

γ -Tocotrienol prevents oxidative stress-induced telomere shortening in human fibroblasts derived from different aged individuals

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Key words: γ -tocotrienol, telomere, telomerase, fibroblasts, oxidative stress, aging

Abbreviations: ALT, alternative lengthening of telomeres; CCM, complete culture media; CO₂, carbon dioxide; DNA, deoxyribonucleic acid; DIG, digoxigenin; EMEM, eagle's minimum essential medium; FCS, fetal calf serum; GTT, γ -tocotrienol; HDF, human diploid fibroblast; H₂O₂, hydrogen peroxide; IC₅₀, inhibitory concentration; IMDM, iscove's modified dulbecco's medium; MF, fibroblasts derived from middle age human; MTS, 3-(4,5-dimethylthiazol-2-yl)-5-carboxymethoxyphenyl 2-(4-sulfophenyl)-2H-tetrazolium; OF, fibroblasts derived from old age human; PCR, polymerase chain reaction; PMS, phenazine methosulphate; ROS, reactive oxygen species; SIPS, stress-induced premature senescence; TRAP, telomeric repeat amplification protocol; TRF, terminal restriction fragment; UV, ultra violet; YF, fibroblasts derived from young human

The effects of palm γ -tocotrienol (GGT) on oxidative stress-induced cellular ageing was investigated in normal human skin fibroblast cell lines derived from different age groups; young (21-year-old, YF), middle (40-year-old, MF) and old (68-year-old, OF). Fibroblast cells were treated with γ -tocotrienol for 24 hours before or after incubation with IC₅₀ dose of H₂O₂ for 2 hours. Changes in cell viability, telomere length and telomerase activity were assessed using the MTS assay (Promega, USA), Southern blot analysis and telomere repeat amplification protocol respectively. Results showed that treatment with different concentrations of γ -tocotrienol increased fibroblasts viability with optimum dose of 80 μ M for YF and 40 μ M for both MF and OF. At higher concentrations, γ -tocotrienol treatment caused marked decrease in cell viability with IC₅₀ value of 200 μ M (YF), 300 μ M (MF) and 100 μ M (OF). Exposure to H₂O₂ decreased cell viability in dose dependent manner, shortened telomere length and reduced telomerase activity in all age groups. The IC₅₀ of H₂O₂ was found to be; YF (700 μ M), MF (400 μ M) and OF (100 μ M). Results showed that viability increased significantly ($p < 0.05$) when cells were treated with 80 μ M and 40 μ M γ -tocotrienol prior or after H₂O₂-induced oxidative stress in all age groups. In YF and OF, pretreatment with γ -tocotrienol prevented shortening of telomere length and reduction in telomerase activity. In MF, telomerase activity increased while no changes in telomere length was observed. However, post-treatment of γ -tocotrienol did not exert any significant effects on telomere length and telomerase activity. Thus, these data suggest that γ -tocotrienol protects against oxidative stress-induced cellular ageing by modulating the telomere length possibly via telomerase.

Introduction

Normal human cells exhibit a limited capacity for proliferation in culture and this finite replicative lifespan has frequently been used as a model of human aging in mitotic tissues and organs.¹ This phenomenon is considered to be associated with reduction in telomere length as an indicator of the number of cell divisions undergone. The human telomere is a simple repeating sequence of six bases, TTAGGG, located at the ends of chromosomes.² It is thought that telomeres have multiple roles, including protection against degeneration, reconstruction, fusion and loss.^{3,4} The telomeric repeat sequences are added on by the enzyme telomerase which, when present, compensates for the loss of DNA from the

end of chromosomes due to incomplete replication. In normal human somatic cells, because of inherent limitations in the mechanics of DNA replication, telomeres shorten at each cell division. In the absence of telomerase, when telomere shortening reaches a critical limit, cells are susceptible to chromosomal aberrations such as end-to-end fusion and aneuploidy. In such a situation, the cells cease to divide and reach replicative senescence.^{5,6} Telomere length may restrict the replicative potential of hemopoietic cells contributing to the decline in immune function with age.⁷ The erosion of chromosome ends, or telomeres, was proposed as a major mechanism that contributes to cellular aging.⁸

While convincing evidence has been obtained that telomere shortening occurs during aging both in vitro and in vivo,

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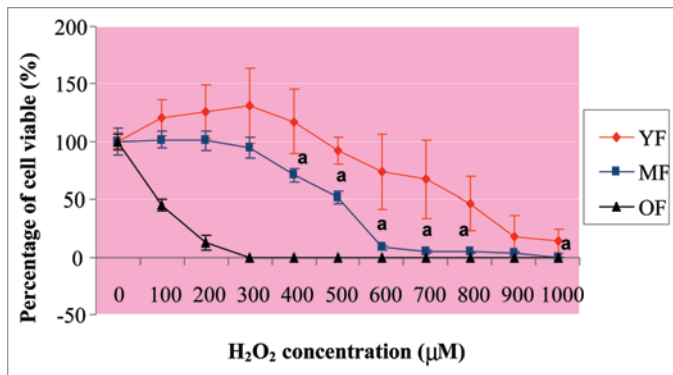


Figure 1. Effects of H₂O₂ toxicity towards viability of fibroblast cell lines derived from young (YF), middle (MF) and old (OF) age humans as assessed by MTS assay. Fibroblast cells were exposed to increasing concentrations of H₂O₂ for 2 hours at 37°C. No significant difference in viable cell number between control and H₂O₂-exposed cultures were observed when the concentration of H₂O₂ was less than 500 μM, 300 μM and 20 μM for fibroblast cells derived from young, middle and old age humans respectively. Cell viability decreased by 50% when exposed to 700 μM, 400 μM and 100 μM of H₂O₂ for fibroblast cells derived from young, middle and old age humans respectively. At more than 1,000 μM, 900 μM and 800 μM for fibroblast cells derived from young, middle and old age humans respectively, almost all cells were dead. The IC₅₀ of H₂O₂ for fibroblast cells derived from young, middle and old age humans were found to be 700 μM, 400 μM and 100 μM respectively. ^aDenotes $p < 0.05$ compared to 0 μM H₂O₂. Data is presented as means \pm SD (n = 3).

additional factors are likely to contribute to aging processes. Thus, it has been hypothesized that oxidative stress is a major determinant of human aging in vivo and that the resistance of animals to oxidative stress may decrease with advancing age.^{9,10} There is increasing evidence that this aspect of the aging process can be reproduced in vitro. For example, it has been shown that increasing the concentration of reactive oxygen species (ROS), either by increasing the oxygen partial pressure, or by treating cells with H₂O₂, can induce premature senescence in young diploid human fibroblasts.¹¹ H₂O₂ is one of the oxidant species and it can be genotoxic. Treatment of human fibroblast with H₂O₂ which induced oxidative damage results in a growth arrest state that is similar to senescence cell.¹²

If free radicals and oxidative stress are at least partly responsible for lifespan and aging, it follows that antioxidant should prolong life and retard aging. The biological activity of vitamin E has generally been associated with its well-defined antioxidant property. Vitamin E is the major chain-breaking antioxidant preventing the propagation of oxidative stress, especially in biological membranes.¹³ Vitamin E also has revealed many important molecular properties, such as the scavenging of ROS to prevent oxidative damage that is associated with many diseases, or the modulation of signal transduction and gene expression in antioxidant and non-antioxidant manners.¹⁴ The other biological functions of vitamin E that are unrelated to its antioxidant properties include its roles in cellular signalling, gene expression, immune response, and apoptosis are now considered to be of importance.^{15,16} In the vitamin E group, α -tocopherol is considered to be the most active form. However, recently, tocotrienol

have gained increasing scientific interest due to their eminent antioxidant effects and a nonantioxidant activity profile that differs somewhat from that of tocopherol.¹³ Tocotrienols are identical in structure to tocopherols except for the degree of saturation in their side chain. Tocopherol has a saturated phytyl tail, while tocotrienols possess an unsaturated isoprenoid side chain. The prenyl side chain of tocotrienol has been postulated to be responsible for the differential membrane distribution and metabolism of tocotrienols when compared with tocopherols. α -Tocotrienol was observed in vitro to possess a remarkably higher antioxidant activity against lipid peroxidation than α -tocopherol. This was due to a more uniform distribution in the membrane lipid bilayer providing a more efficient interaction of the chromanol ring with lipid radicals.¹⁷

In view of this, it is of interest to investigate the effects of tocotrienol in preventing oxidative stress-induced cellular aging by determining telomere lengths and telomerase activities in normal skin fibroblast derived from young, middle and old age humans. Therefore the objective of this study was to determine the protective effects of palm γ -tocotrienol treatment before or after H₂O₂-induced oxidative stress on cell viability, telomere length and telomerase activity in normal human skin fibroblast cell lines derived from different age groups viz; young (21-year-old), middle (40-year-old) and old (68-year-old).

Results

IC₅₀ determination of H₂O₂ and γ -tocotrienol. Results showed that the number of viable cells was not significantly different from control after exposure to less than 500 μM, 300 μM and 200 μM for fibroblast cells derived from young, middle and old age humans respectively (Fig. 1). However, exposure to 700 μM, 400 μM and 100 μM H₂O₂ reduced viable cell number by about 50% (IC₅₀) for fibroblast cells derived from young, middle and old age humans respectively. Almost all the cells were not viable at H₂O₂ concentration of 1,000 μM, 900 μM and 800 μM for fibroblast cells derived from young, middle and old age humans respectively. Incubation with γ -tocotrienol for 24 hours caused an increase in the number of viable cells (Fig. 2) with the highest cell viable at γ -tocotrienol concentration of 80 μM for YF and 40 μM for fibroblast cells derived from middle and old age humans (optimum dose). However incubation with 200 μM, 300 μM and 100 μM γ -tocotrienol reduced viable cell number by about 50% (IC₅₀) for fibroblast cells derived from young, middle and old age humans respectively. Almost all the cells were not viable at γ -tocotrienol concentration of 500 μM for fibroblast cells derived from young and middle age humans and 150 μM for fibroblast cells derived from old age human.

Morphological changes with H₂O₂ and γ -tocotrienol treatment. Control fibroblasts derived from different age humans have live cells that were stained green with Calcein-AM whereas fibroblasts exposed to IC₅₀ H₂O₂ underwent apoptosis and were stained red with propidium iodide (Fig. 3). Fibroblasts derived from young age human showed normal spindle cells with round uniform nuclei and elongated cytoplasm (Fig. 3Ai), while young fibroblasts which were exposed to H₂O₂ showed pyknotic nuclei

with shrunken cytoplasm (Fig. 3Aii). Normal looking fibroblasts but more compact with increased cellularity were observed with γ -tocotrienol treatment (Fig. 3Aiii). Fibroblasts derived from middle age human showed normal nuclei, their cytoplasm were spindled but shorter than young fibroblasts (Fig. 3Bi). Middle age fibroblasts which were exposed to H_2O_2 showed pyknotic nuclei with smudged and elongated cytoplasm (Fig. 3Bii). Fibroblasts treated with γ -tocotrienol however showed similar morphology to untreated control fibroblasts but the cells were more compact (Fig. 3Biii). Old fibroblasts showed mild variations in nuclear size with spindled cytoplasm (Fig. 3Ci) while fibroblasts exposed to H_2O_2 showed pleomorphic nuclei and some pyknotic nuclei, their cytoplasm were thin, elongated and string-like (Fig. 3Cii). Treatment with γ -tocotrienol resulted in similar morphology of old fibroblasts to young fibroblasts (Fig. 3Ciii).

Effect of γ -tocotrienol on H_2O_2 -induced oxidative stress. Treatment of fibroblasts with IC_{50} dose of H_2O_2 resulted in 50% reduction in cell viability (Fig. 4). Pretreatment with optimum concentration of γ -tocotrienol protected fibroblast cells derived from all age groups against H_2O_2 -induced cell loss significantly ($p < 0.05$). The protective effects of γ -tocotrienol was observed much higher in fibroblast cells derived from middle and old age humans compared to fibroblast cells derived from young age human. This protection however was not seen in cells pretreated with high dose (IC_{50} dose) of γ -tocotrienol. Treatment of γ -tocotrienol after exposure to H_2O_2 -induced oxidative stress was also found to increase the number of cell viable in fibroblasts derived from all age groups. The optimum dose of γ -tocotrienol decreased the number of H_2O_2 -induced cell loss significantly ($p < 0.05$). This protective effects was almost similar to the effect of γ -tocotrienol treatment before the cells were exposed to H_2O_2 . However high concentration of γ -tocotrienol (IC_{50} dose) did not provide significant protection against H_2O_2 -induced cell loss in fibroblast cells derived from all age groups.

Estimation of telomere length. Figure 5 shows the Southern blot analysis of fibroblast cells with different treatments. In fibroblast cells derived from old age human, the telomere length is shorter compared to fibroblast cells derived from young age human (Fig. 6). Treatment of fibroblast cells with IC_{50} dose of H_2O_2 resulted in shortening of telomere length. Pretreatment with optimum and IC_{50} doses of γ -tocotrienol protected against H_2O_2 -induced telomere shortening significantly ($p < 0.05$) in fibroblast cells derived from young and old age humans. Treatment of γ -tocotrienol after exposure to H_2O_2 -induced oxidative stress was also found to protect against H_2O_2 -induced telomere shortening significantly ($p < 0.05$) in fibroblast cells derived from young and old age humans.

Estimation of telomerase activity. Figure 7 shows the representative PCR analysis for telomerase activity of fibroblast cells with different treatments. The effects of γ -tocotrienol on telomerase activity is shown in Figure 8. Results showed that telomerase activity is higher in fibroblast cells derived from young age human compared to fibroblast cells derived from middle and old age humans. Treatment of fibroblast cells with IC_{50} dose of H_2O_2 resulted in reduction of telomerase activity. Pretreatment with optimum dose of γ -tocotrienol protected against H_2O_2 -induced

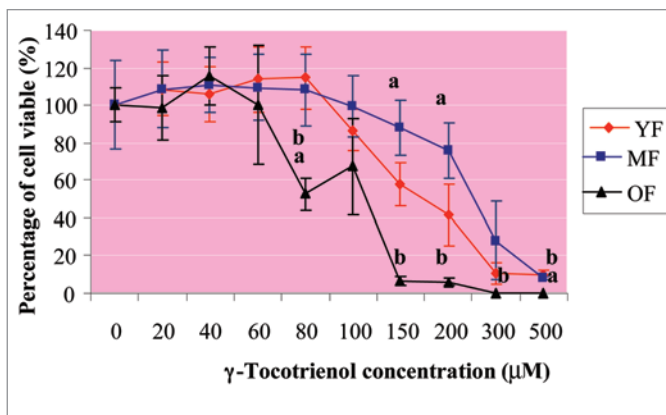


Figure 2. Cytotoxic effects of γ -tocotrienol on fibroblast cell lines as assessed by MTS assay. Percent MTS reduction corresponds to the viable cell number. Fibroblasts were incubated with increasing concentrations of γ -tocotrienol for 24 hours at 37°C. Incubation with γ -tocotrienol caused a significant increase in the number of cell viable. The percentage of the viable cell is highest at γ -tocotrienol concentration of 80 μ M for fibroblast cells derived from young (YF) age human and 40 μ M for fibroblast cells derived from middle (MF) and old (OF) age humans. Cell viability decreased by 50% (IC_{50}) with γ -tocotrienol incubation at 230 μ M, 280 μ M and 110 μ M for fibroblast cells derived from young, middle and old age humans respectively. Almost all cells were dead at γ -tocotrienol more than 500 μ M for fibroblast cells derived from young and middle age humans and 150 μ M for fibroblast cells derived from old age human. ^aDenotes $p < 0.05$ compared to fibroblast cells derived from young age human, ^b $p < 0.05$ compared to fibroblast cells derived from middle age human. Data is presented as means \pm SD, $n = 3$.

telomerase activity loss significantly ($p < 0.05$) in fibroblast cells derived from all age groups. However high concentration of γ -tocotrienol (IC_{50} dose) and treatment of γ -tocotrienol after exposure to H_2O_2 -induced oxidative stress did not provide significant protection against H_2O_2 -induced telomerase activity loss in fibroblast cells derived from all age groups.

Discussion

Accumulation of oxidative damage plays an important role in aging and associated diseases.¹⁸ The pioneering discovery of Hayflick and Moorhead identified the cultured human diploid fibroblast (HDF) as a potential model for aging.¹ There is evidence that in vitro cellular senescence may directly correlate with aging in vivo. In this study, we explored the effect of H_2O_2 on fibroblast cell lines derived from young, middle and old age humans and determined the protective effect of γ -tocotrienol against H_2O_2 -induced oxidative stress by determining cell viability as measured by the MTS assay, and examining the morphology of cells by staining with specific fluorescence dyes that differentiate live from apoptotic cells, determination of telomere length as well as telomerase activity using Southern blot analysis and telomere repeat amplification protocol respectively.

The data showed that H_2O_2 at concentrations 700 μ M, 400 μ M and 100 μ M were cytotoxic to fibroblast cells derived from

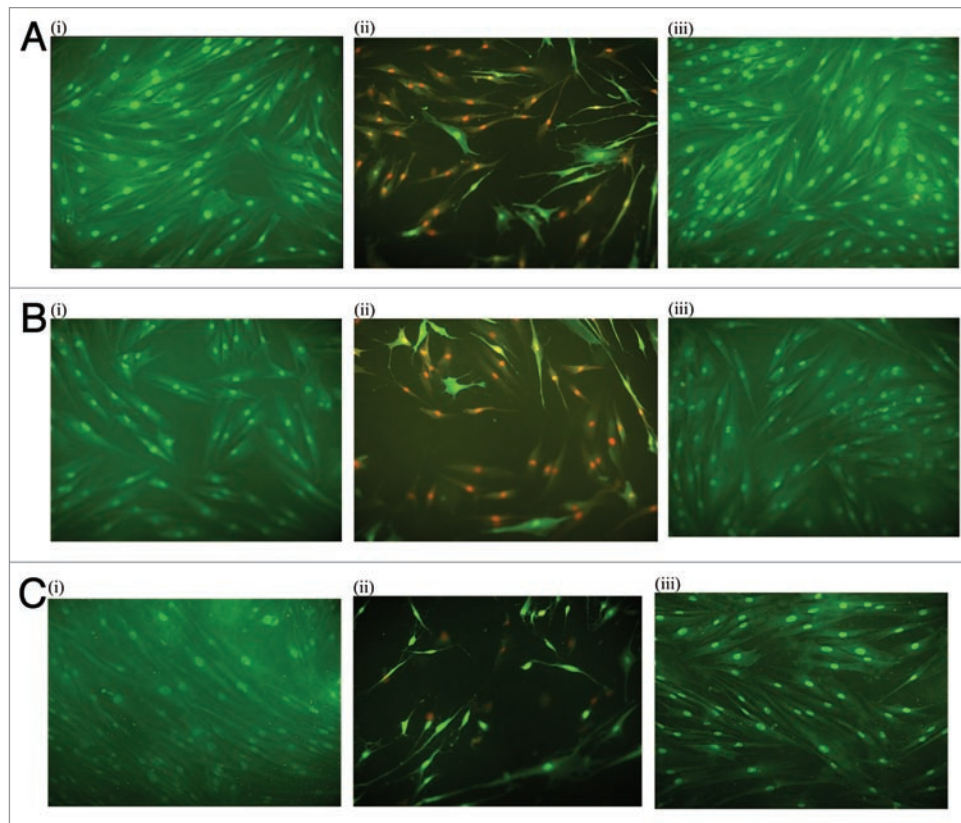


Figure 3. Fluorescence cell death staining for fibroblast cell lines derived from young (A), middle (B) and old (C) age humans, for untreated control fibroblasts (i), fibroblasts exposed to H_2O_2 (ii) at 700 M, 400 M and 100 M for 2 hours respectively and fibroblasts treated with γ -tocotrienol (iii). Live cells were stained with 30 M Calcein-AM (green) and dead cells were stained with 7.5 M propidium iodide (PI, red). Micrographs are shown at X40 magnification.

young, middle and old age humans respectively as it increased the number of cell loss as well as the number of apoptotic cells as shown by the fluorescence dyes staining. Almost all the cells were not viable at H_2O_2 concentration of 1000 μ M, 900 μ M and 800 μ M for fibroblast cells derived from young, middle and old age humans respectively. This is in agreement with earlier observations.¹⁹ H_2O_2 has been reported to directly stimulate endonuclease activity in renal tubular epithelial cells, leading to DNA fragmentation and cell death via apoptosis.²⁰ The present data showed that fibroblast cells derived from old age human was more susceptible to the action of H_2O_2 . This is in agreement with earlier observations.²¹ Old fibroblasts were reported to be more sensitive to oxidative stress due to decreased antioxidant activity, less capability to repair, increased reactive oxygen species (ROS) production, and increased sensitivity of DNA towards the action of ROS.²²

The data showed that at low concentration, γ -tocotrienol increased the number of viable cell. At γ -tocotrienol concentration of 80 μ M the number of cell viable is maximum for young fibroblasts and 40 μ M for middle and old fibroblasts. However, high concentration of γ -tocotrienol is cytotoxic. There was more than 50% reduction in viable cell number at concentrations greater than 230 μ M, 280 μ M and 110 μ M γ -tocotrienol for

young, middle and old fibroblasts respectively. This probably reflects the pro-oxidant activity of γ -tocotrienol.

The present data also showed that γ -tocotrienol provides protection against H_2O_2 -induced cell loss in fibroblast cells derived from all age groups. The protection was more effective in middle and old fibroblasts. Previous study reported that vitamin E reduced H_2O_2 -induced HO^\bullet generation and subsequent DNA base pair modification in human oral epithelial cells.²³ Vitamin E was found to decrease H_2O_2 -induced DNA strand breaks in human skin cell line VH10.²⁴ Konopacka et al. (1998) reported that the addition of vitamin E immediately after radiation treatment to bone marrow polychromatic erythrocytes, reduced radiation-induced micronucleus formation.²⁵ It has been suggested that the inhibitory effect of vitamin E micronucleus formation may be due to modulation of the DNA repair system. Indeed, measurement of DNA repair ability tested in lymphocytes indicated that vitamin E increased the removal rate of damaged DNA compared to cells that are not treated with vitamin E.

Telomere shortening is one of a few mechanisms mediating the development of the senescent phenotype. Telomere effects have been reported to be the predominant trigger of premature senescence in Werner syndrome, a disease that caused many clinical features associated with aging in human. Microarray analyses showed that mRNA expression patterns induced in senescent

Werner syndrome cells appeared similar to those in normal strains.²⁶

Data in the present study seems to suggest that both telomere shortening and decrease telomerase activity to be contributing factors in cellular aging. In the present study, treatment of fibroblast cells with H_2O_2 resulted in shortening of telomere length and reduction of telomerase activity. However, γ -tocotrienol treatment protected against H_2O_2 -induced telomere shortening. γ -tocotrienol was also shown to protect against H_2O_2 -induced telomerase activity loss in fibroblast cells derived from all age groups. Therefore the anti aging effect of tocotrienols may not only be due to their potent antioxidant properties but may also involve its role in maintaining telomere length possible mediated by telomerase. Consistent with the telomere theory of aging, telomerase positive human diploid fibroblasts were resistant to replicative senescence.⁵ Our findings showed similar results whereby increased telomerase activity induced by γ -tocotrienol cause a significantly increased of telomere length in skin fibroblast derived from young and old age humans.

Our data that showed shortening of telomere length with H_2O_2 treatment is consistent with previous findings which reported that telomeres are particularly sensitive to oxidative stress.²⁷ A study by Rubio et al. (2004) showed that telomerase influences the stress response indirectly by altering telomere length, mainly by elongating shorter telomeres.²⁸ Thus, telomere length does not only determine replicative senescence, but also influences the senescence response to genotoxic agents.

Telomere shortening has been proven to be a cell division counter in proliferating fibroblast, or 'replicometer'. Telomere shortening down to a threshold length seems to be the best known predictor of senescence.²⁹ A study by Von Zglinicki (2002) showed that telomere shortening is largely dependent on the interplay of oxidative stress and antioxidant defense rather than to counter clock of numbers of cell divisions.²⁷ H_2O_2 has been the most commonly used inducer of stress-induced premature senescence (SIPS), which shares features of replicative senescence including similar morphology, increase senescence-associated β -galactosidase activity and cell cycle regulation.¹² It has been reported that treating cells exogenously with a certain hydrogen peroxide (H_2O_2) concentration can trigger entry into a senescent-like state.³⁰ Sublethal dose (IC_{50}) of H_2O_2 used in this study induced shortening of telomere length and reduction in telomerase activity of skin fibroblast cells derived from young and old age humans.

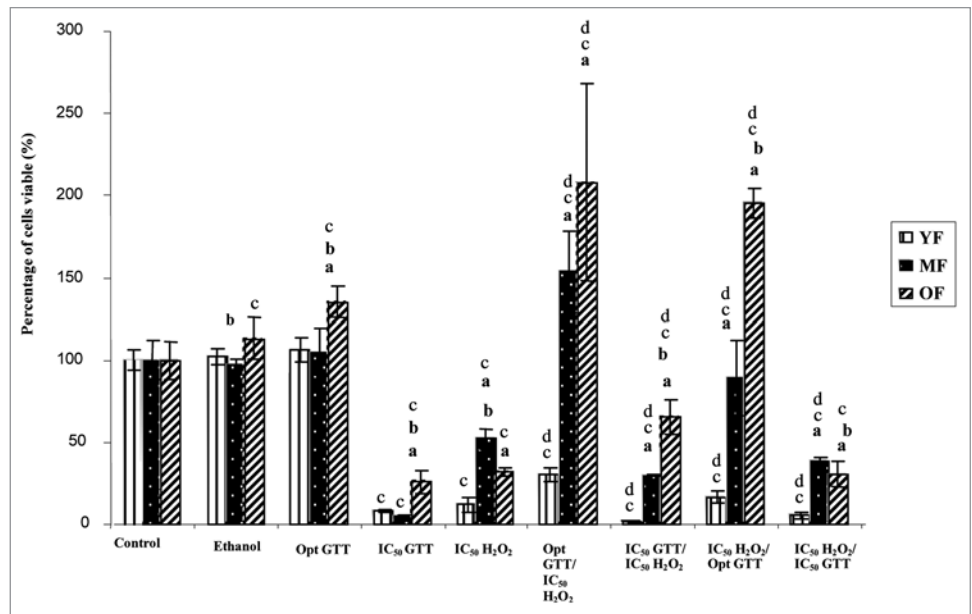


Figure 4. Protective effects of γ -tocotrienol against H_2O_2 -induced cell loss in fibroblasts cultures. Fibroblast cell lines derived from different age groups were treated with γ -tocotrienol (optimum concentration and IC_{50}) for 24 hours at $37^\circ C$ before or after H_2O_2 exposure (IC_{50}) for 2 hours. Cell viability was determined using the MTS reduction assay. Protective effect of γ -tocotrienol (optimum concentration) was seen in fibroblast cells derived from different age groups. ^aDenotes $p < 0.05$ compared to fibroblast cells derived from young age human, ^b $p < 0.05$ compared to fibroblast cells derived from middle age human, ^c $p < 0.05$ compared to control, ^d $p < 0.05$ compared to H_2O_2 -treated cells. Data is presented as means \pm SD, $n = 3$.

Our data that showed γ -tocotrienol protected against H_2O_2 -induced telomere shortening and protected skin fibroblast against H_2O_2 -induced telomerase activity loss suggesting its effect in protecting and recovering the skin fibroblast from oxidative stress. Tocotrienols are capable of scavenging and quenching reactive oxygen species. This antioxidant activity however, remains mainly with its 'chain-breaking' property which neutralizes peroxy and alkoxy radicals generated during lipid peroxidation.¹³ As vitamin E is also involved in cell signaling, we suggested that elongation of telomere length and activation of telomerase activity by γ -tocotrienol may be mediated by cell signaling pathway. Previous study showed that vitamin E supplementation inhibits chromosomal alterations that are measured by chromosome gaps and breaks in c-myc proto-oncogene and transforming growth factor co-overexpressing transgenic mice.³¹ Additional studies have shown that c-myc may also be a 'key switch' for the induction of telomerase activity.³² The c-fos expression is induced when cells are exposed to H_2O_2 and UV, and both H_2O_2 and UV induce DNA damage.³³ The c-fos encodes a nuclear protein that forms heterodimer with jun and binds a common DNA site (AP-1) to regulate various transcriptional activity. Vitamin E treatment also has been shown to induce c-jun expression as well as AP-1 binding activity in human breast cancer cells.^{34,35} These studies suggest that the protective role of vitamin E against DNA damage induced by H_2O_2 and UV is in part, mediated by upregulation of c-fos expression and AP-1 binding activity. Hadshiew et al. (2000) in their study showed that there are homologies between certain protein kinases and telomeric protein suggest a link between

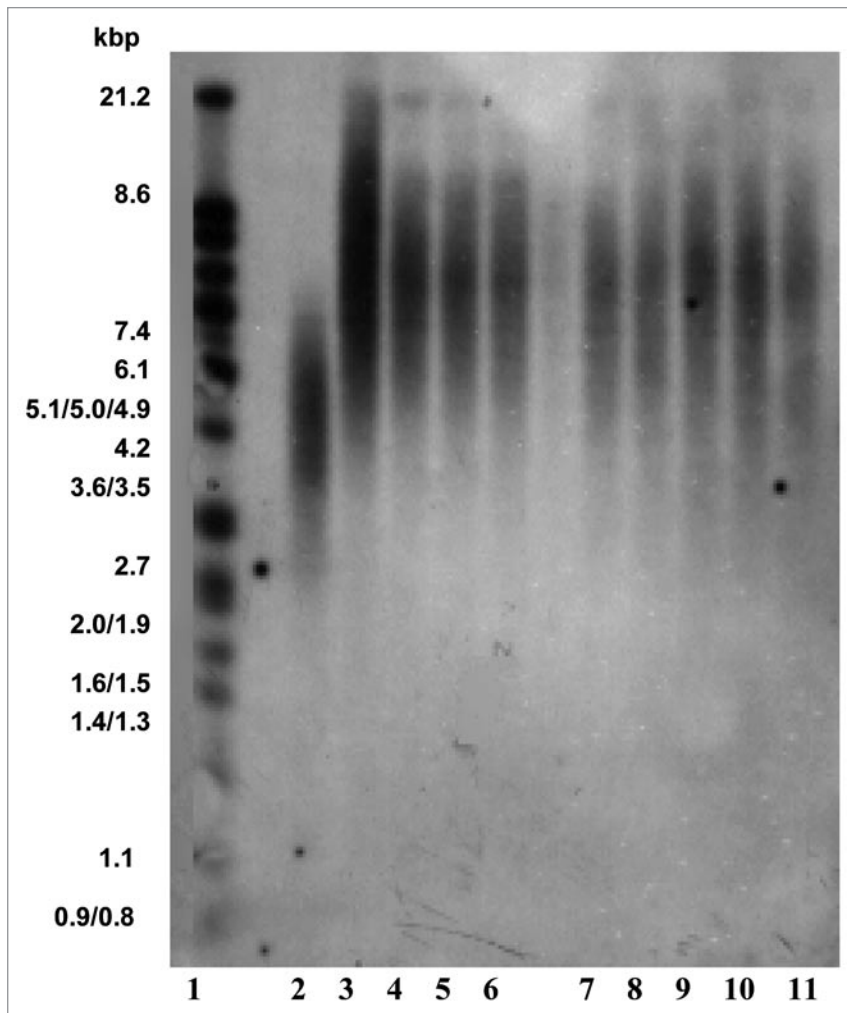


Figure 5. Representative Southern blot for fibroblast cell lines with different treatments. Telomeric DNA shows wide smears in all lanes. Lane 1; molecular weight marker, lanes 2 and 3; control DNA low and high, lane 4; cells without any treatment, lane 5; cells treated with optimum dose of γ -tocotrienol, lane 6; cells treated with IC_{50} dose of γ -tocotrienol, lane 7; cells with H_2O_2 -induced oxidative stress, lane 8; cells treated with optimum dose of γ -tocotrienol prior to H_2O_2 exposure, lane 9; cells treated with IC_{50} dose of γ -tocotrienol prior to H_2O_2 exposure, lane 10; cells treated with optimum dose of γ -tocotrienol after H_2O_2 exposure, lane 11; cells treated with IC_{50} dose of γ -tocotrienol after H_2O_2 exposure.

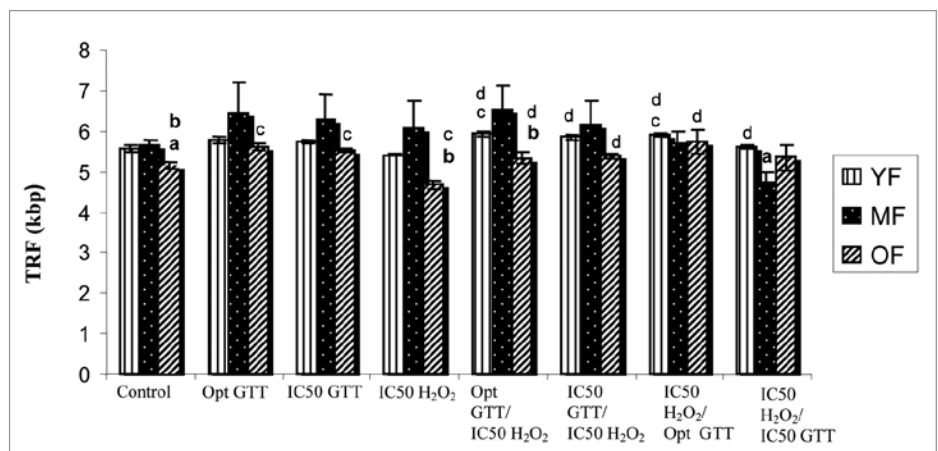
chromosome stability and cell cycle regulation.³⁶ In fact, a critical reduction in telomere length appears to interfere with transcription of sub-telomeric genes and leads to cell cycle arrest or apoptosis and finally the aging phenotype.

Materials and Methods

Materials. γ -tocotrienol extracted from palm oil, was supplied by the Malaysian Palm Oil Board. Eagle's Minimum Essential Medium (EMEM) and Iscove's Modified Dulbecco's Medium (IMDM) were purchased from Flowlab, (Australia) which also provided trypsin, trypan blue, L-glutamin, antibiotics, sodium pyruvate and non-essential amino acids. Fetal calf serum (FCS) from PAA (PAA lab, Austria). H_2O_2 was purchased from BDH (British Drug House). Telomere Length Assay kit was purchased from Roche, Germany while TRAPEze Telomerase Detection kit was purchased from Chemicon, USA. Wizard Genomic Purification Kit was obtained from Promega, USA. Other chemicals and solvents were supplied by Sigma or Merck (Darmstadt, Germany). Fibroblast cells were obtained from American Tissue Culture Collection.

Cell culture and treatment protocols. Normal skin fibroblast cell lines derived from young (21-year-old, YF) and middle age (40-year-old, MF) humans were established in complete culture media (CCM) which contains Iscoves Modified Dulbeccos Media (IMDM) (Flowlab, Australia) containing 3.7 g $NaHCO_3$ supplemented with L-glutamin, 100 unit/ml penicillin, 100 μ g/ml streptomycin, amphotat 0.25 μ g/ml and 10% fetal calf serum (FCS). Cells were maintained at in a humidified atmosphere of 5% CO_2 at 37°C. Normal skin fibroblast cell lines derived from old age human (68-year-old, OF) were

Figure 6. Effect of γ -tocotrienol on telomere length (TRF length) of human skin fibroblast cell lines derived from different age groups. Cells were treated with optimum dose and IC_{50} dose of γ -tocotrienol prior or after H_2O_2 exposure. Protective effects of γ -tocotrienol against H_2O_2 -induced telomere shortening was observed in fibroblast cells derived from young (YF) and old (OF) age humans. ^aDenotes $p < 0.05$ compared to fibroblast cells derived from young age human, ^b $p < 0.05$ compared to fibroblast cells derived from middle age human, ^c $p < 0.05$ compared to control, ^d $p < 0.05$ compared to H_2O_2 -treated cells. Data is presented as means \pm SD, $n = 3$.



established in CCM which contains Eagle's Minimum Essential Medium (EMEM) (Flowlab, Australia) containing 3.7 g NaHCO₃ and Hepes supplemented with L-glutamin, 0.1 mM non-essential amino acids, 1.0 mM sodium pyruvate, 100 unit/ml penicillin, 100 µg/ml streptomycin, amphostat 0.25 µg/ml and 10% FCS. The medium was equilibrated with a humidified atmosphere of 5% CO₂ at 37°C. Cells were subcultured by passing them at a ratio of 1:3 in regular intervals. Cells were passaged such that the monolayers never exceeded 70–80% confluency. Cells at passage 6 (population doubling less than 12) were used for further analysis.

Analysis of cytotoxicity. γ-Tocotrienol was incubated in FCS overnight at 37°C. Dilutions of H₂O₂ and γ-tocotrienol in IMDM or EMEM were made fresh just prior to each experiment. Small volumes of H₂O₂ and γ-tocotrienol were added to each well to make final dilutions of 10–1,000 µM of H₂O₂ and 20–200 µM of γ-tocotrienol, respectively. The cells were then agitated lightly. Cultures were incubated in a humidified incubator at 37°C, 5% CO₂ with H₂O₂ for 2 hours and with γ-tocotrienol for 24 hours. Cell viability was assessed.

Analysis of cell viability. Cell viability was assessed with CellTiter 96* Aqueous Non-Radioactive Cell Proliferation Assay (MTS, Promega, USA). The MTS employs 3-(4,5-dimethylthiazol-2-yl)-5-carboxymethoxyphenyl 2-(4-sulfophenyl)-2H-tetrazolium (MTS) and the electron coupling agent phenazine methosulphate (PMS). The MTS compound is reduced by the dehydrogenase enzymes found in metabolically active cells into a formazan product that is soluble in the medium. The amount of colored formazan product is proportional to the number of viable cells. Briefly, 20 µl MTS solution was added to each well and incubated in humidified incubator at 37°C in 5% CO₂ for 2–4 hours. The quantity of formazan product present was determined by measuring the absorbance at 490 nm with a microtiter plate reader (VeraMax Molecular Devices, USA). Cell death staining make use of the fact that live cells have intracellular esterase that convert non-fluorescent cell-permeable calcein-AM to the intensely

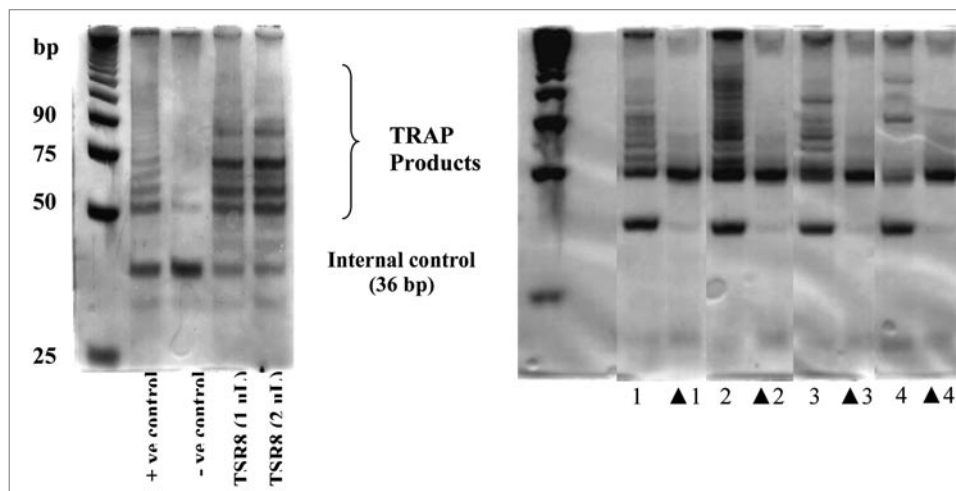


Figure 7. Representative PCR analysis for telomerase activity of fibroblast cell lines with different treatments. Gel A; control set for telomerase activity with molecular weight markers. Gel B; telomerase activity in sample with different treatments. Lane 1, control; lane ▲1, (heat treated) control; lane 2, cells treated with optimum dose of γ-tocotrienol; lane ▲2, cells treated with optimum dose of γ-tocotrienol (heat treated); lane 3, cells treated with IC₅₀ dose of γ-tocotrienol; lane ▲3, cells treated with IC₅₀ dose of γ-tocotrienol (heat treated); lane 4, cells with H₂O₂-induced oxidative stress; lane ▲4, cells with H₂O₂-induced oxidative stress, (heat treated).

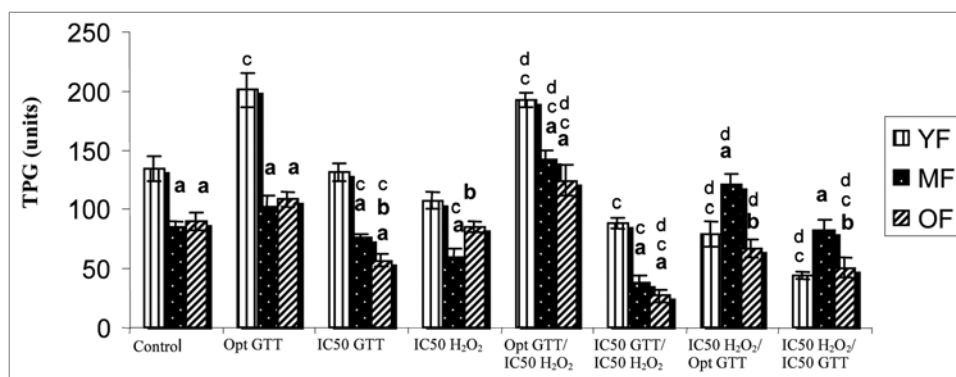


Figure 8. Effect of γ-tocotrienol on telomerase activity (Total Product Generated, TPG) of human skin fibroblast cell lines derived from different age groups. Cells were treated with optimum dose and IC₅₀ dose of γ-tocotrienol prior or after H₂O₂ exposure. Pretreatment with optimum dose of γ-tocotrienol protected against H₂O₂-induced decreased telomerase activity in fibroblast cells derived from young (YF), middle (MF) and old (OF) age humans. ^aDenotes p < 0.05 compared to fibroblast cells derived from young age human, ^bp < 0.05 compared to fibroblast cells derived from middle age human, ^cp < 0.05 compared to control, ^dp < 0.05 compared to H₂O₂-treated cells. Data is presented as means ± SD, n = 3.

fluorescent calcein which is retained within the cells. Viable cell membranes are impermeable to propidium iodide. Dead cells however, allow propidium iodide to enter and bind to nucleic acid. Thus live cells stained with calcein-AM while dead and apoptotic cells stained with propidium iodide to give green and red fluorescence, respectively. Thirty µg/ml calcein-AM and 7.5 µg/ml propidium iodide were added to cultured fibroblasts in chamber slides and incubated for 30 min. Thereafter, cultures were washed with PBS, fixed with fresh 2% paraformaldehyde and then coverslips mounted for microscopic examination.

Analysis of protective effects of γ -tocotrienol. Cells were seeded at 2×10^6 cells/10 ml CCM in petri dish, then incubated with γ -tocotrienol for 24 hours (the concentration of γ -tocotrienol that caused maximum cell viability; optimum dose or the concentration of γ -tocotrienol that caused 50% cell viability; IC_{50} determined earlier) before or after the addition of H_2O_2 for 2 hours (the concentration of H_2O_2 that caused 50% cell viability; IC_{50} determined earlier). Cell viability, telomere length and telomerase activity were determined.

Estimation of telomere length. Genomic DNA was isolated using Wizard Genomic Purification Kit (Promega, USA). The DNA samples were processed separately for each treatment. Three μ g of DNA was digested with 20 units each of *Hinf*I and *Rsa*I for 2 hours at 37°C. Complete cutting is confirmed by electrophoresis of the DNA digests on 0.8% agarose gel. Fractionated DNA fragments were transferred to nylon membranes Hybond-N⁺ (Amersham, UK) by an alkaline transfer technique using capillary blotting. The blotted DNA fragments were hybridized to a digoxigenin (DIG)-labeled probe specific for telomeric repeats and incubated with a DIG-specific antibody covalently coupled to alkaline phosphatase. Finally, the immobilized telomere probe was visualized by virtue of alkaline phosphatase metabolizing CDP-Star, a highly sensitive chemiluminescence substrate. The average terminal restriction fragment (TRF) length was determined by comparing signals relative to a nuclear weight standard on X-ray film. The average TRF length of each sample could be obtained by scanning the exposed X-ray film and analyzed using ImageMaster Total lab software. TRF lengths were recorded for simplicity as telomere lengths.

Estimation of telomerase activity. Detection of telomerase activity using the telomeric repeat amplification protocol (TRAP) in cultured cells involves the addition of TTAGGG repeats by telomerase to an oligonucleotide (TS), and the subsequent PCR amplification of these extension products with both the forward (TS) and reverse (CX) primers (Kim et al. 1994). The TRAPeze telomerase detection kit (Chemicon, USA) was used as recommended by the manufacturer with minor modifications. Briefly, cell pellets were stored at -80°C until lysis was performed. The lysis buffer contained 1% Nonidet P-40 and 0.25 mM sodium deoxycholate to increase the efficiency of extraction. Cells were

lyzed, then left on ice for 30 min, and centrifuged at 14,000 g for 20 min at 4°C. The supernatant was flash-frozen and stored at -80°C. For the PCR reaction, 2 μ l of extract (corresponding to 100–1,000 cells) was added to 48 μ l reaction mixture and two units of Taq DNA polymerase (Promega, USA). After incubation at room temperature for 30 min for the telomerase extension reaction, samples were heated to 92°C for 3 min to inactivate telomerase followed by PCR amplification. PCR products were electrophoresed on 10% polyacrylamide gel, and the gel was further analyzed and quantitated using the ImageMaster Total lab software. Telomerase activity was calculated as the ratio of the intensity of telomerase ladders to the intensity of the 36-bp internal standard.

Statistical data analysis. Each experiment was carried out in triplicates with at least three independent cultures with comparable results. Data are reported as mean \pm SD of at least three experiments. Comparison between groups were made by Student's *t*-test. $p < 0.05$ was considered statistically significant.

Conclusion

The results obtained showed that γ -tocotrienol at low concentration was able to protect against H_2O_2 -induced cell loss in fibroblast cell lines derived from different age groups. γ -tocotrienol was also able to protect against H_2O_2 -induced telomere shortening and telomerase activity loss. Therefore we suggest that γ -tocotrienol protects against oxidative stress-induced cellular aging by modulating the telomere length possibly via telomerase. However more studies need to be done in order to understand the mechanisms.

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