous saccharification and fermentation of cellulose in ionic liquid for efficient production of alpha-ketoglutaric acid by *Yarrowia lipolytica*. *Appl Microbiol Biotechnol* 99:4237–4244. <https://doi.org/10.1007/s00253-015-6521-5>.
- Walker C, Ryu S, Trinh CT. 2019. Exceptional solvent tolerance in *Yarrowia lipolytica* is enhanced by sterols. *Metab Eng* 54:83–95. <https://doi.org/10.1016/j.ymben.2019.03.003>.
- van Rantwijk F, Sheldon RA. 2007. Biocatalysis in ionic liquids. *Chem Rev* 107:2757–2785. <https://doi.org/10.1021/cr050946x>.
- Bokinsky G, Peralta-Yahya PP, George A, Holmes BM, Steen EJ, Dietrich J,

- Lee TS, Tullman-Ereck D, Voigt CA, Simmons BA, Keasling JD. 2011. Synthesis of three advanced biofuels from ionic liquid-pretreated switchgrass using engineered *Escherichia coli*. *Proc Natl Acad Sci U S A* 108:19949–19954. <https://doi.org/10.1073/pnas.1106958108>.
18. Li C, Knierim B, Manisseri C, Arora R, Scheller HV, Auer M, Vogel KP, Simmons BA, Singh S. 2010. Comparison of dilute acid and ionic liquid pretreatment of switchgrass: biomass recalcitrance, delignification and enzymatic saccharification. *Bioresour Technol* 101:4900–4906. <https://doi.org/10.1016/j.biortech.2009.10.066>.
19. Yoo CG, Pu Y, Ragauskas AJ. 2017. Ionic liquids: promising green solvents for lignocellulosic biomass utilization. *Curr Opin Green Sustain Chem* 5:5–11. <https://doi.org/10.1016/j.cogsc.2017.03.003>.
20. Socha AM, Parthasarathi R, Shi J, Pattathil S, Whyte D, Bergeron M, George A, Tran K, Stavila V, Venkatachalam S, Hahn MG, Simmons BA, Singh S. 2014. Efficient biomass pretreatment using ionic liquids derived from lignin and hemicellulose. *Proc Natl Acad Sci U S A* 111:E3587–E3595. <https://doi.org/10.1073/pnas.1405685111>.
21. Wu Y-W, Higgins B, Yu C, Reddy AP, Ceballos S, Joh LD, Simmons BA, Singer SW, VanderGheynst JS. 2016. Ionic liquids impact the bioenergy feedstock-degrading microbiome and transcription of enzymes relevant to polysaccharide hydrolysis. *mSystems* 1:e00120-16. <https://doi.org/10.1128/mSystems.00120-16>.
22. Ouellet M, Datta S, Dibble DC, Tamrakar PR, Benke PI, Li C, Singh S, Sale KL, Adams PD, Keasling JD, Simmons BA, Holmes BM, Mukhopadhyay A. 2011. Impact of ionic liquid pretreated plant biomass on *Saccharomyces cerevisiae* growth and biofuel production. *Green Chem* 13:2743–2749. <https://doi.org/10.1039/c1gc15327g>.
23. Walker C, Ryu S, Na H, Zane M, LaButti K, Lipzen A, Haridas S, Barry K, Grigoriev IV, Quarterman J, Slininger P, Dien B, Trinh CT. 2018. Draft genome assemblies of five robust *Yarrowia lipolytica* strains exhibiting high lipid production, pentose sugar utilization, and sugar alcohol secretion from undetoxified lignocellulosic biomass hydrolysates. *Microbiol Resour Anounc* 7:e01040-18. <https://doi.org/10.1128/MRA.01040-18>.
24. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
25. Pomraning KR, Bredeweg EL, Kerkhoven EJ, Barry K, Haridas S, Hundley H, LaButti K, Lipzen A, Yan M, Magnuson JK, Simmons BA, Grigoriev IV, Nielsen J, Baker SE. 2018. Regulation of yeast-to-hyphae transition in *Yarrowia lipolytica*. *mSphere* 3:e00541-18. <https://doi.org/10.1128/mSphere.00541-18>.
26. Grigoriev IV, Martinez DA, Salamov AA. 2006. Fungal genomic annotation, p 123–142. *In* Arora DK, Berka RM, Singh GB (ed), *Applied mycology and biotechnology*, vol 6. Elsevier, Amsterdam, the Netherlands.
27. Grigoriev IV, Nikitin R, Haridas S, Kuo A, Ohm R, Otilar R, Riley R, Salamov A, Zhao X, Korzeniewski F, Smirnova T, Nordberg H, Dubchak I, Shabalov I. 2014. MycoCosm portal: gearing up for 1000 fungal genomes. *Nucleic Acids Res* 42:D699–D704. <https://doi.org/10.1093/nar/gkt1183>.