

RESEARCH ARTICLE

Open Access

DNA prime and peptide boost immunization protocol encoding the *Toxoplasma gondii* GRA4 induces strong protective immunity in BALB/c mice

Min Meng^{1†}, Aihua Zhou^{2*†}, Gang Lu¹, Lin Wang¹, Guanghui Zhao¹, Yali Han¹, Huaiyu Zhou¹, Hua Cong¹, Qunli Zhao¹, Xing-Quan Zhu³ and Shenyi He^{1*}

Abstract

Background: *Toxoplasma gondii* is a widespread intracellular parasite, which infects most vertebrate animal hosts and causes zoonotic infection in humans. Vaccine strategy remains a promising method for the prevention and control of toxoplasmosis. *T. gondii* GRA4 protein has been identified as a potential candidate for vaccine development. In our study, we evaluated the immune response induced by four different immunization vaccination strategies encoding TgGRA4.

Methods: BALB/c mice were intramuscularly (i.m.) immunized four times according to specific immunization schedules. Generally, mice in experimental groups were immunized with polypeptide, pGRA4, peptide/DNA, or DNA/peptide, and mice in the control groups were injected with PBS or pEGFP. After immunization, the levels of IgG antibodies and cytokine productions were determined by enzyme-linked immunosorbent assays (ELISA). The survival time of mice was also evaluated after challenge infection with the highly virulent *T. gondii* RH strain.

Results: The results showed that mice vaccinated with different immunization regimens (polypeptide, pGRA4, peptide/DNA, or DNA/peptide) elicited specific humoral and cellular responses, with high levels of total IgG, IgG2a isotype and gamma interferon (IFN- γ), which suggested a specific Th1 immunity was activated. After lethal challenge, an increased survival time was observed in immunized mice (11.8 ± 4.8 days) compared to the control groups injected with PBS or pEGFP ($P < 0.05$). Mice injected with PBS or pEGFP died within 8 days, and there was no significant difference in the protection level in two groups ($P > 0.05$).

Conclusions: These results demonstrated that this DNA prime and peptide boost immunization protocol encoding the TgGRA4 can elicit the highest level of humoral and cellular immune responses compared to other immunized groups, which is a promising approach to increase the efficacy of DNA immunization.

Background

Toxoplasma gondii is a widespread intracellular parasite belonging to the phylum Apicomplexa, which infects most vertebrate animal hosts and causes zoonotic infection in humans [1]. In general, an asymptomatic but chronic infection is established in immunocompetent

individuals; However, in selected immunocompromised human, and in particular those with human immunodeficiency virus (HIV) infection, *T. gondii* can result in the extensive and fatal tissue damage [2]. The life cycle of *T. gondii* consists of two phases: the sexual stage only in felines and the asexual stage in human and other intermediate hosts [3]. The asexual component comprises two distinct stages of growth: rapidly growing 'tachyzoites' and latent 'bradyzoite' tissue cysts [4]. Host cell invasion and lysis by the actively dividing tachyzoites are directly responsible for toxoplasmosis, which is

* Correspondence: shenyi.he@hotmail.com; zhouah@sdu.edu.cn

†Equal contributors

¹Department of Parasitology, Shandong University School of Medicine, 250012, Jinan, Shandong Province, Peoples Republic of China

²Department of Pediatrics, Provincial Hospital Affiliated to Shandong University, Shandong University School of Medicine, 250021, Jinan, Shandong Province, Peoples Republic of China

Full list of author information is available at the end of the article

particularly severe in immunocompromised individuals and in the congenitally infected fetus [5,6].

Although drugs are the primary strategy for the treatment of toxoplasmosis, they are poorly tolerated, have severe side effects and drug-resistance, and cannot act against chronic *T. gondii* infection [3]. It is known that individual patients infected with *T. gondii* have serious consequences and the disease burden of congenital toxoplasmosis on a population level is considerably high [7]. Thus, the development of an effective and safe vaccine against *T. gondii* acute and chronic infection is an important and urgent goal. So far, only a commercial vaccine is deployed to control toxoplasma abortion in sheep, which comprises live tachyzoites of the S48 'incomplete' strain of *T. gondii* [8]. In human, an effective vaccine could be valuable for preventing both fetal infection and reactivation in immunocompromised individuals; In livestock such as sheep and goats, it could prevent spontaneous abortion, decrease economic losses and reduce the potential for a major epidemiologic vector for human infection [9,10]. In general, proteins or peptides and DNA vaccines are considered as the third generation subunit vaccines, which are important in preventing intracellular parasites infection [11]. DNA vaccines encoding specific antigen can activate all pathways of immune response, with production of antibody, Th1-dominated CD4⁺ T cells and CTLs (cytotoxic T-cell responses), and provide protection against diseases that require cell-mediated immunity such as intracellular protozoan infections [12]. DNA vaccines against *T. gondii* have been proved that they can induce antibody, specific T-cell responses and protective immunity against acute and chronic challenge in mice [13,14]. In contrast to DNA vaccines, synthetic multiple antigen peptide (MAP) vaccines are an effective and new approach to deliver multiple T-cell and B-cell epitopes as the constituents of a single immunogen, which contains a high concentration of the relevant antigen for inducing immune responses to predefined epitopes [15]. Synthetic MAP vaccines have been shown to elicit better cell-mediated immunity by focusing the host immune response on epitopes known to play a role in protective immunity [16,17]. Recently, several studies have demonstrated the power of synthetic polypeptides vaccines in eliciting protective immunity to intracellular parasites such as *Plasmodium falciparum* [18], *Schistosoma mansoni* [19], *T. gondii* [17], and *Paracoccidioides brasiliensis* [20].

Lots of studies indicate that the family of promising vaccine candidate antigens includes surface antigen (SAGs) [21], dense granule proteins (GRAs) [22], rhoptry proteins (ROPs) [23], and micronemal proteins (MICs) [24]. GRA proteins, highly expressed by *T. gondii*, are involved in parasite survival and virulence and are the major member of the excreted secreted antigens (ESA) [25,26]. Among

the GRA proteins, GRA4 has been identified as the leading candidate for vaccine development [27]. In mice immunized with plasmids encoding for GRA4 antigen or the recombinant GRA4 protein generated a Th1 type immune response and provided partial protection against parasite infection [28,29]. Concerning GRA4 peptides, amino acids 229-242 and 231-245 containing B and T-cell epitopes can induce both humoral and cellular immune responses following oral infection with *T. gondii* [30,31]. Multi-epitope DNA vaccine expressing six antigen segments of SAG1, GRA1 and GRA4 produced stronger humoral and Th1-type cellular immune responses, which is a potential strategy for the control of toxoplasmosis [32]. A multiple antigenic peptide (MAP) vaccine including one B-cell and two T-cell epitopes derived from *T. gondii* antigens (SAG1, GRA4 and GRA1) could trigger stronger humoral and cellular responses against *T. gondii* [17]. Therefore, GRA4 is an attractive and promising vaccine candidate against *T. gondii* and GRA4 231-245 peptide containing B and T-cell determinants has been proved to be immunogenic, and was considered suitable alternative in polypeptide vaccine design.

Furthermore, vaccination strategy is also a key factor in influencing immunity, which is as important as vaccine candidates [33]. One particularly promising approach is the prime-boost strategy, which has been shown to generate high level of T-cell memory in animal models [34]. To increase antibody production following DNA immunizations, prime-boost regimens have been shown to be an effective approach to induce both humoral and cellular immune responses [35,36]. The key strength of this strategy is that greater levels of immunity are established by heterologous prime-boost in which the antigen is applied via different routes and immunization sites [34].

The aim of the present study is to investigate their protective efficacy of different immunization regimens (polypeptide, pGRA4, peptide/DNA, or DNA/peptide) in mice against lethal *T. gondii* challenge. Our results demonstrate that the DNA prime-peptide boost vaccination regime can elicit the highest level of humoral and cellular immune responses, and is an effective strategy in increasing efficacy of DNA immunization.

Methods

Mice and parasite

Specific-pathogen-free (SPF) grade female Kunming and BALB/c mice (6-8 weeks old) were purchased from Shandong University Laboratory Animal Centre (Jinan, China). In this study, the *T. gondii* RH strain (Type I), a highly virulent strain for mice, was used to challenge immunized BALB/c mice. Tachyzoites of the RH strain was maintained in liquid nitrogen and recovered routinely by intraperitoneal passage in Kunming mice in our laboratory according to the method previously described [37].

Tachyzoites of *T. gondii* were harvested from the peritoneal fluid of Kunming mice that had been intraperitoneally infected three days earlier. Soluble tachyzoite antigen (STAg) was obtained from RH strain tachyzoites as previously described [38]. Briefly, parasites obtained from the peritoneal exudates were washed three times by centrifugation, then suspended in sterile phosphate buffered saline (PBS) and sonicated for three 10 min periods at 60 w/s. The toxoplasma sonicate was centrifuged at $2100 \times g$ for 15 min. The supernatant, containing soluble tachyzoite antigens (STAg), was harvested and stored at -80°C until further use.

Construction of the eukaryotic expression plasmid

The total genomic DNA of *T. gondii* RH strain was extracted from purified tachyzoites using a commercial kit (QIAGEN, Germany) and used as a template DNA for the polymerase chain reaction (PCR). The coding sequence of the *T. gondii* GRA4 gene [GenBank: AAA30142.1] was amplified from the *T. gondii* genomic DNA, with a pair of specific synthetic primers (forward primer: 5'-CGGGGTACCATGCAGGGCACTTGGTTTTC-3', reverse primer: 5'-CGCGGGATCCTCACTCTTTGCGCATTCTTT-3'), in which KpnI and BamHI restriction sites were introduced. The PCR conditions were as follows: pre-denaturation at 94°C for 5 min; denaturation at 94°C for 30 s, annealing at 65°C for 30 s and extension at 72°C for 1 min, followed by 30 cycles; final extension at 72°C for 10 min. The sequence of amplified GRA4 DNA fragment was verified by double-stranded sequencing. The GRA4 DNA fragment was subcloned into the eukaryotic expression vector pEGFP-C1 (Clontech, USA) to form the plasmid pEGFP-GRA4 using KpnI and BamHI restriction sites. The recombinant plasmids were then transformed into *Escherichia coli* DH5 α . Positive recombinant plasmids were verified using PCR, double restriction enzyme digestion and double-stranded sequencing. The plasmids were purified using an endotoxin-free plasmid purification kit following the manufacturer's instructions, and stored at -20°C until use. The concentration of pGRA4 was determined by spectrophotometer at OD 260 and OD 280, and the ratios were 1.8-2.0.

Design and synthesis of peptide

Peptide constructs were synthesized, purified, and identified as described previously [17,39]. Briefly, MAP vaccine of GRA4 was designed based on the published peptide sequences containing T-cell epitopes and B-cell epitopes [17,40]. In this study, peptide 231-245 (STEDSGLT-GVKDSSS) was selected and polypeptides were synthesized by Xi'an Bio-scientific Co., Ltd with the Solid-Phase Peptide Synthesizer by adopting four-branched peptide design. The purity of the peptide was confirmed by analytic HPLC.

pEGFP-GRA4 plasmid expression *in vitro*

HEK293T cells were maintained in our laboratory and routinely cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with penicillin (100 IU/ml), streptomycin (100 mg/ml) and 10% fetal bovine serum (FBS) at 37°C with 5% CO_2 . The *in vitro* transfection assay was performed as previously described [41]. Briefly, before transfection, 2×10^4 HEK293T cells were seeded in a 6-well plate. The recombinant pGRA4 plasmid (2 μg /well) was transfected into cells using the Fugene HD transfection reagent as instructed by the manufacturer. pEGFP plasmid-transfected cells and untransfected cells were used as the controls. Plates were incubated for 48 h at 37°C in 5% CO_2 , 95% humidity. The plasmid pGRA4 expression in the cells was detected by western blotting analysis.

Protein productions from HEK293T cells were collected on ice with RIPA Lyses Buffer (50 mM Tris pH 7.4, 150 mM NaCl, 1% Triton X-100, 1% Sodium deoxycholate, 0.1% SDS) containing 1 mM protease inhibitor PMSF (phenylmethanesulfonyl fluoride). The expression of plasmid pGRA4 in HEK cells were demonstrated by western blotting with anti-*T. gondii* polyclonal antibody (Goat) and a HRP (horseradish peroxidase)-labeled rabbit anti-goat IgG antibody (Sigma, USA) as a secondary antibody. The conjugated substrate was visualized with ECL chemiluminescence's reagents as described previously [13], and western blot marker (CWBIO, China) was used as molecular mass standards.

Immunization and challenge

Six groups of BALB/c mice ($n = 12$ each) were individually immunized intramuscularly (i.m.) four times at two weeks interval. The specific immunization schedules and methods for mice are shown in Table 1. Blood samples from each group were collected from the tail vein at the end of each interval, four times in total and sera were separated and stored at -20°C until analyzed for specific antibodies. Two weeks after the last immunization, three mice per group were sacrificed and splenocytes were harvested under aseptic conditions for cytokine assays, and thereafter the remaining mice in all groups were intraperitoneally (i.p.) challenged with 1×10^3 *T. gondii* RH strain tachyzoites suspended in 100 μl phosphate-buffered saline (PBS) to observe the percentage of mice surviving.

Determination of antibodies by ELISA

IgG, IgG1 and IgG2a antibodies in sera samples were determined using an indirect ELISA assay according to the manufacturer's instructions (R&D Systems, USA). In brief, the 96-well microtiter plates were coated over night at 4°C with STAg in 50 mM carbonate buffer (pH 9.6). On the second day, the plates were washed three times with PBS containing 0.05% Tween20 (PBST)

Table 1 Specific immunization regimes of BALB/c mice

Groups ^a	Immunization time			
	1 st	2 nd	3 rd	4 th
PBS	100 µl	100 µl	100 µl	100 µl
pEGFP ^b	100 µg	100 µg	100 µg	100 µg
pGRA4 ^c	100 µg	100 µg	100 µg	100 µg
peptide ^d	100 µg	100 µg	100 µg	100 µg
peptide/DNA	100 µg peptide	100 µg peptide	100 µg pGRA4	100 µg pGRA4
DNA/peptide	100 µg pGRA4	100 µg pGRA4	100 µg peptide	100 µg peptide

^aAll mice of six groups were intramuscularly injected four times at two week interval.

^bEach mouse, 100 µg pEGFP resuspended in 100 µl PBS.

^cEach mouse, 100 µg pGRA4 resuspended in 100 µl PBS.

^dEach mouse, 100 µg peptide resuspended in 100 µl PBS.

and then blocked with 1% Bovine Serum Albumin (BSA) for 1 h at 37°C. Thereafter, the wells were washed and incubated with mice sera diluted in PBS for 1 h at 37°C. HRP-conjugated goat anti-mouse IgG, IgG1 or IgG2a diluted in PBS (Sigma, USA) were used as the secondary antibody and added to the wells for determination of antibody levels and isotype analysis, respectively. Finally, immune complexes were visualized by incubating with orthophenylene diamine (Sigma, USA) and 0.15% H₂O₂ for 30 min. The reaction was stopped by adding 2 M H₂SO₄, and the absorbance was measured at 490 nm with an ELISA reader (BiotekELx800, USA). All samples were run in triplicate.

Cytokine assay

To assay for the levels of cytokine productions, spleens were aseptically removed from three mice per group two weeks after the last injection and splenocytes were obtained as described above and cultured in 96-well microtiter plates. Cell-free supernatants were harvested from microtiter plates and assayed for cytokine levels. The assays were performed for interleukin-4 (IL-4) activity at 24 h, interleukin-10 (IL-10) activity at 72 h, and gamma interferon (IFN-γ) activity at 96 h using a commercial ELISA kit (R&D Systems, USA) following the manufacturer's recommendations. All samples were performed in triplicate.

Statistical analysis

Statistical analysis and graphics were performed using SPSS software. All data, including antibody levels and cytokine productions, were compared between the different groups by one-way ANOVA. Survival time among all the groups was compared using the Kaplan-Meier method. The results in comparisons were considered different if $P < 0.05$.

Ethics statement

This study was approved by the Institutional Animal Care and Use Committee of Shandong University under Contract 2011-0015, and the animals were kept and the experiments were performed in accordance with committee's criteria for the care and use of laboratory animals. All mice were maintained in specific pathogen-free conditions, and all efforts were made to minimize suffering. Humane endpoints to reduce pain or distress in mice were used via euthanasia. Mice were monitored daily for signs of toxoplasmosis, which included difficulties with their food and water intake, lethargy, or severe ascites. Mice with above signs were sacrificed immediately using CO₂ gas. Generally, mice were placed in a chamber and CO₂ was administered at a concentration of 60% to 70% over a 5-minute exposure time, after which the cervical dislocation method was sometimes used to ensure that effective euthanasia had occurred.

Results

Expression of plasmid pGRA4 in HEK293T cell

Forty-eight hours after HEK cells were transfected with pGRA4 or empty plasmid pEGFP, specific green fluorescence was observed under a fluorescence microscope, whereas there was no signal in the untransfected cells (Figure 1A, B & C). In the western blotting analysis (Figure 1D), a specific protein band (about 70 kDa) was detected in cells transfected with pGRA4, but not in cells transfected with empty plasmid or untransfected cells. The fusion protein of about 70 kDa consists of the target protein GRA4 (about 40 kDa) and green fluorescent protein (about 30 kDa). These results indicated that the recombinant plasmid was successfully constructed and expressed *in vitro* and recombinant TgGRA4 protein possessed immunological activity.

Evaluation of the antibody responses by ELISA

To determine the specific antibody titers in all vaccination protocols, the levels of *T. gondii*-specific IgG and IgG subclasses in the sera of mice were assayed by ELISA with STAg as coating antigens. As shown in Figure 2, significantly high levels of IgG antibody appeared following injection of polypeptide, pGRA4, peptide/DNA, and DNA/peptide, as compared to the control groups ($P < 0.05$). As expected, mice injected with PBS and pEGFP did not generate specific antibody response. In all vaccination protocols, the DNA prime-peptide boost regimen leads to remarkable increase in specific IgG antibody production compared to other immunized groups (polypeptide, pGRA4, or peptide/DNA) ($P < 0.05$). However, there was no statistical difference in mice vaccinated with polypeptide, pGRA4, or peptide/DNA ($P > 0.05$).

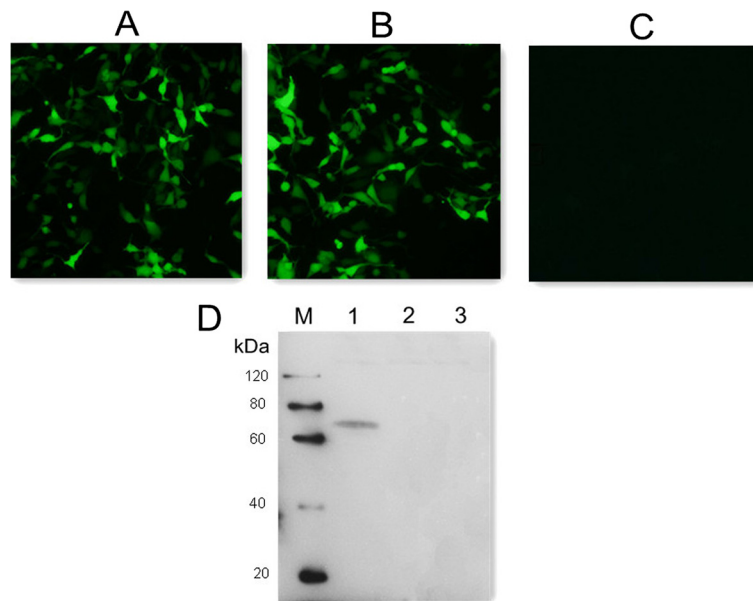


Figure 1 Fluorescence microscopy images of HEK293T cells and western blotting analysis. (A) HEK293 cells transfected with empty vector pEGFP-C1. (B) HEK293 cells transfected with pGRA4. (C) untransfected HEK cells. (D) protein marker (lane M), pGRA4-transfected cells (lane 1), pEGFP-transfected cells (lane 2), untransfected cells (lane 3).

In order to determine whether a Th1 and/or Th2 response was elicited in the immunized mice, we measured by ELISA the specific IgG1 and IgG2a subclasses at the second week after the final immunization. As depicted in Figure 3, an IgG2a predominant production over IgG1

was detected in the sera of vaccinated mice. Apparently, in mice immunized with DNA/peptide, the level of IgG2a was significantly increased ($P < 0.05$), compared with mice immunized with peptide/DNA ($P < 0.05$). This profile of IgG2a over IgG1 suggested that the heterologous DNA

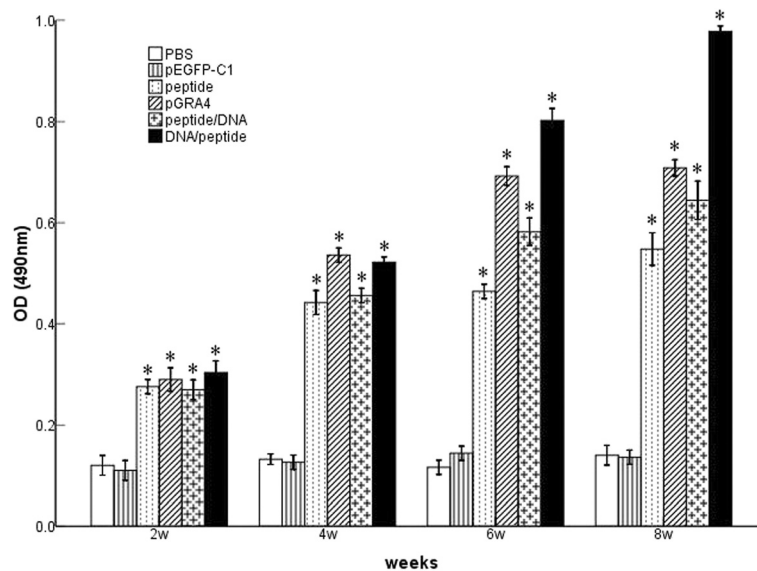


Figure 2 Levels of specific IgG antibodies in BALB/c mice. Sera samples of mice were collected from the tail vein at the end of each interval, four times in total and analyzed by ELISA. Results are shown as mean \pm SD and statistically significant differences ($P < 0.05$) are indicated by (*) as compared to control groups.

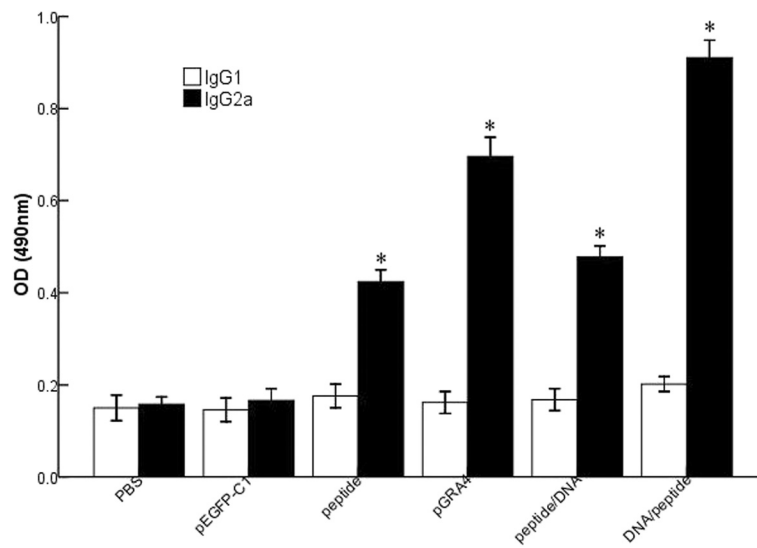


Figure 3 Determination of IgG subclass IgG1 and IgG2a in the sera of BALB/c mice. Sera were taken from mice in all groups two weeks after the last immunization and was analyzed by ELISA. Statistically significant differences ($P < 0.05$) are indicated by an asterisk (*) as compared to control groups.

prime-peptide boost strategy elicited a specific humoral response, and with a Th1-type immune response.

Cytokine production by spleen cells

Culture supernatants of splenocytes from individual mice were harvested two weeks after the final vaccination and were assessed for the production of IFN- γ , IL-4 and IL-10 activities at different times. As shown in Figure 4, large amounts of IFN- γ were detected in the restimulated splenocyte cultures of all the treatment

mice as compared to the control groups ($P < 0.05$). Mice vaccinated with DNA/peptide generated higher level of IFN- γ compared with pGRA4 or polypeptide alone ($P < 0.05$). In the meanwhile, no significant difference was observed in IL-4 and IL-10 levels between these groups ($P > 0.05$). Generally, both Th1 cytokines (IFN- γ and IL-2) and Th2 cytokines (IL-4 and IL-10) are the major parameters to define whether a Th1 and/or Th2 immune response was induced [22]. The high level of IFN- γ and low level of IL-4 and IL-10 suggested that the cellular

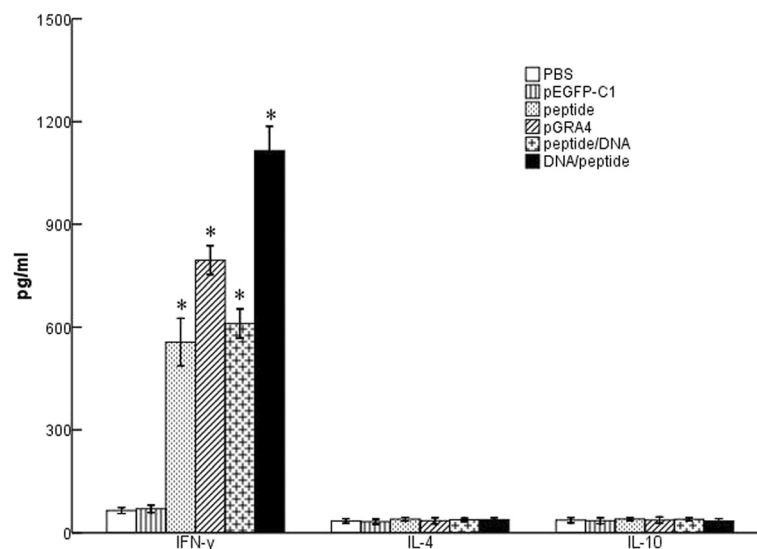


Figure 4 Cytokine productions by spleenocyte cultures from mice. Splenocytes were collected from three mice per group two weeks after the final immunization to detect the level of IFN- γ , IL-4 and IL-10. Data are expressed as mean \pm SD. Statistically significant differences ($P < 0.05$) are indicated by (*) compared with negative control.

immune response induced by the heterologous DNA prime-peptide boost strategy was an enhanced, Th1-biased immunity, further confirming the results of the IgG subclass as shown above.

Evaluation of the survival time in BALB/c mice

To evaluate the protective effect of different vaccination protocols against *T. gondii* infection, mice in all groups were intraperitoneally challenged with 1×10^3 tachyzoites of the virulent RH strain two weeks after last immunization. Survival curves of those different groups of mice are shown in Figure 5. An increased survival time was observed in immunized mice (11.8 ± 4.8 days) compared to the control groups injected with PBS or pEGFP ($P < 0.05$). Mice injected with PBS or pEGFP died within 8 days, and there was no significant difference in the protection level in two groups ($P > 0.05$). Mice immunized with DNA /peptide provide significantly higher protection, with remarkable increase in survival time (16.5 ± 5.4 days), and 40% survival rate was achieved after a lethal challenge, as compare with pGRA4 (11.5 ± 4.1 days), polypeptide (9.0 ± 3.0 days), or peptide /DNA (10.3 ± 3.2 days) ($P < 0.05$). These results demonstrated DNA prime-peptide boost protocol is an effective approach to produce protection against *T. gondii* acute infection.

Discussion

In the present study, we have compared the protective efficacy of different immunization strategies in BALB/c mice. Our results suggested that all the vaccination regimens can trigger a significantly higher level of humoral and cellular immune responses than the control groups

injected with PBS or pEGFP. Especially, the regime with DNA vaccine priming followed by peptide boosting elicited significantly high levels of IgG, IgG2a and IFN- γ secretion, which is an effective approach to provide protection against lethal *T. gondii* infection.

Immunotherapy may be one of the most promising approaches to prevent the infection of *T. gondii*. DNA vaccines encoding TgGRA4 often induce a specific Th1-polarized immune response, but DNA vaccine is often poorly immunogenic, and cannot produce enough protection against *T. gondii* challenge [28,42]. Heterologous prime-boost strategies have emerged as a powerful vaccination approach for inducing very strong cellular immunity, which involve the administration of two different vaccines expressing the same antigen, given several weeks apart [43]. The mechanism of prime-boost regimen is that the first immunization initiates the priming of the immune response and subsequent immunizations trigger further expansion of antigen specific cells and selection of cells with high antigen avidity to boost the specific responses [11]. Recently, several reports have demonstrated the efficacy of heterologous prime-boost vaccination strategies in generating protective immune response to a variety of pathogens, including malaria [44], *Leishmania* [45], and hepatitis C virus [46]. To overcome a significantly lower antibody titer following DNA immunization, prime-boost strategies have been used to increase immune responses to a number of DNA vaccines [47]. Concerning GRA4, DNA prime and a vaccinia virus boost immunization regimen has been evidenced and can induce effective immune responses [35]. Priming with DNA and boosting with polypeptide is another new and promising approach. In our present

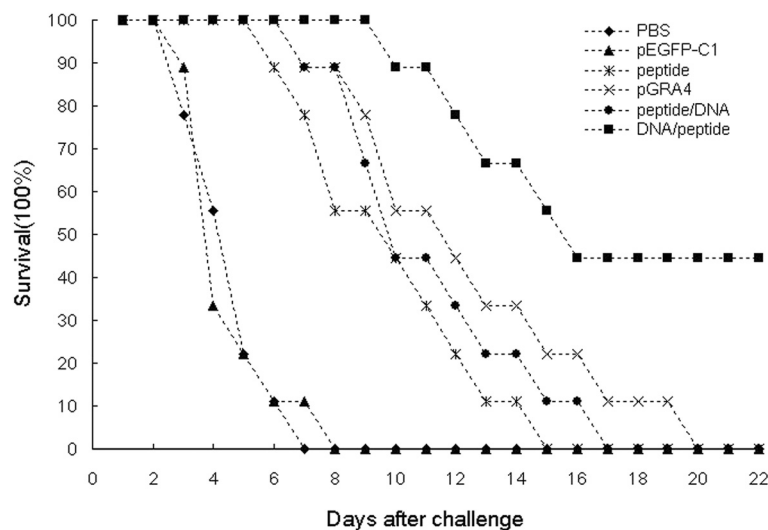


Figure 5 Survival curves of BALB/c mice in all groups after *T. gondii* acute challenge. Each mouse was challenged intraperitoneally with 1×10^3 *T. gondii* RH strain tachyzoites at two weeks after the last immunization. Survival times were monitored daily after the challenge. Each group was composed of nine mice.

study, DNA prime-peptide boost immunization regimen encoding TgGRA4 has been proved to be very effective for eliciting humoral and cellular responses in BALB/c mice.

Humoral and cellular immunity play an important role in host resistance against *T. gondii*. Primary infection with *T. gondii* results in a strong and persistent Th1-type immune response, defined by the high level of IgG antibody response and large amounts of cytokines, which are essential for the control of infection [48,49]. Their findings have suggested that a good immunization protocol should be able to direct the T-helper cells toward a Th1 rather than a Th2-type response [50]. Regarding the roles of humoral immunity, specific IgG antibodies play a protective role in controlling *T. gondii* chronic infection in collaboration with macrophages and preventing reactivation because IgG antibodies can inhibit the attachment of the parasite to the host cell receptors [51]. In this study, only the immunized mice had remarkable levels of serum antigen-specific IgG antibodies, with a high ratio of IgG2a to IgG1 antibody titers. A significant higher level of IgG antibodies response was detected in mice immunized with DNA/peptide as compared with other groups immunized with peptide, pGRA4 or peptide/DNA ($P < 0.05$). These results indicated that specific Th1 cells were mainly activated, which was further confirmed by analysis of the cytokine productions.

To further characterize the polarization of the immune response, the levels of cytokine productions (IFN- γ , IL-4 and IL-10) from spleen cells of mice were evaluated. Generally, cytokines play an important role in host resistance against *T. gondii*. IFN- γ -dependent cell-mediated immunity was important in early infection and control the replication of the protozoan [52]. Meanwhile, IFN- γ is of primary importance in restricting the growth of *T. gondii* in the acute phase of the infection and preventing reactivation of parasites from dormant cysts [53]. Furthermore, a previous study suggested that IFN- γ correlates with the differentiation of Th1 cells and IL-4 induces the development of Th2 cells [54]. Therefore, IFN- γ can be considered as a marker for protective immunity against *T. gondii*. Meanwhile, studies suggested the level of Th2-associated cytokines (IL-4 and IL-10) produced in *T. gondii* infection play an important role in immune response. IL-4 appear relatively late after infection, which perhaps prevents the host from succumbing to early Th1-polarized hyperactive immune response that can be detrimental to the host, and high level of IL-4 was known to antagonize the production of IFN- γ [55,56]. Under our experimental conditions, high level of IFN- γ production was induced in the experimental mice as compared to the controls, whereas the productions of IL-4 and IL-10 were maintained at the same levels

and no significant difference was observed among all the groups. It is obvious that the level of IFN- γ were significantly higher than those of IL-4 and IL-10 especially in DNA/peptide regimen, which suggested the findings are typical of Th1-polarized signal, since the presence of IL-4 is a potent stimulus for Th2 differentiation. Therefore, these findings indicated specific Th1-type immune response was activated in response to TgGRA4, which are consistent with previous studies [17,28].

The main purpose of vaccination is to generate protective immune response against the infection of *T. gondii* and prolong the survival time of mice. In this study, 1×10^3 RH tachyzoites was used to challenge immunized BALB/c mice. The RH strain of *T. gondii* is highly virulent in mice and no previous vaccines have been reported to provide entirely protect against intraperitoneal challenge with *T. gondii* [38,57]. In contrast to the non-immunized mice, a significantly prolonged survival time was obtained in all immunized mice. In addition, the survival rate of the DNA/peptide group reached to 40%, whereas mice in other groups died within 20 days. In all vaccination strategies, DNA/peptide produces an effective and significant degree of protection, and the effective and strong immunity acquired in the vaccination is capable of protecting the mice from *T. gondii* lethal acute challenge infection. Although the immunizations did not provide complete protection against acute challenge with *T. gondii*, the survival time of mice is longer and the survival rate was significantly increased than control groups. In this study, we mainly analyze a Th1-biased response to evaluate the efficiency of vaccine, which is the major limitation of our current study design. The immune response elicited by vaccine in mice was complex, and it is not sufficient to merely analyze a Th1-biased response. In the future study, evaluating the efficiency of vaccine from different aspects could be a new and promising strategy. In addition, many factors, such as *T. gondii* strain, the dose of inoculum, the inoculation route, and the mouse strain, might hinder an efficient evaluation of *T. gondii* vaccines [58]. It is known that intraperitoneal infection with tachyzoites is an unnatural route though it can easily be performed. Thus, in some immunization studies, oral challenge, the natural route of infection, were made with cystogenic strains [59]. Therefore, in our study, many parameters might hinder the effective evaluation of protective immunity. The current study is the lack of a full and systematic evaluation of the immune response generated by different experimental parameters. In further studies, considering different aspects and optimizing challenge route could achieve more encouraging results and effectively evaluate the immune response elicited by *T. gondii* vaccine.

Conclusions

In the present study, we have demonstrated that TgGRA4 can induce significant humoral and cellular Th1 immune responses using a heterologous prime-boost vaccination strategy. The results suggested DNA prime-peptide boost vaccination elicited stronger humoral and cellular immune responses against *T. gondii* as well as increased the survival rate significantly compared with the controls. Therefore, this DNA prime-peptide boost immunization regime based on GRA4 may be an important approach to prevent *T. gondii* infection, particularly with respect to generating an efficient protective immune response.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

SH and AZ conceived and designed the study, and contributed to the revision of the manuscript. MM carried out the experiments and drafted the manuscript. GL and LW participated in mouse immunization and challenge. GZ, YH, HZ, HC, QZ and XQZ helped in study implementation and data collection. All authors read and approved the final manuscript.

Acknowledgments

This work was supported, in part, by grants from the National Natural Science Foundation of China (Grant Nos. 81071373 and 81271857), the State Key Laboratory of Veterinary Etiological Biology (Grant No. SKLVEB2011KFKT005) and the Shandong Provincial Natural Science Foundation (Grant No. ZR2009CM079). XQZ is supported by the National Natural Science Foundation of China (Grant Nos. 31230073, 31172316 and 31101812).

Author details

¹Department of Parasitology, Shandong University School of Medicine, 250012, Jinan, Shandong Province, Peoples Republic of China. ²Department of Pediatrics, Provincial Hospital Affiliated to Shandong University, Shandong University School of Medicine, 250021, Jinan, Shandong Province, Peoples Republic of China. ³State Key Laboratory of Veterinary Etiological Biology, Key Laboratory of Veterinary Parasitology of Gansu Province, Lanzhou Veterinary Research Institute, Chinese Academy of Agricultural Sciences, 730046, Lanzhou, Gansu Province, Peoples Republic of China.

Received: 6 May 2013 Accepted: 14 October 2013

Published: 23 October 2013

References

1. Figueras MJ, Martin OA, Echeverria PC, De Miguel N, Naguleswaran A, Sullivan WJ Jr, Corvi MM, Angel SO: *Toxoplasma gondii* Sis1-like J-domain protein is a cytosolic chaperone associated to HSP90/HSP70 complex. *Int J Biol Macromol* 2012, **50**(3):725–733.
2. Niedelman W, Gold DA, Rosowski EE, Sprockholt JK, Lim D, Farid Arenas A, Melo MB, Spooner E, Yaffe MB, Saeji JP: The rhoptry proteins ROP18 and ROP5 mediate *Toxoplasma gondii* evasion of the murine, but not the human, interferon-gamma response. *PLoS Pathog* 2012, **8**(6):e1002784.
3. Kurokawa H, Kato K, Iwanaga T, Sugi T, Sudo A, Kobayashi K, Gong H, Takemae H, Recuenco FC, Horimoto T, et al: Identification of *Toxoplasma gondii* cAMP dependent protein kinase and its role in the tachyzoite growth. *PLoS One* 2011, **6**(7):e22492.
4. Black MW, Boothroyd JC: Lytic cycle of *Toxoplasma gondii*. *Microbiol Mol Biol Rev* 2000, **64**(3):607–623.
5. Carey KL, Jongco AM, Kim K, Ward GE: The *Toxoplasma gondii* rhoptry protein ROP4 is secreted into the parasitophorous vacuole and becomes phosphorylated in infected cells. *Eukaryot Cell* 2004, **3**(5):1320–1330.
6. Israelski DM, Remington JS: Toxoplasmosis in patients with cancer. *Clin Infect Dis* 1993, **17**(Suppl 2):S423–S435.
7. Havelaar AH, Kemmeren JM, Kortbeek LM: Disease burden of congenital toxoplasmosis. *Clin Infect Dis* 2007, **44**(11):1467–1474.
8. Buxton D, Innes EA: A commercial vaccine for ovine toxoplasmosis. *Parasitology* 1995, **110**(Suppl):S11–S16.
9. Mohamed RM, Aosai F, Chen M, Mun HS, Norose K, Belal US, Piao LX, Yano A: Induction of protective immunity by DNA vaccination with *Toxoplasma gondii* HSP70, HSP30 and SAG1 genes. *Vaccine* 2003, **21**(21–22):2852–2861.
10. Haumont M, Delhaye L, Garcia L, Jurado M, Mazzu P, Daminet V, Verlant V, Bollen A, Biemans R, Jacquet A: Protective immunity against congenital toxoplasmosis with recombinant SAG1 protein in a guinea pig model. *Infect Immun* 2000, **68**(9):4948–4953.
11. Lambrecht-Washington D, Qu BX, Fu M, Anderson LD Jr, Eagar TN, Stuve O, Rosenberg RN: A peptide prime-DNA boost immunization protocol provides significant benefits as a new generation Abeta42 DNA vaccine for Alzheimer disease. *J Neuroimmunol* 2013, **254**(1–2):63–68.
12. Stevenson FK, Rosenberg W: DNA vaccination: a potential weapon against infection and cancer. *Vox Sang* 2001, **80**(1):12–18.
13. Cui X, Lei T, Yang D, Hao P, Li B, Liu Q: *Toxoplasma gondii* immune mapped protein-1 (TgIMP1) is a novel vaccine candidate against toxoplasmosis. *Vaccine* 2012, **30**(13):2282–2287.
14. Yuan ZG, Zhang XX, Lin RQ, Petersen E, He S, Yu M, He XH, Zhou DH, He Y, Li HX, et al: Protective effect against toxoplasmosis in mice induced by DNA immunization with gene encoding *Toxoplasma gondii* ROP18. *Vaccine* 2011, **29**(38):6614–6619.
15. Ciesielski MJ, Kazim AL, Barth RF, Fenstermaker RA: Cellular antitumor immune response to a branched lysine multiple antigenic peptide containing epitopes of a common tumor-specific antigen in a rat glioma model. *Cancer Immunol Immunother* 2005, **54**(2):107–119.
16. Joshi MB, Gam AA, Boykins RA, Kumar S, Sacci J, Hoffman SL, Nakhasi HL, Kenney RT: Immunogenicity of well-characterized synthetic plasmodium falciparum multiple antigen peptide conjugates. *Infect Immun* 2001, **69**(8):4884–4890.
17. Wang Y, Wang M, Wang G, Pang A, Fu B, Yin H, Zhang D: Increased survival time in mice vaccinated with a branched lysine multiple antigenic peptide containing B- and T-cell epitopes from *T. gondii* antigens. *Vaccine* 2011, **29**(47):8619–8623.
18. Mahajan B, Berzofsky JA, Boykins RA, Majam V, Zheng H, Chattopadhyay R, de la Vega P, Moch JK, Haynes JD, Belyakov IM, et al: Multiple antigen peptide vaccines against plasmodium falciparum malaria. *Infect Immun* 2010, **78**(11):4613–4624.
19. El Ridi R, Montash M, Tallima H: Immunogenicity and vaccine potential of dipeptidic multiple antigen peptides from *Schistosoma mansoni* glyceraldehyde 3-phosphate dehydrogenase. *Scand J Immunol* 2004, **60**(4):392–402.
20. Travassos LR, Rodrigues EG, Iwai LK, Taborda CP: Attempts at a peptide vaccine against paracoccidiodomycosis, adjuvant to chemotherapy. *Mycopathologia* 2008, **165**(4–5):341–352.
21. Angus CW, Klivington-Evans D, Dubej JP, Kovacs JA: Immunization with a DNA plasmid encoding the SAG1 (P30) protein of *Toxoplasma gondii* is immunogenic and protective in rodents. *J Infect Dis* 2000, **181**(1):317–324.
22. Golkar M, Shokrgozar MA, Rafati S, Musset K, Assmar M, Sadaie R, Cesbron-Delauw MF, Mercier C: Evaluation of protective effect of recombinant dense granule antigens GRA2 and GRA6 formulated in monophosphoryl lipid a (MPL) adjuvant against *Toxoplasma* chronic infection in mice. *Vaccine* 2007, **25**(21):4301–4311.
23. Mishima M, Xuan X, Yokoyama N, Igarashi I, Fujisaki K, Nagasawa H, Mikami T: Recombinant feline herpesvirus type 1 expressing *Toxoplasma gondii* ROP2 antigen inducible protective immunity in cats. *Parasitol Res* 2002, **88**(2):144–149.
24. Liu MM, Yuan ZG, Peng GH, Zhou DH, He XH, Yan C, Yin CC, He Y, Lin RQ, Song HQ, et al: *Toxoplasma gondii* microneme protein 8 (MIC8) is a potential vaccine candidate against toxoplasmosis. *Parasitol Res* 2010, **106**(5):1079–1084.
25. Mercier C, Adjogbe KD, Daubener W, Delauw MF: Dense granules: are they key organelles to help understand the parasitophorous vacuole of all apicomplexa parasites? *Int J Parasitol* 2005, **35**(8):829–849.
26. Cesbron-Delauw MF: Dense-granule organelles of *Toxoplasma gondii*: their role in the host-parasite relationship. *Parasitol Today* 1994, **10**(8):293–296.
27. Mevelec MN, Chardes T, Mercereau-Pujjalon O, Bourguin I, Achbarou A, Dubremetz JF, Bout D: Molecular cloning of GRA4, a *Toxoplasma gondii* dense granule protein, recognized by mucosal IgA antibodies. *Mol Biochem Parasitol* 1992, **56**(2):227–238.

28. Desolme B, Mevelec MN, Buzoni-Gatel D, Bout D: **Induction of protective immunity against toxoplasmosis in mice by DNA immunization with a plasmid encoding *Toxoplasma gondii* GRA4 gene.** *Vaccine* 2000, **18**(23):2512–2521.
29. Mevelec MN, Bout D, Desolme B, Marchand H, Magne R, Bruneel O, Buzoni-Gatel D: **Evaluation of protective effect of DNA vaccination with genes encoding antigens GRA4 and SAG1 associated with GM-CSF plasmid, against acute, chronic and congenital toxoplasmosis in mice.** *Vaccine* 2005, **23**(36):4489–4499.
30. Chardes T, Bourguin I, Mevelec MN, Dubremetz JF, Bout D: **Antibody responses to *Toxoplasma gondii* in sera, intestinal secretions, and milk from orally infected mice and characterization of target antigens.** *Infect Immun* 1990, **58**(5):1240–1246.
31. Meisel R, Stachelhaus S, Mevelec MN, Reichmann G, Dubremetz JF, Fischer HG: **Identification of two alleles in the GRA4 locus of *Toxoplasma gondii* determining a differential epitope which allows discrimination of type I versus type II and III strains.** *Mol Biochem Parasitol* 1996, **81**(2):259–263.
32. Liu S, Shi L, Cheng YB, Fan GX, Ren HX, Yuan YK: **Evaluation of protective effect of multi-epitope DNA vaccine encoding six antigen segments of *Toxoplasma gondii* in mice.** *Parasitol Res* 2009, **105**(1):267–274.
33. Shedlock DJ, Weiner DB: **DNA vaccination: antigen presentation and the induction of immunity.** *J Leukoc Biol* 2000, **68**(6):793–806.
34. Woodland DL: **Jump-starting the immune system: prime-boosting comes of age.** *Trends Immunol* 2004, **25**(2):98–104.
35. Zhang G, Huong VT, Battur B, Zhou J, Zhang H, Liao M, Kawase O, Lee EG, Dautu G, Igarashi M, *et al*: **A heterologous prime-boost vaccination regime using DNA and a vaccinia virus, both expressing GRA4, induced protective immunity against *Toxoplasma gondii* infection in mice.** *Parasitology* 2007, **134**(Pt 10):1339–1346.
36. Min J, Qu D, Li C, Song X, Zhao Q, Li XA, Yang Y, Liu Q, He S, Zhou H: **Enhancement of protective immune responses induced by *Toxoplasma gondii* dense granule antigen 7 (GRA7) against toxoplasmosis in mice using a prime-boost vaccination strategy.** *Vaccine* 2012, **30**(38):5631–5636.
37. Meng M, He S, Zhao G, Bai Y, Zhou H, Cong H, Lu G, Zhao Q, Zhu XQ: **Evaluation of protective immune responses induced by DNA vaccines encoding *Toxoplasma gondii* surface antigen 1 (SAG1) and 14-3-3 protein in BALB/c mice.** *Parasit Vectors* 2012, **5**:273.
38. Qu DF, Yu HJ, Liu Z, Zhang DF, Zhou QJ, Zhang HL, Du AF: **Ginsenoside Rg1 enhances immune response induced by recombinant *Toxoplasma gondii* SAG1 antigen.** *Vet Parasitol* 2011, **179**(1–3):28–34.
39. Zhang J, Yang JM, Wang HJ, Ru GQ, Fan DM: **Synthesized multiple antigenic polypeptide vaccine based on B-cell epitopes of human heparanase could elicit a potent antimetastatic effect on human hepatocellular carcinoma in vivo.** *PLoS One* 2013, **8**(1):e52940.
40. Cong H, Gu QM, Yin HE, Wang JW, Zhao QL, Zhou HY, Li Y, Zhang JQ: **Multi-epitope DNA vaccine linked to the A2/B subunit of cholera toxin protect mice against *Toxoplasma gondii*.** *Vaccine* 2008, **26**(31):3913–3921.
41. Gong P, Huang X, Yu Q, Li Y, Huang J, Li J, Yang J, Li H, Zhang G, Ren W, *et al*: **The protective effect of a DNA vaccine encoding the *Toxoplasma gondii* cyclophilin gene in BALB/c mice.** *Parasite Immunol* 2013, **35**(3–4):140–146.
42. Chen R, Lu SH, Tong QB, Lou D, Shi DY, Jia BB, Huang GP, Wang JF: **Protective effect of DNA-mediated immunization with liposome-encapsulated GRA4 against infection of *Toxoplasma gondii*.** *J Zhejiang Univ Sci B* 2009, **10**(7):512–521.
43. Kim HD, Jin JJ, Maxwell JA, Fukuchi K: **Enhancing Th2 immune responses against amyloid protein by a DNA prime-adenovirus boost regimen for Alzheimer's disease.** *Immunol Lett* 2007, **112**(1):30–38.
44. Gilbert SC, Schneider J, Hannan CM, Hu JT, Plebanski M, Sinden R, Hill AV: **Enhanced CD8 T cell immunogenicity and protective efficacy in a mouse malaria model using a recombinant adenoviral vaccine in heterologous prime-boost immunisation regimes.** *Vaccine* 2002, **20**(7–8):1039–1045.
45. Gonzalo RM, Del Real G, Rodriguez JR, Rodriguez D, Heljasvaara R, Lucas P, Larraga V, Esteban M: **A heterologous prime-boost regime using DNA and recombinant vaccinia virus expressing the *Leishmania infantum* P36/LACK antigen protects BALB/c mice from cutaneous leishmaniasis.** *Vaccine* 2002, **20**(7–8):1226–1231.
46. Pancholi P, Perkus M, Tricoche N, Liu Q, Prince AM: **DNA immunization with hepatitis C virus (HCV) polycistronic genes or immunization by HCV DNA priming-recombinant canarypox virus boosting induces immune responses and protection from recombinant HCV-vaccinia virus infection in HLA-A2.1-transgenic mice.** *J Virol* 2003, **77**(1):382–390.
47. Alekseeva E, Sominskaya I, Skrastina D, Egorova I, Starodubova E, Kushners E, Mihailova M, Petrakova N, Bruverer R, Kozlovskaya T, *et al*: **Enhancement of the expression of HCV core gene does not enhance core-specific immune response in DNA immunization: advantages of the heterologous DNA prime, protein boost immunization regimen.** *Genet Vaccines Ther* 2009, **7**:7.
48. Gazzinelli RT, Hieny S, Wynn TA, Wolf S, Sher A: **Interleukin 12 is required for the T-lymphocyte-independent induction of interferon gamma by an intracellular parasite and induces resistance in T-cell-deficient hosts.** *Proc Natl Acad Sci USA* 1993, **90**(13):6115–6119.
49. Machado AV, Caetano BC, Barbosa RP, Salgado AP, Rabelo RH, Garcia CC, Bruna-Romero O, Escriou N, Gazzinelli RT: **Prime and boost immunization with influenza and adenovirus encoding the *Toxoplasma gondii* surface antigen 2 (SAG2) induces strong protective immunity.** *Vaccine* 2010, **28**(18):3247–3256.
50. Zheng B, Lu S, Tong Q, Kong Q, Lou D: **The virulence-related rhoptry protein 5 (ROP5) of *Toxoplasma gondii* is a novel vaccine candidate against toxoplasmosis in mice.** *Vaccine* 2013, **31**(41):4578–4584.
51. Kang H, Remington JS, Suzuki Y: **Decreased resistance of B cell-deficient mice to infection with *Toxoplasma gondii* despite unimpaired expression of IFN-gamma, TNF-alpha, and inducible nitric oxide synthase.** *J Immunol* 2000, **164**(5):2629–2634.
52. Sher A, Denkers EY, Gazzinelli RT: **Induction and regulation of host cell-mediated immunity by *Toxoplasma gondii*.** *Ciba Found Symp* 1995, **195**:95–104. discussion 104–109.
53. Suzuki Y, Orellana MA, Schreiber RD, Remington JS: **Interferon-gamma: the major mediator of resistance against *Toxoplasma gondii*.** *Science* 1988, **240**(4851):516–518.
54. Maggi E, Parronchi P, Manetti R, Simonelli C, Piccinni MP, Rugiu FS, De Carli M, Ricci M, Romagnani S: **Reciprocal regulatory effects of IFN-gamma and IL-4 on the in vitro development of human Th1 and Th2 clones.** *J Immunol* 1992, **148**(7):2142–2147.
55. Gazzinelli RT, Wysocka M, Hieny S, Scharton-Kersten T, Cheever A, Kuhn R, Muller W, Trinchieri G, Sher A: **In the absence of endogenous IL-10, mice acutely infected with *Toxoplasma gondii* succumb to a lethal immune response dependent on CD4+ T cells and accompanied by overproduction of IL-12, IFN-gamma and TNF-alpha.** *J Immunol* 1996, **157**(2):798–805.
56. Dawson HD, Beshah E, Nishi S, Solano-Aguilar G, Morimoto M, Zhao A, Madden KB, Ledbetter TK, Dubey JP, Shea-Donohue T, *et al*: **Localized multigene expression patterns support an evolving Th1/Th2-like paradigm in response to infections with *Toxoplasma gondii* and *Ascaris suum*.** *Infect Immun* 2005, **73**(2):1116–1128.
57. Liu Q, Shang L, Jin H, Wei F, Zhu XQ, Gao H: **The protective effect of a *Toxoplasma gondii* SAG1 plasmid DNA vaccine in mice is enhanced with IL-18.** *Res Vet Sci* 2010, **89**(1):93–97.
58. Jongert E, Roberts CW, Gargano N, Forster-Waldl E, Petersen E: **Vaccines against *Toxoplasma gondii*: challenges and opportunities.** *Mem Inst Oswaldo Cruz* 2009, **104**(2):252–266.
59. Hiramoto RM, Galisteo AJ, DoNascimento N, DeAndrade HF Jr: **200 Gy sterilised *Toxoplasma gondii* tachyzoites maintain metabolic functions and mammalian cell invasion, eliciting cellular immunity and cytokine response similar to natural infection in mice.** *Vaccine* 2002, **20**(16):2072–2081.

doi:10.1186/1471-2334-13-494

Cite this article as: Meng *et al*: DNA prime and peptide boost immunization protocol encoding the *Toxoplasma gondii* GRA4 induces strong protective immunity in BALB/c mice. *BMC Infectious Diseases* 2013 **13**:494.