




# Genome Sequence of *Streptomyces cavourensis* BUU135, Isolated from Soil from a Tropical Fruit Farm in Thailand

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**ABSTRACT** *Streptomyces cavourensis* BUU135 is a bacterial species isolated from the soil of a tropical fruit farm. The genome of *S. cavourensis* BUU135 comprises a gene encoding nebramycin 5' synthase, which produces nebramycin 5' by catalyzing the O-carbamoylation reaction of tobramycin. The newly sequenced 7.66-Mb draft genome of *S. cavourensis* BUU135 may contribute to the discovery of novel natural products derived from this organism.

*Streptomyces cavourensis* is a Gram-positive bacterium found abundantly in soil (1). The type strain of *S. cavourensis* was first reported as a contaminant in marine fungal culture (2). Colonies of *S. cavourensis* are yellow or red and can produce spore chains (2). The temperature for the optimal growth of *S. cavourensis* is 28°C (2).

The bacterial strain BUU135 was isolated from soil from a tropical fruit farm in Chanthaburi Province, Thailand (12°45'31.8"N, 102°01'58.7"E). One gram of the soil sample was diluted in 10 ml of sterile phosphate-buffered saline (PBS) buffer (pH 7.4), and 100  $\mu$ l of the dilution was spread onto International *Streptomyces* Project 2 medium (ISP-2). The culture was incubated at 25°C for 48 h. A single colony was picked and cultured in ISP-2 broth with shaking at 200 rpm at 25°C overnight. Genomic DNA (gDNA) from strain BUU135 was extracted by the phenol-chloroform method described by Sambrook and Russell (3). The NanoDrop spectrophotometer (Thermo Fisher Scientific, USA) was used to determine the quality and quantity of the gDNA. A sequencing library was prepared using the Ion PI Hi-Q OT2 template and Ion Plus fragment library kits. The library was placed on an Ion PI chip, and sequencing was performed using an Ion Proton sequencer (Thermo Fisher Scientific). The average read length was 83 bp. There were 6,289,427 raw reads (depth of coverage, 106 $\times$ ) generated by the sequencing run. The draft genome sequence of strain BUU135 was identified using the Type Strain Genome Server (TYGS) (4). The bacterial strain BUU135 was affiliated with *Streptomyces cavourensis* (DSM 41795) with a digital DNA-DNA hybridization (dDDH) value of 95.4% (4).

The quality of the raw reads was determined using AfterQC version 0.9.6 with default parameters (5). *De novo* genome assembly was performed using the raw reads and SPAdes version 3.13.1 in --careful mode (6). The genome assembly metric was determined using QUAST version 5.0.2 with default parameters (7). The draft genome sequence of *S. cavourensis* BUU135 comprises 7,657,683 bp in 2,754 contigs with an  $N_{50}$  value of 4,468 bp and 71.73% GC content. Genome quality assessment was performed using CheckM version 1.1.3 with default parameters (8). The CheckM analysis revealed

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that the completeness of the *S. cavourensis* BUU135 genome was 96.28%, with 5.34% contamination. Genome annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) with default parameters (9). The annotated genome sequence of *S. cavourensis* BUU135 contains 8,398 total genes, 8,326 protein-coding sequences, 53 tRNA genes, 16 rRNA genes, 3 noncoding RNAs (ncRNAs), and 4 CRISPR regions.

One annotated locus of the *tobZ* gene in the *S. cavourensis* BUU135 genome was determined using Prokka version 1.13.7 with default parameters (10). The *tobZ* gene encodes nebramycin 5' synthase, an enzyme responsible for the production of nebramycin 5' (6''-O-carbamoyltobramycin) by exhibiting ATP-dependent carbamoylation of tobramycin (11, 12). Nebramycin 5' is an aminoglycoside antibiotic with activities against various types of bacterial infections (11, 12). Therefore, *S. cavourensis* BUU135 may be responsible for producing other derivatives of nebramycin 5' with potential antimicrobial activities.

**Data availability.** The whole-genome shotgun sequence of *S. cavourensis* BUU135 has been deposited at DDBJ/ENA/GenBank under accession number [JADZLU000000000](https://www.ncbi.nlm.nih.gov/nuclseq/JADZLU000000000) and SRA accession number [SRR13123741](https://www.ncbi.nlm.nih.gov/sra/SRR13123741).

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## REFERENCES

- Hodgson DA. 2000. Primary metabolism and its control in *Streptomyces*: a most unusual group of bacteria. *Adv Microb Physiol* 42:47–238. [https://doi.org/10.1016/s0065-2911\(00\)42003-5](https://doi.org/10.1016/s0065-2911(00)42003-5).
- Skarbek JD, Brady LR. 1978. *Streptomyces cavourensis* sp. nov. (nom. rev.) and *Streptomyces cavourensis* subsp. *washingtonensis* subsp. nov., a chromomycin-producing subspecies. *Int J Syst Evol Microbiol* 28:45–53. <https://doi.org/10.1099/00207713-28-1-45>.
- Sambrook J, Russell DW. 2001. Preparation and analysis of eukaryotic genomic DNA, p 6.1–6.62. *In* Sambrook J, Russell DW (ed), *Molecular cloning: a laboratory manual*, 3rd ed, vol 1. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>.
- Chen S, Huang T, Zhou Y, Han Y, Xu M, Gu J. 2017. AfterQC: automatic filtering, trimming, error removing and quality control for fastq data. *BMC Bioinformatics* 18:80. <https://doi.org/10.1186/s12859-017-1469-3>.
- Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Pribelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. *J Comput Biol* 20:714–737. <https://doi.org/10.1089/cmb.2013.0084>.
- Mikheenko A, Pribelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUASt-LG. *Bioinformatics* 34:i142–i150. <https://doi.org/10.1093/bioinformatics/bty266>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
- Park JW, Park SR, Han AR, Ban YH, Yoo YJ, Kim EJ, Yoon YJ. 2010. The nebramycin aminoglycoside profiles of *Streptomyces tenebrarius* and their characterization using an integrated liquid chromatography-electrospray ionization-tandem mass spectrometric analysis. *Anal Chim Acta* 661:76–84. <https://doi.org/10.1016/j.aca.2009.12.014>.
- Parthier C, Görlich S, Jaenecke F, Breithaupt C, Bräuer U, Fandrich U, Clausnitzer D, Wehmeier UF, Böttcher C, Scheel D, Stubbs MT. 2012. The O-carbamoyltransferase TobZ catalyzes an ancient enzymatic reaction. *Angew Chem Int Ed Eng* 51:4046–4052. <https://doi.org/10.1002/anie.201108896>.