

REVIEW LETTER

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# The role of macrophage subtypes and exosomes in immunomodulation

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## Abstract

Macrophages are influential members of the innate immune system that can be reversibly polarized by different microenvironment signals. Cell polarization leads to a wide range of features, involving the migration, development, and organization of the cells. There is mounting evidence that macrophage polarization plays a key role in the initiation and development of a wide range of diseases. This study aims to give an overview of macrophage polarization, their different subtypes, and the importance of alternatively activated M2 macrophage and classically activated M1 macrophage in immune responses and pathological conditions. This review provides insight on the role of exosomes in M1/M2-like macrophage polarization and their potential as a promising therapeutic candidate.

**Keywords:** Exosomes, Inflammation, Macrophages, Macrophage polarization

## Introduction

Macrophages and their phagocytosis activity were first discovered by Elie Metchnikoff and Paul Ehrlich in 1908 [1–3]. The mononuclear phagocyte system (MPS) is a professional phagocyte that comprises dendritic cells (DCs), blood monocytes, and tissue macrophages [4]. Macrophages are mononuclear cells that are the most plentiful and widespread immune cells and are involved in phagocytosis, homeostasis, and remodeling after injury and are necessary during organ development [5–7]. Macrophages can originate from local tissue-resident macrophages with a self-renewed ability or blood monocytes. Bone marrow progenitor-derived monocytes migrate to tissue by receiving stimuli signals and then become macrophages [4, 8]. Macrophages have plastic characteristics. These characteristics enable them to switch their phenotypes and functions in connection with various microenvironmental signals [9].

Macrophages have been observed in many tissues. They are categorized on the basis of their location and function, for instance, microglial cells in the central nervous system (CNS) with the ability to clear defective neurons; alveolar macrophages in the lung, which are needed for lung homeostasis; osteoclasts in bone with bone remodeling activity [10–12]; and Kupffer cells, which are the most lavish macrophages present in liver [13–15]. Macrophages are innate immune cells that can affect a variety of processes,



including tissue repair, angiogenesis, and immunomodulation [16, 17]. Macrophages gain different phenotypes and functions under normal condition or during disease. Macrophages' ability to change their functions in response to different signals is known as polarization, which is a multifactorial process [18]. This is a key mediator of different diseases, including autoimmune diseases [19], glycolipid metabolic disorders [20], neurology disorders [21], cardiovascular diseases [3], and cancers [22]. Different polarized macrophages, M1 (classically activated macrophages) and M2 (alternatively activated macrophages), express diverse cell surface markers and factors (Table 1) [23].

The polarization of macrophages is a dynamic process, so it can be reversed by different microenvironment conditions in different biological conditions and diseases. Macrophages can polarize with more than two forms [24]. M2 macrophage, also called activated or healing macrophage, was first described in the 1990s [25]. Subsets of M2 macrophages include M2a, M2b, M2c, and M2d, with different properties such as cell markers, cytokines, and functions, i.e., M2a enhances cell growth and tissue repair while M2b, M2c, and M2d play crucial roles in inflammation, phagocytosis, and tumor progression, respectively [16, 26]. M2d macrophages are the main element of tumor microenvironment and are generally called tumor-associated macrophage (TAM). They could promote cancer-related processes such as progression and invasion of cancerous

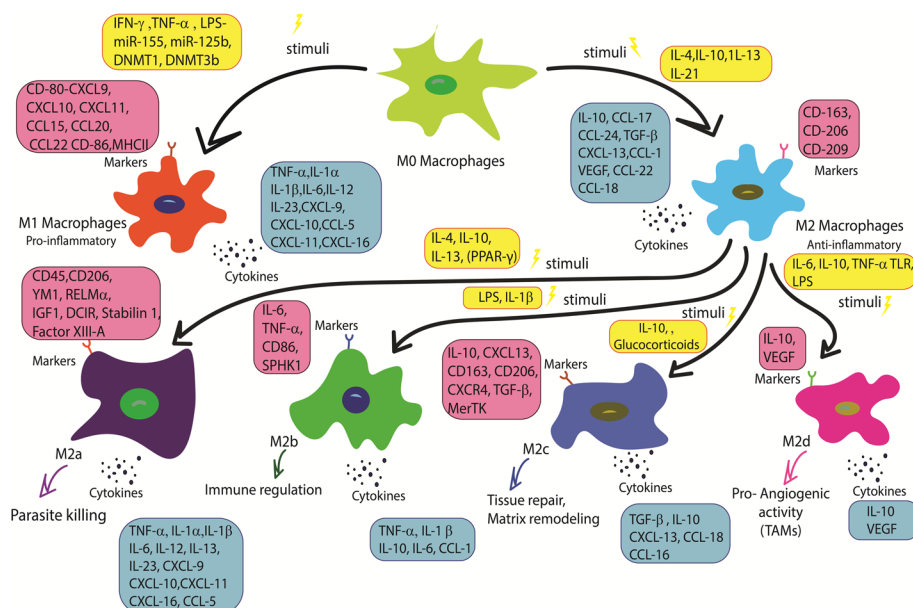
**Table 1** Macrophage subtypes and their characteristic markers and stimuli

Macrophage types	Suggested roles	Markers	Different stimulator factors	References
M1	Pro-inflammation, microbicidal effect, tumor resistance	IL-6, IL-10 (low), IL-12 (high), iNOS, CD80, CD86, CXCL9, CXCL10, CXCL11, CCL15, CCL20, CCL22, TLR2, TLR4, MHCII, TNF- $\alpha$	IL-1, IL-6, IL-12, CXCL1-3, CXCL-5, CXCL8-10, CCL2, Type I IFN, IFN- $\gamma$ , TNF- $\alpha$ , STAT1, iNOS, LPS, M-CSF, NF- $\kappa$ B, IRF5 miR-155, miR-125b, DNMT1, DNMT3b, HDAC3	[19, 28–31]
M2a	Allergy, profibrotic, anti-inflammatory, wound healing	IL-10, IL-1R, IL-27R $\alpha$ , CCL1, CCL17, CCL18, CCL22, CD11b, CD45, CD206, YM1, RELM $\alpha$ , IGF1, DCIR, Stabilin 1, Factor XIII-A, Ly6C, TREM-2, DC-SIGN	IL-4, IL-10, IL-13, (PPAR- $\gamma$ )	[19, 28, 32–35]
M2b	Th2 activation, immune regulation, promoting infection, tumor progression	IL-6, TNF- $\alpha$ , CD86, SPHK1	IL-1 $\beta$ , LPS	[28, 31, 36]
M2c	Immunosuppression, phagocytosis, tissue repair, matrix remodeling	IL-10, CXCL13, CD163, CD206, CXCR4, TGF- $\beta$ , MerTK,	IL-10, glucocorticoids IL-6, IL-10, TNF- $\alpha$ , TLR,	[28, 36, 37]
M2d	Tumor progression, angiogenesis, clearance of apoptotic tissue	IL-10, VEGF, TGF- $\beta$ ,	LPS	[28, 36, 38]

CCL, C–C chemokine ligand; CXCL, C–X–C chemokine ligand; CXCR, C–X–C chemokine receptor; DCIR, dendritic cell immunoreceptor; IFN, interferon; IFN $\gamma$ , interferon- $\gamma$ ; IL, interleukin; IL-1R, IL-1 receptor; IL-27R $\alpha$ , IL-27 receptor  $\alpha$ -chain; iNOS, inducible nitric-oxide synthase; RELM $\alpha$ , resistin-like molecule- $\alpha$ ; SPHK1, sphingosine kinase 1; TLR, Toll-like receptor; CD, cluster of differentiation; TGF- $\beta$ , transforming growth factor- $\beta$ ; STATs, signal transducer and activator of transcription; PPAR- $\gamma$ , peroxisome proliferator-activated receptor gamma; TNF- $\alpha$ , tumor necrotic factor- $\alpha$ ; YM1, chitinase-like protein 3; LPS, lipopolysaccharides; VEGF, vascular endothelial growth factor; ICAM, intercellular adhesion molecule; DC-SIGN, dendritic cell-specific ICAM-grabbing non integrin; IRF5, interferon regulatory factor 5; HDAC3, histone deacetylase 3; MerTK, myeloid epithelial reproductive tyrosine kinase; M-CSF, macrophage-colony-stimulating factor; DNMT, DNA methyl transferase; miRs, microRNAs; NF- $\kappa$ B, nuclear factor- $\kappa$ B; MHCII, major histocompatibility complex II; IGF, insulin-like growth factor; Ly6C, lymphocyte antigen 6 complex; TREM-2, triggering receptor expressed on myeloid cells 2

cells (Fig. 1) [27, 28]. There is a dynamic balance between different types of macrophages that could cause a variety of diseases when it is disturbed. The number of tissue-resident macrophages is regulated by colony-stimulating factor-1 (CSF-1) or macrophage-colony-stimulating factor (M-CSF), interleukin-34 (IL-34), and colony-stimulating factor-1 receptor (CSF-1R) [4]. M1 macrophages are the first agents of protection in blocking the intracellular pathogens, and their activation can promote T-helper lymphocytes type 1 (Th1) polarization. M1 macrophages release great amounts of some cytokines that have pro-inflammatory roles, including tumor necrotic factor- $\alpha$  (TNF- $\alpha$ ), monocyte chemo attractant protein-1 (MCP-1), IL-6, IL-1, IL-12, type 1 interferons (IFNs), inducible nitric oxide synthase (iNOS), and C-X-C motif chemokine ligands (CXCLs) such as CXCL1-3, CXCL5, and CXCL8-10.

M2 macrophages are involved in infections caused by fungal, parasitic, or helminthic pathogens and conversely express high level of dectin-1, DC-SIGN (CD209), mannose receptor (CD206), CD163, scavenger receptor A and B-1, C-C chemokine receptor 2 (CCR2), C-X-C motif chemokine receptor 1 (CXCR1), and CXCR2. M2 macrophages produce materials that play a role in tissue remodeling and repair, such as IL-10, chitinase-like protein 3 (YM1), macrophage and granulocyte inducer-form 1 (MgI1), and arginase-1 [4, 16]. Arginine metabolism pathways play a central role in macrophage

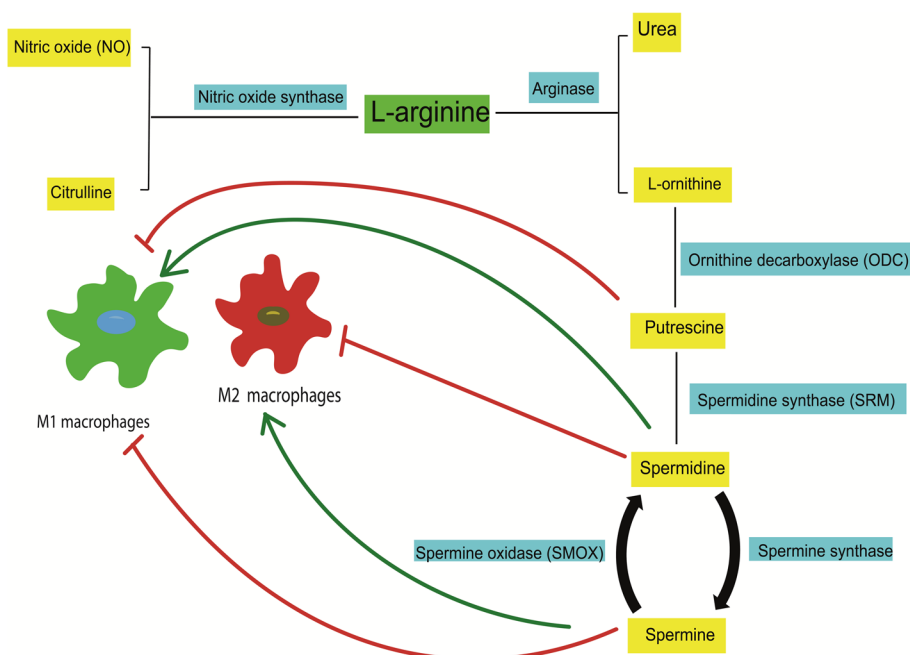


**Fig. 1** Macrophage polarization phenotypes and subtypes according to their different characteristics have many roles in immune responses. M1 macrophages have a pro-inflammatory role by their cytokines, but M2 macrophages, which are divided into four subtypes, have many different roles. For example, M2a macrophages play an important role in parasite killing, M2b macrophages function as immune system regulators, M2c macrophages assist in the wound healing process, and M2d macrophages have a pro-angiogenic role and are very important in tumor progression. CCL, C-C chemokine ligand; CXCL, C-X-C chemokine ligand; CXCR, C-X-C chemokine receptor; DCIR, dendritic cell immunoreceptor; IFN, interferon; IFN $\gamma$ , interferon- $\gamma$ ; IL, interleukin; RELM $\alpha$ , resistin-like molecule- $\alpha$ ; SPHK1, sphingosine kinase 1; TLR, Toll-like receptor; CD, cluster of differentiation; TGF- $\beta$ , transforming growth factor- $\beta$ ; PPAR- $\gamma$ , peroxisome proliferator-activated receptor gamma; TNF- $\alpha$ , tumor necrotic factor- $\alpha$ ; YM1, chitinase-like protein 3; LPS, lipopolysaccharides; VEGF, vascular endothelial growth factor; MerTK, myeloid epithelial reproductive tyrosine kinase; DNMT, DNA methyl transferase; miRs, microRNAs; MHCII, major histocompatibility complex II; IGF, insulin-like growth factor

polarization (Fig. 2) [29, 30]. Macrophage polarization is governed by the surrounding microenvironment, including cytokines and other components such as oligosaccharides, or by exosomes [22, 31]. On the other hand, epigenetic mechanisms such as chromatin remodeling, DNA methylation (DNAm), and histone modifications can control this process in connection with different factors. It was demonstrated that different levels of DNA methyl transferase 1 (DNMT 1), 3a and b, are expressed in M1 and M2 macrophages [4].

### The importance of macrophage polarization balance

Macrophage activation is necessary for the appropriate response against pathogen spreading in infected tissue. The process begins with pathogen-associated molecules releasing active danger signals that induce most tissue-resident macrophages to M1 polarization with nitrogen/oxygen-reactive agents and pro-inflammatory cytokine production ability. The next step is clearing cellular debris, then wound-healing signals commence M2 polarization with anti-inflammatory activity [26, 32–34]. Data validate the association between various diseases and the balance of M1/M2 macrophage polarization [35]. M1/M2 ratio and the regulation of macrophage polarization are very



**Fig. 2** Mammalian arginine metabolism pathways and M1 and M2 macrophage polarization. Arginine metabolism can be derived via NOS or arginase. NOS, which is expressed in M1 macrophages, causes arginine metabolism to release NO and citrulline, and M2 macrophages synthesize arginase, which causes arginine metabolism to release ornithine and urea. Downstream pathways of ornithine include putrescine, spermidine, and spermine, which hydrolyze products of ODC, SRM, and spermine synthase, respectively. Spermine can be resynthesized to spermidine by SMOX. As shown in the figure, putrescine and spermine downregulate the polarization of the M1 macrophages. Spermidine upregulates M1 macrophage polarization. In the case of M2 macrophage polarization, spermidine and spermine have inhibitor and enhancer effects, respectively. M1 macrophages increase NOS, but M2 macrophages upregulate arginase and ODC. Both arginine metabolic pathways arrest each other. Enzymes are shown by blue boxes and metabolites by yellow boxes, respectively. NOS, nitric oxide synthase; NO, nitric oxide; ODC, ornithine decarboxylase; SRM, spermidine synthase; SMOX, spermine oxidase

important. This ratio can signal progression of many inflammation-related diseases, for example, in psoriasis, which is a chronic inflammatory skin disorder. M1 macrophage markers are more abundant than M2 macrophage markers in psoriatic tissues [36]. Some pathogens and even tumors are able to reduce the M1/M2 ratio to avoid inflammation response [37–39]. For instance, some *Lactobacillus* are able to inhibit the formation of foam cells, which are a type of macrophage with lipoprotein ingestion activity [40]. Macrophage polarization is linked with some clinical conditions, including diabetes and obesity [3, 41], rheumatoid arthritis (RA) [42–44], chronic obstructive pulmonary disease [45], atherosclerosis [46, 47], non-alcoholic fatty liver disease (NAFLD) [48], osteoclastogenesis [49, 50], asthma [51], hypertension, and cardiovascular disease [52].

Balanced M1/M2 ratios are necessary for appropriate inflammatory response [16, 53]. Different stimuli factors or pathways involved in macrophage polarization can be promising candidates for therapeutic targets (Table 2) [4, 8, 54]. For instance, thiazolidinediones (TZDs), which target a member of the M2-like macrophage pathway, peroxisome proliferator-activated receptor gamma (PPAR- $\gamma$ ), are used for patients with type 2 diabetes (T2D) [55]. In a clinical study, it was observed that reducing the ratio of M1/M2 macrophages by blocking T-cell death-associated gene 8 (TDAG8), which has pro-inflammatory role, can attenuate RA progression (NSC745885) [56].

Macrophages possess phagocytosis activity, which makes them able to capture nanoscale particles, and thus are appropriate candidates for targeting macrophages [57–61]. In a study, a bioactive nanodevice was designed to convert the M1 phenotype to the M2 phenotype. A nanodevice is a peptide-coated gold nanoparticle (GNP) that promotes inflammation resolution. It also can be used as a novel therapeutic agent for patients with acute lung injury (ALI) and acute respiratory distress syndrome (ARDS) [62]. It has been shown that nanoparticles, including designed miRNAs or siRNAs, switch macrophage polarization in some disease conditions [63, 64]. Clinical and experimental data

**Table 2** Immunomodulatory drugs that target macrophage polarization

immunomodulatory drugs	Functions	References
Thapsigargin	Promote M2 polarization	[95]
Glucocorticoids	Trigger M2 polarization	[96]
Azithromycin	Promotes polarization from M1 to M2	[97]
5-Aminosalicylates (5-ASAs)	Inhibitory role in macrophage activation and inflammation suppressor	[98]
[6-(1-Methyl-4-nitroimidazol-5-yl)thiopurine]	Repressed nitric oxide synthase (iNOS) expression	[99]
Imiquimod	Restore pro-inflammatory of TAMs	[100]
PLX3397, PLX108-01 (pexidartinib)	Deplete macrophages	[101–103]
Trabectedin	Deplete macrophages	[104]
CP-690,550 (tofacitinib)	Inhibit modulate gene expression in macrophages	[105]
Hyaluronic acid oligosaccharides	Modulate macrophage polarity	[106]
Tocilizumab	M1 macrophage suppression	[107, 108]

have shown that M2 macrophages, which are TAMs, improve the growth, migration, invasion, and immunosuppressive activities of tumor cells [65–67]. Various therapeutic strategies targeting TAMs, depletion of the M2 macrophage ratio, or converting M2 macrophages to M1 macrophages are highlighted as potential strategies for suppressing tumor progression [65, 68–70]. TAMs can be used as novel targets in cancer therapy. For instance, a saponin component isolated from *Astragali radix*, called astragaloside IV (AS-IV), is reported to reduce tumor growth and metastasis by arresting the polarization of M2 macrophages through the adenosine monophosphate-activated protein kinase (AMPK) signaling pathway [65, 71].

Another study has demonstrated that M1 macrophage induction can increase cellular apoptosis and decrease tumor metastasis and chemotherapy resistance in mice with hepatocellular carcinoma (HCC) [72]. Macrophage targeting strategies in combination with chemotherapies exhibit more antitumor activity [73]. Among the autoimmune diseases caused by imbalanced M1/M2 ratio are systemic lupus erythematosus (SLE) [74], inflammatory bowel diseases (IBD) [75], autoimmune myocarditis [76], and autoimmune neuritis [77, 78]. Chronic inflammation is associated with age-related diseases such as cardiovascular disease, diabetes, and Alzheimer's disease [5]. Much research has revealed that macrophages participate in the process of pregnancy [79] and could have important effects on preeclampsia, miscarriage, and preterm birth [32]. Different phenotypes and functions of macrophages are essential for each phase of pregnancy to establish and maintain pregnancy. Decidual macrophages participate in implantation, spiral artery remodeling, and angiogenesis of embryo, and they also protect the embryo from pathogens and maternal immune responses [80].

According to various studies, different polarized states of macrophages have been associated with diseases, including cystic fibrosis (CF) and asthma; high level of M2-polarized macrophages is correlated with higher-severity asthma [81]. Accumulating data have shown indispensable roles for M2b macrophages in cardiovascular diseases [82]. In addition to M1 and M2 macrophages, a “chimeric” M1–M2 type with mixed biological function and phenotype that can cause impaired inflammation conditions is described in some cases such as rheumatoid arthritis [83]. In fact, recent evidence shows a continuum of different macrophages with different markers and phenotypes that strongly depend on their microenvironment. Therefore, they can be considered as a spectrum [84–86], although simplified macrophage classification (M1/M2) is used for better understanding. M1 and M2 macrophages can be reprogrammed by different stimuli signals. These reversible changes are essential during inflammation and its resolution phases [87, 88]. Of note, the tumor microenvironment (TME) has a vital effect on cancer progression [89, 90]. Macrophages are able to alter their features according to the TME. Therefore, macrophage polarization can be considered as a therapeutic strategy for cancer treatment [66].

### **Inflammation and the role of macrophage polarization in this process**

Inflammation is a physiological condition that occurs in response to various situations, such as injury and infection. Acute inflammation is the first mechanism in responding to these conditions [91]. Uncontrolled acute inflammation may lead to chronic inflammation, which has been related to many diseases [92]. Sterile inflammation (SI) occurs



by nonmicrobial factors such as chemical, physical, or metabolic stimuli, while nonsterile inflammation occurs by infection [93, 94]. Regenerative inflammation takes place in cases of low-grade damage or in highly regenerative tissues, such as the liver, and this type of inflammation plays a critical role in regeneration and repair [95]. Fibrotic inflammation occurs as a response to extensive damage or in poorly regenerative tissues, such as the myocardium. Macrophages play a crucial role in fibrotic inflammation [96]. Inflammation can lead to different immune responses via releasing molecular mediators. These mediators have roles in the direction of vascular responses, immune cell recruitment, macrophage polarization, pathogen clearance, repair of damaged tissue, and restoration of homeostasis. The balance of signal transducer and activator of transcription (STATs) activation has a very important role in macrophage polarization. The activation of STAT1 can lead to M1 macrophage polarization. This is important in the process of cytotoxicity and pro-inflammatory functions. On the contrary, some cytokines such as IL-4/IL-13 and IL-10 and activation of STAT3 and STAT6 can increase M2 polarization. Some other factors such as DNA methylation, chromatin remodeling, histone modification, and meta-inflammation, caused by chronic overnutrition and obesity, are involved in macrophage polarization. There are many clinical trials on macrophage polarization and inflammation (Table 3) [16, 97, 98]. Cardiac macrophages (CMs) are tissue-resident macrophages that are critical agents in the generation and development of cardiac inflammation, tissue remodeling, and repair. CMs are activated by the recognition of DAMPs or PAMPs. via cytokines released from inflammatory cells in the myocardium. An example of this is a promotion of M2 phenotype in dead cell clearance processes and an increase in the level of IL-10 and transforming growth factor- $\beta$  (TGF- $\beta$ ), but pro-inflammatory cytokines such as TNF- $\alpha$  promote the M1 phenotype [99]. Unregulated immune responses mediated by macrophages may lead to chronic kidney disease (CKD). The balance of macrophage polarization between M1 phenotype and M2 phenotype is important in tissue injury.

M1 macrophages play a key role in CKD. These macrophages increase plasma pro-inflammatory biomarkers such as TNF- $\alpha$  in patients, which may lead to chronic renal insufficiency. Furthermore, M2 macrophages are involved in chronic renal inflammation, especially in the repair phase, where M1 macrophages switch to M2 macrophages and secrete anti-inflammatory cytokines such as IL-10, IL-22, and TGF- $\beta$ . M2 macrophages are involved more in the progression of fibrosis than M1 macrophages because they secrete profibrotic factors such as TGF- $\beta$  [100]. Adipocytes can play an important role in the management of macrophage polarization in adipose tissue. In healthy conditions, adipocytes promote M2-like polarization, but in obesity, adipocytes may favor the prevalence of M1-like macrophage polarization [101, 102]. Imbalance of inflammatory responses can cause many inflammatory insufficiencies, like what happens in IBD. In this case, lamina propria macrophages from patients with IBD have M1 phenotype rather than M2 phenotype, and as mentioned previously, these macrophages can produce a large amount of pro-inflammatory cytokines [103]. The polarization of macrophages is tied to glycolytic changes and oxidative phosphorylation (OXPHOS) metabolism. Indeed, M1 macrophages rely on glycolytic changes but M2 macrophages rely on fatty-acid-fueled OXPHOS. Many risk factors change macrophage polarization, including obesity

**Table 3** Clinical trials on macrophage polarization and inflammation

Status	Study title	Intervention	ClinicalTrials.gov identifier
Recruiting	Treatment of macrophage activation syndrome (MAS) with anakinra	•Drug: kineret •Drug: placebo	NCT02780583
Completed	Effect of liraglutide (Victoza) on inflammation in human adipose tissue and blood	•Drug: Victoza (liraglutide) with dietitian monitoring •Other: placebo with dietitian monitoring	NCT02650206
Completed	The effect of gut sterilization on macrophage activation in patients with alcoholic hepatitis	•Drug: combined vancomycin and gentamycin and meropenem	NCT03157388
Completed	Macrophage activation markers during sofosbuvir-based treatment regimens of chronic hepatitis C	•Drug: galactose •Procedure: gastroscopy •Procedure: liver biopsy •Procedure: FibroScan •Procedure: liver vein catheterization •Drug: sofosbuvir	NCT02528461
Unknown status	New candidate criteria for diagnosis of macrophage activation syndrome	–	NCT01095146
Completed	Exploration of immunity in Gaucher disease	–	NCT01358188
Completed	A study to investigate the safety and efficacy of emapalumab, an anti-IFN-gamma mAb in patients with systemic juvenile idiopathic arthritis (sJIA) or adult-onset Still's disease (AOSD) developing macrophage activation syndrome/secondary HLH (MAS/sHLH)	•Drug: emapalumab	NCT03311854
Recruiting	sCD163 in patients with PBC—assessment of disease severity and prognosis	•Other: blood samples, FibroScan, and questionnaires	NCT02924701
Completed	A role for RAGE/TXNIP/inflammasome axis in alveolar macrophage activation during ARDS (RIAMA): a proof-of concept clinical study	•Other: RAGE/TXNIP/inflammasome axis	NCT02545621
Recruiting	sCD163 in patients with PBC—assessment of treatment response	•Other: blood samples •Device: FibroScan •Other: questionnaires •Biological: liver biopsy	NCT02931513
Completed	Downmodulating monocyte activation for HIV-1-associated neurocognitive disorders (HAND)	•Drug: atorvastatin (Lipitor) •Drug: placebo	NCT01600170
Completed	A trial of validation and restoration of immune dysfunction in severe infections and sepsis	•Drug: anakinra •Drug: recombinant human interferon gamma •Drug: placebo	NCT03332225

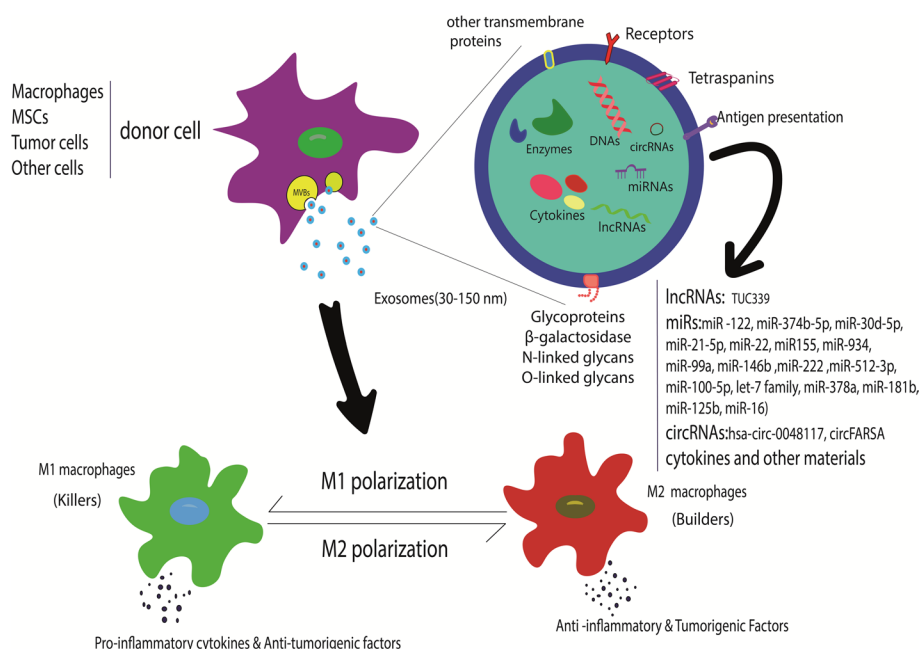
and hypertension. These factors can lead to chronic and systemic inflammation by M1 macrophage activation. Obesity and hypertension can change target-organ damage by hormones, local inflammatory signals, and hypoxia-induced signaling or alteration in glycolytic- OXPHOS paradigm in macrophages [104]. Macrophage polarization processes and their regulation are very important for sufficient immune responses, and any skew in these processes may lead to some inflammation disorders [105].



### Exosomes involve in macrophage polarization

Exosomes are a subset of nanoscale (30–150 nm) extracellular vesicles (EVs) that are released from almost every eukaryotic cell. Exosomes carry and deliver biological material, including proteins, lipids, saccharides, and genetic signals such as messenger ribonucleic acid (mRNAs), microRNAs (miRs), long noncoding RNAs (lncRNAs), deoxyribonucleic acid (DNAs), and circular RNAs (circRNAs). They have a very important role in cell communication. Exosomes have various effects on immune cell responses, stromal cells, and extracellular matrix (ECM), and can even alter them and their behaviors [106–108]. Exosomes derived from different cells play a key role in macrophage polarization processes. They are also able to change macrophage polarization. Recently, a research study demonstrated that M2-Exo causes a reprogramming of the M1 phenotype to the M2 phenotype (Fig. 3) [109]. Mesenchymal stromal cells (MSCs) have great potential to differentiate into many cell types. Numerous studies have demonstrated that exosomes secreted by MSCs have an important role in macrophage polarization [110, 111]. Exosomes present in serum have demonstrated involvement in IBD. A study reported that pregnancy zone protein (PZP) can be used as a biomarker in IBD [78].

Exosomes derived from MSCs of human bone marrow assist in the regulation of inflammatory responses. The systemic administration of these exosomes could substantially mitigate colitis in various models of IBD [112]. MSC exosomes have an important role in neuroinflammatory conditions. These exosomes can regulate macrophage polarization and change it toward an anti-inflammatory phenotype [113]. Also, MSC exosomes have a tumor-growth-suppressive effect by increasing inflammatory infiltration [111]. Pro-inflammatory bone-marrow-derived mesenchymal stem cells (BMMSCs) secrete



**Fig. 3** Exosomes are derived from different cells, including MSCs, tumor cells, or other cells that are present in the microenvironment such as macrophages. These exosomes can switch macrophage polarization according to their cargos. MSCs, mesenchymal stromal cells; lncRNA, long noncoding RNAs; miR, microRNAs; circRNA, circular RNA

exosomes that potentially promote macrophage M2 polarization. BMMSCs exosomes can reduce macrophage M1 polarization by regulation of the protein kinase B1/protein kinase B2 (AKT1/AKT2) signaling pathway and relieve myocardial injury [114, 115]. Fibronectin type III domain-containing protein 5 (FNDC5) is a transmembrane protein located in the cytoplasm that can increase BMMSC exosome secretion. This mechanism can promote M2 polarization by the nuclear factor- $\kappa$ B (NF- $\kappa$ B) signaling pathway [116, 117]. A study reported that administration of BMMSC-derived exosomes can induce macrophage M2 polarization, improve the inflammatory microenvironment, and promote fibrocartilage regeneration, especially at the tendon–bone interface [118].

Exosomes of adipose-derived stem cells (ADSCs) have an important role in obesity-associated inflammation and other metabolic disorders. They can induce anti-inflammation M2 macrophage polarization by carrying active STAT3 and inhibiting macrophage inflammatory responses [119]. ADSC exosomes have a critical role in myocardial repair after myocardial infarction (MI). These exosomes can decrease lipopolysaccharides (LPS)-induced inflammation by activating sphingosine 1-phosphate (S1P), sphingosine kinase 1 (SphK1 or SK1), and sphingosine-1-phosphate receptor 1 (S1PR1) signaling, which leads to promotion of macrophage M2 polarization [120].

Exosomes derived from human umbilical cord mesenchymal stem cells (hUCMSCs) have involvement in regulation of macrophage polarization. They can inhibit M1 polarization and promote M2 polarization. These exosomes can inhibit the expression of tumor necrosis factor receptor-associated factor 1 (TRAF1), which has been shown to be involved in the macrophage M1 polarization mechanism and ameliorate steroid-resistant asthma (SSRA), an important clinical problem in asthma management [121]. Exosomes carry genetic signals such as miRs, which play a critical role in macrophage polarization. For example, mammary epithelial cells (MECs) can regulate immune system responses, secreting exosomes carrying exosomal miR-122. This miR can promote polarization of M1 macrophages by suppressing cytokine signaling 1 (SOCS1), STAT1, and STAT3 [122].

It has been demonstrated that exosomal miR-21-5p, originated from MSCs, can stimulate the polarization of M2 macrophages. It can reduce the inflammatory response and promote heart cell repair after myocardial ischemia–reperfusion injury [123]. It is reported that exosomal miR-21-5p released from MSCs has an important role in macrophage polarization. miR-21-5p induces M2 polarization by mediating phosphatase and tensin homolog (PTEN) downregulation. This mechanism can support lung cancer cell growth and facilitate their invasion [110]. Upregulation of miR-374b-5p in exosomes, derived from hypoxic tubular epithelial cells (TECs), reduces SOCS1 expression and promotes M1 macrophage activation during renal ischemia–reperfusion injury (RIRI) conditions [124, 125]. Exosomes derived from polymorph nuclear neutrophils (PMNs) have an important role in sepsis-related ALI. These exosomes promote M1 macrophage activation by miR-30d-5p, which targets SOCS-1 and sirtuin 1 (SIRT1) in macrophages [126]. Atherosclerosis is an inflammatory disease that leads to clogging of blood vessels by the accumulation of lipids.

Exosomes, which originate from different cells, have a very important role in atherosclerosis. For example, exosomal miR-155 can enlarge inflammatory cytokines and M1 polarization markers such as cluster of differentiation 80 (CD80) and CD86,

which are involved in the process of atherosclerosis through pro-inflammatory M1 polarization [127]. Some exosomal miRs such as miR-100-5p, miR-512-3p, let-7 family, and miR-21a-5p are derived from MSCs and have various properties. As these exosomal miRs can induce M2 macrophage polarization, they are capable of suppressing atherosclerosis [128, 129]. Bone-marrow-derived macrophages (BMDMs) can release exosomal miRs with anti-inflammatory properties, including miR-99a, miR-146b, and miR-378a, which are capable of promoting M2 polarization in BMDMs [127, 130]. Wu et al. generated exosomes with anti-inflammatory functions in atherosclerosis. These exosomes are derived from M2 macrophages and contain hexyl 5-aminolevulinate hydrochloride (HAL), which is FDA-approved and has an anti-inflammatory effect. This makes HALM2 exosomes a promising candidate for atherosclerosis therapy applying macrophage-derived exosomes [127, 131].

Exosomes can modulate immune responses in tumor cells. Tumor-derived exosomes (TEs) can change macrophage polarization, and could activate anti-inflammatory pro-tumorigenic M2 macrophage phenotypes or pro-inflammatory anti-tumorigenic M1 macrophage phenotypes, change the M1/M2 ratio in the TME, and promote tumor growth [132]. Melanoma-derived exosomes can upregulate specific macrophage polarization factors and promote mixed M1 and M2 macrophage phenotypes [128]. Lung tumor cell-derived exosomes can reprogram macrophage metabolism and promote M2 macrophage polarization [133]. Exosomal miR-222 derived from adriamycin-resistant breast cancer cells can target phosphatases and PTEN gene, and activate the Akt pathway, so it can switch macrophage polarization to M2 phenotype and stimulate tumor growth [134, 135]. Exosomal miR-222 derived from adriamycin-resistant breast cancer cells can directly target phosphatase and the PTEN gene, activate the Akt pathway, convert macrophage polarization to M2 phenotype, and stimulate tumor growth by M2 macrophage polarization [136]. It has been reported that exosomes derived from hypoxic tumor cells can elevate oxidative phosphorylation in macrophages that originate from bone marrow by let-7a exosomal miR, which suppresses insulin-Akt-mammalian target of rapamycin (mTOR) signaling pathway and promotes M2-like macrophages [137].

Exosomal lncRNAs participate in macrophage polarization. For example, HCC-derived exosomes contain different levels of exosomal lncRNA TUC339 that regulate macrophage polarization [138]. It has been identified that circRNA, which is carried by exosomes, has important regulatory roles in different pathophysiological processes. For example, exosomal hsa-circ-0048117 has been upregulated in esophageal squamous cell carcinoma (ESCC). Upregulation of hsa-circ0048117 can promote M2 macrophage polarization and regulate ESCC progression. Another exosomal circRNA is circFARSA, which is highly expressed in tumor cells and is capable of inducing the promotion of M2 phenotypes and facilitating metastasis in non-small cell lung cancer (NSCLC) cells [139]. M1 macrophage-derived exosomes can be used as paclitaxel (PTX) nanocarriers and enhance antitumor activity of PTX [140]. These can also be tumor biomarkers such as exosomal miR-16, which are received by macrophages. These act as metastasis biomarkers in breast cancer [141], showing that exosomes can be employed for the treatment of cancer [59].

## Conclusion

Recently, it was confirmed that inflammation is a sign of chronic disease such as cancers, diabetes, cardiovascular, and neurologic system disorders. This review points out that the plasticity features of macrophages give them a critical role in inflammatory conditions. Macrophage polarization can be switched from pro-inflammatory with anti-tumorigenic macrophages (M1-like) phenotype to anti-inflammatory with pro-tumorigenic (M2-like) phenotype. In addition, the balance between different types of macrophages plays a key role in immune system dysfunctions. Exosomes as vehicles in cell communication have serious involvement in macrophage polarization. Cancerous cells can modulate immune cell responses by their secreted exosomes. There are many immune-based strategies that have been established for cancer therapy such as cancer vaccines. Various agents have been used for delivering of medicines.

Exosomes can change macrophage polarization and promote or prevent different subtypes of macrophage population via their cargos, such as miRs, cricRNAs, and lncRNAs. Numerous studies have reported that these nano-sized vesicles could be engineered for medical aims and developed as delivery systems for immune system modulation. However, there are several unsolved problems in the clinical application of exosomes as biologically derived nanovesicles. Examples of this are selecting the appropriate exosome isolation method, ensuring purity of exosomes, and identifying an efficient method for exosome modification. In conclusion, further studies on macrophage polarization mechanisms and its related pathways are needed to elucidate exosomes' roles in these pathways and their therapeutic potential in the development of immunotherapies for various medical aims.

## Abbreviations

MPS	Mononuclear phagocyte system
DCs	Dendritic cells
CNS	Central nervous system
TAM	Tumor-associated macrophages
CSF-1	Colony-stimulating factor-1
M-CSF	Macrophage-colony-stimulating factor
CSF-1R	Colony-stimulating factor-1 receptor
Th1	T-helper lymphocytes type 1
TNF- $\alpha$	Tumor necrotic factor- $\alpha$
MCP-1	Monocyte chemo attractant protein-1
IL	Interleukin
IFNs	Interferons
iNOS	Inducible nitric oxide synthase
CXCLs	C-X-C motif chemokine ligands
CCR	C-C chemokine receptor
CXCR	C-X-C motif chemokine receptor
YM1	Chitinase-like protein 3
MgI1	Macrophage and granulocyte inducer-form 1
DNMT	DNA methyl transferase
DNAm	DNA methylation
RA	Rheumatoid arthritis
NAFLD	Non-alcoholic fatty liver disease
TZDs	Thiazolidinediones
PPAR- $\gamma$	Peroxisome proliferator-activated receptor gamma
T2D	Type 2 diabetes
TDAG8	T-cell death-associated gene 8
GNP	Gold nanoparticle
ALI	Acute lung injury
ARDS	Acute respiratory distress syndrome
AS-IV	Astragaloside IV
AMPK	Adenosine monophosphate-activated protein kinase
HCC	Hepatocellular carcinoma

SLE	Systemic lupus erythematosus
IBD	Inflammatory bowel diseases
CF	Cystic fibrosis
TME	Tumor microenvironment
STATs	Signal transducer and activator of transcription
CMs	Cardiac macrophages
TGF- $\beta$	Transforming growth factor- $\beta$
CKD	Chronic kidney disease
OXPHOS	Oxidative phosphorylation
EVs	Extracellular vesicles
DNA	Deoxyribonucleic acid
miRs	MicroRNAs
lncRNA	Long noncoding RNAs
circRNAs	Circular RNAs
mRNA	Messenger ribonucleic acid
ECM	Extracellular matrix
MSCs	Mesenchymal stromal cells
BMMSCs	Bone marrow-derived mesenchymal stem cells
AKT	Protein kinase B
FNDC5	Fibronectin type III domain-containing protein 5
NF- $\kappa$ B	Nuclear factor- $\kappa$ B
ADSCs	Adipose derived stem cells
MI	Myocardial infarction
LPS	Lipopolysaccharides
S1P	Sphingosine 1-phosphate
SI	Sterile inflammation
SphK1 or SK1	Sphingosine kinase 1
S1PR1	Sphingosine-1-phosphate receptor 1
hUCMSCs	Human umbilical cord mesenchymal stem cells
TRAF1	Tumor necrosis factor receptor-associated factor 1
SSRA	Steroid-resistant asthma
MECs	Mammary epithelial cells
SOCS1	Suppressor of cytokine signaling 1
PMNs	Polymorph nuclear neutrophils
PTEN	Phosphate and tensin homolog
PZP	Pregnancy zone protein
TECs	Tubular epithelial cells
RIRI	Renal ischemia–reperfusion injury
SIRT1	Sirtuin 1
BMDMs	Bone-marrow-derived macrophages
CD	Cluster of differentiation
HAL	Hexyl 5-aminolevulinate hydrochloride
TEs	Tumor-derived exosomes
mTOR	Mammalian target of rapamycin
ESCC	Esophageal squamous cell carcinoma
NSCLC	Non-small cell lung cancer
PTX	Paclitaxel
CCL	Chemokine (C–C motif) ligand
TLR	Toll-like receptor
VEGF	Vascular endothelial growth factor
ICAM	Intercellular adhesion molecule
DC-SIGN	Dendritic cell-specific ICAM-grabbing non integrin
IRF5	Interferon regulatory factor 5
HDAC3	Histone deacetylase 3
MerTK	Myeloid epithelial reproductive tyrosine kinase
NOS	Nitric oxide synthase
NO	Nitric oxide
ODC	Ornithine decarboxylase
SRM	Spermidine synthase
SMOX	Spermine oxidase
siRNA	Small interfering RNA
DAMP	Danger-associated molecular patterns
PAMP	Pathogen-associated molecular patterns
M2-Exo	M2-derived exosomes
FDA	US Food and Drug Administration

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**References**

1. Nathan C. Metchnikoff's legacy in 2008. *Nat Immunol.* 2008;9(7):695–8.
2. Kaufmann SH. Immunology's foundation: the 100-year anniversary of the Nobel Prize to Paul Ehrlich and Elie Metchnikoff. *Nat Immunol.* 2008;9(7):705–12.
3. Chylikova J, Dvorackova J, Tauber Z, Kamarad V. M1/M2 macrophage polarization in human obese adipose tissue. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub.* 2018;162(2):79–82.
4. Parisi L, Gini E, Baci D, Tremolati M, Fanuli M, Bassani B, et al. Macrophage polarization in chronic inflammatory diseases: killers or builders? *J Immunol Res.* 2018;2018:9.
5. Oishi Y, Manabe I. Macrophages in age-related chronic inflammatory diseases. *NPJ Aging Mech Dis.* 2016;2(1):1–8.
6. Dzik JM. The ancestry and cumulative evolution of immune reactions. *Acta Biochim Pol.* 2010;57:4.
7. Guo F, Zhang H, Chen C, Hu S, Wang Y, Qiao J, et al. Autophagy favors *Brucella melitensis* survival in infected macrophages. *Cell Mol Biol Lett.* 2012;17(2):249–57.
8. Vannella KM, Wynn TA. Mechanisms of organ injury and repair by macrophages. *Annu Rev Physiol.* 2017;79:593–617.
9. Peter J. Macrophage polarization. *Annu Rev Physiol.* 2017;79(1):541–66.
10. Xie J, Huang Z, Yu X, Zhou L, Pei F. Clinical implications of macrophage dysfunction in the development of osteoarthritis of the knee. *Cytokine Growth Factor Rev.* 2019;46:36–44.
11. Liang Y, Yang N, Pan G, Jin B, Wang S, Ji W. Elevated IL-33 promotes expression of MMP2 and MMP9 via activating STAT3 in alveolar macrophages during LPS-induced acute lung injury. *Cell Mol Biol Lett.* 2018;23(1):52.
12. Asaga H, Ishigami A. Microglial expression of peptidylarginine deiminase 2 in the prenatal rat brain. *Cell Mol Biol Lett.* 2007;12(4):536–44.
13. Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. *Nature.* 2013;496(7446):445–55.
14. Yang X, Chang Y, Wei W. Emerging role of targeting macrophages in rheumatoid arthritis: focus on polarization, metabolism and apoptosis. *Cell Prolif.* 2020;53(7): e12854.
15. Li X, Li M, Huang S, Qiao S, Qin Z, Kang C, et al. The effect of buffalo CD14 shRNA on the gene expression of TLR4 signal pathway in buffalo monocyte/macrophages. *Cell Mol Biol Lett.* 2014;19(4):623–37.
16. Atri C, Guerfali FZ, Laouini D. Role of human macrophage polarization in inflammation during infectious diseases. *Int J Mol Sci.* 2018;19(6):1801.
17. O'Reilly C, Doroudian M, Mawhinney L, Donnelly SC. Targeting MIF in cancer: therapeutic strategies, current developments, and future opportunities. *Med Res Rev.* 2016;36(3):440–60.
18. Murray PJ, Allen JE, Biswas SK, Fisher EA, Gilroy DW, Goerdt S, et al. Macrophage activation and polarization: nomenclature and experimental guidelines. *Immunity.* 2014;41(1):14–20.
19. Li F, Yang Y, Zhu X, Huang L, Xu J. Macrophage polarization modulates development of systemic lupus erythematosus. *Cell Physiol Biochem.* 2015;37(4):1279–88.
20. Lee B-C, Lee J. Cellular and molecular players in adipose tissue inflammation in the development of obesity-induced insulin resistance. *Biochem Biophys Acta.* 2014;1842(3):446–62.
21. Donnelly DJ, Popovich PG. Inflammation and its role in neuroprotection, axonal regeneration and functional recovery after spinal cord injury. *Exp Neurol.* 2008;209(2):378–88.
22. Shapouri-Moghaddam A, Mohammadian S, Vazini H, Taghadosi M, Esmaeili SA, Mardani F, et al. Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol.* 2018;233(9):6425–40.
23. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol.* 2004;25(12):677–86.



24. Linton MF, Moslehi JJ, Babaev VR. Akt signaling in macrophage polarization, survival, and atherosclerosis. *Int J Mol Sci.* 2019;20(11):2703.
25. Stein M, Keshav S, Harris N, Gordon S. Interleukin 4 potentially enhances murine macrophage mannose receptor activity: a marker of alternative immunologic macrophage activation. *J Exp Med.* 1992;176(1):287–92.
26. Funes SC, Rios M, Escobar-Vera J, Kalergis AM. Implications of macrophage polarization in autoimmunity. *Immunology.* 2018;154(2):186–95.
27. Wu H, Xu JB, He YL, Peng JJ, Zhang XH, Chen CQ, et al. Tumor-associated macrophages promote angiogenesis and lymphangiogenesis of gastric cancer. *J Surg Oncol.* 2012;106(4):462–8.
28. Tamura R, Tanaka T, Yamamoto Y, Akasaki Y, Sasaki H. Dual role of macrophage in tumor immunity. *Immunotherapy.* 2018;10(10):899–909.
29. Latour YL, Gobert AP, Wilson KT. The role of polyamines in the regulation of macrophage polarization and function. *Amino Acids.* 2020;52(2):151–60.
30. Rath T, Baker K, Pyzik M, Blumberg RS. Regulation of immune responses by the neonatal Fc receptor and its therapeutic implications. *Front Immunol.* 2015;5:664.
31. Arabpour M, Saghazadeh A, Rezaei N. Anti-inflammatory and M2 macrophage polarization-promoting effect of mesenchymal stem cell-derived exosomes. *Int Immunopharmacol.* 2021;97: 107823.
32. Porta C, Riboldi E, Ippolito A, Sica A, editors. Molecular and epigenetic basis of macrophage polarized activation. In: *Seminars in immunology*; 2015: Elsevier.
33. Zhang X, Mosser D. Macrophage activation by endogenous danger signals. *J Pathol.* 2008;214(2):161–78.
34. O'Neill A, Tynan A, Mawhinney L, Kennedy S, Caraher E, Poynton F, et al., editors. Macrophage migration inhibitory factor enhances *Pseudomonas aeruginosa* biofilms, a potential novel therapeutic for cystic fibrosis patients. *Irish J Med Sci*; 2017: Springer London Ltd 236 grays inn rd, 6th floor, London wc1x 8hl, England.
35. Gabrilovich DL, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. *Nat Rev Immunol.* 2012;12(4):253–68.
36. Lu C-H, Lai C-Y, Yeh D-W, Liu Y-L, Su Y-W, Hsu L-C, et al. Involvement of M1 macrophage polarization in endosomal toll-like receptors activated psoriatic inflammation. *Mediat Inflamm.* 2018;2018:9.
37. Benoit M, Desnues B, Mege J-L. Macrophage polarization in bacterial infections. *J Immunol.* 2008;181(6):3733–9.
38. Domínguez-Soto Á, Usategui A, Casas-Engel M, Simón-Fuentes M, Nieto C, Cuevas VD, et al. Serotonin drives the acquisition of a profibrotic and anti-inflammatory gene profile through the 5-HT7R-PKA signaling axis. *Sci Rep.* 2017;7(1):1–15.
39. Mostafaei S, Kazemnejad A, Azimzadeh Jamalkandi S, Amirhashchi S, Donnelly SC, Armstrong ME, et al. Identification of novel genes in human airway epithelial cells associated with chronic obstructive pulmonary disease (COPD) using machine-based learning algorithms. *Sci Rep.* 2018;8(1):1–20.
40. Ding Y-H, Qian L-Y, Pang J, Lin J-Y, Xu Q, Wang L-H, et al. The regulation of immune cells by Lactobacilli: a potential therapeutic target for anti-atherosclerosis therapy. *Oncotarget.* 2017;8(35):59915.
41. Lumeng CN, Bodzin JL, Saltiel AR. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J Clin Investig.* 2007;117(1):175–84.
42. Wang Y, Han C-C, Cui D, Li Y, Ma Y, Wei W. Is macrophage polarization important in rheumatoid arthritis? *Int Immunopharmacol.* 2017;50:345–52.
43. Kinne RW, Stuhl Müller B, Burmester G-R. Cells of the synovium in rheumatoid arthritis. *Macrophages Arthritis Res Ther.* 2007;9(6):1–16.
44. Tardito S, Martinelli G, Soldano S, Paolino S, Pacini G, Patane M, et al. Macrophage M1/M2 polarization and rheumatoid arthritis: a systematic review. *Autoimmun Rev.* 2019;18(11): 102397.
45. Shaykhi R, Krause A, Salit J, Strulovici-Barel Y, Harvey B-G, O'Connor TP, et al. Smoking-dependent reprogramming of alveolar macrophage polarization: implication for pathogenesis of chronic obstructive pulmonary disease. *J Immunol.* 2009;183(4):2867–83.
46. De Gaetano M, Crean D, Barry M, Belton O. M1-and M2-type macrophage responses are predictive of adverse outcomes in human atherosclerosis. *Front Immunol.* 2016;7:275.
47. Stöger JL, Gijbels MJ, van der Velden S, Manca M, van der Loos CM, Biessen EA, et al. Distribution of macrophage polarization markers in human atherosclerosis. *Atherosclerosis.* 2012;225(2):461–8.
48. Kazankov K, Jørgensen SMD, Thomsen KL, Møller HJ, Vilstrup H, George J, et al. The role of macrophages in nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. *Nat Rev Gastroenterol Hepatol.* 2019;16(3):145–59.
49. Fukui S, Iwamoto N, Takatani A, Igawa T, Shimizu T, Umeda M, et al. M1 and M2 monocytes in rheumatoid arthritis: a contribution of imbalance of M1/M2 monocytes to osteoclastogenesis. *Front Immunol.* 2018;8:1958.
50. Cross M, Smith E, Hoy D, Nolte S, Ackerman I, Fransen M, et al. The global burden of hip and knee osteoarthritis: estimates from the global burden of disease 2010 study. *Ann Rheum Dis.* 2014;73(7):1323–30.
51. Madore A-M, Perron S, Turmel V, Laviolette M, Bissonnette EY, Laprise C. Alveolar macrophages in allergic asthma: an expression signature characterized by heat shock protein pathways. *Hum Immunol.* 2010;71(2):144–50.
52. Harwani SC. Macrophages under pressure: the role of macrophage polarization in hypertension. *Transl Res.* 2018;191:45–63.
53. Dehbidi MY, Goodarzi N, Azhdari MH, Doroudian M. Mesenchymal stem cells and their derived exosomes to combat Covid-19. *Rev Med Virol.* 2021;34:2.
54. Galeotti C, Kaveri SV, Bayry J. iVIG-mediated effector functions in autoimmune and inflammatory diseases. *Int Immunol.* 2017;29(11):491–8.
55. Consoli A, Formoso G. Do thiazolidinediones still have a role in treatment of type 2 diabetes mellitus? *Diabetes Obes Metab.* 2013;15(11):967–77.
56. Kung C-C, Dai S-P, Chiang H, Huang H-S, Sun W-H. Temporal expression patterns of distinct cytokines and M1/M2 macrophage polarization regulate rheumatoid arthritis progression. *Mol Biol Rep.* 2020;47(5):3423–37.
57. Parayath NN, Parikh A, Amiji MM. Repolarization of tumor-associated macrophages in a genetically engineered non-small cell lung cancer model by intraperitoneal administration of hyaluronic acid-based nanoparticles encapsulating microRNA-125b. *Nano Lett.* 2018;18(6):3571–9.

58. Mehanna MM, Mohyeldin SM, Elgindy NA. Respirable nanocarriers as a promising strategy for antitubercular drug delivery. *J Control Release*. 2014;187:183–97.
59. Hanjani NA, Esmaelizad N, Zanganeh S, Gharavi AT, Heidarizadeh P, Radfar M, et al. Emerging role of exosomes as biomarkers in cancer treatment and diagnosis. *Crit Rev Oncol Hematol*. 2022;169: 103565.
60. Doroudian M, O'Neill A, O'Reilly C, Tynan A, Mawhinney L, McElroy A, et al. Aerosolized drug-loaded nanoparticles targeting migration inhibitory factors inhibit *Pseudomonas aeruginosa*-induced inflammation and biofilm formation. *Nanomedicine*. 2020;15(30):2933–53.
61. Doroudian M, O'Reilly C, Crosbie-Staunton K, MacLoughlin R, Prina-Mello A, Volkov Y, et al., editors. Good things come in small packages: An aerosolized delivery system for small molecule inhibitors of macrophage migration inhibitory factor. *Irish J Med Sci*; 2016: springer London Ltd 236 grays inn Rd, 6th floor, London wc1x 8hl, England.
62. Wang L, Zhang H, Sun L, Gao W, Xiong Y, Ma A, et al. Manipulation of macrophage polarization by peptide-coated gold nanoparticles and its protective effects on acute lung injury. *J Nanobiotechnol*. 2020;18(1):1–16.
63. Bejerano T, Etzion S, Elyagon S, Etzion Y, Cohen S. Nanoparticle delivery of miRNA-21 mimic to cardiac macrophages improves myocardial remodeling after myocardial infarction. *Nano Lett*. 2018;18(9):5885–91.
64. Kim B, Pang H-B, Kang J, Park J-H, Ruoslahti E, Sailor MJ. Immunogene therapy with fusogenic nanoparticles modulates macrophage response to *Staphylococcus aureus*. *Nat Commun*. 2018;9(1):1–13.
65. Xu F, Cui W-Q, Wei Y, Cui J, Qiu J, Hu L-L, et al. Astragaloside IV inhibits lung cancer progression and metastasis by modulating macrophage polarization through AMPK signaling. *J Exp Clin Cancer Res*. 2018;37(1):1–16.
66. Kimura YN, Watari K, Fotovati A, Hosoi F, Yasumoto K, Izumi H, et al. Inflammatory stimuli from macrophages and cancer cells synergistically promote tumor growth and angiogenesis. *Cancer Sci*. 2007;98(12):2009–18.
67. Mu X, Shi W, Xu Y, Xu C, Zhao T, Geng B, et al. Tumor-derived lactate induces M2 macrophage polarization via the activation of the ERK/STAT3 signaling pathway in breast cancer. *Cell Cycle*. 2018;17(4):428–38.
68. Ostuni R, Kratochvill F, Murray PJ, Natoli G. Macrophages and cancer: from mechanisms to therapeutic implications. *Trends Immunol*. 2015;36(4):229–39.
69. Sica A, Schioppa T, Mantovani A, Allavena P. Tumour-associated macrophages are a distinct M2 polarised population promoting tumour progression: potential targets of anti-cancer therapy. *Eur J Cancer*. 2006;42(6):717–27.
70. Yu H, Yin S, Zhou S, Shao Y, Sun J, Pang X, et al. Magnolin promotes autophagy and cell cycle arrest via blocking LIF/Stat3/Mcl-1 axis in human colorectal cancers. *Cell Death Dis*. 2018;9(6):1–13.
71. Singh Y, Pawar VK, Meher JG, Raval K, Kumar A, Shrivastava R, et al. Targeting tumor associated macrophages (TAMs) via nanocarriers. *J Control Release*. 2017;254:92–106.
72. Hu X, Wang H, Han C, Cao X. Src promotes anti-inflammatory (M2) macrophage generation via the IL-4/STAT6 pathway. *Cytokine*. 2018;111:209–15.
73. Mehla K, Singh P. Metabolic regulation of macrophage polarization in cancer. *Trends Cancer*. 2019;5:822–34.
74. Triantafyllopoulou A, Franzke C-W, Seshan SV, Perino G, Kalliolias GD, Ramanujam M, et al. Proliferative lesions and metalloproteinase activity in murine lupus nephritis mediated by type I interferons and macrophages. *Proc Natl Acad Sci USA*. 2010;107(7):3012–7.
75. Steinbach EC, Plevy SE. The role of macrophages and dendritic cells in the initiation of inflammation in IBD. *Inflamm Bowel Dis*. 2014;20(1):166–75.
76. Su Z, Zhang P, Yu Y, Lu H, Liu Y, Ni P, et al. HMGB1 facilitated macrophage reprogramming towards a proinflammatory M1-like phenotype in experimental autoimmune myocarditis development. *Sci Rep*. 2016;6:8.
77. Han R, Xiao J, Zhai H, Hao J. Dimethyl fumarate attenuates experimental autoimmune neuritis through the nuclear factor erythroid-derived 2-related factor 2/hemoxygenase-1 pathway by altering the balance of M1/M2 macrophages. *J Neuroinflamm*. 2016;13(1):1–14.
78. Shao J, Jin Y, Shao C, Fan H, Wang X, Yang G. Serum exosomal pregnancy zone protein as a promising biomarker in inflammatory bowel disease. *Cell Mol Biol Lett*. 2021;26(1):36.
79. Hazan AD, Smith SD, Jones RL, Whittle W, Lye SJ, Dunk CE. Vascular-leukocyte interactions: mechanisms of human decidual spiral artery remodeling in vitro. *Am J Pathol*. 2010;177(2):1017–30.
80. Wang L-L, Li Z-H, Wang H, Kwak-Kim J, Liao A-H. Cutting edge: the regulatory mechanisms of macrophage polarization and function during pregnancy. *J Reprod Immunol*. 2022;151: 103627.
81. Gharib SA, McMahan RS, Eddy WE, Long ME, Parks WC, Aitken ML, et al. Transcriptional and functional diversity of human macrophage repolarization. *J Allergy Clin Immunol*. 2019;143(4):1536–48.
82. Wang L, Zhang S, Wu H, Rong X, Guo J. M2b macrophage polarization and its roles in diseases. *J Leukocyte Biol*. 2019;106(2):345–58.
83. Quero L, Hanser E, Manigold T, Tladen AN, Kyburz D. TLR2 stimulation impairs anti-inflammatory activity of M2-like macrophages, generating a chimeric M1/M2 phenotype. *Arthritis Res Ther*. 2017;19(1):1–13.
84. Palma A, Jarrah AS, Tieri P, Cesareni G, Castiglione F. Gene regulatory network modeling of macrophage differentiation corroborates the continuum hypothesis of polarization states. *Front Physiol*. 2018;9:1659.
85. Gao T, Huang F, Wang W, Xie Y, Wang B. Interleukin-10 genetically modified clinical-grade mesenchymal stromal cells markedly reinforced functional recovery after spinal cord injury via directing alternative activation of macrophages. *Cell Mol Biol Lett*. 2022;27(1):1–27.
86. Babazadeh S, Nassiri SM, Siavashi V, Sahlabadi M, Hajinasrollah M, Zamani-Ahmadm Mahmudi M. Macrophage polarization by MSC-derived CXCL12 determines tumor growth. *Cell Mol Biol Lett*. 2021;26(1):1–15.
87. Tarique AA, Logan J, Thomas E, Holt PG, Sly PD, Fantino E. Phenotypic, functional, and plasticity features of classical and alternatively activated human macrophages. *Am J Respir Cell Mol Biol*. 2015;53(5):676–88.
88. Xu W, Zhao X, Daha MR, van Kooten C. Reversible differentiation of pro- and anti-inflammatory macrophages. *Mol Immunol*. 2013;53(3):179–86.
89. Korniluk A, Koper O, Kemona H, Dymicka-Piekarska V. From inflammation to cancer. *Irish J Med Sci*. 2017;186(1):57–62.
90. Mostafavi S, Zalpoor H, Hassan ZM. The promising therapeutic effects of metformin on metabolic reprogramming of cancer-associated fibroblasts in solid tumors. *Cell Mol Biol Lett*. 2022;27(1):58.

91. Chimenti I, Sattler S, Del Monte-Nieto G, Forte E. Editorial: fibrosis and inflammation in tissue pathophysiology. *Front Physiol.* 2021;12: 830683.
92. Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, et al. Inflammatory responses and inflammation-associated diseases in organs. *Oncotarget.* 2018;9(6):7204–18.
93. Rubartelli A, Lotze MT, Latz E, Manfredi A. Mechanisms of sterile inflammation. *Front Immunol.* 2013;4:398.
94. Zindel J, Kubers P. DAMPs, PAMPs, and LAMPs in immunity and sterile inflammation. *Annu Rev Pathol.* 2020;15(1):493–518.
95. Cooke JP. Inflammation and its role in regeneration and repair. *Circ Res.* 2019;124(8):1166–8.
96. Mack M. Inflammation and fibrosis. *Matrix Biol.* 2018;68–69:106–21.
97. Li C, Xu MM, Wang K, Adler AJ, Vella AT, Zhou B. Macrophage polarization and meta-inflammation. *Transl Res.* 2018;191:29–44.
98. Parisi L, Gini E, Baci D, Tremolati M, Fanuli M, Bassani B, et al. Macrophage polarization in chronic inflammatory diseases: killers or builders? *J Immunol Res.* 2018;2018:8917804.
99. Lafuse WP, Wozniak DJ, Rajaram MVS. Role of cardiac macrophages on cardiac inflammation, fibrosis and tissue repair. *Cells.* 2021;10(1):51.
100. Lee H, Fessler MB, Qu P, Heymann J, Kopp JB. Macrophage polarization in innate immune responses contributing to pathogenesis of chronic kidney disease. *BMC Nephrol.* 2020;21(1):270.
101. Liu X, Chu H, Ji Y, Bosnjak Z, Ao H, Li T. Which BMI for diabetes patients is better? from the view of the adipose tissue macrophage-derived exosome. *Diabetes Metab Syndr Obes.* 2022;15:141–53.
102. Zhang J, Liu Y, Yin W, Hu X. Adipose-derived stromal cells in regulation of hematopoiesis. *Cell Mol Biol Lett.* 2020;25(1):16.
103. Moreira Lopes TC, Mosser DM, Gonçalves R. Macrophage polarization in intestinal inflammation and gut homeostasis. *Inflamm Res.* 2020;69(12):1163–72.
104. Mouton AJ, Li X, Hall ME, Hall JE. Obesity, hypertension, and cardiac dysfunction: novel roles of immunometabolism in macrophage activation and inflammation. *Circ Res.* 2020;126(6):789–806.
105. Liu W, Yu M, Chen F, Wang L, Ye C, Chen Q, et al. A novel delivery nanobiotechnology: engineered miR-181b exosomes improved osteointegration by regulating macrophage polarization. *J Nanobiotechnology.* 2021;19(1):269.
106. Tai Y-L, Chen K-C, Hsieh J-T, Shen T-L. Exosomes in cancer development and clinical applications. *Cancer Sci.* 2018;109(8):2364–74.
107. Letafati A, Najafi S, Mottahedi M, Karimzadeh M, Shahini A, Garousi S, et al. MicroRNA let-7 and viral infections: focus on mechanisms of action. *Cell Mol Biol Lett.* 2022;27(1):14.
108. He X-Y, Yu H-M, Lin S, Li Y-Z. Advances in the application of mesenchymal stem cells, exosomes, biomimetic materials, and 3D printing in osteoporosis treatment. *Cell Mol Biol Lett.* 2021;26(1):47.
109. Kim H, Wang SY, Kwak G, Yang Y, Kwon IC, Kim SH. Exosome-guided phenotypic switch of M1 to M2 macrophages for cutaneous wound healing. *Adv Sci (Weinh).* 2019;6(20):1900513.
110. Zhang F, Guo J, Zhang Z, Qian Y, Wang G, Duan M, et al. Mesenchymal stem cell-derived exosome: a tumor regulator and carrier for targeted tumor therapy. *Cancer Lett.* 2022;526:29–40.
111. Liang W, Chen X, Zhang S, Fang J, Chen M, Xu Y, et al. Mesenchymal stem cells as a double-edged sword in tumor growth: focusing on MSC-derived cytokines. *Cell Mol Biol Lett.* 2021;26(1):3.
112. Liu H, Liang Z, Wang F, Zhou C, Zheng X, Hu T, et al. Exosomes from mesenchymal stromal cells reduce murine colonic inflammation via a macrophage-dependent mechanism. *JCI Insight.* 2019;4:24.
113. Bischoff JP, Schulz A, Morrison H. The role of exosomes in intercellular and inter-organ communication of the peripheral nervous system. *FEBS Lett.* 2022;8:56.
114. Xu R, Zhang F, Chai R, Zhou W, Hu M, Liu B, et al. Exosomes derived from pro-inflammatory bone marrow-derived mesenchymal stem cells reduce inflammation and myocardial injury via mediating macrophage polarization. *J Cell Mol Med.* 2019;23(11):7617–31.
115. Li Q, Yu P, Wang W, Zhang P, Yang H, Li S, et al. Gene expression profiles of various cytokines in mesenchymal stem cells derived from umbilical cord tissue and bone marrow following infection with human cytomegalovirus. *Cell Mol Biol Lett.* 2014;19(1):140–57.
116. Ning H, Chen H, Deng J, Xiao C, Xu M, Shan L, et al. Exosomes secreted by FNDC5-BMMSCs protect myocardial infarction by anti-inflammation and macrophage polarization via NF- $\kappa$ B signaling pathway and Nrf2/HO-1 axis. *Stem Cell Res Ther.* 2021;12(1):519.
117. Ghashghaeinia M, Toulany M, Saki M, Rodemann HP, Mrowietz U, Lang F, et al. Roles of the NF $\kappa$ B and glutathione pathways in mature human erythrocytes. *Cell Mol Biol Lett.* 2012;17(1):11–20.
118. Shi Y, Kang X, Wang Y, Bian X, He G, Zhou M, et al. Exosomes derived from bone marrow stromal cells (BMSCs) enhance tendon–bone healing by regulating macrophage polarization. *Med Sci Monit.* 2020;26: e923328.
119. Zhao H, Shang Q, Pan Z, Bai Y, Li Z, Zhang H, et al. Exosomes from adipose-derived stem cells attenuate adipose inflammation and obesity through polarizing m2 macrophages and being in white adipose tissue. *Diabetes.* 2018;67(2):235–47.
120. Deng S, Zhou X, Ge Z, Song Y, Wang H, Liu X, et al. Exosomes from adipose-derived mesenchymal stem cells ameliorate cardiac damage after myocardial infarction by activating S1P/SK1/S1PR1 signaling and promoting macrophage M2 polarization. *Int J Biochem Cell Biol.* 2019;114: 105564.
121. Dong B, Wang C, Zhang J, Zhang J, Gu Y, Guo X, et al. Exosomes from human umbilical cord mesenchymal stem cells attenuate the inflammation of severe steroid-resistant asthma by reshaping macrophage polarization. *Stem Cell Res Ther.* 2021;12(1):204.
122. Cai M, Shi Y, Zheng T, Hu S, Du K, Ren A, et al. Mammary epithelial cell derived exosomal MiR-221 mediates M1 macrophage polarization via SOCS1/STATs to promote inflammatory response. *Int Immunopharmacol.* 2020;83: 106493.

123. Shen D, He Z. Mesenchymal stem cell-derived exosomes regulate the polarization and inflammatory response of macrophages via miR-21-5p to promote repair after myocardial reperfusion injury. *Ann Transl Med*. 2021;9(16):1323.
124. Ding C, Zheng J, Wang B, Li Y, Xiang H, Dou M, et al. Exosomal MicroRNA-374b-5p from tubular epithelial cells promoted m1 macrophages activation and worsened renal ischemia/reperfusion injury. *Front Cell Dev Biol*. 2020;8:587693.
125. Zhao S, Wu W, Liao J, Zhang X, Shen M, Li X, et al. Molecular mechanisms underlying the renal protective effects of coenzyme Q10 in acute kidney injury. *Cell Mol Biol Lett*. 2022;27(1):57.
126. Jiao Y, Zhang T, Zhang C, Ji H, Tong X, Xia R, et al. Exosomal miR-30d-5p of neutrophils induces M1 macrophage polarization and primes macrophage pyroptosis in sepsis-related acute lung injury. *Crit Care*. 2021;25(1):356.
127. Heo J, Kang H. Exosome-dosis. *Int J Mol Sci*. 2022;23(2):1002.
128. Li J, Xue H, Li T, Chu X, Xin D, Xiong Y, et al. Exosomes derived from mesenchymal stem cells attenuate the progression of atherosclerosis in ApoE<sup>-/-</sup> mice via miR-let7 mediated infiltration and polarization of M2 macrophage. *Biochem Biophys Res Commun*. 2019;510(4):565–72.
129. Ma J, Chen L, Zhu X, Li Q, Hu L, Li H. Mesenchymal stem cell-derived exosomal miR-21a-5p promotes M2 macrophage polarization and reduces macrophage infiltration to attenuate atherosclerosis. *Acta Biochim Biophys Sin*. 2021;53(9):1227–36.
130. Bouchareychas L, Duong P, Covarrubias S, Alsop E, Phu TA, Chung A, et al. Macrophage exosomes resolve atherosclerosis by regulating hematopoiesis and inflammation via MicroRNA cargo. *Cell Rep*. 2020;32(2): 107881.
131. Wu G, Zhang J, Zhao Q, Zhuang W, Ding J, Zhang C, et al. Molecularly engineered macrophage-derived exosomes with inflammation tropism and intrinsic heme biosynthesis for atherosclerosis treatment. *Angew Chem*. 2020;132(10):4097–103.
132. Baig MS, Roy A, Rajpoot S, Liu D, Savai R, Banerjee S, et al. Tumor-derived exosomes in the regulation of macrophage polarization. *Inflamm Res*. 2020;69(5):435–51.
133. Bardi GT, Smith MA, Hood JL. Melanoma exosomes promote mixed M1 and M2 macrophage polarization. *Cytokine*. 2018;105:63–72.
134. Pritchard A, Tousif S, Wang Y, Hough K, Khan S, Strenkowski J, et al. Lung tumor cell-derived exosomes promote M2 macrophage polarization. *Cells*. 2020;9:5.
135. Nahand JS, Vandchali NR, Darabi H, Doroudian M, Banafshe HR, Moghoofoei M, et al. Exosomal microRNAs: novel players in cervical cancer. *Epigenomics*. 2020;12(18):1651–60.
136. Chen WX, Wang DD, Zhu B, Zhu YZ, Zheng L, Feng ZQ, et al. Exosomal miR-222 from adriamycin-resistant MCF-7 breast cancer cells promote macrophages M2 polarization via PTEN/Akt to induce tumor progression. *Aging (Albany NY)*. 2021;13(7):10415–30.
137. Park JE, Dutta B, Tse SW, Gupta N, Tan CF, Low JK, et al. Hypoxia-induced tumor exosomes promote M2-like macrophage polarization of infiltrating myeloid cells and microRNA-mediated metabolic shift. *Oncogene*. 2019;38(26):5158–73.
138. Li X, Lei Y, Wu M, Li N. Regulation of macrophage activation and polarization by HCC-derived exosomal lncRNA TUC339. *Int J Mol Sci*. 2018;19(10):2958.
139. Li J, Zhang G, Liu C-G, Xiang X, Le MT, Sethi G, et al. The potential role of exosomal circRNAs in the tumor microenvironment: insights into cancer diagnosis and therapy. *Theranostics*. 2022;12(1):87.
140. Wang P, Wang H, Huang Q, Peng C, Yao L, Chen H, et al. Exosomes from M1-polarized macrophages enhance paclitaxel antitumor activity by activating macrophages-mediated inflammation. *Theranostics*. 2019;9(6):1714–27.
141. Vannini I, Fanini F, Fabbri M. Emerging roles of microRNAs in cancer. *Curr Opin Genet Dev*. 2018;48:128–33.

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