## **ECOLOGY**

## Comment on Trophic strategy and bleaching resistance in reef-building corals

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In an era of major environmental changes, understanding corals' resistance to bleaching is as crucial as it is challenging. A promising framework for inferring corals' trophic strategies from Stable Isotope Bayesian Ellipses has been recently proposed to this end. As a contribution to this framework, we quantify a risk of bias inherent in its application and propose three alternative adjustments.

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It is generally accepted that the symbiotic association between corals and their endosymbiotic algae (Symbiodiniaceae) is fundamental to the development of coral reefs as they transfer the major part of their photosynthates to the coral host (autotrophic nutrition) (1). However, corals are considered as mixotrophs, also acquiring energy through host consumption of exogenous organic resources (heterotrophy). Over the past decades, rising seawater temperatures have destabilized the symbiosis between corals and their algal endosymbionts, inducing massive bleaching events. Understanding the mechanisms underlying coral species' resistance to environmental changes or resilience to bleaching is therefore a major research and conservation challenge (2). Previous works have suggested that heterotrophy was one determinant of corals' mortality levels during and following bleaching events [e.g., (3)]. Conti-Jerpe et al. (4) recently provided a groundbreaking contribution to this field, showing that resistance to temperature rise is correlated with trophic strategy in symbiotic corals. Their mesocosm experiment demonstrated that heterotrophic corals were more tolerant to increasing temperatures. To evaluate the trophic strategy in a panel of coral genera, they proposed an innovative method relying on Stable Isotope Bayesian Ellipses [SIBER; (5)]. From both a theoretical and technical point of view, we think that some adjustments would increase the confidence associated with further applications of this promising method to the original mixotrophic nature of corals.

Inferring metabolic processes from statistics unavoidably smooths the path to approximations, whether purposeful or unintended. One risk in comparing trophic status from nitrogen and carbon isotopic niches is the missing of a potential nonhomogeneous distribution of the resource base in lower trophic levels (6, 7). Spatiotemporal variability in the stable isotope values of primary producers, i.e., nitrogen isoscapes, propagates up the food web and is reflected in the isotope values of consumers (8). In other words, different isotopic niches among consumers sampled at different places or times may not reflect differences in diet if their food sources had different isotope values (9). Similarly, divergent symbiont communities between closely located colonies of the same species could result in variable isotope values (10). The study of Conti-Jerpe et al. (4) relies on two distinct datasets: the isotope niche of corals sampled at 23 sites up to 45 km apart and a "temperature-resistance"

mesocosm experiment conducted at one of these locations. SIBER Corrected Standard Ellipses (SEAcs), as applied to the first dataset, were designed specifically for isotopic niche comparisons and to deal with small sample sizes by including 40% of observations around the mean (5). This application of the maximum likelihood hypothesis to multivariate normal distributions is helpful for removing extreme values and to bring broad ecological patterns to light. However, extreme values may also represent ecologically relevant processes (5), particularly in datasets covering large spatiotemporal extents.

In corals, the consumer (host) and the producer (symbiont) belong to the same holobiont. This infra-individual scale implies a maximum difference in mean  $\delta^{15}N$  values of the two fractions around  $3.4 \pm 1.1\%$ , i.e., an average trophic step (11, 12). Technically, this limit reinforces the risk of missing ecologically relevant patterns when using SIBER's SEAc (40%) for coral metapopulations. This risk can be illustrated using the summary statistics of stable isotope analyses for the genus *Platygyra* presented by Conti-Jerpe *et al.* (4). For the 40 individuals of this genus, sampled at 12 stations, located along a steep environmental gradient (13),  $\delta^{13}$ C values of host and symbionts were nearly identical, but  $\delta^{15}N$  means [ $\Delta$ Mean  $\delta^{15}N_{(Host-Symb)}$ ] differed by +2.5% with cumulated SDs of 2%. In such a situation, the overlap between SEAc would be lower than 10% in 95% of cases (Fig. 1), and the Hotelling test would be significant in 100% of cases (2000) simulations). As a consequence, Conti-Jerpe et al. (4) defined this genus as heterotrophic. However, simulations suggest an 85% chance that one or more subgroups do not present distinct niches in the isotope biplot according to the Hotelling  $T^2$  test and may thus be considered autotrophic (Fig. 2). This risk of masking groups that do not have distinct niches is obviously reduced with higher Δmean and ΣSD (Fig. 2). Our example highlights a risk from spatial variations in one dimension of the isotopic biplot, the  $\delta^{15}N$  values. Spatiotemporal variations in  $\delta^{13}$ C values of corals are also plausible (14-16) and would likely affect the bidimensional segregation of trophic niches in the same way. Note that this risk may be considered while applying SIBER to the isotope niche differentiation in other symbiotic organisms such as sponges (17).

From a theoretical perspective, the distribution of corals across a wide range of environmental conditions is certainly the result of adaptive trophic strategies (18). Many studies showed that shifts from autotrophy to heterotrophy can occur within species at varying depth (19) or turbidity (20) or depending on resource availability (21). We are thus convinced that accounting for the adaptive potential of species/genera toward a range of environmental conditions will be determinant in predicting the fate of symbiotic corals.

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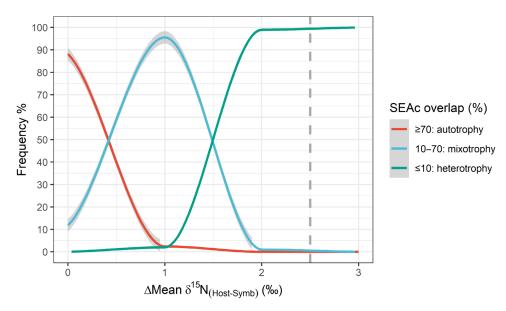


Fig. 1. Probability of the three cutoff values [suggested by (1)] resulting from the use of SIBER's SEAc as a function of the difference between the mean  $\delta^{15}N$  isotopic values of the two considered groups (Host and Symbionts) [N = 40;  $\Sigma SD_{(Host-Symb)} = 2\infty$ ]. Dotted line represents the values obtained for Platygyra sp. [ $\Delta Mean \ \delta^{15}N_{(Host-Symb)} = 2.5\%$ ;  $\Sigma SD_{(Host-Symb)} = 2\%$ ]. Results were obtained from 3500 simulations.

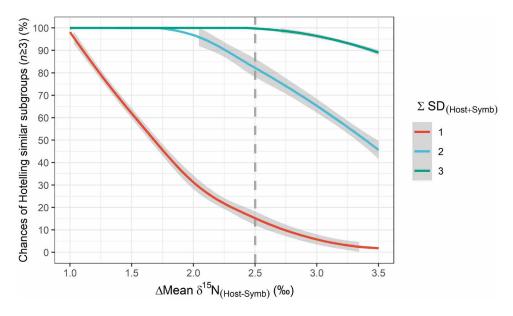


Fig. 2. Chances that a pooled dataset contains at least one subgroup with overlapping isotopic niches as a function of the difference between the mean isotopic values of the two considered groups (Host and Symbionts). The curves correspond to different levels of SD associated with the means of the pooled fractions. Niche segregation is deduced from the P value of Hotelling tests (P > 0.05) for 182,000 simulated data per SD. Dotted line represents the values obtained for P latygyra sp. [ $\Delta$ Mean  $\delta$ <sup>15</sup>N<sub>(Host-Symb)</sub> = 2.5% and  $\Sigma$ SD<sub>(Host-Symb)</sub> = 2].

To better reflect this potential at a fine spatiotemporal scale, we propose three adjustments to the analytical tools proposed by Conti-Jerpe *et al.* (4).

1) Hotelling tests should be conducted at the highest spatiotemporal resolution when possible. This would allow for applying SEAc to remove the least frequent behaviors while controlling for a potential plasticity in the study taxon. This approach may also be used to identify contrasting subgroups and help to consider the best scale for pooling data and designing SEAc. Note that Hotelling tests may be used to compare two groups of data from n = 2 but would be more reliable as much as the sample size increases, hence our choice of  $n \ge 3$  in the presented simulations (Fig. 2).

2) Subgrouping repeated measures demonstrated that SEAcs substantially help to deal with small sample sizes, while both standard ellipses (40% and 95% of observations) produce fair estimates of isotopic niche width for sample sizes >30 (5, 22). Thereby, for spatiotemporally heterogeneous datasets with n > 30, drawing ellipses including 95% of observations may be a reasonable compromise to

lower the risk of mischaracterization due to spatiotemporal variability in isotope values.

3) Conti-Jerpe *et al.* (4) calculated the overlap between SEAc as a proportion of the host SEAc. Considering this overlap as the proportion of the nonoverlapping area may produce a metric more independent from variations of the host isotope niche width (5). While this third adjustment alone would tend to lower the overlap metric between host and symbiont fractions, combination of adjustment two along with this adjustment three will likely produce larger overlap metrics. The cutoff (10% and 70%) values proposed by Conti-Jerpe *et al.* (4) to this overlap metric would thereby make mixotrophic profiles more common. This sounds like a fair adjustment considering the established mixotrophic nature of most symbiotic coral genera, with species/colonies more autotrophic than others.

It is worth mentioning that these SIBER-derived metrics are insensitive to the sign of the  $\Delta$ Mean  $\delta^{15}N_{(Host-Symb)}$ . This difference is found to be positive in most studies, including that of Conti-Jerpe *et al.* (4). However, some reported that host tissues can be 15N-depleted relative to the symbionts, with varying interpretations depending on the context (23, 24). Investigations beyond SIBER-derived inferences may thus be needed in such particular cases.

These "refinements" of the method would undoubtedly account for the ability of symbiotic corals to complement or temporarily replace autotrophic nutrition with heterotrophy, depending on fluctuations of their environment. Conti-Jerpe *et al.*'s (4) approach likely prioritized the characterization of broad trends that matched their observations at the mesocosm study site. We are confident that these comments could contribute to further developments of their promising analytical framework toward comprehensive predictions regarding the fate of coral reefs and a facilitated response to management stakes.

## **REFERENCE AND NOTES**

- L. Muscatine, J. W. Porter, Reef corals: Mutualistic symbioses adapted to nutrient-poor environments. Bioscience 27, 454–460 (1977).
- A. G. Grottoli, L. J. Rodrigues, J. E. Palardy, Heterotrophic plasticity and resilience in bleached corals. *Nature* 440, 1186–1189 (2006).
- K. R. N. Anthony, M. O. Hoogenboom, J. A. Maynard, A. G. Grottoli, R. Middlebrook, Energetics approach to predicting mortality risk from environmental stress: A case study of coral bleaching. Funct. Ecol. 23, 539–550 (2009).
- I. E. Conti-Jerpe, P. D. Thompson, C. W. M. Wong, N. L. Oliveira, N. N. Duprey, M. A. Moynihan, D. M. Baker, Trophic strategy and bleaching resistance in reef-building corals. Sci. Adv. 6, eaaz5443 (2020).
- L. Jackson, R. Inger, A. C. Parnell, S. Bearhop, Comparing isotopic niche widths among and within communities: SIBER—Stable Isotope Bayesian Ellipses in R. J. Anim. Ecol. 80, 595–602 (2011).
- C. A. Layman, D. M. Post, Can stable isotope ratios provide for community-wide measures of trophic structure? Reply. *Ecology* 89, 2358–2359 (2008).
- D. J. Hoeinghaus, S. C. Zeug, Can stable isotope ratios provide for community-wide measures of trophic structure? Comment. Ecology 89, 2353–2357 (2008).
- K. W. McMahon, L. L. Hamady, S. R. Thorrold, Ocean ecogeochemistry: A review. Oceanogr. Mar. Biol. 51, 327–374 (2013).

- J. M. Heikoop, J. J. Dunn, M. J. Risk, T. Tomascik, H. P. Schwarcz, I. M. Sandeman, P. W. Sammarco, δ15 N and δ13 C of coral tissue show significant inter-reef variation. Coral Reefs 19, 189–193 (2000).
- C. B. Wall, M. Kaluhiokalani, B. N. Popp, M. J. Donahue, R. D. Gates, Divergent symbiont communities determine the physiology and nutrition of a reef coral across a lightavailability gradient. *ISME J.* 14, 945–958 (2020).
- M. Minagawa, E. Wada, Stepwise enrichment of 15N along food chains: Further evidence and the relation between δ15N and animal age. Geochim. Cosmochim. Acta 48, 1135–1140 (1984).
- O. Hoegh-Guldberg, L. Muscatine, C. Goiran, D. Siggaard, G. Marion, Nutrient-induced perturbations to δ13C and δ15N in symbiotic dinoflagellates and their coral hosts. *Mar. Ecol. Prog. Ser.* 280, 105–114 (2004).
- N. N. Duprey, M. Yasuhara, D. M. Baker, Reefs of tomorrow: Eutrophication reduces coral biodiversity in an urbanized seascape. Glob. Change Biol. 22, 3550–3565 (2016).
- C. Orejas, A. Gori, C. Rad-Menéndez, K. S. Last, A. J. Davies, C. M. Beveridge, D. Sadd, K. Kiriakoulakis, U. Witte, J. M. Roberts, The effect of flow speed and food size on the capture efficiency and feeding behaviour of the cold-water coral *Lophelia pertusa*. J. Exp. Mar. Biol. Ecol. 481, 34–40 (2016).
- C. Treignier, I. Tolosa, R. Grover, S. Reynaud, C. Ferrier-Pagès, Carbon isotope composition of fatty acids and sterols in the scleractinian coral *Turbinaria reniformis*: Effect of light and feeding. *Limnol. Oceanogr.* 54, 1933–1940 (2009).
- Y. Tanaka, A. Suzuki, K. Sakai, The stoichiometry of coral-dinoflagellate symbiosis: Carbon and nitrogen cycles are balanced in the recycling and double translocation system. ISME J. 12, 860–868 (2018).
- C. J. Freeman, D. M. Baker, C. G. Easson, R. W. Thacker, Shifts in sponge-microbe mutualisms across an experimental irradiance gradient. *Mar. Ecol. Prog. Ser.* 526, 41–53 (2015).
- T. F. Goreau, N. I. Goreau, C. M. Yonge, Reef corals: Autotrophs or heterotrophs? *Biol. Bull.* 141, 247–260 (1971).
- J. E. Palardy, A. G. Grottoli, K. A. Matthews, Effects of upwelling, depth, morphology and polyp size on feeding in three species of Panamanian corals. *Mar. Ecol. Prog. Ser.* 300, 79–89 (2005).
- K. R. Anthony, K. E. Fabricius, Shifting roles of heterotrophy and autotrophy in coral energetics under varying turbidity. J. Exp. Mar. Biol. Ecol. 252, 221–253 (2000).
- M. D. Fox, E. A. Elliott Smith, J. E. Smith, S. D. Newsome, Trophic plasticity in a common reef-building coral: Insights from δ13C analysis of essential amino acids. Funct. Ecol. 33, 2203–2214 (2019).
- J. Syväranta, A. Lensu, T. J. Marjomäki, S. Oksanen, R. I. Jones, An empirical evaluation
  of the utility of convex hull and standard ellipse areas for assessing population niche
  widths from stable isotope data. PLOS ONE 8, e56094 (2013).
- L. J. Rodrigues, A. G. Grottoli, Calcification rate and the stable carbon, oxygen, and nitrogen isotopes in the skeleton, host tissue, and zooxanthellae of bleached and recovering Hawaiian corals. *Geochim. Cosmochim. Acta* 70, 2781–2789 (2006).
- C. Ferrier-Pagès, A. Peirano, M. Abbate, S. Cocito, A. Negri, C. Rottier, P. Riera,
   R. Rodolfo-Metalpa, S. Reynaud, Summer autotrophy and winter heterotrophy in the temperate symbiotic coral *Cladocora caespitosa*. *Limnol. Oceanogr.* 56, 1429–1438 (2011).

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