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# PERSPECTIVES



# Donor genetic backgrounds contribute to the functional heterogeneity of stem cells and clinical outcomes

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## Abstract

Stable and sustainable stem cell sources for stem cell-based therapies are scarce and a key bottleneck for clinical applications. The regenerative potential of stem cells is usually attributed to several allogeneic or even autologous donor-related factors. Genetic background and epigenetic variations in different individuals may significantly affect the functional heterogeneity of stem cells. Particularly, singlenucleotide polymorphisms (SNPs) have been implicated in diseases with monogenetic or multifactorial and complex genetic etiologies. However, the possible effects of individual SNPs on donor stem cells remain far from fully elucidated. In this Perspective, we will discuss the roles played by donor genetic traits in the functional heterogeneity of induced pluripotent stem cells, mesenchymal stem cells, and hematopoietic stem cells and their implications for regenerative medicine and therapy.

#### KEYWORDS

cellular therapy, clinical translation, genomics, hematopoietic stem cells (HSCs), hematopoietic stem cell transplantation, induced pluripotent stem cells (iPSCs), mesenchymal stem cells (MSCs), stem cells

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Ting Wang and Juan Zhang contributed equally to this study.

# 1 | INTRODUCTION

Rapid advances in stem cell research have brought revolutionary progress to modern biology and medicine. These include the understanding of stem cell function, tissue development and growth, maintenance of tissue homeostasis, tissue regeneration, aging/degeneration, and therapies for diseases that cannot be cured with conventional medicine.<sup>1,2</sup> As of March 2020, the number of stem cell-based registered clinical trials (www.ClinicalTrials.gov) has exceeded 5000, which is greater than any other single therapeutic technique or approach. As expected, the majority of these trials use hematopoietic stem cells (HSCs) and mesenchymal stem cells (MSCs) that can be acquired from various tissue sources.

Several major stem cell types, including ESCs, induced pluripotent stem cells (iPSCs), adult stem cells, and stem cells harvested from the fetus and related tissues (eg, umbilical cord blood, Wharton jelly from the umbilical cord, amnion, and placenta), have been comprehensively described and are being studied or used in the clinic.<sup>3</sup> Whereas ESCs and iPSCs display pluripotent differentiation capabilities, most adult stem cell types, such as HSCs and MSCs derived from the bone marrow or other sources, have limited differentiation potential. Stem cells used in the clinic can be either allogeneic or autologous, depending on donor or tissue source availability. All stem cell types have relative advantages and limitations, and no single cell type meets all the optimal criteria for clinical applications.<sup>4</sup> Thus, a wide variety of parameters should be considered for optimizing therapeutic efficacy while minimizing the potential for serious adverse events. This is particularly important in light of the goal of precision medicine; an initiative and effort that aims to make advances in tailoring medical care to the individual on a genetic and molecular basis.<sup>5</sup>

It is generally thought that major stem cell functions are induced by functional and/or architectural replacement of damaged tissue, paracrine signaling, or cellular mechanisms supporting self-restoration of the diseased tissue or organ. Remarkably, these mechanisms are poorly defined for stem cell-based therapy. In these cases, the modeof-action of stem cells at the genetic and molecular levels should be elaborated for an in-depth understanding of therapeutic mechanisms. Additionally, stem cell functionality and therapeutic efficacy may be influenced by several donor-originated factors such as age, metabolic status, disease condition, genotype, and disease susceptibility-related genetic background.<sup>6-9</sup>

Evidence supporting the importance of donor genetic backgrounds underlying stem cell functional heterogeneity is emerging. In this Perspective, we summarize the current research and discuss the potential contribution of genetic backgrounds in order to advance the field of stem cell-based therapy.

# 2 | GENETIC BACKGROUNDS DRIVE iPSC FUNCTIONAL HETEROGENEITY

Emerging data have demonstrated that human iPSC technology has tremendous potential as cellular models for human diseases. However,

### Significance statement

In the past decades, thousands of genetic variations termed single-nucleotide polymorphisms (SNPs) have been identified, many of which are likely associated with complex human diseases that were previously hypothesized to have other unique genetic drivers. Genetic studies are rapidly being extended to stem cell research and regenerative medicine models. Considering the impact of SNPs in the etiology of diseases, it is reasonable to consider that stem cells carrying disease-associated SNPs should not be transplanted onto the recipients with the same disease.

variable genetic and phenotypic traits could restrict human iPSC applications. One previous study reported that 25 human iPSC lines were constructed from the same set of somatic tissues derived from different individuals.<sup>10</sup> Single-cell RNA-sequencing (RNA-seq) was performed for each cell line and resultant data were evaluated to identify the main source of heterogeneity. These comprehensive experiments indicated that genetic variations between different donors were the primary source of unique transcriptional differences between lines and that genetic background differences accounted for the majority of human iPSC functional heterogeneity. Additionally, genome-wide profiling was carried out on 711 human iPSC lines derived from 301 healthy donors by the Human Induced Pluripotent Stem Cells Initiative.<sup>11</sup> These data suggested that 5% to 46% of the variations in different human iPSC phenotypes, including in the epigenome, transcriptome, proteome, cell differentiation capacity, and cellular morphology, stemmed from genetic differences between individuals. Furthermore, RNA-seg analysis and linear mixed models identified the genetic origins affecting gene expression changes in 317 human iPSC lines derived from 101 donors and suggested that  $\sim$ 50% of genomewide expression variations came from donor individuals.<sup>12</sup> In light of the recent advance of understanding on the variant-to-function issue, that is, the causative role of genetic variants in diseases or phenotypic traits,<sup>13,14</sup> it would be highly speculable that a large proportion of variable iPSC phenotypes are likely a result of particular genetic variations.

# 3 | DISEASE-ASSOCIATED VARIATIONS ORIGINATING IN DONORS MAY CONSTRAIN THE APPLICATION OF MSCs

MSC heterogeneity exists among individuals, ages, tissue sources, cultural and cryopreservative conditions, passages, and even inter- or intracell colonies, resulting in their varied functional ability in vivo, which may affect the MSC use in regenerative medicine and their therapeutic efficacy.<sup>6-9,15</sup> Interestingly, varied ex vivo growth kinetics of MSCs from different donors may also be an important affecting factor for processing and manufacturing cellular products.<sup>16,17</sup> A latest

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study using global transcriptomic analysis revealed the genetic role of glutathione S-transferase theta 1 (GSTT1) gene present/null variations in the growth-capacity of human MSC and GSTT1 was proposed as a novel genomic DNA biomarker for human MSC scalability.<sup>17</sup> In the meanwhile, tremendous efforts have been made to reduce MSC heterogeneity and select certain "pure" or functional subpopulations of MSCs, by using clone selection, cell surface markers, single cell analysis, and other technologies.<sup>18-21</sup> On top of these, optimization and selection of donors by a broad genetic analysis may at least in part benefit therapeutic applications in which selected genetic factors can serve as general indicators of successful stem cell therapy.

Bone, cartilage, and joint surrounding tissues all have a mesenchymal origin. Incidentally, MSCs appear to be the most mature and the widely used stem cell for clinical therapies and research related to bone and cartilage diseases. Among them is osteoarthritis (OA), the most common and severe degenerative disease of the joints.<sup>22</sup> MSC transplantation has successfully been used to treat OA in several animal models and has emerged as one of the most promising methods for OA therapy.<sup>23,24</sup> In addition, more than 50 proof-ofconcept clinical trials using MSCs are registered on www. ClinicalTrials.gov. Many trials involve the intra-articular injection of autologous bone marrow- or adipose-derived MSCs. However, some studies showed that proliferation, chondrogenic differentiation, and adipogenic differentiation capacity of MSCs isolated from OA patients were significantly decreased.<sup>25</sup> These results raise the possibility that the genetic background of OA patient-derived MSCs may impair their therapeutic efficacy. The genetic architecture of OA is complex and may involve hundreds of genes. Growth differentiation factor 5 (GDF5), which is thought to be indispensable for cartilage development and homeostasis, is a susceptibility gene.<sup>26-28</sup> Mutations in GDF5 can lead to severely impaired joints and skeletons in both mice and humans. Single-nucleotide polymorphism (SNP) (rs144383) (+104T/C), which lies in the 5' untranslated promoter region of GDF5, is strongly associated with OA and such a strong correlation between rs143383 and OA exists in multiple ethnic groups.<sup>26</sup> The T allele of rs143383 is more likely to result in a 27% reduction of GDF5 expression relative to C allele.<sup>29</sup> About 80% of patients with OA carry the T allele, while the C allele exerts a lower risk of OA (30-40%). Therefore, a minor but persistent imbalance of GDF5 expression throughout life may render an individual more susceptible to OA.

MSCs originating either from the bone marrow or residing in cartilage tissue are responsible for the homeostasis and regeneration of the cartilage. One may speculate that the precise balance of *GDF5* is a key factor affecting the physiological function of MSCs in cartilage and that rs143383 might be used as a selection parameter for MSC donors. This hypothesis should be tested to determine its effect of on autologous or allogeneic MSC proliferation, chondrogenic differentiation, and repair efficacy. If validated, it would be reasonable to recommend that rs143383 should be accounted for when identifying MSC donors for OA patients. This idea could theoretically be extended to any other SNPs already experimentally proven to exert an adverse/ advantageous effect on MSCs.

Many degenerative or metabolic diseases are classified as complex disease with multiple genetic components,<sup>30,31</sup> such as type 2 diabetes and obesity and neurodegenerative diseases, and have proven to be perfect candidate diseases of stem cell treatment.<sup>32-36</sup> From multiple GWAS projects for years, tens to hundreds of genetic variants have been suggested to possibly attribute to each of these diseases.<sup>37-40</sup> SNPs associated with or adjacent to those key genes with a proven or putative function in regulating specific cell types are particularly attracting intense attention. Such examples include rs7903146 (the TCF7L2 gene) with type 2 diabetes,<sup>40</sup> rs356219 (the SNCA gene) with Parkinson's disease, 35,37,39 and SNP rs2075650 (the TOMM40 gene) and rs4420638 (the APOE gene) with Alzheimer's disease.<sup>38</sup> Therefore, considering the importance of SNPs in the etiology of many complex and multifactorial diseases, it is reasonable for us to recommend that stem cells carrying known disease-causing or susceptibility SNPs should not be transplanted into recipients to treat the same disease. Such a recommendation of course does not mean to neglect many other sources of MSC heterogeneity and the top task would still be the optimization of stem cell dose, the route of cell administration, and the selection of most suitable disease stages, in order to obtain the optimal clinical outcomes.

# 4 | CRUCIAL ROLES OF SNP-ASSOCIATED HSC HETEROGENEITY IN CLINICAL OUTCOMES

Hematopoietic stem cell transplantation (HSCT), either allogeneic or autologous, remains the most commonly applied clinical stem cellbased therapy that is broadly used to treat and even cure leukemias and other disorders of the blood and immune system.<sup>41,42</sup> In most cases, allogeneic transplantation may be the only available choice. The outcome of HSCT strongly depends on the quality of the human leukocyte antigen (HLA) match between the donor and recipient identified by high-resolution HLA genotype and match techniques. Transplantation with suboptimally matched donor stem cells exerts an unacceptably high risk of immune rejection and other life-threatening complications, such as disease relapse, graft vs host disease, and viral infection.

Despite stringent procedures to secure the best HLA matching, life-threatening complications continue to occur following HSCT. Several studies have found that SNPs in genes related to innate and adaptive immune responses, residual leukemia, allo-antigens, and drug metabolism are associated with HSCT outcomes and should be applied to identify patients at risk for post-HSCT complications.<sup>43-45</sup> A recent cohort analysis of 1157 HLA-matched cases found that patients with donors homozygous for the C variant of rs10912564 in the key costimulatory molecule TNFSF4 had better disease-free survival, overall survival with less treatment-related mortality.<sup>46</sup> The study further demonstrated that the TNFSF4C variant had a higher affinity for the transcription factor Myb and increased percentage of TNFSF4-positive B cells compared with CT or TT genotypes, suggesting that the TNFSF4C variant may associate with outcomes. 1498

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The CCR5 coreceptor on CD4<sup>+</sup> T cells is critical for R5-tropic HIV viral entry. In 2009, the "Berlin patient" presented with relapsed acute myelogenous leukemia and human immunodeficiency virus (HIV-1) symptoms and underwent allogeneic HSCT derived from a donor lacking the CCR5 coreceptor (homozygous for the  $\Delta$ 32 deletion).<sup>47</sup> Since then, HIV in the patient's blood and tissues has remained undetectable and antiretroviral therapy has been halted. Ten years later, a second individual (the "London patient") with HIV-1 infection and concomitant leukemia was cured by allogeneic HSCT with stem cells bearing the CCR5 $\Delta$ 32/ $\Delta$ 32 mutation.<sup>48</sup> With the aim of mimicing these results while avoiding ethics and safety issues, the CRISPR/ Cas9 gene editing technology was recently adopted to edit CCR5 in HSCs.<sup>49</sup> The approach is expected to benefit more HIV patients and produce more "Berlin patients" and "London patients" in the future.

# 5 | CONCLUSIONS

Stem cell and genomics research are two tightly connected areas, both of which advance rapidly and can provide perfect opportunities and examples for precision and personalized therapies. New technologies and facilities are now readily accessible in hospitals and laboratories and enable the goal. These include the availability of multiple stem cell sources, extensive next-generation sequencing, as well as efficient and safe gene modification techniques.

Broad or even whole genome sequencing-based gene testing has nowadays become acceptable more than ever before.<sup>50</sup> Knowing the genetic traits may not only be benefit recipients but also a long-term interest for donors. This may however be a dilemma considering that there may be potential ethical issues regarding the disclosure of personal information and that additional procedures are taken when screening a particular genetic background of donors. In addition, for most species and cell types, the amount of genomics data keeps accumulating daily and has reached multi-Gigabyte levels. It would be ideal to assign functional significance to each genetic variation identified in a clinical context for a specific tissue or cell type. This requires functional assays in animal or stem cell models in addition to populationlevel replication studies in designated clinical settings. As multiple SNPs are implicated in most clinical conditions, it would be necessary to establish a high-throughput screening assay to assess the functional significance of a large number of genetic variants. For these purposes, CRISPR/Cas9 editing in iPSCs may serve as a good experimental model system to define the contribution of a gene or its variant to cell differentiation and regeneration.

Ultimately, any formal recommendations regarding the impact of SNPs carried by donor stem cells on the outcomes of stem cell therapy will require investigation in patients in the context of clinical trials, as performed for HSCT. Furthermore, we should acknowledge that attributing interdonor variations solely to genetics is difficult, given that different donors will have confounding variables such as unmatched lifestyle choices and experiences. Critics may argue that it is hard enough to find a match based on HLA, let alone including additional genetic criteria. Research is just beginning to address these questions and we think that the benefits to improved therapeutic outcome following stem-cell transplantation will far outweigh these concerns.

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## CONFLICT OF INTEREST

The authors declared no potential conflicts of interest.

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T.W., J.Z.: conception and design, manuscript writing; J.Q.L.: reference selection and manuscript drafting; G.Q.Z., F.Z.: conception and design, manuscript writing, financial support, and final approval of the manuscript.

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#### REFERENCES

- Chandel NS, Jasper H, Ho TT, Passegué E. Metabolic regulation of stem cell function in tissue homeostasis and organismal ageing. *Nat Cell Biol.* 2016;18(8):823-832.
- Knoepfler PS. From bench to FDA to bedside: US regulatory trends for new stem cell therapies. *Adv Drug Deliv Rev.* 2015;82-83: 192-196.
- Zakrzewski W, Dobrzyński M, Szymonowicz M, Rybak Z. Stem cells: past, present, and future. Stem Cell Res Ther. 2019;10(1):68.
- Varghese J, Griffin M, Mosahebi A, Butler P. Systematic review of patient factors affecting adipose stem cell viability and function: implications for regenerative therapy. *Stem Cell Res Ther.* 2017;8 (1):45.
- Sankar PL, Parker LS. The Precision Medicine Initiative's All of Us Research Program: an agenda for research on its ethical, legal, and social issues. *Genet Med.* 2017;19(7):743-750.
- Huang S, Leung V, Peng S, et al. Developmental definition of MSCs: new insights into pending questions. *Cell Reprogram.* 2011;13(6): 465-472.
- Liu Y, Muñoz N, Bunnell BA, et al. Density-dependent metabolic heterogeneity in human mesenchymal stem cells. *STEM CELLS*. 2015;33(11): 3368-3381.
- Prall WC, Saller MM, Scheumaier A, et al. Proliferative and osteogenic differentiation capacity of mesenchymal stromal cells: influence of harvesting site and donor age. *Injury*. 2018;49(8):1504-1512.
- Widman A, Reshef R. Precision in donor selection: identifying ideal stem-cell donors through their T cells. *Exp Hematol.* 2016;44(11): 1020-1023.
- Rouhani F, Kumasaka N, de Brito MC, Bradley A, Vallier L, Gaffney D. Genetic background drives transcriptional variation in human induced pluripotent stem cells. *PLoS Genet*. 2014;10(6):e1004432.
- Kilpinen H, Goncalves A, Leha A, et al. Common genetic variation drives molecular heterogeneity in human iPSCs. *Nature*. 2017;546 (7658):370-375.
- Carcamo-Orive I, Hoffman GE, Cundiff P, et al. Analysis of transcriptional variability in a large human iPSC library reveals genetic and non-genetic determinants of heterogeneity. *Cell Stem Cell*. 2017;20 (4):518-532.

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- Nandakumar SK, Liao X, Sankaran VG. In the blood: connecting variant to function in human hematopoiesis. *Trends Genet*. 2020;36(8): 563-576.
- 14. Shastry BS. SNPs: impact on gene function and phenotype. *Methods Mol Biol*. 2009;578:3-22.
- 15. Rennerfeldt DA, Van Vliet KJ. Concise review: when colonies are not clones: evidence and implications of Intracolony heterogeneity in mesenchymal stem cells. *STEM CELLS*. 2016;34(5):1135-1141.
- Detela G, Bain OW, Kim HW, et al. Donor variability in growth kinetics of healthy hMSCs using manual processing: considerations for manufacture of cell therapies. *Biotechnol J.* 2018;13(2). https://doi. org/10.1002/biot.201700085.
- Sathiyanathan P, Samsonraj RM, Tan CLL, et al. A genomic biomarker that identifies human bone marrow-derived mesenchymal stem cells with high scalability. *STEM CELLS*. 2020. https://doi.org/10.1002/stem. 3203.
- McLeod CM, Mauck RL. On the origin and impact of mesenchymal stem cell heterogeneity: new insights and emerging tools for single cell analysis. *Eur Cell Mater*. 2017;34:217-231.
- Rosu-Myles M, McCully J, Fair J, et al. The globoseries glycosphingolipid SSEA-4 is a marker of bone marrow-derived clonal multipotent stromal cells in vitro and in vivo. *Stem Cells Dev.* 2013;22 (9):1387-1397.
- Stüdle C, Occhetta P, Geier F, Mehrkens A, Barbero A, Martin I. Challenges toward the identification of predictive markers for human mesenchymal stromal cells chondrogenic potential. STEM CELLS TRANSLA-TIONAL MEDICINE. 2019;8(2):194-204.
- Wilson NK, Kent DG, Buettner F, et al. Combined single-cell functional and gene expression analysis resolves heterogeneity within stem cell populations. *Cell Stem Cell*. 2015;16(6):712-724.
- 22. Cushnaghan J, Dieppe P. Study of 500 patients with limb joint osteoarthritis. I. Analysis by age, sex, and distribution of symptomatic joint sites. *Ann Rheum Dis.* 1991;50(1):8-13.
- Horie M, Choi H, Lee RH, et al. Intra-articular injection of human mesenchymal stem cells (MSCs) promote rat meniscal regeneration by being activated to express Indian hedgehog that enhances expression of type II collagen. Osteoarthr Cartil. 2012;20(10):1197-1207.
- 24. Jo CH, Lee YG, Shin WH, et al. Intra-articular injection of mesenchymal stem cells for the treatment of osteoarthritis of the knee: a proof-of-concept clinical trial. *STEM CELLS*. 2014;32(5):1254-1266.
- Murphy JM, Dixon K, Beck S, Fabian D, Feldman A, Barry F. Reduced chondrogenic and adipogenic activity of mesenchymal stem cells from patients with advanced osteoarthritis. *Arthritis Rheum*. 2002;46(3): 704-713.
- Miyamoto Y, Mabuchi A, Shi D, et al. A functional polymorphism in the 5' UTR of GDF5 is associated with susceptibility to osteoarthritis. *Nat Genet*. 2007;39(4):529-533.
- Southam L, Rodriguez-Lopez J, Wilkins JM, et al. An SNP in the 5'-UTR of GDF5 is associated with osteoarthritis susceptibility in Europeans and with in vivo differences in allelic expression in articular cartilage. *Hum Mol Genet*. 2007;16(18):2226-2232.
- Valdes AM, Evangelou E, Kerkhof HJ, et al. The GDF5 rs143383 polymorphism is associated with osteoarthritis of the knee with genomewide statistical significance. *Ann Rheum Dis.* 2011;70(5):873-875.
- Syddall CM, Reynard LN, Young DA, Loughlin J. The identification of trans-acting factors that regulate the expression of GDF5 via the osteoarthritis susceptibility SNP rs143383. *PLoS Genet*. 2013;9(6): e1003557.
- Cano-Gamez E, Trynka G. From GWAS to function: using functional genomics to identify the mechanisms underlying complex diseases. *Front Genet*. 2020;11:424.
- Claussnitzer M, Cho JH, Collins R, et al. A brief history of human disease genetics. *Nature*. 2020;577(7789):179-189.
- Kang JM, Yeon BK, Cho SJ, Suh YH. Stem cell therapy for Alzheimer's disease: a review of recent clinical trials. J Alzheimers Dis. 2016;54(3): 879-889.

- Mendes Filho D, Ribeiro PDC, Oliveira LF, et al. Therapy with mesenchymal stem cells in Parkinson disease: history and perspectives. *Neurologist*. 2018;23(4):141-147.
- Moreira A, Kahlenberg S, Hornsby P. Therapeutic potential of mesenchymal stem cells for diabetes. J Mol Endocrinol. 2017;59(3): R109-R120.
- Oh SH, Lee SC, Kim DY, et al. Mesenchymal stem cells stabilize axonal transports for autophagic clearance of α-Synuclein in Parkinsonian models. STEM CELLS. 2017;35(8):1934-1947.
- Zhang Y, Chen W, Feng B, Cao H. The clinical efficacy and safety of stem cell therapy for diabetes mellitus: a systematic review and metaanalysis. *Aging Dis.* 2020;11(1):141-153.
- Edwards TL, Scott WK, Almonte C, et al. Genome-wide association study confirms SNPs in SNCA and the MAPT region as common risk factors for Parkinson disease. Ann Hum Genet. 2010;74(2):97-109.
- Horwitz T, Lam K, Chen Y, Xia Y, Liu C. A decade in psychiatric GWAS research. *Mol Psychiatry*. 2019;24(3):378-389.
- Kalia LV, Lang AE. Parkinson's disease. Lancet. 2015;386(9996): 896-912.
- Xue A, Wu Y, Zhu Z, et al. Genome-wide association analyses identify 143 risk variants and putative regulatory mechanisms for type 2 diabetes. *Nat Commun.* 2018;9(1):2941.
- Gonsalves WI, Buadi FK, Ailawadhi S, et al. Utilization of hematopoietic stem cell transplantation for the treatment of multiple myeloma: a Mayo Stratification of Myeloma and Risk-Adapted Therapy (mSMART) consensus statement. *Bone Marrow Transplant*. 2019;54 (3):353-367.
- 42. Singh AK, McGuirk JP. Allogeneic stem cell transplantation: a historical and scientific overview. *Cancer Res.* 2016;76(22):6445-6451.
- Ayuk F, Beelen DW, Bornhäuser M, et al. Relative impact of HLA matching and non-HLA donor characteristics on outcomes of allogeneic stem cell transplantation for acute myeloid leukemia and myelodysplastic syndrome. *Biol Blood Marrow Transplant*. 2018;24 (12):2558-2567.
- Dickinson AM, Norden J. Non-HLA genomics: does it have a role in predicting haematopoietic stem cell transplantation outcome? Int J Immunogenet. 2015;42(4):229-238.
- Ritari J, Hyvärinen K, Koskela S, et al. Genomic prediction of relapse in recipients of allogeneic haematopoietic stem cell transplantation. *Leukemia*. 2019;33(1):240-248.
- 46. Jindra PT, Conway SE, Ricklefs SM, et al. Analysis of a genetic polymorphism in the costimulatory molecule TNFSF4 with hematopoietic stem cell transplant outcomes. *Biol Blood Marrow Transplant*. 2016;22 (1):27-36.
- Hutter G, Nowak D, Mossner M, et al. Long-term control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation. N Engl J Med. 2009;360(7):692-698.
- Gupta RK, Abdul-Jawad S, McCoy LE, et al. HIV-1 remission following CCR5Delta32/Delta32 haematopoietic stem-cell transplantation. *Nature*. 2019;568(7751):244-248.
- Xu L, Wang J, Liu Y, et al. CRISPR-edited stem cells in a patient with HIV and acute lymphocytic leukemia. N Engl J Med. 2019;381(13): 1240-1247.
- Marzuillo C, De Vito C, D'Andrea E, et al. Predictive genetic testing for complex diseases: a public health perspective. QJM. 2014;107(2): 93-97.

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