

Extraction, characterization, and anticancer potential of extracellular polymeric substances from marine actinobacteria of *Streptomyces* species

Z. Mohamed Noufal,
Pitchiah Sivaperumal¹, P. Elumalai

Department of Pharmacology, Saveetha Dental College and Hospital, Saveetha Institute of Medical and Technical Sciences, Saveetha University,
¹Cellular and Molecular Research Centre, Marine Biomedical Research Lab and Environmental Toxicology Unit, Saveetha Dental College and Hospital, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai, Tamil Nadu, India

J. Adv. Pharm. Technol. Res.

ABSTRACT

To check the ability of the anticancer activity, the extracted extracellular polymeric substances (EPS) from marine actinobacteria were done. Potential of the anticancer activity of EPS which was extracted from marine actinobacteria of *Streptomyces* species through an assay called MTT. Marine actinobacterial isolation, identification and micromorphology of the strain, and biochemical analysis were performed (Shirling and Gottlieb, 1966). The production of EPS from marine actinobacteria was quantified (P. Sivaperumal *et al.*, 2018). Carbohydrate content in the EPS was quantified, and MCF-7 cell proliferation was done using an MTT assay. EPS-producing marine *Streptomyces* was isolated and identified. The production of EPS and their protein, carbohydrate, lipid, and other parameters were estimated. Further, the EPS showed more than 50% of inhibition after 72 h using the MTT assay in the MCF-7 cancer cell line. The present study exhibited that EPS from marine *Streptomyces* species has significant anticancer activity.

Key words: Eco-friendly, extracellular polymeric substance, green synthesis, innovative technique, innovative technology, *Streptomyces*

INTRODUCTION

Anticancer is employed against or tending to arrest or prevent cancer. The drugs used for preventing cancer are called anticancer drugs which have an anticancer activity toward cancer-causing stimulation and cancer

cells.^[1] To check the ability of the anticancer activity of the extracted extracellular polymeric substances (EPS) from marine actinobacteria of *Streptomyces* species through an assay called MTT.^[2,3] *In vitro*, the MTT assay was the most abundantly used one to accounting the basic anticancer activity of natural products.^[4,5]

Microbial natural polymers such as EPS are huge molecular weight in nature which establish the structural integrity of biofilms and functions. Furthermore, the basic constituent of EPS will define the physical and biochemical properties of biofilm.^[6] Protein and polysaccharides are the major constituents of EPS, followed by lipids, nucleic acids, humic substances, and other molecules.^[7,8] Microbes are used to produce a wide range of polysaccharides that are

Address for correspondence:

Dr. Pitchiah Sivaperumal,
Cellular and Molecular Research Centre, Marine Biomedical Research Lab and Environmental Toxicology Unit, Saveetha Dental College and Hospital, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai - 600 077, Tamil Nadu, India.
E-mail: sivaperumalp.sdc@saveetha.com

Submitted: 09-May-2022

Revised: 30-Jun-2022

Accepted: 09-Jul-2022

Published: 30-Nov-2022

Access this article online

Quick Response Code:



Website:

www.japtr.org

DOI:

10.4103/japtr.japtr_331_22

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

How to cite this article: Noufal ZM, Sivaperumal P, Elumalai P. Extraction, characterization, and anticancer potential of extracellular polymeric substances from marine actinobacteria of *Streptomyces* species. *J Adv Pharm Technol Res* 2022;13:S125-9.

varied in functional and structural basis. These sugar-based substances from microbes are produced in extracellular and intracellular activity.^[9,10] In previous studies, the addition of cyanobacterial EPS can significantly remove heavy metals such as copper, cadmium, and lead and stimulate the biosorption processes in wastewater treatment.^[11]

Melanins are termed macromolecules made by oxidative polymerization consisting of indolic or phenolic compounds.^[12] Regularly, the next shades are earthy colored or dark in shading; however, numerous different tones have additionally been noticed. Melanins are additionally hydrophobic and contrarily charged.^[13] The melanin production is started from L-tyrosine by means of progression an enzymatic as well as nonenzymatic responses through protein tyrosinase. Tyrosinases from various natural sources are used for the amalgamation of (Levodopa and l-3,4-dihydroxyphenylalanine) L-DOPA, and therefore, the expulsion of phenolic compounds from wastewaters.^[14,15] There are three sorts of melanins, for instance, eumelanin, pheomelanin, and allomelanins, and their description is also done in microbes.^[16-18]

Actinobacteria are foremost biotechnologically and commercially important prokaryotes by the contribution of 50% bioactive metabolites out of 23000 overall microbial secondary metabolites.^[19-21] These compounds display an in-depth variety of industrially helpful activities, such as cytotoxic, medicament, antifungal, antiprotozoal, anticancer, medicinal drug, anti-inflammatory drug, anthelmintic, and herbicide.^[22] Our team has done research on biomedical aspects and experiences that have been interpreted to a high level of publications.^[12,23-41] The present study aimed to analyze the anticancer properties of EPS from marine actinobacterium of *Streptomyces* species.

MATERIALS AND METHODS

Sample processing

The marine sediment collection was done around the Parangipettai coast, Tamilnadu, by Van Veen grab. The collected sediments were carefully transferred into a sterile container and reached to the laboratory. After reaching the laboratory, the sample was air-dried for 48 h then sundried for 12 h. The air-dried samples are macerated through mortar and pestle.

Actinobacterial isolation and identification

Actinobacterial was enumerated on KUA (Kuster's agar medium) supplemented with 10 µg/ml of cycloheximide and nalidixic acid as an antibacterial and antifungal agent.^[42] The sediment sample was serially diluted for spread plate on KUA and incubated at ambient temperature for a week. The total population density of actinobacteria from the sediment sample was expressed in colony-forming units per gram. Morphologically, distinct colonies

were picked for pure culture and further analysis. The conventional identification of aerial mass color, melanoid pigments,^[43] reverse side and soluble pigments, spore chain morphology, and utilization of carbon sources was done by International Streptomyces Project (ISP) method. Further, chemotaxonomical characteristics were also done to identify the marine actinobacteria.

Extracellular polymeric substances production and quantification

The marine actinobacterial EPS production was estimated by the method of Sivaperumal *et al.*,^[6] with slight modification.

Estimation of extracellular polymeric substances components

Total carbohydrate in EPS was assessed by the method of phenol sulfuric acid method.^[44] Protein content was done by bicinchoninic assay^[45], and the nucleic acid content was estimated.^[46]

MTT assay

The anti-proliferating activity of marina actinobacteria EPS was analyzed by MTT assay on breast cancer cell line (MSF-7). Morphological observation and statistical analysis also were done, followed by the method of Ciapetti.

RESULTS AND DISCUSSION

Isolation of marine *Streptomyces* species

The present study, conventional identification was done to identify the marine actinobacteria *Streptomyces* species. The chemotaxonomic characteristics such as cell wall analysis, sugar pattern, and cell wall type also have been done to identify the *Streptomyces* species. The spore chain has shown the actinobacteria rectiflexibles. On the basis of assimilation of carbon source, it has shown positive utilization for arabinose, xylose, inositol, fructose, rhamnose, and sucrose also; it has shown negative

Table 1: Conventional findings of marine actinobacteria *Streptomyces* species

Color of aerial mycelium	Grey
Melanoid pigment	-
Reverse side pigment	-
Soluble pigment	-
Spore chain	RF
Assimilation of carbon source	
Arabinose	+
Xylose	+
Inositol	+
Mannitol	-
Fructose	+
Rhamnose	+
Sucrose	+
Raffinose	-

Table 2: Chemotaxonomic characteristic of *Streptomyces* sp.

Cell wall amino acids			Cell wall sugar		Cell wall type	Index
LL-DAP	MesoDAP	Glycine	Arabinose	Galactose		
+	-	+	-	-	I	<i>Streptomyces</i>

Table 3: Extracellular polymeric substance components of marine actinobacteria *Streptomyces* species

EPS Components	Percentage of composition
Carbohydrate	38
Protein	41
Nucleic acid	11
Unidentified	10

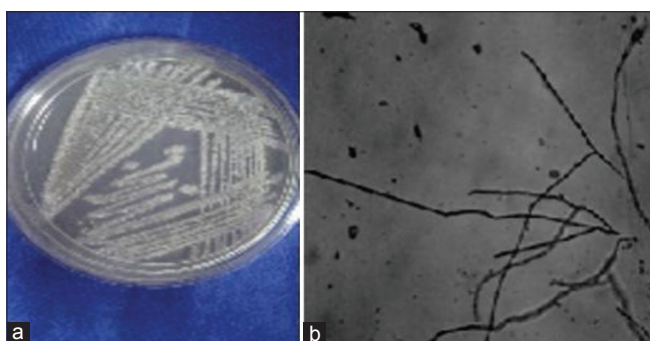


Figure 1: (a and b) Marine *Streptomyces* species strain and their spore chain morphology

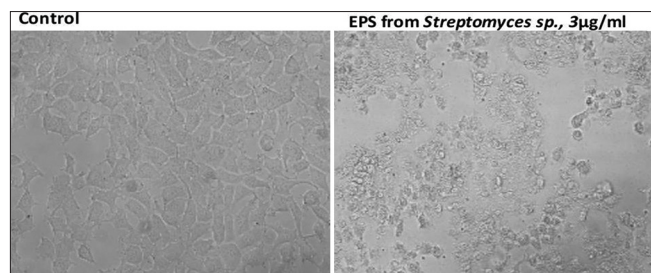


Figure 2: The anticancer potential of EPS obtained from *Streptomyces* sp. against the MCF-7 cell line, the control group used was DMSO. The obtained EPS from *Streptomyces* species was 3 µg/ml. EPS: Extracellular polymeric substances, DMSO: Dimethyl sulfoxide

utilization for mannitol and raffinose [Table 1]. It has been shown positive for LL-DAP and glycine and negative for MesoDAP for cell wall amino acids. In cell wall sugar, both arabinose and galactose were negative. It was found to be a type I cell wall index [Table 2].

Extracellular polymeric substances components

The EPS obtained from *Streptomyces* sp. contains 38% of carbohydrates, 41% of protein, 11% of nucleic acid, and 10% of unidentified compounds [Table 3].

Anticancer potential through MTT assay

Numerous antitumor marine active compounds derived from marine actinobacteria, among them EPS have shown a significant successive part in recognition of bioactive compounds for pharmaceutical use.^[47,48] Actinobacteria from marine sources have numerous potential in providing beneficial leads with specific biological and chemical properties such as anticancer, antimicrobial, antimalarial, antiviral, and anti-inflammatory.^[49] A novel actinobacteria of *Salinispora* has produced a novel compound salinisporamide A actively hinder the growth of cancer cells.^[50]

This current original study has clearly shown that the effect of EPS from marine *Streptomyces* species on the culture plate of cancer cells has displayed a reasonable degree of anticancer potential, causing approximately over inhibition of 50% was observed after 72 h. On using the MCF-7 cell line, the capacity of the confirmed compound has shown almost over 50% of inhibition in 72 h using the MTT assay^[51] [Figures 1a, b and 2].

In a corresponding clinical study, it has been reported that *Streptomyces* species, which are isolated and contaminated from humus soils, the Western Ghats, have exhibited anticancer potentials against the selected cancer cells. Secondary metabolites compounds from actinobacteria, particularly the *Streptomyces* could be the greatest significant marine source for acting as an anticancer reagent and has high anticancer potential.^[52] There are previous studies indicating that the marine actinobacterium of *Streptomyces* species has anticancer potential.^[53]

CONCLUSION

In the current study, we thought of finding the anticancer potential of marine actinobacterial (*Streptomyces* species) EPS. It has revealed that the species have over 50% of inhibition after 72 h at 3 µg/ml using the MCF-7. Hence, we concluded that the EPS from marine *Streptomyces* species has potential anticancer properties, and it might be useful for marine drug research in future.

Acknowledgment

The authors would like to thank Saveetha Dental College and Saveetha Institute of Medical and Technical Sciences for their kind support to utilize the facilities for the study.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Kamala K, Sivaperumal P, Thilagaraj R, Natarajan E. Bioremediation of Sr²⁺-ion radionuclide by using marine *Streptomyces* sp. CuOff24 extracellular polymeric substances. *J Chem Technol Biotechnol* 2019;95:893-903.
- Prakash OM, Kumar A, Kumar P, Ajeet. Anticancer potential of plants and natural products. *Am J Pharmacol Sci* 2013;1:104-15.
- Kittakoop P. Anticancer drugs and potential anticancer leads inspired by natural products. In: *Studies in Natural Products Chemistry*. Edited by Atta-ur-Rahman, Elsevier, Amsterdam, Netherlands; 2015. p. 251-307.
- van Meerloo J, Kaspers GJ, Cloos J. Cell sensitivity assays: The MTT assay. In: Cree IA, editor. *Cancer Cell Culture: Methods and Protocols*. Totowa, NJ: Humana Press; 2011. p. 237-45.
- Ciapetti G, Cenni E, Pratelli L, Pizzoferrato A. *In vitro* evaluation of cell/biomaterial interaction by MTT assay. *Biomaterials* 1993;14:359-64.
- Sivaperumal P, Kamala K, Rajaram R. Adsorption of cesium ion by marine actinobacterium *Nocardopsis* sp. 13H and their extracellular polymeric substances (EPS) role in bioremediation. *Environ Sci Pollut Res Int* 2018;25:4254-67.
- Sheng GP, Yu HQ, Li XY. Extracellular polymeric substances (EPS) of microbial aggregates in biological wastewater treatment systems: A review. *Biotechnol Adv* 2010;28:882-94.
- Wingender J, Neu TR, Flemming HC. What are bacterial extracellular polymeric substances? In: Wingender J, Neu TR, Flemming HC, editors. *Microbial Extracellular Polymeric Substances: Characterization, Structure and Function*. Berlin, Heidelberg: Springer Berlin Heidelberg; 1999. p. 1-19.
- Sutherland IW. Bacterial exopolysaccharides. *Adv Microb Physiol* 1972;8:143-213.
- Sivaperumal P, Kamala K, Rajaram R. Biosorption of long half-life radionuclide of strontium ion (Sr⁺) by marine actinobacterium *Nocardopsis* sp. 13H. *Geomicrobiol J* 2018;35:300-10.
- Carmichael WW. The toxins of cyanobacteria. *Sci Am* 1994;270:78-86.
- Gomathi M, Prakasam A, Rajkumar PV, Rajeshkumar S, Chandrasekaran R, Anbarasan PM. Green synthesis of silver nanoparticles using *Gymnema sylvestre* leaf extract and evaluation of its antibacterial activity. *South Afr J Chem Eng* 2020;32:1-4.
- Perumal S, Thirunavukkarasu AR, Pachiappan P. *Advances in Marine and Brackishwater Aquaculture*. Springer, New Delhi; 2015.
- Lerner AB, Fitzpatrick TB. Biochemistry of melanin formation. *Physiol Rev* 1950;30:91-126.
- Riley PA. Melanin. *Int J Biochem Cell Biol* 1997;29:1235-9.
- Sivaperumal P, Kamala K, Rajaram R. Bioactive DOPA melanin isolated and characterised from a marine actinobacterium *Streptomyces* sp. MVCS6 from Versova coast. *Nat Prod Res* 2015;29:2117-21.
- Meredith P, Sarna T. The physical and chemical properties of eumelanin. *Pigment Cell Res* 2006;19:572-94.
- Anton AH, Sayre DF. The distribution of dopamine and dopa in various animals and a method for their determination in diverse biological material. *Pharmacol Exp Ther* 1964;145:326-36.
- Sivaperumal P, Kamala K, Rajaram R, Mishra SS. Melanin from marine *Streptomyces* sp. (MVCS13) with potential effect against ornamental fish pathogens of *Carassius auratus* (Linnaeus, 1758). *Biocatal Agric Biotechnol* 2014;3:134-41.
- Sharaf MH, El-Sherbiny GM, Moghannem SA, Abdelmonem M, Elsehemy IA, Metwaly AM, *et al.* New combination approaches to combat methicillin-resistant *Staphylococcus aureus* (MRSA). *Sci Rep* 2021;11:4240.
- Oya R, Ikemura K. Can flow cytometrically determined DNA ploidy and S-phase fraction predict regional metastasis in squamous cell carcinoma of the oral cavity? *Head Neck* 2002;24:136-42.
- Thiemann T. Isolation of phthalates and terephthalates from plant material – Natural products or contaminants? *Open Chem J* 2021;8:1-36.
- Rajeshkumar S, Venkat Kumar S, Ramaiah A, Agarwal H, Lakshmi T, Roopan SM. Biosynthesis of zinc oxide nanoparticles using *Mangifera indica* leaves and evaluation of their antioxidant and cytotoxic properties in lung cancer (A549) cells. *Enzyme Microb Technol* 2018;117:91-5.
- Nandhini NT, Rajeshkumar S, Mythili S. The possible mechanism of eco-friendly synthesized nanoparticles on hazardous dyes degradation. *Biocatal Agric Biotechnol* 2019;19:101138.
- Vairavel M, Devaraj E, Shanmugam R. An eco-friendly synthesis of *Enterococcus* sp.-mediated gold nanoparticle induces cytotoxicity in human colorectal cancer cells. *Environ Sci Pollut Res* 2020;27:8166-75.
- Rajasekaran S, Damodharan D, Gopal K, Rajesh Kumar B, De Poures MV. Collective influence of 1-decanol addition, injection pressure and EGR on diesel engine characteristics fueled with diesel/LDPE oil blends. *Fuel* 2020;277:118166.
- Santhoshkumar J, Sowmya B, Venkat Kumar S, Rajeshkumar S. Toxicology evaluation and antidermatophytic activity of silver nanoparticles synthesized using leaf extract of *Passiflora caerulea*. *South Afr J Chem Eng* 2019;29:17-23.
- Kathiswar RR, Ezhilarasan D, Rajeshkumar S. β -Sitosterol-assisted silver nanoparticles activates Nrf2 and triggers mitochondrial apoptosis via oxidative stress in human hepatocellular cancer cell line. *J Biomed Mater Res Part A* 2020;108:1899-908.
- Saravanan M, Arokiyaraj S, Lakshmi T, Pugazhendhi A. Synthesis of silver nanoparticles from *Phanerochaete chrysosporium* (MTCC-787) and their antibacterial activity against human pathogenic bacteria. *Microb Pathog* 2018;117:68-72.
- Gheena S, Ezhilarasan D. Syringic acid triggers reactive oxygen species-mediated cytotoxicity in HepG2 cells. *Hum Exp Toxicol* 2019;38:694-702.
- Ezhilarasan D, Sokal E, Najimi M. Hepatic fibrosis: It is time to go with hepatic stellate cell-specific therapeutic targets. *Hepatobiliary Pancreat Dis Int* 2018;17:192-7.
- Ezhilarasan D. Oxidative stress is bane in chronic liver diseases: Clinical and experimental perspective. *Arab J Gastroenterol* 2018;19:56-64.
- Gomathi AC, Xavier Rajarathinam SR, Mohammed Sadiq A, Rajeshkumar S. Anticancer activity of silver nanoparticles synthesized using aqueous fruit shell extract of *Tamarindus indica* on MCF-7 human breast cancer cell line. *J Drug Deliv Sci Technol* 2020;55:101376.
- Dua K, Wadhwa R, Singhvi G, Rapalli V, Shukla SD, Shastri MD, *et al.* The potential of siRNA based drug delivery in respiratory disorders: Recent advances and progress. *Drug Dev Res* 2019;80:714-30.
- Ramesh A, Varghese S, Jayakumar ND, Malaiappan S. Comparative estimation of sulfiredoxin levels between chronic periodontitis and healthy patients-A case-control study. *J Periodontol* 2018;89:1241-8.
- Arumugam P, George R, Jayaseelan VP. Aberrations of m6A regulators are associated with tumorigenesis and metastasis in head and neck squamous cell carcinoma. *Arch Oral Biol* 2021;122:105030.
- Joseph B, Prasanth CS. Is photodynamic therapy a viable antiviral weapon against COVID-19 in dentistry? *Oral Surg Oral Med Oral Pathol Oral Radiol* 2021;132:118-9.

38. Ezhilarasan D, Apoorva VS, Vardhan NA. Syzygium cumini extract induced reactive oxygen species-mediated apoptosis in human oral squamous carcinoma cells. *J Oral Pathol Med* 2019;48:115-21.
39. Duraisamy R, Krishnan CS, Ramasubramanian H, Sampathkumar J, Mariappan S, Navarasampatti Sivaprakasam A. Compatibility of nonoriginal abutments with implants: Evaluation of microgap at the implant – Abutment interface, with original and nonoriginal abutments. *Implant Dent* 2019;28:289.
40. Gnanavel V, Roopan SM, Rajeshkumar S. Aquaculture: An overview of chemical ecology of seaweeds (food species) in natural products. *Aquaculture* 2019;507:1-6.
41. Markov A, Thangavelu L, Aravindhana S, Zekiy AO, Jarahian M, Chartrand MS, *et al.* Mesenchymal stem/stromal cells as a valuable source for the treatment of immune-mediated disorders. *Stem Cell Res Ther* 2021;12:192.
42. Kathiresan K, Balagurunathan R, Selvam MM. Fungicidal activity of marine actinomycetes against phytopathogenic fungi. *Indian J Biotechnol* 2005;4:271-6.
43. Shirling EB, Gottlieb D. Methods for characterization of *Streptomyces* species. *Int J Syst Evol Microbiol* 1966;16: 313-340.
44. Dubois M, Gilles KA, Hamilton JK, Rebers PA, Smith F. Colorimetric method for determination of sugars and related substances. *Anal Chem* 1956;28:350-6.
45. Smith PK, Krohn RI, Hermanson GT, Mallia AK, Gartner FH, Provenzano MD, *et al.* Measurement of protein using bicinchoninic acid. *Anal Biochem* 1985;150:76-85.
46. Sheng JJ, Georgi D, Lee J. Dynamic flow analysis of probe-type formation tests. All Days 2005. Paper presented at the SPE Annual Technical Conference and Exhibition, Dallas, Texas, October 2005. Paper Number: SPE-96891-MS.
47. Manivasagan P, Venkatesan J, Sivakumar K, Kim SK. Pharmaceutically active secondary metabolites of marine actinobacteria. *Microbiol Res* 2014;169:262-78.
48. Ravikumar S, Fredimoses M, Gnanadesigan M. Anticancer property of sediment actinomycetes against MCF-7 and MDA-MB-231 cell lines. *Asian Pac J Trop Biomed* 2012;2:92-6.
49. Ravikumar S, Gnanadesigan M, Saravanan A, Monisha N, Brindha V, Muthumari S. Antagonistic properties of seagrass associated *Streptomyces* sp. RAUACT-1: A source for anthraquinone rich compound. *Asian Pac J Trop Med* 2012;5:887-90.
50. Manivasagan P, Kang KH, Sivakumar K, Li-Chan EC, Oh HM, Kim SK. Marine actinobacteria: An important source of bioactive natural products. *Environ Toxicol Pharmacol* 2014;38:172-88.
51. Abd-Elnaby H, Abo-Elala G, Abdel-Raouf U, Abd-elwahab A, Hamed M. Antibacterial and anticancer activity of marine *Streptomyces parvus*: Optimization and application. *Biotechnol Biotechnol Equip* 2016;30:180-91.
52. Ahmad MS, El-Gendy AO, Ahmed RR, Hassan HM, El-Kabbany HM, Merdash AG. Exploring the antimicrobial and antitumor potentials of *Streptomyces* sp. AGM12-1 isolated from Egyptian soil. *Front Microbiol* 2017;8:438.
53. Khalifa SA, Elias N, Farag MA, Chen L, Saeed A, Hegazy MF, *et al.* Marine natural products: A source of novel anticancer drugs. *Mar Drugs* 2019;17:E491.