

Rapid on-site evaluation (ROSE) with EUS-FNA: The ROSE looks beautiful

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INTRODUCTION

Since EUS-FNA was first reported in 1992, it has become the first-line technique for the acquisition of tissue from gastrointestinal and adjacent organs with high safety and reliability.^[1,2] EUS-FNA is a multistep procedure that is affected by various uncertain factors,^[3-8] including rapid on-site evaluation (ROSE), whose use has been under debate for many years.

The rationale for using ROSE of EUS-guided tissue acquisition is the real-time evaluation of sample adequacy and diagnostic yield. This technique was expected to decrease the period of diagnosis with fewer needle passes as well as achieve a real-time and accurate diagnosis of digestive diseases.^[9] In addition, during the ROSE procedure, cytopathologists can determine whether additional sampling is required for further auxiliary diagnosis.^[10-13]

THE DEBATE LASTED 20 YEARS: RAPID ON-SITE EVALUATION OR NOT?

Early in this century, although experts advocated using ROSE to acquire adequate tissue samples, there are few data supporting this recommendation. Klapman

et al.^[14] compared EUS-guided tissue acquisition results from two medical centers and found that ROSE increased the diagnostic yield of EUS-FNA. In a single-center prospective study reported in 2005,^[15] high accuracy in a series of EUS-FNA with ROSE was again demonstrated. Consequently, EUS centers were recommended to be equipped with ROSE.

Eloubeidi *et al.*^[16] evaluated a series of EUS-FNA specimens sampled by one endoscopist in 2006. The ROSE outcomes were consistent with the final cytological evaluation (kappa score: 84.0%) in this prospective study.

In subsequent years, observational studies were repeated, and the diagnostic accuracy and tissue adequacy of EUS-FNA were demonstrated to improve significantly with the use of ROSE in gastrointestinal lesions. The greatest improvement was seen in solid pancreatic lesions. One meta-analysis^[17] of 34 studies was designed to evaluate whether ROSE affects the diagnostic accuracy of EUS-FNA in solid pancreatic lesions. In that study, the meta-regression model showed that ROSE remained an independent determinant of EUS-FNA accuracy ($P = 0.001$). Although the

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sensitivity was relatively low in many studies which were short of ROSE, only 2 of 14 studies that did not use ROSE had a sensitivity of 95% or higher. The authors concluded that the accuracy of EUS-FNA would be higher with the availability of ROSE.^[17]

Another meta-analysis reported in 2014 has also shown that ROSE increases the adequacy rate of EUS-FNA in solid pancreatic lesions by 3.5%.^[18]

Less information is available on the adverse events in EUS-FNA associated with ROSE. In 2011, Iglesias-Garcia *et al.*^[19] reported a study to evaluate the influence of ROSE on EUS-FNA in the differential diagnosis of solid pancreatic lesions in an unselected series of consecutive patients. The incidence of complications was significantly lower in the ROSE group, a result possibly associated with the lower number of passes required to obtain sufficient samples.^[20-22]

If this were the end of the research journey, the method of ROSE would seem highly favorable. However, studies are increasingly showing the contrary.

The effects of EUS-FNA with ROSE have been estimated in two randomized controlled trials involving patients with solid lesions of the pancreas.^[23,24] In the two studies, seven passes were performed in randomized patients without ROSE. Therefore, the number of needle passes was significantly smaller in ROSE group than seven. There was no significant difference in other outcomes, including diagnostic yield, sample quality, and adequacy between the two groups.^[23] In contrast to findings from a previous study,^[19] the lower number of needle passes in the ROSE group did not indicate a lower incidence of complications or a shorter procedural duration.^[23] The use of ROSE did not decrease the expense of EUS-guided biopsy and has been suggested to increase it.^[23-25]

In a multicentric randomized controlled study^[26] evaluating whether ROSE might improve the diagnostic yield rate of EUS-FNA in lymph node lesions, ninety patients were divided into two groups: ROSE+ and ROSE-. The slides' review time was shorter, and postprocedural pain was found less in the ROSE+ group. There was no statistical difference in the procedural times, complication rates, and mean costs between the two groups ($P = 0.06$, $P = 0.99$, and $P = 0.91$, respectively). These findings indicated that the

diagnostic yield of EUS-FNA in lymph node lesions had no relationship with the application of ROSE. These results do not support the recommendation to apply ROSE in EUS-FNA for lymph node lesions.

Hewitt *et al.*^[27] performed a meta-analysis and found that the pooled sensitivity of EUS-FNA with and without ROSE was 88 (87%–90%) and 80 (78%–82%), respectively, thus suggesting an 8% improvement in the sensitivity of EUS-FNA with ROSE. However, there was no statistical difference in this improvement ($P = 0.077$).

Kong *et al.*^[28] reported a systematic review including seven studies with 1299 patients. In that study, EUS-FNA with ROSE did not enhance the diagnostic yield or the diagnostic adequacy. There was no statistically significant difference in the number of needle passes between the ROSE and non-ROSE groups. The pooled sensitivity and specificity of the two groups were comparable.

The results of four meta-analyses,^[17,18,27,28] most of which were studies of patients with pancreatic masses, were contradictory. Two of the meta-analyses concluded that there was an improvement in the specimen adequacy and diagnostic yield associated with ROSE.^[17,18] However, the other two meta-analyses did not support these advantages.^[27,28]

Given that the current evidence is not concordant, the European Society of Gastrointestinal Endoscopy panel recommends EUS-FNA with or without ROSE equally (moderate-quality evidence, strong recommendation).^[25]

Therefore, evidence of whether ROSE might improve the results of EUS-FNA remains conflicting. In the debate on whether to use this method, the lack of on-site cytopathologists is always the key problem. According to a study on the practice patterns in EUS-FNA published in 2016, ROSE was available in 48% of European centers, 55% of Asian centers, and almost all centers (98%) from the USA.^[29] The obstacles to expanding ROSE include limited cytopathologist staffing, cost-effective performance, longer procedural duration, and a lack of belief in its added value.^[29]

Nevertheless, the diagnostic efficiency of EUS-FNA is dependent on the EUS techniques and experience of the endosonographers. Considering that not every

endoscopist works in a big center, ROSE seems to be a helpful tool for making up for the lack of EUS experience and technology in improving the diagnostic yield of EUS-FNA. ROSE, performed by a cytopathologist, provided a highly accurate diagnosis, which was highly consistent with the final results.^[16] ROSE may improve the adequacy of FNA specimens by 10%–30%, reduce the number of passes, decrease the duration of diagnosis, and lessen adverse events such as abdominal pain.^[30] After analyzing all studies, the clinical benefits of ROSE are obvious, and the ROSE should be applied, especially for the endosonographers, in the learning stage of EUS-FNA and for centers in which specimen adequacy rates are not high enough.^[30]

COULD ENDOSONOGRAPHERS LEARN TO PERFORM RAPID ON-SITE EVALUATION?

Cytopathologist staffing is not possible for all EUS centers, let alone all EUS-FNA procedures. Thus, it may be a possible solution to train endosonographers to evaluate the specimen themselves during the EUS-FNA procedure. Experienced endosonographers are thought to be able to assess the adequacy of the specimen obtained by EUS-FNA. Some researchers have begun to verify this possibility. However, a double-blind prospective controlled trial^[31] indicated that even well-trained endosonographers were less accurate than a cytopathologist in evaluating the specimen adequacy ($P = 0.004$) and in the preliminary estimate of malignancy ($P < 0.001$).^[31]

In a prospective double-blinded study on gross visual inspection during FNA between cytotechnologists and endoscopic technologists, neither cytotechnologists nor trained endoscopic technologists were able to evaluate the specimen adequacy accurately just by gross visual inspection of the slide.^[32]

Hayashi *et al.*^[33] retrospectively evaluated patients who underwent EUS-FNA for solid pancreatic lesions. Before the study, two endosonographers were trained for cytological interpretation, especially the four cytological features of pancreatic cancer, namely anisonucleosis, nuclear membrane irregularity, overlapping, and enlargement. The authors concluded that in this case, sample evaluation by trained endosonographers using a simple cytology grading system was very helpful.^[30]

TELECYTOPATHOLOGY

In recent years, advances in digital imaging technology have made it possible to remotely evaluate FNA specimens through telecytology. Khurana *et al.*^[34] first evaluated telecytology for ROSE of EUS-FNA samples of pancreas in 2012. Real-time images of Diff-Quik™ stained cytology smears were transmitted and evaluated by a cytopathologist. They found that the accuracies of preliminary diagnosis for pancreatic cancer between telecytology and conventional microscopy were comparable.

Two years later, Khurana *et al.*^[35] performed on-site telecytology evaluation on 95 cases. This study indicated that the telecytology could reduce the nondiagnostic rate, especially in the characteristics of solid lesions, and may serve as an adequate substitute for ROSE.

The current development of sharing economy model offers us a possibility that the on-site evaluation available in one center could be shared online across multiple centers to improve the efficiency of ROSE, which may alleviate the shortage of cytopathologists.

ARTIFICIAL INTELLIGENCE AND RAPID ON-SITE EVALUATION

In recent years, because of improvements in deep-learning techniques and increasing computational power, artificial intelligence (AI) has made impressive progress in interpreting complex images. Researchers have begun applying AI to learn and analyze pathology images.^[36-38]

Inoue *et al.*^[39] reported an automatic visual inspection method based on supervised machine learning for ROSE in EUS-FNA. This approach aims to clarify the relationship between the content of cellular structures, including tumor cells, and the image quality of specimens sampled by FNA. A stationary Gaussian mixture model is used to classify the local statistics of sample images because it can sufficiently estimate the universal mode. The results indicated that the method is helpful for EUS-FNA in aiding on-site visual inspection of cellular tissue, thus indicating areas highly likely to include tumor cells.

CONCLUSION

EUS-FNA has been developed as an analytical technique to sample digestive system lesions. The

requirement for ROSE remains one of the most controversial topics in the field of EUS-FNA. Based on the consideration that EUS-FNA combined with ROSE must not be inferior to non-ROSE, we still recommend implementing ROSE in the centers with enough staffing of the cytopathologists. The rapid development of electronic communication technology and AI technology has the potential to fundamentally change this issue, in terms of both cytopathologist time and clinical cost, and we look forward to this becoming a reality in future.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Michl P, Pauls S, Gress TM. Evidence-based diagnosis and staging of pancreatic cancer. *Best Pract Res Clin Gastroenterol* 2006;20:227-51.
2. Petrone MC, Archibugi L. The resectable pancreatic ductal adenocarcinoma: To FNA or not to FNA? A diagnostic dilemma, FNA pros. *Endosc Ultrasound* 2017;6:S71-3.
3. Cazacu IM, Luzuriaga Chavez AA, Saftoiu A, *et al.* A quarter century of EUS-FNA: Progress, milestones, and future directions. *Endosc Ultrasound* 2018;7:141-60.
4. Baysal B, Masri OA, Eloubeidi MA, *et al.* The role of EUS and EUS-guided FNA in the management of subepithelial lesions of the esophagus: A large, single-center experience. *Endosc Ultrasound* 2017;6:308-16.
5. Hussain I, Ang TL. Cystic pancreatic lymphangioma diagnosed with endoscopic ultrasound-guided fine needle aspiration. *Dig Endosc* 2017;6:136-9.
6. Ge N, Hu J, Sun S, *et al.* Endoscopic ultrasound-guided pancreatic pseudocyst drainage with lumen-apposing metal stents or plastic double-pigtail stents: A multifactorial analysis. *J Transl Int Med* 2017;5:213-9.
7. Dietrich CF. The resectable pancreatic ductal adenocarcinoma: To FNA or not to FNA? A diagnostic dilemma, introduction. *Endosc Ultrasound* 2017;6:S69-70.
8. Sahai AV. The resectable pancreatic lesion: To FNA or not to FNA? A diagnostic dilemma, FNA cons. *Endosc Ultrasound* 2017;6:S74-5.
9. Porfyridis I, Georgiadis G, Michael M, *et al.* Rapid on-site evaluation with the hemacolor rapid staining method of medical thoracoscopy biopsy specimens for the management of pleural disease. *Respirology* 2016;21:1106-12.
10. Hocke M, Topalidis T, Braden B, *et al.* "Clinical" cytology for endoscopists: A practical guide. *Endosc Ultrasound* 2017;6:83-9.
11. Lopes CV, Dedavid E Silva TL, Coelho NH, *et al.* The value of endoscopic ultrasound-fine needle aspiration in the suspicion of pancreatic hydatid cyst in endemic areas with negative serology (with video). *Endosc Ultrasound* 2017;6:350-1.
12. Wang Y, Chai N, Feng J, *et al.* A prospective study of endoscopic ultrasonography features, cyst fluid carcinoembryonic antigen, and fluid cytology for the differentiation of small pancreatic cystic neoplasms. *Endosc Ultrasound* 2018;7:335-42.
13. Biermann K, Lozano Escario MD, Hébert-Magee S, *et al.* How to prepare, handle, read, and improve EUS-FNA and fine-needle biopsy for solid pancreatic lesions: The pathologist's role. *Endosc Ultrasound* 2017;6:S95-8.
14. Klapman JB, Logrono R, Dye CE, *et al.* Clinical impact of on-site cytopathology interpretation on endoscopic ultrasound-guided fine needle aspiration. *Am J Gastroenterol* 2003;98:1289-94.
15. Tournoy KG, Praet MM, Van Maele G, *et al.* Esophageal endoscopic ultrasound with fine-needle aspiration with an on-site cytopathologist: High accuracy for the diagnosis of mediastinal lymphadenopathy. *Chest* 2005;128:3004-9.
16. Eloubeidi MA, Tamhane A, Jhala N, *et al.* Agreement between rapid onsite and final cytologic interpretations of EUS-guided FNA specimens: Implications for the endosonographer and patient management. *Am J Gastroenterol* 2006;101:2841-7.
17. Hébert-Magee S, Bae S, Varadarajulu S, *et al.* The presence of a cytopathologist increases the diagnostic accuracy of endoscopic ultrasound-guided fine needle aspiration cytology for pancreatic adenocarcinoma: A meta-analysis. *Cytopathology* 2013;24:159-71.
18. Matynia AP, Schmidt RL, Barraza G, *et al.* Impact of rapid on-site evaluation on the adequacy of endoscopic-ultrasound guided fine-needle aspiration of solid pancreatic lesions: A systematic review and meta-analysis. *J Gastroenterol Hepatol* 2014;29:697-705.
19. Iglesias-Garcia J, Dominguez-Munoz JE, Abdulkader I, *et al.* Influence of on-site cytopathology evaluation on the diagnostic accuracy of endoscopic ultrasound-guided fine needle aspiration (EUS-FNA) of solid pancreatic masses. *Am J Gastroenterol* 2011;106:1705-10.
20. Chong CC, Teoh AY, Tang RS, *et al.* EUS-FNA using 22G nitinol or proCore needles without on-site cytopathology. *Endosc Ultrasound* 2018;7:56-60.
21. Kawakami H, Kubota Y. New curved linear echoendoscope for endoscopic ultrasonography-guided fine-needle aspiration in patients with Roux-en-Y reconstruction (with videos). *Endosc Ultrasound* 2018;7:128-9.
22. Wu D, Li JN, Qian JM. Endoscopic diagnosis and treatment of precancerous colorectal lesions in patients with inflammatory bowel disease: How does the latest SCENIC international consensus intersect with our clinical practice? *J Transl Int Med* 2017;5:4-7.
23. Wani S, Mullady D, Early DS, *et al.* The clinical impact of immediate on-site cytopathology evaluation during endoscopic ultrasound-guided fine needle aspiration of pancreatic masses: A prospective multicenter randomized controlled trial. *Am J Gastroenterol* 2015;110:1429-39.
24. Lee LS, Nieto J, Watson RR, *et al.* Randomized noninferiority trial comparing diagnostic yield of cytopathologist-guided versus 7 passes for EUS-FNA of pancreatic masses. *Dig Endosc* 2016;28:469-75.
25. Polkowski M, Jenssen C, Kaye P, *et al.* Technical aspects of endoscopic ultrasound (EUS)-guided sampling in gastroenterology: European Society of Gastrointestinal Endoscopy (ESGE) technical guideline – March 2017. *Endoscopy* 2017;49:989-1006.
26. Kappelle WF, Van Leerdam ME, Schwartz MP, *et al.* Rapid on-site evaluation during endoscopic ultrasound-guided fine-needle aspiration of lymph nodes does not increase diagnostic yield: A randomized, multicenter trial. *Am J Gastroenterol* 2018;113:677-85.
27. Hewitt MJ, McPhail MJ, Possamai L, *et al.* EUS-guided FNA for diagnosis of solid pancreatic neoplasms: A meta-analysis. *Gastrointest Endosc* 2012;75:319-31.
28. Kong F, Zhu J, Kong X, *et al.* Rapid on-site evaluation does not improve endoscopic ultrasound-guided fine needle aspiration adequacy in pancreatic masses: A Meta-analysis and systematic review. *PLoS One* 2016;11:e0163056.
29. van Riet PA, Cahen DL, Poley JW, *et al.* Mapping international practice patterns in EUS-guided tissue sampling: Outcome of a global survey. *Endosc Int Open* 2016;4:E360-70.
30. Iglesias-Garcia J, Lariño-Noia J, Abdulkader I, *et al.* Rapid on-site evaluation of endoscopic-ultrasound-guided fine-needle aspiration diagnosis of pancreatic masses. *World J Gastroenterol* 2014;20:9451-7.
31. Savoy AD, Raimondo M, Woodward TA, *et al.* Can endosonographers evaluate on-site cytologic adequacy? A comparison with cytotechnologists. *Gastrointest Endosc* 2007;65:953-7.
32. Nguyen YP, Maple JT, Zhang Q, *et al.* Reliability of gross visual assessment of specimen adequacy during EUS-guided FNA of pancreatic masses. *Gastrointest Endosc* 2009;69:1264-70.
33. Hayashi T, Ishiwatari H, Yoshida M, *et al.* Rapid on-site evaluation by endosonographer during endoscopic ultrasound-guided fine needle aspiration for pancreatic solid masses. *J Gastroenterol Hepatol* 2013;28:656-63.

34. Khurana KK, Rong R, Wang D, *et al.* Dynamic telecytology for on-site preliminary diagnosis of endoscopic ultrasound-guided fine needle aspiration of pancreatic masses. *J Telemed Telecare* 2012;18:253-9.
35. Khurana KK, Graber B, Wang D, *et al.* Telecytology for on-site adequacy evaluation decreases the nondiagnostic rate in endoscopic ultrasound-guided fine-needle aspiration of pancreatic lesions. *Telemed J E Health* 2014;20:822-7.
36. Landau MS, Pantanowitz L. Artificial intelligence in cytopathology: A review of the literature and overview of commercial landscape. *J Am Soc Cytopathol* 2019;8:230-41.
37. Lariño-Noia J, Iglesias-Garcia J, de la Iglesia-Garcia D, *et al.* EUS-FNA in cystic pancreatic lesions: Where are we now and where are we headed in the future? *Endosc Ultrasound* 2018;7:102-9.
38. Rimbaş M, Crino SF, Gasbarrini A, *et al.* EUS-guided fine-needle tissue acquisition for solid pancreatic lesions: Finally moving from fine-needle aspiration to fine-needle biopsy? *Endosc Ultrasound* 2018;7:137-40.
39. Inoue H, Ogo K, Tabuchi M, *et al.* An Automatic Visual Inspection Method Based on Supervised Machine Learning for Rapid on-Site Evaluation in EUS-FNA. Paper Presented at: 2014 Proceedings of the SICE Annual Conference (SICE); 2014.