



Complete Genome Sequence of *Acinetobacter pittii* BHS4, Isolated from Air-Conditioning Condensate in Hong Kong

B. H. Saunders,^a G. K. K. Lai,^a  S. D. J. Griffin,^a  F. C. C. Leung^a

^aShuyuan Molecular Biology Laboratory, The Independent Schools Foundation Academy, Hong Kong, Hong Kong SAR, China

ABSTRACT *Acinetobacter pittii* is widespread in the environment, and the *Acinetobacter calcoaceticus-baumannii* complex, to which it belongs, is a major cause of hospital-acquired pneumonia and bacteraemia. *A. pitti* BHS4 was isolated from an air-conditioning unit in Hong Kong and its complete genome sequence (3,901,980 bp; GC content, 38.79%) established through hybrid assembly.

The *Acinetobacter calcoaceticus-baumannii* complex is a major cause of hospital-acquired pneumonia and bacteraemia, with *A. baumannii* designated priority 1 on the WHO Priority Pathogens List (PPL) (1, 2). *Acinetobacter* species are known to accommodate a range of environmental conditions, and multidrug-resistant strains are increasingly being encountered (3–7). Recently, outbreaks of carbapenem-resistant *Acinetobacter* infections have been reported in COVID-19 intensive care unit (ICUs) (8, 9), with contaminated aerosols and ventilation units the likely sources (10, 11). *Acinetobacter pittii* is a Gram-negative, aerobic, non-motile coccobacillus widely distributed in water, soil, and occasionally food stocks (12–16). While *A. baumannii* is the most common species in most regions, *A. pitti* and *A. nosocomialis* appear more prevalent in Southeast Asia (17).

BHS4 was isolated from water dripping from an air-conditioning unit in Mong Kok, Hong Kong. A 100-μl aliquot of the water sample was initially spread onto LB agar and incubated overnight at 37°C. Selected cream-colored colonies were passaged 10 times on LB agar and finally overnight in LB broth before genomic DNA (gDNA) extraction using Invitrogen's PureLink genomic DNA minikit. Paired-end short-read sequencing libraries were prepared using the Nextera XT DNA library preparation kit and sequenced using the Illumina MiSeq platform with v3 chemistry (2 × 300 bp). Adapter sequences were removed using Trimmomatic v0.32 (18) and the reads quality filtered and trimmed. The resulting Illumina data set contained 1,204,982 read pairs with an average length of 297 bp (approximately 358 Mbp). Long-read libraries, prepared from the same extracted DNA using a gDNA rapid barcoding kit (SQK-RBK004), were sequenced using a SpotON flow cell vR9 and MinION sequencer, with data acquisition using MinKNOW v3.1.8 software and base calling using Guppy v2.1.3 (all from Oxford Nanopore). The final long-read data set, trimmed using Porechop v0.2.4 (19, 20), totaled 488,644 reads (3.07 Gbp) with a median length of 15,833 bp (N_{50} , 60,789 bp). Default parameters were used for all software unless otherwise specified.

Assembly of the short reads using Newbler v2.7 (Roche Diagnostics) suggested a draft genome of ~3.85 Mbp, based on 217 contigs at an average depth of 96×. However, a complete genome sequence was generated by combining the Illumina and MinION data sets using Unicycler v0.4.3 (21), yielding a circular chromosome of 3,894,835 bp and a circular plasmid of 7,145 bp, which were submitted to the NCBI PGAP v5.0 for annotation (22).

Multilocus sequence typing (MLST) of the BHS4 chromosome found proximity to *Acinetobacter pittii* IEC338SC (GenBank accession number CP015145.1), with average nucleotide identity of 96.9% (23, 24). In the plasmid, the nucleotide identity of the *repB* gene corresponds with *Acinetobacter baumannii* plasmid homology group GR3 (25) and with the recently defined plasmid lineage LN-16 (26). It encodes the *higBA2* toxin-antitoxin operon,

Citation Saunders BH, Lai GKK, Griffin SDJ, Leung FCC. 2021. Complete genome sequence of *Acinetobacter pittii* BHS4, isolated from air-conditioning condensate in Hong Kong. *Microbiol Resour Announc* 10:e00880-21. <https://doi.org/10.1128/MRA.00880-21>.

Editor David A. Baltrus, University of Arizona

Copyright © 2021 Saunders et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to S. D. J. Griffin, sgriffin@isf.edu.hk, or F. C. C. Leung, fleung@isf.edu.hk.

Received 4 September 2021

Accepted 5 October 2021

Published 21 October 2021

shown to enable a response to environmental stress (27). In antimicrobial susceptibility tests, BHS4 demonstrated resistance to vancomycin and cephalothin (30- μ g disks; Liofilchem) but not to ampicillin, Augmentin, cefepime, or ertapenem, despite the presence of the beta-lactamases OXA-421 and ADC-20 (28).

Data availability. The complete genome sequences and raw sequence data for *A. pittii* BHS4 are available through NCBI under BioProject accession number PRJNA729870, GenBank accession numbers CP075323 (chromosome) and CP075324 (plasmid), and SRA accession numbers SRX11971290 (Illumina MiSeq) and SRX11971291 (MinION).

REFERENCES

- Mancilla-Rojano J, Ochoa SA, Reyes-Grajeda JP, Flores V, Medina-Contreras O, Espinosa-Mazariego K, Parra-Ortega I, Rosa-Zamboni D, Castellanos-Cruz MDC, Arellano-Galindo J, Cevallos MA, Hernández-Castro R, Xicohtencatl-Cortes J, Cruz-Córdova A. 2020. Molecular epidemiology of *Acinetobacter calcoaceticus*-*Acinetobacter baumannii* complex isolated from children at the Hospital Infantil de México Federico Gómez. *Front Microbiol* 11:576673. <https://doi.org/10.3389/fmicb.2020.576673>.
- World Health Organization. 2017. Global priority list of antibiotic-resistant bacteria to guide research, discovery, and development of new antibiotics. https://www.who.int/medicines/publications/WHO-PPL-Short_Summary_25Feb-ET_NM_WHO.pdf. Accessed 27 July 2021.
- Shi J, Sun T, Cui Y, Wang C, Wang F, Zhou Y, Miao H, Shan Y, Zhang Y. 2020. Multidrug resistant and extensively drug resistant *Acinetobacter baumannii* hospital infection associated with high mortality: a retrospective study in the pediatric intensive care unit. *BMC Infect Dis* 20:597. <https://doi.org/10.1186/s12879-020-05321-y>.
- Wisplinghoff H, Paulus T, Lungenheim M, Stefanik D, Higgins PG, Edmond MB, Wenzel RP, Seifert H. 2012. Nosocomial bloodstream infections due to *Acinetobacter baumannii*, *Acinetobacter pittii* and *Acinetobacter nosocomialis* in the United States. *J Infect* 64:282–290. <https://doi.org/10.1016/j.jinf.2011.10.008>.
- Lolans K, Rice TW, Munoz-Price LS, Quinn JP. 2006. Multicity outbreak of carbapenem-resistant *Acinetobacter baumannii* isolates producing the carbapenemase OXA-40. *Antimicrob Agents Chemother* 50:2941–2945. <https://doi.org/10.1128/AAC.00116-06>.
- Chusri S, Chongsuvivatwong V, Rivera JL, Silpapojakul K, Singhamanan K, McNeil E, Doi Y. 2014. Clinical outcomes of hospital-acquired infection with *Acinetobacter nosocomialis* and *Acinetobacter pittii*. *Antimicrob Agents Chemother* 58:4172–4179. <https://doi.org/10.1128/AAC.02992-14>.
- Salto IP, Torres Tejerizo G, Wibberg D, Pühler A, Schlüter A, Pistorio M. 2018. Comparative genomic analysis of *Acinetobacter* spp. plasmids originating from clinical settings and environmental habitats. *Sci Rep* 8:7783. <https://doi.org/10.1038/s41598-018-26180-3>.
- Pascale R, Bussini L, Gaibani P, Bovo F, Fornaro G, Lombardo D, Ambretti S, Pensalfine G, Appolloni L, Bartoletti M, Tedeschi S, Tumietto F, Lewis R, Viale P, Giannella M. 16 April 2021. Carbapenem-resistant bacteria in an intensive care unit during the coronavirus disease 2019 (COVID-19) pandemic: a multi-center before-and-after cross-sectional study. *Infect Control Hosp Epidemiol*: 1–6. <https://doi.org/10.1017/ice.2021.144>.
- Gaibani P, Viciani E, Bartoletti M, Lewis RE, Tonetti T, Lombardo D, Castagnetti A, Bovo F, Horna CS, Ranieri M, Viale P, Re MC, Ambretti S. 2021. The lower respiratory tract microbiome of critically ill patients with COVID-19. *Sci Rep* 11:10103. <https://doi.org/10.1038/s41598-021-89516-6>.
- Mousa M, Schwartz D, Carmeli Y, Nutman A. 2019. Droplet aerosol dissemination of carbapenem-resistant *Acinetobacter baumannii* surrounding ventilated patients. *Infect Control Hosp Epidemiol* 40:365–367. <https://doi.org/10.1017/ice.2018.335>.
- Munoz-Price LS, Fajardo-Aquino Y, Arheart KL, Cleary T, DePascale D, Pizano L, Namias N, Rivera JL, O'Hara JA, Doi Y. 2013. Aerosolization of *Acinetobacter baumannii* in a trauma ICU. *Crit Care Med* 41:1915–1918. <https://doi.org/10.1097/CCM.0b013e31828a39c0>.
- Pailhoriès H, Tiry C, Eveillard M, Kempf M. 2018. *Acinetobacter pittii* isolated more frequently than *Acinetobacter baumannii* in blood cultures: the experience of a French hospital. *J Hosp Infect* 99:360–363. <https://doi.org/10.1016/j.jhin.2018.03.019>.
- Erdem G, Leber A. 2018. *Acinetobacter* species, p 851–853.e2. In Long S, Prober C, Fischer M (ed), *Principles and practice of pediatric infectious diseases*, 5th ed. Elsevier, Amsterdam, The Netherlands. <https://doi.org/10.1016/b978-0-323-40181-4.00149-3>.
- Nemec A, Krizova L, Maixnerova M, van der Reijden TJK, Deschaght P, Passet V, Vaneechoutte M, Brisse S, Dijkshoorn L. 2011. Genotypic and phenotypic characterization of the *Acinetobacter calcoaceticus*-*Acinetobacter baumannii* complex with the proposal of *Acinetobacter pittii* sp. nov. (formerly *Acinetobacter* genomic species 3) and *Acinetobacter nosocomialis* sp. nov. (formerly *Acinetobacter* genomic species 13TU). *Res Microbiol* 162:393–404. <https://doi.org/10.1016/j.resmic.2011.02.006>.
- Malta RCR, Ramos GLDPA, Nascimento JDS. 2020. From food to hospital: we need to talk about *Acinetobacter* spp. *Germs* 10:210–217. <https://doi.org/10.18683/germs.2020.1207>.
- Larcher R, Pantel A, Arnaud E, Sotto A, Lavigne J-P. 2017. First report of cavitary pneumonia due to community-acquired *Acinetobacter pittii*, study of virulence and overview of pathogenesis and treatment. *BMC Infect Dis* 17:477. <https://doi.org/10.1186/s12879-017-2589-0>.
- Lynch JP, III, Zhanell GG, Clark NM. 2017. Infections due to *Acinetobacter baumannii* in the ICU: treatment options. *Semin Respir Crit Care Med* 38: 311–325. <https://doi.org/10.1055/s-0037-1599225>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Wick RR. 2017. Porechop. <https://github.com/rwick/Porechop>.
- Wick RR, Judd LM, Holt KE. 2018. Deepbinner: demultiplexing barcoded Oxford Nanopore reads with deep convolutional neural networks. *PLoS Comput Biol* 14:e1006583. <https://doi.org/10.1371/journal.pcbi.1006583>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaudeau-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. *Nucleic Acids Res* 46:D851–D860. <https://doi.org/10.1093/nar/gkx1068>.
- Alanjary M, Steinke K, Ziemert N. 2019. AutoMLST: an automated Web server for generating multi-locus species trees highlighting natural product potential. *Nucleic Acids Res* 47:W276–W282. <https://doi.org/10.1093/nar/gkz282>.
- Yoon S-H, Ha S-M, Lim J, Kwon S, Chun J. 2017. A large-scale evaluation of algorithms to calculate average nucleotide identity. *Antonie Van Leeuwenhoek* 110:1281–1286. <https://doi.org/10.1007/s10482-017-0844-4>.
- Bertini A, Poirel L, Mugnier PD, Villa L, Nordmann P, Carattoli A. 2010. Characterization and PCR-based replicon typing of resistance plasmids in *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 54:4168–4177. <https://doi.org/10.1128/AAC.00542-10>.
- Salgado-Camargo AD, Castro-Jaimes S, Gutierrez-Rios R-M, Lozano LF, Altamirano-Pacheco L, Silva-Sánchez J, Pérez-Oseguera Á, Volkow P, Castillo-Ramírez S, Cevallos MA. 2020. Structure and evolution of *Acinetobacter baumannii* plasmids. *Front Microbiol* 11:1283. <https://doi.org/10.3389/fmicb.2020.01283>.
- Armalytė J, Jurénas D, Krasauskas R, Čepauskas A, Sužiedėliūnė E. 2018. The *higBA* toxin-antitoxin module from the opportunistic pathogen *Acinetobacter baumannii*—regulation, activity, and evolution. *Front Microbiol* 9:732. <https://doi.org/10.3389/fmicb.2018.00732>.
- McArthur AG, Wagglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, Kalan L, King AM, Kotova K, Morar M, Mulvey MR, O'Brien JS, Pawlowski AC, Piddock LJ, Spanogiannopoulos P, Sutherland AD, Tang I, Taylor PL, Thaker M, Wang W, Yam M, Yu T, Wright GD. 2013. The Comprehensive Antibiotic Resistance Database. *Antimicrob Agents Chemother* 57:3348–3357. <https://doi.org/10.1128/AAC.00419-13>.