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## REVIEW

# Stepping forward: T-cell redirecting bispecific antibodies in cancer therapy



Xiaojing Qin<sup>a,b,†</sup>, Wenjing Ning<sup>a,b,†</sup>, Han Liu<sup>a,b</sup>, Xue Liu<sup>a,b,\*</sup>,  
Wenxin Luo<sup>a,b,\*</sup>, Ningshao Xia<sup>a,b</sup>

<sup>a</sup>State Key Laboratory of Vaccines for Infectious Diseases, Xiang An Biomedicine Laboratory, School of Public Health, Xiamen University, Xiamen 361102, China

<sup>b</sup>National Institute of Diagnostics and Vaccine Development in Infectious Diseases, State Key Laboratory of Molecular Vaccinology and Molecular Diagnostics, National Innovation Platform for Industry–Education Integration in Vaccine Research, the Research Unit of Frontier Technology of Structural Vaccinology of Chinese Academy of Medical Sciences, Xiamen University, Xiamen 361102, China

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### KEY WORDS

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Tumor microenvironment;  
Cancer immunotherapy;  
Combination therapy strategies

**Abstract** T cell-redirecting bispecific antibodies are specifically designed to bind to tumor-associated antigens, thereby engaging with CD3 on the T cell receptor. This linkage between tumor cells and T cells actively triggers T cell activation and initiates targeted killing of the identified tumor cells. These antibodies have emerged as one of the most promising avenues within tumor immunotherapy. However, despite success in treating hematological malignancies, significant advancements in solid tumors have yet to be explored. In this review, we aim to address the critical challenges associated with T cell-redirecting bispecific antibodies and explore novel strategies to overcome these obstacles, with the ultimate goal of expanding the application of this therapy to include solid tumors.

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\*Corresponding authors.

E-mail addresses: [wxluo@xmu.edu.cn](mailto:wxluo@xmu.edu.cn) (Wenxin Luo), [liuxue1108@xmu.edu.cn](mailto:liuxue1108@xmu.edu.cn) (Xue Liu).

†These authors made equal contributions to this work.

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## 1. Introduction

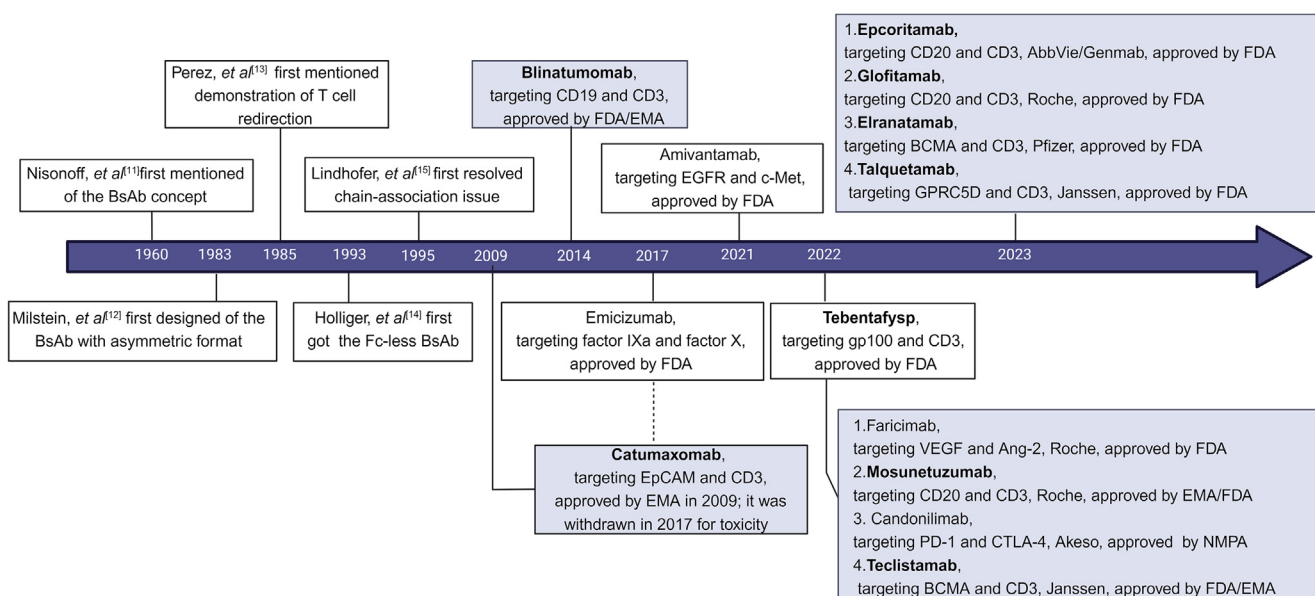
Cancer has an extremely high incidence and mortality rate worldwide, making it one of the significant public health issues currently, posing a severe threat to human health<sup>1</sup>. Immunotherapy, one of the most promising strategies for tumor treatment that harnesses the body's immune system to combat tumor cells selectively, has received significant attention internationally<sup>2–5</sup>. T-cell-based therapies hold a prominent position in the scope of immunotherapy, which mainly involves three primary categories: immune checkpoint inhibitors (ICIs), chimeric antigen receptor (CAR) T-cell therapy, and bispecific antibodies, each addressing the disease through distinct mechanisms. ICIs could block T cell activation negative regulators, such as PD-1 and CTLA-4. By inhibiting the interaction between these checkpoint molecules and their ligands, these therapies rejuvenate the immune system's ability to combat cancer. However, ICIs induce long-term remission only in patients with specific cancer types and only in a minority of patients. CAR T therapy is to couple the potency of a T cell with the specificity of an antibody to kill diseased cells precisely. However, the production of CAR T cells necessitates *ex vivo* genetic modification and proliferation, typically of autologous T cells, which is a time-consuming, relatively inefficient, and costly process. Bispecific antibodies (BsAbs) are composed of two distinct antibodies, enabling them to bind to either two epitopes of the same antigen or two distinct antigens<sup>6,7</sup>. Due to the complexity of the tumor's immunosuppressive microenvironment and the immune escape mechanisms, single immune-modulating strategies still have significant limitations, such as their capacity only to solely target individual molecules, insufficient treatment, or side effects<sup>8,9</sup>. BsAbs, especially T cell-redirecting BsAbs, have emerged as a promising and progressive avenue in T-cell immunotherapy<sup>10</sup>.

So far, over 100 BsAbs are undergoing clinical research. Among these, thirteen BsAbs have been granted market approval, with nine targeting the CD3 receptor (Figs. 1 and 2, and Table 1)<sup>11–15</sup>. In the architecture of a T-cell redirecting BsAb, one arm is designed to bind to the tumor cell surface antigen tumor-

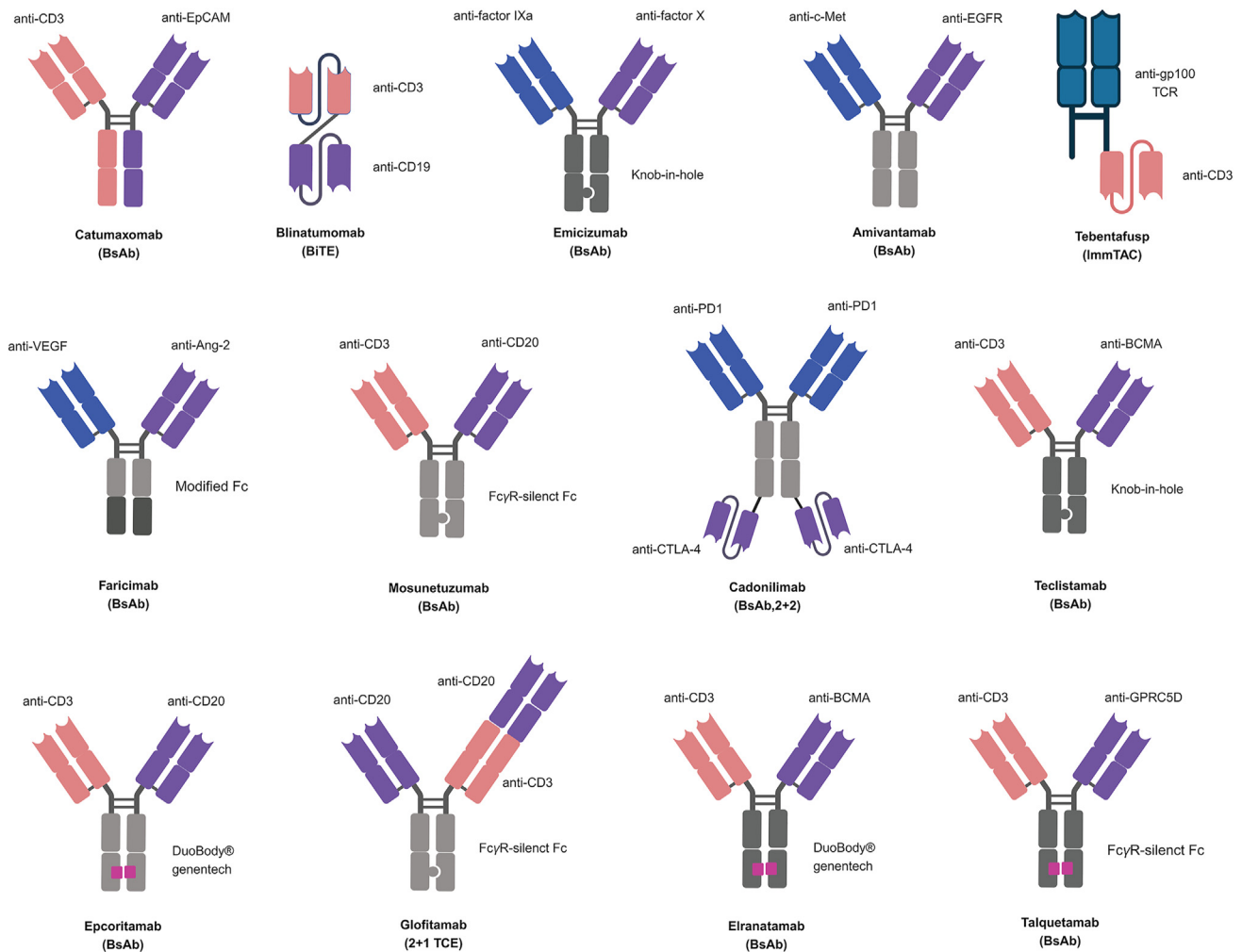
associated antigens (TAA), while the other arm is connected to the CD3 $\epsilon$  subunit of the T cell receptor complex (TCR)<sup>16–19</sup>. These antibodies can remarkably bridge the tumor and T cells, facilitate interactions with T cells, and trigger cytotoxic responses<sup>20,21</sup>. In a concise overview, the T-cell redirecting BsAb establishes an immune synapse upon binding the target and T cells<sup>21</sup>. This synapse enables T cells to transport cytolytic proteins such as perforin and apoptosis-inducing proteins like granzymes) to the target cell membrane, effectively leading to tumor cell killing. The cytolytic activity of CD3<sup>+</sup> T cells is redirected towards tumor cells, promoting their elimination independent of major histocompatibility complex (MHC) molecules, thus bypassing the constraints of MHC limitation<sup>6</sup>. The synapse formation also triggers the cross-linking of TCR, subsequent activation of T cells, releasing pro-inflammatory cytokines, and induction of T cell proliferation. This review provides a broad overview of the current challenges and future perspectives for developing T-cell redirecting BsAbs for cancer treatment, which provides insights into the research and development of novel cancer therapeutics using BsAbs.

## 2. Challenges of T-cell redirecting BsAbs

T-cell redirecting BsAbs face numerous challenges in cancer immunotherapy, mainly toxicity, loss of TAA, the tumor microenvironment, infections, and tumor-lysis syndrome. Toxicity includes on-target off-tumor toxicities (target expression in healthy tissues), increased cytokine release leads to cytokine release syndrome (CRS), and immune effector cell-associated neurotoxicity syndrome (ICANS). The release of tumor cells after lysis also causes metabolic abnormalities. Furthermore, tumor cells can suppress T cell activation and evade immune surveillance through various mechanisms, such as antigen expression loss, antigen endocytosis, or shedding. Immunosuppression of the TME and infection also enhances the side effects of BsAbs (Fig. 3). The clinical presentation ranges from mild symptoms, such as fever, to severe symptoms, such as hypoxia and hypotension.



**Figure 1** Timeline of some major advances in bispecific antibodies.



**Figure 2** Schematic structures of thirteen bispecific antibodies that have been approved. Bispecific antibodies recognize antigens (blue and purple) by two different antigen-binding arms, and most bispecific antibodies are established on IgG1/4 scaffolds. IgG: immunoglobulin G; CD3: cluster of differentiation 3; EpCAM: epithelial cell adhesion molecule; CD19: cluster of differentiation 19; EGFR: estimated glomerular filtration rate; c-Met: cellular-mesenchymal epithelial transition factor; VEGF: vascular endothelial growth factor; Ang-2: angiopoietin-2; PD-1: programmed cell death protein 1; CTLA-4: cytotoxic T lymphocyte associate protein-4; BCMA: B cell maturation antigen; CD20: cluster of differentiation 20.

## 2.1. Toxicity

### 2.1.1. On-target, off-tumor toxicity

The first challenge in T-cell redirecting BsAbs immunotherapy is identifying and differentiating therapeutic targets in normal cells from those in tumor cells. Most membrane proteins are conserved between tumor and healthy cells<sup>22</sup>. Ideal TAAs are limited to a small fraction of tumors, and solid tumor TAAs are usually expressed on tissues of healthy organs<sup>23,24</sup>, resulting in varying degrees of off-tumor toxicity<sup>25</sup>. Some proteins with significant expression differences in solid tumors are intracellular proteins, but traditional antibodies cannot penetrate the cells<sup>26,27</sup>. These intracellular proteins require recognition through HLA class I molecules, such as ImmTACs or TCR-like antibodies<sup>28–30</sup>. TCR-like CD3 BsAbs still exhibit on-target-off-tumor side effects in clinical trials. For example, dose-limiting toxicity was observed in clinical trials of tebentafusp. Tebentafusp is a bispecific protein composed of a soluble T-cell receptor incorporating single-chain variable fragments targeting the CD3 domain<sup>31–33</sup>. This high-affinity, high-

specificity T-cell receptor targets a 9-amino acid peptide derived from intracellular gp100 protein through proteolytic degradation, presented by HLA molecules on the surface of target cells (including melanocytes and melanoma cells). By targeting tumor-associated antigens specifically shared by tumors, these T-cell receptor bispecific molecules can recruit T cells and direct them toward tumors. Patients treated with tebentafusp experienced skin-related adverse events, possibly due to the recognition of gp100-expressing melanocytes. In order to improve the selectivity and specificity of tumor antibodies and reduce off-tumor toxicity, many researchers have designed novel T-cell redirecting BsAbs. For example, they enhance the binding to tumor cells by adding a second tumor-associated antigen (TAA) binding domain. If the affinity to the TAA is too high, it may lead to binding to low-expressing TAAs on healthy cells. On the other hand, if the affinity to the TAA is too low, it may affect the efficiency of T-cell redirecting BsAbs. Besides, the design of antibodies can be improved to develop BsAbs with a modulatory activity that is only activated in the acidic tumor microenvironment. The acidic pH is due to the

**Table 1** Overview of clinical advances in T-cell redirecting bispecific antibodies in the treatment of tumors.

Drug	Target	Indication	Phase	Clinical trial	Status	Last update posted	Research institution
Cibisatamab	CD3 × CEA	Metastatic colorectal cancer	II	NCT04826003	Recruiting	2023/8/1	Roche Holding AG
Nivatrotamab	CD3 × GD2	Neuroblastoma, small cell carcinoma, osteosarcoma	II	NCT03033303	Active, not recruiting	2023/7/5	Y-mAbs Therapeutics, Inc. Memorial Sloan Kettering Cancer Center
BNT-142	CD3 × CLDN6	Solid tumor	I/II	NCT05262530	Recruiting	2023/6/22	BioNTech SE
GEN-1047	CD3 × B7-H4	Breast cancer, endometria cancer, ovarian cancer	I/II	NCT05180474	Recruiting	2023/4/21	Genmab A/S
GEN1044	CD3 × 5T4	Locally advanced or metastatic solid tumor(s), prostate cancer, esophageal cancer	I/II	NCT04424641	Terminated	2023/2/1	Genmab A/S
CD30 biAb-AATC	CD3 × CD30	Pediatric cancer, Hodgkin disease, CD30-positive diffuse large B-cell lymphoma	I/II	NCT05544968	Not yet recruiting	2022/9/19	Medical College of Wisconsin Research Foundation
CM-350	CD3 × GPC3	Advanced solid tumor	I/II	NCT05263960	Recruiting	2022/6/9	Keymed Biosciences Inc.
RO-7444973	CD3 × MAGEA4	Solid tumor	I	NCT05129280	Terminated	2023/8/4	Roche Holding AG
A-337	CD3 × EpCAM	EpCAM expressing solid tumours	I	CTR20232278	Not yet recruiting	2023/8/2	Evive Biotech
Runimotamab	CD3 × HER2	Solid tumor	I	NCT03448042	Recruiting	2023/7/13	Genentech, Inc.
ERY-974	CD3 × GPC3	Hepatocellular carcinoma (HCC)	I	NCT05022927	Recruiting	2023/7/10	Chugai Pharmaceutical Co., Ltd.
AMG-199	CD3 × MUC17	MUC17-positive solid tumors	I	NCT04117958	Terminated	2023/6/22	BeiGene Pharmaceuticals (Guangzhou) Co., Ltd.
BA-1202	CD3 × CEA	Advanced solid tumor	I	NCT05909241	Not yet recruiting	2023/6/22	Luye Pharma AG
Vixtimotamab	CD3 × CD33	AML, childhood	I	NCT05077423	Terminated	2023/5/30	Amphivena Therapeutics, Inc.
BA-3182	CD3 × EpCAM	Advanced adenocarcinoma	I	NCT05808634	Not yet recruiting	2023/4/11	BioAtla, Inc.
EMB-07	CD3 × ROR1	Advanced/metastatic solid tumors	I	NCT05607498	Recruiting	2023/3/17	EpimAb Biotherapeutics, Inc.
CX-904	CD3 × EGFR	Solid tumor	I	NCT05387265	Recruiting	2023/2/28	Amgen, Inc.
JNJ-63898081	CD3 × PSMA	Neoplasms	I	NCT03926013	Completed	2022/11/7	Janssen Research & Development LLC
GBR-1342	CD3 × CD38	HER2 expressing solid tumors	I	NCT02829372	Terminated	2020/10/9	Ichnos Sciences, Inc.
PF-6671008	CD3 × CDH3	Neoplasms	I	NCT02659631	Terminated	2020/5/6	MacroGenics, Inc.

CEA: carcinoembryonic antigen; GD2: disialoganglioside GD2; CLDN6: claudin-6; GPC3: glypican-3; MAGEA4: melanoma-associated antigen A4; HER2: human epidermal growth factor receptor 2; ROR1: inactive tyrosine-protein kinase transmembrane receptor; EGFR: epidermal growth factor receptor; PSMA: prostate-specific membrane antigen; CDH3: cadherin-3.

lactic acid produced by the glycolytic pathway used by tumor cells. The acidic environment creates an immunosuppressive microenvironment that helps tumor cells evade the immune system while also affecting the bioavailability of classical antibodies. Researchers have developed selective antibodies and CAR-T cell therapy technologies based on acidic pH, which can increase the TME targeting of antibodies and CAR-T cells and reduce on-target, off-tumor toxicity. BA3182, a conditionally active biologic, possesses two binding domains for EpCAM and CD3ε. *In vitro* studies reveal that BA3182 can specifically and reversibly bind to the target proteins EpCAM and CD3 under the acidic conditions characteristic of the tumor microenvironment. Conversely, at the physiological alkaline pH of 7.4 found in normal cells, its binding affinity is considerably diminished. *In vivo* research indicates that BA3182 offers a therapeutic index surpassing non-CAB variants by over a hundredfold,

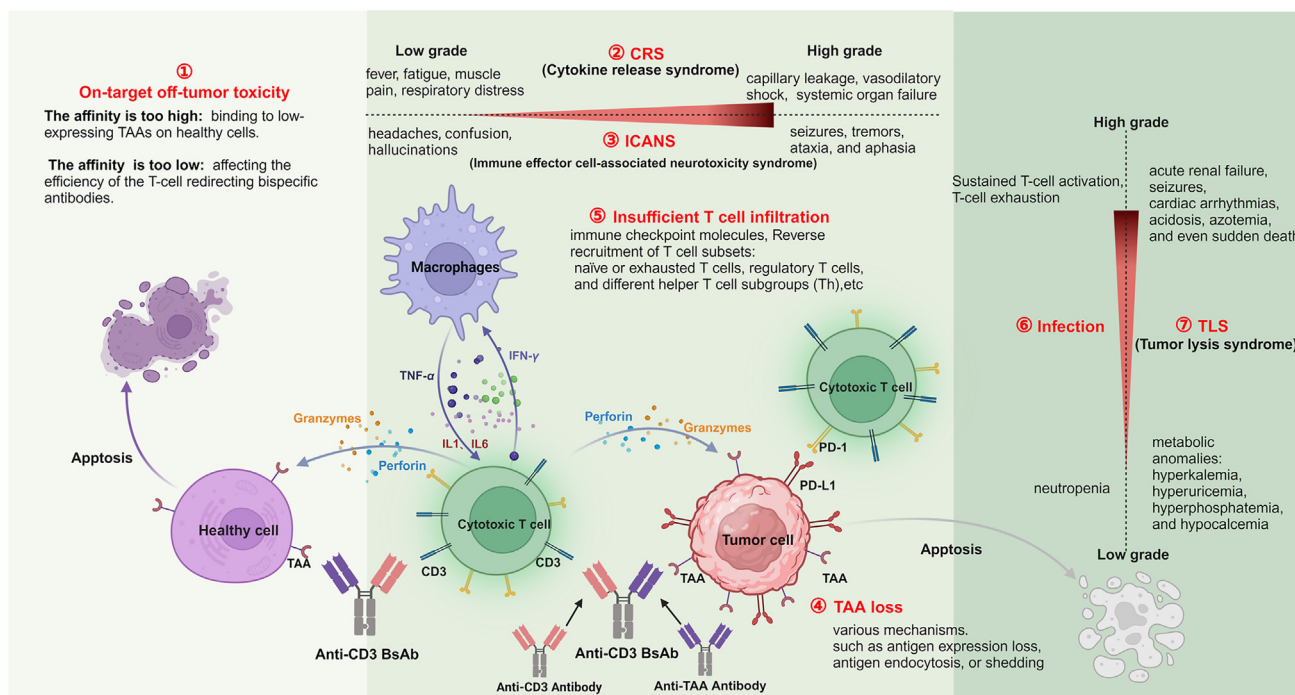
underscoring the immense potential of CAB BsAbs compared to their traditional counterparts (NCT05808634).

Furthermore, by combining bispecific antibodies with other forms of treatment, such as immunomodulators and small molecule drugs, comprehensive strategies can significantly enhance the selectivity and specificity of bispecific antibodies, maximize their therapeutic effects, and simultaneously reduce off-target toxicity to normal tissues.

### 2.1.2. CRS

Cytokine release syndrome (CRS) emerges due to excessive activation of immune cells, marking a severe immune-related adverse reaction<sup>33</sup>. CRS is characterized by a substantial surge in pro-inflammatory cytokines within the bloodstream, including IL-6, IL-10, TNF-α, and IFN-γ, leading to a cytokine storm.





**Figure 3** Challenges of T-cell redirecting bispecific antibodies in cancer immunotherapy. ① On-target, off-tumor toxicity. A subset of solid tumor antigens is expressed in tissues of healthy organs, resulting in varying degrees of off-tumor toxicity. ② Cytokine release syndrome (CRS). A significant surge of proinflammatory cytokines due to overactivation of immune cells leads to a cytokine storm. ③ Immune Effector Cell-Associated Neurotoxicity Syndrome (ICANS). ④ TAA loss. Tumor cells can suppress T cell activation and evade immune surveillance through various mechanisms, such as antigen expression loss, antigen endocytosis, or shedding. ⑤ Tumors microenvironment (TME). It includes insufficient T cell infiltration and reverse recruitment of T cell subsets. A significant feature of solid tumors is the presence of an immunosuppressive TME. Secondly, in addition to recruiting effector T cells, T-cell redirecting bispecific antibodies also recruit other immune cells, which may lead to drug resistance. ⑥ Infection. Infections are common in patients receiving bispecific antibody therapy, and combination therapy increases the risk of infection. ⑦ Tumor-lysis syndrome (TLS) is attributed to the breakdown of tumor cells, which release their contents into the systemic circulation, resulting in metabolic abnormalities.

Clinical manifestations of CRS include fever, fatigue, muscle pain, respiratory distress, capillary leakage, vasodilatory shock, and systemic organ failure<sup>34</sup>.

The pioneering BsAb drug, Catumaxomab, showed promising efficacy in treating malignant ascites. However, its strong immunogenicity gave rise to pronounced hepatotoxicity and triggered a high-intensity cytokine storm when it went off-target, leading to its withdrawal from the market in 2017<sup>35</sup>. CRS is the predominant toxicity concern in therapies involving BsAbs that bridge T cells<sup>36</sup>. Existing pharmacological interventions, such as the immunosuppressant agent dexamethasone and the IL-6 receptor inhibitor tocilizumab, offer promising avenues for mitigating CRS. In 2017, due to insufficient endogenous immune response to clear patients' cancer, Ishiguro and colleagues designed a BsAb targeting CD3/GPC3 for solid tumors. In a monkey model, the main adverse event was cytokine storm, mitigated by pre-treatment with dexamethasone without compromising anti-tumor efficacy. In the dose-escalation phase I trial, 29 subjects were enrolled, with a dose range of 0.003–0.81  $\mu\text{g}/\text{kg}$ <sup>37–39</sup>. The results showed that more than 20% of the patients experienced CRS reactions and fever. The dose of 0.81  $\mu\text{g}/\text{kg}$  was found to be intolerable; however, doses lower than 0.81  $\mu\text{g}/\text{kg}$  were generally well-tolerated, and adverse events could be managed with corticosteroids<sup>40</sup> and anti-IL-6 receptor inhibitors (NCT05022927).

TNB-383B, co-developed by Tenebio and AbbVie, is a novel BsAb featuring a unique structural configuration: two  $\alpha\text{BCMA}$

molecules on one arm, a singular  $\alpha\text{CD3}$  arm, and a silent IgG4 Fc segment<sup>41</sup>. This design endows it with a potent T-cell activating capacity and a low-affinity anti-CD3 activity, which balances robust cytotoxic effects with minimized cytokine release. In phase I clinical trials, TNB-383B is evaluated for safety, tolerability, and pharmacokinetics, including its maximum concentration and half-life in the circulatory system. Preliminary data suggest a dosage-dependent therapeutic response, with an impressive 80% overall response rate (ORR) at doses equal to or exceeding 40  $\text{mg}$ <sup>42</sup>. Common side effects encountered in the trials include CRS, fatigue, headaches, reduced blood cell counts, infections, and nausea.

In November 2023, the BsAb targeting CD3/CD20, Glofitamab, was approved by the NMPA for marketing in China. Before that, in March 2023, Glofitamab was approved for marketing in Canada; in June, it was approved by the US Food and Drug Administration (FDA) for marketing. Glofitamab, a 2 + 1 CD3/CD20 BsAb, targets T cells to malignant B cells to exert its anti-tumor effects<sup>43,44</sup>. In a Phase II clinical trial for the treatment of aggressive lymphoma, CRS was the most common adverse reaction, occurring in the majority (approximately 63%) of patients, mainly at Grade 1 (47.4%) and Grade 2 (11.7%), and one patient discontinued the trial due to CRS.

BsAbs currently demonstrate potential effectiveness in treating certain malignant tumors; however, CRS, as a significant adverse effect issue, remains to be addressed. As a result, future research may focus more on the design and improvement of BsAbs. These

antibodies aim to maximize the targeted effects on tumor cells while minimizing the impact on healthy tissues and controlling the host's inflammatory response. Additionally, drugs such as dexamethasone used in pre-treatment have shown potential in reducing CRS induced by BsAbs treatment. Future therapies may adopt more of this kind of strategy, which includes using more advanced immunosuppressants or inflammation pathway-specific inhibitors to decrease the toxic side effects associated with treatment.

### 2.1.3. Neurotoxicity

Neurotoxicity of BsAbs in tumor immunotherapy is often an indirect consequence, transpiring concurrently with CRS or shortly after CRS<sup>45–49</sup>. Nevertheless, it can also occur independently of CRS<sup>41,42</sup>. This form of neurotoxicity is referred to as ICANS and is characterized by a range of symptoms, including headaches, confusion, hallucinations, seizures, tremors, ataxia, and aphasia<sup>39</sup>. The risk of ICANS can be mitigated through dose escalation and premedication<sup>45</sup>. Supportive care interventions for ICANS-affected patients involve the administration of tocilizumab (when CRS is concurrent), corticosteroids, or antiepileptic drugs.

Neurotoxicity has been observed in a subset of patients with B-cell hematological malignancies who underwent T-cell engager therapy<sup>50–56</sup>. Blinatumomab, a T-cell redirecting BsAb, has been developed to target CD19, an antigen expressed on B-cell acute lymphoblastic leukemia cells. In a multicenter, single-arm, phase 2 clinical trial, blinatumomab rarely induced adverse events in tumor immunotherapy; they were largely transient and resolved effectively. However, a majority of these adverse events exhibited a notable association with a high rate of neurotoxicity (all grades at 47%–53%; grade 3 or higher at 7%–13%)<sup>57–60</sup>. Compared to treatments targeting other antigens, CD19-directed BsAbs demonstrated a heightened frequency of neurotoxicity, a phenomenon potentially attributed to the presence of CD19 on cells in the blood–brain barrier<sup>61</sup>.

Clinicians can formulate precision therapeutic strategies by deepening our understanding of the risk factors and mechanisms underlying neurotoxicity. These strategies involve adjusting dosages and selecting appropriate antibody treatments contingent on a patient's clinical symptoms, genetic predispositions, and immune system status. Enriched knowledge of ICANS paves the way for innovative support treatments. The future holds promise for BsAbs and related immunotherapies to reduce side effects and enhance patient safety while effectively combatting cancer.

### 2.2. TAA loss

The development of T-cell redirecting BsAbs is predicated on identifying and TAAs. Nevertheless, tumor cells can inhibit T cell activation and circumvent immune detection through several strategies, including losing antigen expression, antigen endocytosis, or shedding. The immune system's elimination of cells exhibiting tumor-specific antigens can give rise to variations of the tumor that lack these antigens. Additionally, analogous to viral "antigenic drift", tumor cells can evade T cell-mediated destruction by modifying their antigenic profile through epitope mutations. The CD19 antigen has emerged as an effective target for the immunotherapeutic treatment of acute lymphoblastic leukemia, yet resistance to the BsAb blinatumomab, targeting CD3/CD19, has been observed during treatment<sup>62</sup>. Flow cytometry analysis of tumor tissues from resistant patients revealed mutations in the CD19 antigen, indicative of TAA loss. Further analysis showed no structural anomalies in the CD19 gene, suggesting the presence of

complete antigen loss escape variants, as opposed to single-epitope loss or splice variants.

BsAb research faces obstacles in treating mature solid tumors such as gastrointestinal cancer, melanoma, and glioma due to the scarcity of identifiable tumor-specific antigens and the underlying heterogeneity of solid tumor cells. Therefore, selecting optimal targets predominantly found on tumor cells and displaying minimal expression on healthy cells to reduce collateral damage to normal tissue while enhancing treatment specificity is crucial. Given tumors' diverse nature and variability, therapies concentrating on a single antigen may prove insufficient to eradicate cancer cells. Future research should pivot towards combination therapies that concurrently target multiple antigens, thereby increasing the likelihood of therapeutic success and mitigating the risk of resistance emergence.

### 2.3. Tumors microenvironment

#### 2.3.1. Insufficient T cell infiltration

Compared to hematologic malignancies, one distinguishing feature of solid tumors is the presence of an immunosuppressive TME<sup>63,64</sup>. The TME encompasses a complex interplay of elements, including immune suppressor cells, stromal cells, immune suppressor cytokines, and soluble molecules, all inhibiting anti-tumor immunity<sup>29</sup>. Both tumor cells and immune suppressor cells express immune checkpoint molecules, which upregulation often leads to tumor cell immune evasion<sup>64–66</sup>. Moreover, tumor cells employ other tactics to evade immune system surveillance. Under normal circumstances, there exists a dynamic balance between molecules of immune activation and immune suppression<sup>67</sup>, collectively maintaining the stability of the body's immune system<sup>68</sup>. However, excessive expression of immune suppressor factors on tumor or T cells disrupts T cell activity, leading to exhaustion and restraining the infiltration of CD8<sup>+</sup> T cells into the tumor microenvironment<sup>69–71</sup>. Inadequate tumor-infiltrating lymphocytes into tumors may also be attributed to prolonged antigen stimulation in the TME<sup>71</sup>, causing a decline in the frequency of tumor-specific T cell clones and a reduction in effector T cells, culminating in T cell exhaustion<sup>72,73</sup>.

#### 2.3.2. Reverse recruitment of T-cell subsets

T-cell redirecting BsAbs are designed to interact with tumor target cells and effector T cells, as brief contact can promote the formation of an immune synapse between T cells and tumor cells<sup>74</sup>. Afterward, the presence of perforin and granzyme in the immune synapse is observed<sup>75</sup>, subsequently facilitating the elimination of tumor cells by T cells. Most cytotoxic T cells (CTL), such as CD8<sup>+</sup> T cells, CD4<sup>+</sup> T cells,  $\gamma\delta$ T cells, and NKT cells, can participate in tumor cell lysis through this mechanism<sup>76</sup>. CD8<sup>+</sup> T cells within CTL are the primary effector cells<sup>77</sup>. Intravenous administration of T cell redirecting bispecific antibodies has shown the ability to recruit or activate CTL cells directly, alongside recruiting other immune cells<sup>78</sup>. However, this recruitment is not exclusive and may encompass various T cell subgroups, including those with suppressive functions such as naïve or exhausted T cells, regulatory T cells, and different helper T cell subgroups (Th). This indiscriminate recruitment could lead to drug resistance, weakening the therapeutic potential of tumor cell killing by CD8<sup>+</sup> T cells. Notably, a clinical study has indicated that resistance to Binatumomab arises from CD3<sup>+</sup> CD4<sup>+</sup> CD25<sup>+</sup> hiFoxP3<sup>+</sup> Treg cells<sup>20</sup>, which hinder the dissolution of CD8<sup>+</sup> T cells and suppress their tumor-killing capabilities. Additionally, the recruited CD4<sup>+</sup> T cells,

predominantly Th1 and Th17, often release cytokines and spark a cytokine storm. Therefore, enhancing the specificity of BsAbs can lead to better cell killing and reduce cytotoxic effects. Furthermore, exploring alternative targets for T cell recruitment or activation, such as CD28, emerges as a strategic avenue to address the non-selective recruitment of T cells by BsAbs in immune therapy.

#### 2.4. Infection

Infections are frequent in patients treated with BsAbs, although their frequency varies considerably across diverse clinical studies<sup>79,80</sup>. Apart from tumor type, this could be associated with the extent of prior immunosuppressive therapy. The frequency of neutropenia caused by BsAbs may also fluctuate among hematological malignancies<sup>81–83</sup>. Furthermore, sustained T cell activation mediated by BsAbs may lead to T cell exhaustion, thereby increasing the risk of infection. Additionally, the combinations of T-cell redirecting BsAbs with other treatment strategies may also increase the incidence and severity of infections.

#### 2.5. Tumor-lysis syndrome (TLS)

TLS represents a metabolic emergency attributed to the breakdown of a substantial number of tumor cells, which release their contents into the systemic circulation. This phenomenon results in metabolic anomalies like hyperkalemia, hyperuricemia, hyperphosphatemia, and hypocalcemia<sup>84</sup>. These metabolic abnormalities can lead to acute renal failure, seizures, cardiac arrhythmias, acidosis, azotemia, and even sudden death. Although BsAbs can potentially reduce tumor burden, the development of TLS may also occasionally occur. Patients at risk of TLS, particularly those with elevated tumor burden or impaired renal function, should receive prophylaxis measures such as uric acid-lowering medications and adequate hydration<sup>85–88</sup>.

Building on our understanding of TLS, future research and clinical practices should prioritize the identification of reliable biomarkers for predicting TLS risk, thereby enabling prophylactic interventions before its onset. Given that acute renal failure is one of the principal complications associated with TLS, developing strategies and pharmaceuticals to preserve renal function represents a critical area of investigation.

### 3. Implications for treatment strategies

#### 3.1. Development of T-cell redirecting BsAbs in different formats

##### 3.1.1. Antibody potency

The design of BsAbs configurations can impact the target binding site<sup>89</sup>. A bivalent BsAb featuring one binding site for each target is called 1 + 1; introducing additional binding sites can result in tri-specific antibodies (2 + 1) or quadrivalent BsAb (2 + 2 or 1 + 3). Although traditional BsAbs maintain a 1:1 ratio of CD3 and TAA<sup>90</sup>, enhancing the binding capacity through a 1 + 2 design may bolster the recognition of tumor cells while avoiding activation of CD3 on T cells in the absence of TAA. IGM2323 is a 10+1 type targeting CD20/CD3<sup>91</sup>. IGM-2323 is structured by the fusion of anti-CD3 scFv to the J chain, forming a 10:1 antibody form of anti-CD20: anti-CD3. IGM-2323 superimposed the TDCC effect on the action of CDC and played a significantly enhanced B-cell clearance effect, demonstrating the synergistic effect of

BsAb (NCT04082936). Compared to rituximab, IGM-2323 has a 1000-fold increase in killing activity.

Among the BsAbs that have been marketed, Roche has two BsAbs targeting CD20/CD3—Mosunetuzumab and Glofitamab<sup>92–96</sup>. Although the two new drugs are dual antibodies with the same target, their structure and indications differ. Mosunetuzumab has a 1 + 1 structure, while Glofitamab has a 2 + 1 structure. The former is approved for follicular lymphoma and the latter for treating DLBCL. This 2 + 1 structure design makes it more binding to CD20 on the surface of B cells, induces rapid T cell activation and cytokine release, and leads to target cell lysis. The activity is 10–1000 times higher than traditional 1 + 1 BsAbs. In the Phase Ib/II GO40516 study, Mosunetuzumab and Polivy achieved an ORR of 65.0% in severely treated R/R NHL patients, with a CR of 48.3%. CRS occurred in 18%, and all adverse events occurred in the first treatment cycle and were grade 1 to 2. In the Phase I/Ib dose-escalation NP30179 study, Glofitamab alone achieved an ORR of 81.0% for patients with R/R NHL who received severe treatment and 100% ORR in the glofitamab plus Gazyva group. In the Phase Ib/II NP39488 study, Glofitamab and Polivy had an ORR of 73% over a 3.2-month follow-up period, including a CR of 51.5%, and no CRS above grade 3 was observed.

##### 3.1.2. Tri-specific antibody

Among the successful cancer immunotherapies, approved BsAbs such as Blinatumomab have stimulated researchers' interest in other multispecific antibody therapies<sup>97</sup>. T-cell redirected tri-specific antibodies represent a promising multi-targeting immune intervention; some are currently in the clinical research stage (see Table 2). Tri-specific antibodies extend from BsAbs by introducing an extra antigen binding site, totaling three sites. This enables simultaneous interaction with target and effector cells (typically T cells or NK cells), enhancing the killing of target cells<sup>97,98</sup>. Compared to monoclonal and BsAbs, some tri-specific antibodies have two additional specific antigen binding sites, which are advantageous for increasing binding specificity, improving the accuracy of drug targeting to tumor cells, and reducing off-target toxicity<sup>99,100</sup>. Furthermore, some tri-specific antibodies bind to tumor antigens through one binding site; the second binding site on the other arm can specifically bind to the TCR on T cells, thereby activating the first signal pathway and guiding T cells to the vicinity of target cells to achieve the aggregation/localization of immune cells<sup>101</sup>. The third binding site on the arm triggers the second signal pathway, further activating immune cells. Those tri-specific antibodies have a strong cytotoxic effect on target cells through bridging and activating immune cells simultaneously or bridging immune cells and blocking dual signal pathways.

For example, Wu et al.<sup>100</sup> and Sun et al.<sup>102</sup> developed a T-cell redirected tri-specific antibody, SAR-442257, that interacts with CD38, CD3 and CD28. The engagement of CD3 and low-affinity CD28 affords efficient T cell activation ability, whereas the anti-CD38 antibody directs T cells to certain lymphomas, leukemias, and myeloma cells. *In vitro*, SAR-442257 showed 1000–10,000-fold higher cytotoxic activity against myeloma cells than CD38 monoclonal antibody. This promising approach is being explored for relapsed/refractory multiple myeloma (MM) and non-classical Hodgkin's lymphoma patients within phase I clinical trials (NCT04401020). Sun et al.<sup>102</sup> and Seung et al.<sup>103</sup> developed a HER2/CD3/CD28 tri-specific antibody based on the HER2/CD3 BsAb. Coupling an anti-CD28 variable domain to the anti-CD3 variable region in a humanized mouse model, regression of breast cancer cells mediated by CD4<sup>+</sup> and CD8<sup>+</sup> T cells was

**Table 2** Representative clinical trials of the combination strategy.

Combination strategy	Antibody	Target	Phase	Clinical trial	Adaptive symptom	Source of information
Antibody potency	IGM2323, 1 + 10	CD3/CD20	I/II	NCT04082936	Non-Hodgkin lymphoma, follicular lymphoma, DLBCL	<a href="https://www.clinicaltrials.gov/">https://www.clinicaltrials.gov/</a>
Trispecific antibody	Glofitamab, 2 + 1	CD3/CD20	Marketing	–	DLBCL	2023, FDA
	HPN-424	CD3/PSMA/albumin	I/II	NCT03577028	Prostatic cancer	<a href="https://www.clinicaltrials.gov/">https://www.clinicaltrials.gov/</a>
	CMG1A46	CD3/CD19/CD20	I/II	NCT05348889	Acute lymphoblastic leukemia, non-Hodgkin lymphoma, hematologic neoplasms	
	PIT-565	CD2/CD3/CD19	I	NCT05397496	B-cell lymphoma, adult lymphoblastic lymphoma, non-hodgkin lymphoma	
	SAR-442257	CD3/CD28+/CD38	I	NCT04401020	Neoplasms, non-Hodgkin lymphoma, multiple myeloma	
Pro-drug	JNJ-80948543	CD3/CD20/CD79B	I	NCT05424822	B-cell chronic lymphocytic leukemia, non-Hodgkin lymphoma	
	AMX-818	CD3+HER2/EGFR	I	NCT05356741	Locally advanced or metastatic HER2-expressing cancers	Ref [180]
	ADG138	CD3+HER2	IND-enabling	–	Solid tumors	AACR poster, Adagene's official website
Mobilization of other immune cells	CX-904	CD3+EGFR	I	NCT05387265	Solid tumours	AACR poster, Adagene's official website
	AFM13	CD30/CD16	I/II	NCT04074746	Anaplastic large cell lymphoma, B-cell non-Hodgkin lymphoma, classic Hodgkin lymphoma	<a href="https://www.clinicaltrials.gov/">https://www.clinicaltrials.gov/</a>
	Reducing affinity of CD3 arm	Elranatamab	CD3/BCMA	IV	NCT06057402	Multiple myeloma
TNB-585		CD3/PSMA	I	NCT04104607	Castration resistant metastatic prostate cancer	Ref [126]
MCLA-117		CD3/CLL	I	NCT03038230	Acute myeloid leukemia	Ref [130]
Combining with chemotherapy drugs	NILK-2301	CD3/CEACAM5	Drug discovery	–	CEACAM5-expressing cancers	Ref [181]
	Chemotherapy+ Mosunetuzumab	CD3+CD20	I	NCT05464329	Diffuse large B cell lymphoma, high-grade B-cell lymphoma, transformed B-cell lymphoma	<a href="https://www.clinicaltrials.gov/">https://www.clinicaltrials.gov/</a>
	Combining with radiation therapy	p53-SADA-BsAb	GD2	I	–	Non-small cell lung cancer, melanoma, Sarcoma
Combining with oncolytic viruses	EphA2 TEA VV	EphA2	Drug discovery	–	Solid tumors	Ref [140]
	OAd-MUC16-BiTE	MUC16	Drug discovery	–	Ovarian cancer	Ref [165]
Combining with targeted therapy	BHKis + CD3/CD19 BsAb	CD3+CD19	Drug discovery	–	Chronic lymphocytic leukemia	Ref [170]
Combining immune checkpoint inhibitors	M7824	PD-L1+TGF $\beta$	II	NCT03840902	Non-small cell lung cancer	<a href="https://www.clinicaltrials.gov/">https://www.clinicaltrials.gov/</a>
	SHR1701	PD-L1+TGF $\beta$	I/II	NCT04407741	Solid tumors, lymphomas	



observed. The single-valent anti-CD28 utilized in this tri-specific antibody did not induce the severe cytokine release observed previously with the bivalent superagonist anti-CD28 agents. This antibody is currently undergoing phase I clinical trials (NCT05013554). T-cell redirecting BsAbs emerge as an ideal anti-tumor therapy, representing a multitargeted immunotherapeutic intervention with promising prospects for immunotherapy against human cancers.

### 3.1.3. Pro-drug

T-cell redirected bispecific antibodies, exemplified by Blinatumomab (CD19/CD3), have succeeded in hematologic malignancies<sup>104–107</sup>. However, in solid tumors, the development of such BsAbs faces challenges, primarily due to T-cell overstimulation or on-target off-tumor effects causing damage to normal tissues<sup>20,108</sup>. Researchers are addressing these issues from multiple angles, one approach being tumor-specific activation of bispecific antibodies. The pro-drug design is used to reduce the specific killing of normal tissue cells by BsAb drugs<sup>109</sup>, significantly improving the effectiveness and safety of BsAb drugs<sup>110</sup>. In normal tissues, BsAb drugs exist in the form of pro-body without activity<sup>111</sup>. The TME harbors abundant matrix metalloproteinases<sup>112</sup>. Upon reaching specific sites in the tumor area, they undergo enzymatic cleavage by certain enzymes, removing a small peptide segment that masks the binding site, thereby enabling the BsAb to exert its function<sup>111,112</sup>.

Tianyan Pharmaceuticals' SAFEbody BsAb platform technology effectively employs this concept in design. ADG138 is a BsAb targeting HER2 × CD3, where the epitopes of HER2 and CD3 are masked<sup>113</sup>. Upon reaching the target site, the masking peptide is removed, exposing the Fab region of the antibody and revealing the binding sites for anti-HER2 and anti-CD3. This allows for the normal binding of the antibody to the antigens on tumor cells, recruiting and activating T cells to kill tumor cells selectively. CI107, a BsAb, consists of the cetuximab antibody targeting EGFR and the humanized SP34 antibody targeting CD3, fused in scFv form at the N-terminus of the cetuximab antibody heavy chain<sup>114</sup>. CI107 utilizes Tianyan Pharmaceuticals' SAFEbody platform to mask the epitopes of EGFR and CD3. Short masking peptides that can be specifically cleaved are fused at the N-termini of both the cetuximab antibody light chain and the scFv. *In vivo*, CI107 exhibited a potent, dose-dependent induction of tumor regression in established colon cancer xenografts mice. CI107 significantly reduced cytokine release and showed no elevation in AST levels. In contrast, administering the activated T cell bispecific antibody (TCB) increased IL-6, IFN- $\gamma$ , and AST levels. These discernible differences underscore the compelling effect of dual masking on the EGFR and CD3 binding domains within CI107. This innovative approach substantially enhances the tolerability and safety profile of the probody TCB. Besides, the maximum tolerated dose of CI107 was determined in cynomolgus monkeys and was over 60 times higher than that of the unmasked TCB. Animals administered with CI107 exhibited significantly lower toxicity levels, further reinforcing the notion of an expanded safety margin for this novel therapeutic. This design prevents damage to normal tissues and side effects. CI107 has demonstrated good safety and efficacy in pre-clinical stages as well.

### 3.2. Mobilization of other immune cells to precisely eradicate tumors

Cell engagers could bridge tumor and immune cells to form an immune synapse-mediated cytotoxic effect<sup>21,115</sup>. In addition to

bispecific T cell engager (BiTE), it can also mobilize other immune cells, such as NK cells<sup>116</sup>. The bispecific NK cell engager antibody redirects CD16A<sup>+</sup> NK cells to tumor cells and activates NK cells. Compared to BiTE, a noteworthy attribute of the bispecific NK cell engager lies in its ability to reduce CRS and neurotoxicity<sup>117,118</sup>. AFM13 is the pioneering tetravalent BsAb to target CD30 and CD16<sup>119</sup>. This dual targeting empowers AFM13 to bind CD16A on NK cells and CD30 on lymphoma cells, thereby mediating NK cell killing and clearance of cancer cells<sup>120</sup>. Interventions utilizing modified immune cells AFM13-NK and BsAbs AFM13 for the treatment of relapsed or refractory CD30<sup>+</sup> Hodgkin's or non-Hodgkin's lymphoma patients are currently in Phase I/II (NCT04074746)<sup>121–123</sup>. The application of AFM13 immunotherapy may benefit patients by enabling their immune system to attack tumor cells and potentially inhibit tumor growth and metastasis. Combining AFM13 with NK cells may potentiate tumor cell demise and suppression of tumor growth beyond the scope of AFM13 monotherapy.

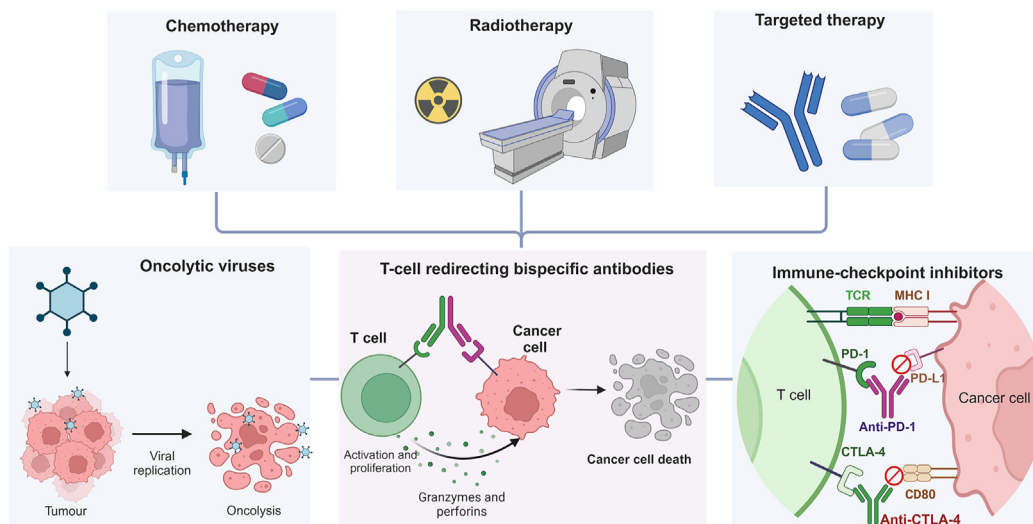
### 3.3. Reducing affinity of CD3 arm

Studies have highlighted a significant trend in T-cell redirecting BsAbs associating affinity for CD3 with pharmacokinetic properties<sup>124,125</sup>. Intriguingly, the antibodies with a low-affinity CD3 arm exhibited slower clearance and higher drug exposure than a higher-affinity CD3 arm<sup>126,127</sup>. However, this correlation presents considerable challenges<sup>21</sup>, such as the affinity for CD3 exerts a multifaceted influence on various properties crucial for therapeutic efficacy, including biodistribution, potency, and toxicity<sup>128,129</sup>. For instance, high CD3 affinity correlates with CRS and might result in a lack of potency and proper biodistribution. Ideally, optimizing the performance of a T-cell redirecting BsAb, such as the appropriate affinity of the CD3 and TAA arms, could result in the desired balance between potency, pharmacokinetics, and safety for meeting therapeutic objectives.

CD3/CLL1 TDB, a BsAb binding CD3 and CLL-1, is engineered to treat acute myeloid leukemia<sup>130</sup>. Notably, results in monkeys revealed that only the low-affinity CD3/CLL1 TDB was well tolerated and able to deplete target cells, compared to the high-affinity TDBs<sup>130</sup>. They provide evidence that using a low-affinity CD3 arm could circumvent toxicities related to CRS and provide acceptable safety while maintaining robust activity. Consequently, one of the most promising strategies to generate a promising BsAb involves pairing low-affinity CD3 antibodies with high-affinity antibodies targeting the TAA<sup>130–133</sup>. Besides, the target selection of antibodies depends on the different expression levels of TAAs, while varying structural formats yield different effects. For instance, compared to tandem scFv-based BsAbs, those based on Fab exhibit superior biophysical properties. Furthermore, Fab-based BsAbs retain the activity of their tandem scFv counterparts and possess unique biological activity<sup>134</sup>. Sebag et al.<sup>135</sup> developed a novel BsAb, Elranatamab, which optimizes affinity for binding to CD3 and the B cell maturation antigen BCMA for treating multiple myeloma<sup>135–139</sup>. Decreasing antibody affinity promotes T cell activation and enhances their anti-tumor activity against multiple myeloma, reducing off-target cell toxicity to some extent.

### 3.4. Combining T-cell redirecting BsAb with other antitumor therapies

T-cell redirecting BsAbs has demonstrated remarkable efficacy in tumor treatment<sup>140</sup>. However, the effectiveness achieved through a



**Figure 4** T cell-redirecting BsAbs combination therapy strategies. These include systemic chemotherapies, radiotherapy, targeted therapies, oncolytic viruses and immune-checkpoint inhibitors, which can create a synergistic effect, further enhancing the efficacy of tumor therapy.

single treatment approach is limited, and it is challenging to eliminate tumors solely relying on this treatment approach. Building upon the antitumor mechanisms of T-cell redirecting BsAbs, combining them with other treatment modalities, including systemic chemotherapies, radiotherapy, targeted therapies, and immunotherapies, creates a synergistic effect, further enhancing tumor therapy's effectiveness (Fig. 4).

#### 3.4.1. Combining with chemotherapy drugs

One of the challenges in utilizing T-cell redirecting BsAbs for treating solid tumors lies in the limited infiltration of T-cells within the TME<sup>20,141</sup>. Traditional cytotoxic agents, like DNA intercalators<sup>142,143</sup>, nucleotide analogs<sup>144</sup>, or alkylating agents<sup>145</sup>, are also being considered for combination with T-cell redirecting BsAbs. Chemotherapies are designed to target rapidly dividing cells directly<sup>146</sup>; thus, combining chemotherapy and T-cell redirecting BsAbs might improve therapeutic responses. Preliminary data substantiating this hypothesis have been reported. For example, ERY974, a T cell-redirecting antibody that targets glypican-3 and CD3, demonstrates modest antitumor effectiveness when used as a monotherapy in non-inflamed NCI-H446 xenograft tumors. This is due to the limited infiltration of ERY974-redirected T cells, primarily at the tumor-stromal boundary. Meanwhile, the cytotoxic agent paclitaxel monotherapy initially showed strong efficacy but could not prevent eventual tumor regrowth. However, ERY974 with chemotherapy synergistically and reciprocally increases antitumor efficacy in non-inflamed tumors by promoting T-cell infiltration into the tumor center and increasing ERY974 distribution in the tumor. This combination therapy may be effective for patients with non-inflamed tumors and those resistant to existing therapies. Chemotherapeutic cyclophosphamide is a potent antitumor drug as an alkylating agent that can alkylate the DNA of tumor cells within their cells<sup>147–149</sup>. At the same time, cyclophosphamide alone can inhibit tumor growth during the initial treatment phase; tumors often resurge within 2–3 weeks<sup>150</sup>. BCMA/CD3 BsAb monotherapy causes an increase in the number of CD8<sup>+</sup> T-cells. However, this excessive activation of T-cells can result in T-cell depletion, leading to temporary tumor regression followed by tumor recurrence after 3–4 weeks of treatment. Combining of

BsAbs and cyclophosphamide can remarkably eradicate the tumor<sup>150</sup>. Experimental data proved that cyclophosphamide can inhibit the excessive activation of T cells and prevent T cell depletion. It can also inhibit the up-regulation of PD-L1 on the tumor surface, promote the renewal of memory T cells, and ensure that the T cells exhibit prolonged and highly effective antitumor activity. Therefore, the therapeutic strategy of combining T-cell redirecting BsAbs with chemotherapeutic drugs holds significant promise for further development<sup>151–153</sup>.

#### 3.4.2. Combining with radiation therapy

T-cell redirecting BsAbs and radiotherapy are two distinct domains in cancer treatment, each possessing non-overlapping cytotoxic effects<sup>154,155</sup>. A synergistic approach materializes through the combination of targeted immunotherapy, exemplified by tumor-targeting agents (BsAbs) and potent payloads (radioisotopes). This multifaceted strategy employs multiple stages, with BsAbs administered over 24 h, followed by a free radical scavenger, and a radioactive metal injection 4 h later. This orchestrated sequence seeks to overcome the limitations of conventional radiation therapy, including inadequate radioisotope uptake and elevated kidney retention. An article has reported a self-assembling and disassembling (SADA) BsAbs that fuses with p53 protein to form a p53–SADA–BsAbs tetramer. After administration, it forms dimers or monomers in the bloodstream, which can be cleared and reduce immunogenicity in rats after kidney filtration, with no short-term or long-term toxicity to the bone marrow, kidneys, or liver<sup>137</sup>. Notably, using anti-GD2 BsAbs in pretargeted radioimmunotherapy for neuroblastoma has showcased improved therapeutic effects in terms of tumor shrinkage, elimination of micrometastases, and prevention of relapse<sup>156–158</sup>.

Radiation therapy operates by stimulating the immune system, thereby bolstering immune-stimulating cell infiltration within tumors, amplifying the immunogenicity of tumor cells, and augmenting antitumor immune responses. Nonetheless, it also engenders local or systemic immune suppression, occasionally even at low doses, culminating in temporary bone marrow inhibition and diminished lymphocyte counts. Radiation therapy can inadvertently impair CD8<sup>+</sup> T cells<sup>159</sup>, pivotal for effective immune therapy response. Hence, in some instances, radiation

therapy might not accentuate the efficacy of combined immunotherapy and may attenuate its potency.

### 3.4.3. Combining with oncolytic viruses

Oncolytic viruses can infect, replicate, and lyse tumor cells while enhancing immune therapy and inducing increased permeability of the immune-tumor microenvironment, allowing for sufficient infiltration of CD8<sup>+</sup> T cells in the tumor microenvironment<sup>160</sup>. However, due to their non-targeted impact on normal cells, oncolytic viruses are often used in combination therapy strategies in clinical trials, resulting in promising advancements<sup>161–164</sup>. Yu et al.<sup>140</sup> engineered an oncolytic vaccinia virus (VV) that carries a BiTE antibody targeting the tumor cell surface antigen EphA2 and named it EphA2 TEA VV. In a mouse model of lung cancer xenografts, EphA2 TEA VV triggered CTL activation and redirection, outperforming conventional oncolytic vaccinia virus in achieving more potent anti-tumor outcomes. In another study, Wang et al.<sup>165</sup> designed an oncolytic adenovirus (OAd-MUC16-BiTE) carrying a BiTE antibody targeting MUC16 (mucin 16). Stable BiTE expression within tumor tissues was achieved post-local administration, mitigating systemic toxicity from systemic application. OAd-MUC16-BiTE mediated antigen-specific T cell activation and target cell lysis. Furthermore, it demonstrated the ability to “reverse” the tumor microenvironment, fostering CTL infiltration, amplifying T cell-mediated tumor cell elimination, and significantly enhancing anti-tumor efficacy.

The functional mechanism of oncolytic measles viruses harboring bispecific T-cell engagers (MV-BiTEs) hinges on the production and release of BiTE antibodies from tumor cells infected with BiTEs<sup>166</sup>. This triggers the assembly of multinucleated syncytia, culminating in the destruction of the tumor cells. Concurrently, BiTEs mobilize T cells to target and eliminate non-infected tumor cells, manifesting additional bystander effects. The therapeutic efficacy of MV-BiTEs was assessed in immunocompetent mouse models. Specifically, in the MC38-CEA model, MV-BiTE treatment extended the survival rates and effectively curbed tumor growth compared to the control group. Within the B16-CD20-CD46 model, MV-BiTE therapy augmented tumor-infiltrating T cells' number and activation levels, thereby bolstering anti-tumor immunity. These findings affirm that MV-BiTEs offer a potent, relapse-free treatment for solid tumors while inducing protective immunity.

Merging BsAbs with oncolytic viruses presents a promising strategy that supersedes the current limitations of individual BiTE treatments, offering a compelling avenue for solid tumor therapy. These studies underscore that combining oncolytic viruses and BiTE achieves a synergistic effect that overcomes the limitations of single-agent therapy.

### 3.4.4. Combining with targeted therapy

*Targeted therapy* is a precision treatment that designs corresponding targeted therapeutic drugs at the cellular and molecular levels to target specific cancer sites that have been identified. These drugs typically exert their effects predominantly on tumor cells, minimizing collateral damage to normal cells.

In small molecule targeted inhibitors, the primary targets are often vital kinases involved in cell signaling pathways, such as the mammalian target of rapamycin (mTOR), mitogen-activated protein kinase, and anaplastic lymphoma kinase. An exhaustive examination of 52 FDA-approved small molecule tyrosine kinase

inhibitors revealed that select drugs aimed at mTOR, JAK, and Src kinases effectively suppress T cell proliferation<sup>167</sup>. When co-administered with T-cell redirecting BsAbs *in vitro*, mTOR and JAK kinase inhibitors diminished cytokine release related to CRS without undermining the tumor-killing potential of T-cell bispecific antibodies. In contrast, Src kinase inhibitor dasatinib successfully suppressed CRS-related cytokines but at the cost of impaired T cell cytotoxicity. Therefore, mTOR and JAK kinase inhibitors are preferable for mitigating CRS reactions in clinical settings without sacrificing anti-tumor effectiveness.

Bruton tyrosine kinase inhibitors are the preferred therapeutic agents for patients with chronic lymphocytic leukemia (CLL)<sup>168,169</sup>. The efficacy of BHK in CLL can be enhanced by combining it with CD19/CD3 BsAbs<sup>170</sup>. Notably, BTKis actively disrupts CLL cells' immunosuppressive mechanisms, irrespective of ITK inhibition, thereby bolstering the cytotoxic activity of autologous T cells<sup>171–174</sup>. Elevated CTLA-4 expression in peripheral blood CLL cells has been documented, and its subsequent downregulation upon BTKi treatment suggests a mechanistic link that may enhance T-cell cytotoxic responses. Thus, combining BTKis with BsAbs is a robust strategy for achieving sustained CLL remission and counteracting drug resistance, which may have broader applications in treating other mature B-cell malignancies.

Targeted therapy has made substantial strides in cancer care, offering personalized treatments by engaging tumor-associated molecules. This not only heightens therapeutic efficacy but also minimizes adverse side effects. The combination of BsAbs can markedly elevate treatment success rates, underscoring the vast potential of this approach.

### 3.4.5. Combining immune checkpoint inhibitors

Even after BsAbs and T cells navigate multiple barriers to penetrate tumor sites, a host of elements can dampen their therapeutic efficacy. Exhaustion of T cells owing to upregulation of immune-checkpoint proteins during therapy has been described with various BiTEs, *in vitro* and patients. It is a potential immune escape mechanism underlying both intrinsic and acquired resistance. Combinatorial immunotherapy approaches have been designed to counteract T-cell dysfunction/exhaustion mechanisms using inhibitory checkpoint-blocking antibodies to improve the clinical outcomes of individuals receiving CD3<sup>+</sup> bispecific T-cell redirection therapies. Interestingly, resistance to blinatumomab treatment has been attributed to increased PD-L1 expression in CD19<sup>+</sup> leukemic cells<sup>175</sup>. In order to antagonize PD-L1 expressed by the leukemic cells, blinatumomab in combination with pembrolizumab (anti-PD-1 therapy) resulted in enhanced T cell-mediated cytotoxicity relative to that observed with the parental BiTE alone<sup>176</sup>. A large number of clinical trials are currently evaluating the safety and efficacy of combinatorial approaches for the treatment of relapsed/refractory acute leukemia or lymphoma patients (NCT03160079, NCT03605589, NCT03340766, NCT03512405).

Belmontes et al.<sup>177</sup> have found that combining PD-1 antibodies, CTLA-4 antibodies, and 4-1BB agonist antibodies with CD3 BsAbs can enhance the efficacy of CD3 BsAbs. Among them, 4-1BB has demonstrated the most substantial impact, particularly in amplifying CD8<sup>+</sup> T cells within tumor tissues and overcoming treatment resistance in solid tumors. One of the most rapidly advancing BsAbs, SMET12 (targeting EGFR/CD3), combined with PD-1 antibodies, is now in Phase II clinical trials.

Cibisatamab, TCB targeting CEA/CD3, is a novel T-cell redirecting BsAbs targeting CEA on tumor cells and CD3 on T cells<sup>178</sup>. Cibisatamab displays potent anti-tumor activity in pre-clinical models, leading to increased intra-tumoral T-cell infiltration and activation and upregulating PD-1/PD-L1. In the ongoing dose-escalation phase I studies (NCT02324257 and NCT02650713), activity appeared to be enhanced with doses in combination with atezolizumab, with a manageable safety profile.

An EGFR/CD3 molecule, SMET12, is currently in preclinical development for treating EGFR-positive advanced/metastatic solid tumors. Pharmacological studies demonstrate that SMET12 combined with Treprinumab (anti-PD-1 therapy) can effectively eliminate the immune suppressive environment caused by the expression of inhibitory receptors such as PD-1 after T cell activation<sup>179</sup>, further exerting the anti-tumor function of SMET12 to achieve synergistic enhancement. It has significant advantages and potential good prospects for treating solid tumors.

#### 4. Conclusions and prospects

T-cell immunotherapy constitutes a dynamic area in cancer treatment, with three main approaches currently in the spotlight: ICIs, T-cell redirecting BsAbs, and CAR-T cell therapy. Notably, T-cell redirecting BsAbs have achieved a revolutionary breakthrough in cancer therapy. Unlike ICIs, which release the general suppression of the cellular immune system and may trigger adverse reactions to non-tumor cells, T-cell redirecting BsAbs precisely engage T cells with tumor cells, enhancing T cell activation and proliferation. Their mechanism of action enables potentially quicker responses than ICIs by inducing direct T cell-mediated cytotoxicity. In contrast to personalized therapies like CAR-T, which are time-consuming and expensive due to the need for individualized collection, modification, and expansion of a patient's T cells, bispecific antibodies offer a more straightforward and cost-effective "off-the-shelf" solution<sup>182</sup>. They can be manufactured and stockpiled on a large scale, reducing treatment costs. Nevertheless, the progression of T-cell redirecting BsAbs in solid tumors is relatively slower, and there are still many challenges and areas for optimization, including CRS, targeted and non-targeted cytotoxicity, TAA loss, limited T-cell infiltration and suppression of T-cell activity in the tumor microenvironment.

CRS can be observed in patients treated with T-cell redirecting BsAbs such as Binatumomab, and the severity depends on the treatment type and target<sup>89,150</sup>. Preclinical studies using humanized mouse models have found that TNF- $\alpha$  produced by activated T cells is the primary mediator of CRS induced by T-cell redirecting BsAbs. This leads to massive secretion of inflammatory cytokines by monocytes<sup>183</sup>. To overcome this challenge, T-cell redirecting BsAbs could be designed using a low-affinity anti-CD3 to reduce the potential for dose-limiting toxicities associated with CRS. Besides, blocking upstream TNF- $\alpha$  and downstream IL-1 $\beta$  or IL-6 can induce tumor killing with reduced cytokine release<sup>184,185</sup>.

The challenges posed by targeted and non-targeted cytotoxicity, the limited number of effector cells, and the suppressed T-cell activity within tumors are deeply intertwined<sup>186,187</sup>. These three obstacles are indispensable in tackling the limitations associated with T-cell redirecting BsAbs<sup>188</sup>. Most antibodies currently undergoing development are centered around differentiation markers<sup>189</sup>. Many investigational solid tumor targets, like CEA, EGFR, EpCAM, and HER2, fall within this differentiation marker category<sup>190</sup>. Noteworthy among these can be expressed in normal

cells, albeit at limited levels, thereby narrowing the therapeutic window and circumscribing the potential for T-cell redirecting BsAbs design. To enhance the targeting of antibodies to tumor cells and reduce drug resistance, simultaneously targeting two different tumor antigens and tri-specific antibody development can be employed to address these challenges.

Many strategies have been devised to tackle the challenge of on-target off-tumor effects; one of the innovative approaches is utilizing a "mask" to shield the antibody epitope until it reaches the tumor site for conditional activation. The stimuli involved in this strategy include light, temperature, enzymes, pH, ions, small effect molecules, or rare antigens. In addition to the pro-drugs and cleavage enzymes mentioned earlier, structural modification of the antibody Fc domain has achieved significant breakthroughs. The TME caused by changes in tumor cell metabolism is characterized by acidic pH, a widely used parameter in targeted TME research. Low pH significantly impacts the TME, leading to immune suppression, immune escape, and disease progression. This acidic extracellular environment significantly affects the bioavailability of therapeutic antibodies. Harnessing this characteristic as a trigger for therapeutic antibody activation not only affords precise control but also mitigates drug toxicity linked to off-target effects. For instance, Sulea et al.<sup>191</sup> developed a pH-dependent HER2 antibody (bH1-P5P8) that exhibits heightened antigen binding in acidic pH compared to neutral pH, bolstering tissue-specific distribution and diminishing distribution-related side effects in normal tissues.

The objective of achieving a durable cure with limited toxicity is also a driving force in designing and developing new tumor immunotherapies. Efficacy in tumor treatment relies mainly on immune activation strategies, especially the cytotoxic activity of immune T cells within the tumor. However, the TME can affect the infiltrating T cells, curbing their proliferation and function, curtailing effector T-cell quantity and activity. Which may result in reduced or lost activity such as impaired proliferation and immune function. The emergence of T cell exhaustion, characterized by the expression of inhibitory receptors, represents a pivotal mechanism in tumor immune evasion. By countering these signals, interventions like ICIs have demonstrated considerable antitumor potency by revitalizing T-cell functionality within the tumor microenvironment. With the ongoing advancement of science and technology, researchers have proposed innovative approaches combining T-cell redirection bispecific antibodies with other strategies, including ICIs, immune activators, and oncolytic viruses. Current and upcoming clinical trials must prove the value of these combinatory approaches. As the number of BsAbs undergoing clinical trials increases, our understanding of the optimal structures and constructs for targeting particular tumor types is expanding. Such knowledge will improve the antitumor efficacy of T cell-engaging therapies, prevent disease relapse, and drive future advancements in this field.

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## Author contributions

Xiaojing Qin, Wenjing Ning reviewed the literature and drafted the manuscript. Wenxin Luo and Xue Liu conceived and supervised the project. Other authors participated in the search and collation of literature. All of the authors have read and approved the final manuscript.

## Conflicts of interest

The authors declare no conflicts of interest.

## References

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA A Cancer J Clin* 2021;**71**:209–49.
- Zhang Y, Zhang Z. The history and advances in cancer immunotherapy: understanding the characteristics of tumor-infiltrating immune cells and their therapeutic implications. *Cell Mol Immunol* 2020;**17**:807–21.
- Pilard C, Ancion M, Delvenne P, Jerusalem G, Hubert P, Herfs M. Cancer immunotherapy: it's time to better predict patients' response. *Br J Cancer* 2021;**125**:927–38.
- Wang DR, Wu XL, Sun YL. Therapeutic targets and biomarkers of tumor immunotherapy: response versus non-response. *Signal Transduct Targeted Ther* 2022;**7**:331.
- Chehelgerdi M, Chehelgerdi M, Allela OQB, Pecho RDC, Jayasankar N, Rao DP, et al. Progressing nanotechnology to improve targeted cancer treatment: overcoming hurdles in its clinical implementation. *Mol Cancer Mol Cancer* 2023;**22**:169.
- Ma J, Mo Y, Tang M, Shen J, Qi Y, Zhao W, et al. Bispecific antibodies: from research to clinical application. *Front Immunol* 2021;**12**:626616.
- Sziji P, Chudasama V. The renaissance of chemically generated bispecific antibodies. *Nat Rev Chem* 2021;**5**:78–92.
- Zhu Y, Yu X, Thamphiwatana SD, Zheng Y, Pang Z. Nanomedicines modulating tumor immunosuppressive cells to enhance cancer immunotherapy. *Acta Pharm Sin B* 2020;**10**:2054–74.
- Wu SY, Wu FG, Chen XY. Antibody-incorporated nanomedicines for cancer therapy. *Adv Mater* 2022;**34**:2109210.
- Wei J, Yang Y, Wang G, Liu M. Current landscape and future directions of bispecific antibodies in cancer immunotherapy. *Front Immunol* 2022;**13**:1035276.
- Nisonoff A, Wissler FC, Lipman LN. Properties of the major component of a peptic digest of rabbit antibody. *Science* 1960;**132**:1770–1.
- Milstein C, Cuello AC. Hybrid hybridomas and their use in immunohistochemistry. *Nature* 1983;**305**:537–40.
- Perez P, Hoffman RW, Shaw S, Bluestone JA, Segal DM. Specific targeting of cytotoxic T cells by anti-T3 linked to anti-target cell antibody. *Nature* 1985;**316**:354–6.
- Holliger P, Prospero T, Winter G. "Diabodies": small bivalent and bispecific antibody fragments. *Proc Natl Acad Sci U S A* 1993;**90**:6444–8.
- Lindhofer H, Mocikat R, Steipe B, Thierfelder S. Preferential species-restricted heavy/light chain pairing in rat/mouse quadromas. Implications for a single-step purification of bispecific antibodies. *J Immunol* 1995;**155**:219–25.
- Bargou RC. The expanding success of T cell-engaging bispecific antibodies. *Nat Cancer* 2023;**4**:1054–5.
- Haber L, Olson K, Kelly MP, Crawford A, DiLillo DJ, Tavaré R, et al. Generation of T-cell-redirecting bispecific antibodies with differentiated profiles of cytokine release and biodistribution by CD3 affinity tuning. *Sci Rep* 2021;**11**:14397.
- van de Donk NWJ, Zweegman S. T-cell-engaging bispecific antibodies in cancer. *Lancet* 2023;**402**:142–58.
- Kamakura D, Asano R, Yasunaga M. T cell bispecific antibodies: an antibody-based delivery system for inducing antitumor immunity. *Pharmaceuticals* 2021;**14**:1172.
- Singh A, Dees S, Grewal IS. Overcoming the challenges associated with CD3<sup>+</sup> T-cell redirection in cancer. *Br J Cancer* 2021;**124**:1037–48.
- Xiong W, Chen Y, Kang X, Chen Z, Zheng P, Hsu YH, et al. Immunological synapse predicts effectiveness of chimeric antigen receptor Cells. *Mol Ther* 2021;**29**:1349–51.
- Lin CY, Lee CH, Chuang YH, Lee JY, Chiu YY, Wu Lee YH, et al. Membrane protein-regulated networks across human cancers. *Nat Commun* 2019;**10**:3131.
- Jhunjunwala S, Hammer C, Delamarre L. Antigen presentation in cancer: insights into tumour immunogenicity and immune evasion. *Nat Rev Cancer* 2021;**21**:298–312.
- Andersen MH. Tumor microenvironment antigens. *Semin Immunopathol* 2023;**45**:253–64.
- Peri A, Salomon N, Wolf Y, Kreiter S, Diken M, Samuels Y. The landscape of T cell antigens for cancer immunotherapy. *Nat Cancer* 2023;**4**:937–54.
- Raman V, Van Dessel N, Hall CL, Wetherby VE, Whitney SA, Kolewe EL, et al. Intracellular delivery of protein drugs with an autonomously lysing bacterial system reduces tumor growth and metastases. *Nat Commun* 2021;**12**:6116.
- Pishesha N, Harmand TJ, Ploegh HL. A guide to antigen processing and presentation. *Nat Rev Immunol* 2022;**22**:751–64.
- Xu Y, Salazar GT, Zhang N, An Z. T-cell receptor mimic (TCRm) antibody therapeutics against intracellular proteins. *Antib Ther* 2019;**2**:22–32.
- Tian Z, Liu M, Zhang Y, Wang X. Bispecific T cell engagers: an emerging therapy for management of hematologic malignancies. *J Hematol Oncol* 2021;**14**:75.
- Passariello M, Yoshioka A, Takahashi K, Hashimoto SI, Inoue T, Nakamura K, et al. Novel tri-specific tribodies induce strong T cell activation and anti-tumor effects *in vitro* and *in vivo*. *J Exp Clin Cancer Res* 2022;**41**:269.
- Damato BE, Dukes J, Goodall H, Carvajal RD. Tebentafusp: T cell redirection for the treatment of metastatic uveal melanoma. *Cancers* 2019;**11**:971.
- Middleton MR, McAlpine C, Woodcock VK, Corrie P, Infante JR, Steven NM, et al. Tebentafusp, a TCR/anti-CD3 bispecific fusion protein targeting gp100, potently activated antitumor immune responses in patients with metastatic melanoma. *Clin Cancer Res* 2020;**26**:5869–78.
- Morris EC, Neelapu SS, Giavridis T, Sadelain M. Cytokine release syndrome and associated neurotoxicity in cancer immunotherapy. *Nat Rev Immunol* 2022;**22**:85–96.
- Frey N. Cytokine release syndrome: who is at risk and how to treat. *Best Pract Res Clin Haematol* 2017;**30**:336–40.
- Frampton JE. Catumaxomab: in malignant ascites. *Drugs* 2012;**72**:1399–410.
- Cao Y, Wang J, Jiang S, Lyu M, Zhao F, Liu J, et al. JAK1/2 inhibitor ruxolitinib promotes the expansion and suppressive action of polymorphonuclear myeloid-derived suppressor cells via the JAK/STAT and ROS–MAPK/NF- $\kappa$ B signalling pathways in acute graft-versus-host disease. *Clin Transl Immunology* 2023;**12**:e1441.
- Sano Y, Azuma Y, Tsunenari T, Kayukawa Y, Shinozuka J, Fujii E, et al. Combination of T cell-redirecting bispecific antibody ERY974 and chemotherapy reciprocally enhances efficacy against non-inflamed tumours. *Nat Commun* 2022;**13**:5265.
- Safran H, Druta M, Morse M, Lynce F, Pintova S, Almhanna K, et al. Abstract CT111: results of a phase 1 dose escalation study of ERY974, an anti-glypican 3 (GPC3)/CD3 bispecific antibody, in patients with advanced solid tumors. *Cancer Res* 2021;**81**:CT111.

39. Moreau P, Garfall AL, van de Donk NWCJ, Nahi H, San-Miguel JF, Oriol A, et al. Teclistamab in relapsed or refractory multiple myeloma. *N Engl J Med* 2022;**387**:495–505.
40. Ishiguro T, Sano Y, Komatsu SI, Kamata-Sakurai M, Kaneko A, Kinoshita Y, et al. An anti-glypican 3/CD3 bispecific T cell-redireciting antibody for treatment of solid tumors. *Sci Transl Med* 2017;**9**:eaal4291.
41. Moreau P, Kumar SK, San Miguel J, Davies F, Zamagni E, Bahlis N, et al. Treatment of relapsed and refractory multiple myeloma: recommendations from the International Myeloma Working Group. *Lancet Oncol* 2021;**22**:e105–18.
42. Shah N, Chari A, Scott E, Mezzi K, Usmani SZ. B-cell maturation antigen (BCMA) in multiple myeloma: rationale for targeting and current therapeutic approaches. *Leukemia* 2020;**34**:985–1005.
43. Minson A, Dickinson M. Glofitamab CD20-TCB bispecific antibody. *Leuk Lymphoma* 2021;**62**:3098–108.
44. Dickinson MJ, Carlo-Stella C, Morschhauser F, Bachy E, Corradini P, Iacoboni G, et al. Glofitamab for relapsed or refractory diffuse large B-cell lymphoma. *N Engl J Med* 2022;**387**:2220–31.
45. Lee DW, Gardner R, Porter DL, Louis CU, Ahmed N, Jensen M, et al. Current concepts in the diagnosis and management of cytokine release syndrome. *Blood* 2014;**124**:188–95.
46. Ludwig H, Terpos E, van de Donk N, Mateos MV, Moreau P, Dimopoulos MA, et al. Prevention and management of adverse events during treatment with bispecific antibodies and CART cells in multiple myeloma: a consensus report of the European Myeloma Network. *Lancet Oncol* 2023;**24**:e255–69.
47. Cohen AD, Mateos MV, Cohen YC, Rodriguez-Otero P, Paiva B, van de Donk NWCJ, et al. Efficacy and safety of cilta-cel in patients with progressive multiple myeloma after exposure to other BCMA-targeting agents. *Blood* 2023;**141**:219–30.
48. Gaballa MR, Banerjee P, Milton DR, Jiang X, Ganesh C, Khazal S, et al. Blinatumomab maintenance after allogeneic hematopoietic cell transplantation for B-lineage acute lymphoblastic leukemia. *Blood* 2022;**139**:1908–19.
49. Roth P, Winklhofer S, Müller AMS, Dummer R, Mair MJ, Gramatzki D, et al. Neurological complications of cancer immunotherapy. *Cancer Treat Rev* 2021;**97**:102189.
50. San-Miguel J, Dhakal B, Yong K, Spencer A, Anguille S, Mateos MV, et al. Cilta-cel or standard care in lenalidomide-refractory multiple myeloma. *N Engl J Med* 2023;**389**:335–47.
51. Foà R, Bassan R, Vitale A, Elia L, Piciocchi A, Puzzolo MC, et al. Dasatinib-blinatumomab for pH-positive acute lymphoblastic leukemia in adults. *N Engl J Med* 2020;**383**:1613–23.
52. Trudel S, Cohen AD, Krishnan AY, Fonseca R, Spencer A, Berdeja JG, et al. Cevostamab monotherapy continues to show clinically meaningful activity and manageable safety in patients with heavily pre-treated relapsed/refractory multiple myeloma (RRMM): updated results from an ongoing phase I study. *Blood* 2021;**138**:157.
53. van der Sluis IM, de Lorenzo P, Kotecha RS, Attarbaschi A, Escherich G, Nysom K, et al. Blinatumomab added to chemotherapy in infant lymphoblastic leukemia. *N Engl J Med* 2023;**388**:1572–81.
54. Brown PA, Ji L, Xu X, Devidas M, Hogan LE, Borowitz MJ, et al. Effect of postinduction therapy consolidation with blinatumomab vs chemotherapy on disease-free survival in children, adolescents, and young adults with first relapse of B-cell acute lymphoblastic leukemia: a randomized clinical trial. *JAMA* 2021;**325**:833–42.
55. Locatelli F, Zugmaier G, Rizzari C, Morris JD, Gruhn B, Klingebiel T, et al. Effect of blinatumomab vs chemotherapy on event-free survival among children with high-risk first-relapse B-cell acute lymphoblastic leukemia: a randomized clinical trial. *JAMA* 2021;**325**:843–54.
56. Jabbour E, Short NJ, Jain N, Huang X, Montalban-Bravo G, Banerjee P, et al. Ponatinib and blinatumomab for Philadelphia chromosome-positive acute lymphoblastic leukaemia: a US, single-centre, single-arm, phase 2 trial. *Lancet Haematol* 2023;**10**:e24–34.
57. Topp MS, Gökbuget N, Stein AS, Zugmaier G, O'Brien S, Bargou RC, et al. Safety and activity of blinatumomab for adult patients with relapsed or refractory B-precursor acute lymphoblastic leukaemia: a multicentre, single-arm, phase 2 study. *Lancet Oncol* 2015;**16**:57–66.
58. Kantarjian H, Stein A, Gökbuget N, Fielding AK, Schuh AC, Ribera JM, et al. Blinatumomab versus chemotherapy for advanced acute lymphoblastic leukemia. *N Engl J Med* 2017;**376**:836–47.
59. Gökbuget N, Dombret H, Bonifacio M, Reichle A, Graux C, Faul C, et al. Blinatumomab for minimal residual disease in adults with B-cell precursor acute lymphoblastic leukemia. *Blood* 2018;**131**:1522–31.
60. Stein AS, Schiller G, Benjamin R, Jia C, Zhang A, Zhu M, et al. Neurologic adverse events in patients with relapsed/refractory acute lymphoblastic leukemia treated with blinatumomab: management and mitigating factors. *Ann Hematol* 2019;**98**:159–67.
61. Parker KR, Migliorini D, Perkey E, Yost KE, Bhaduri A, Bagga P, et al. Single-cell analyses identify brain mural cells expressing CD19 as potential off-tumor targets for CAR-T immunotherapies. *Cell* 2020;**183**:126–42.
62. Braig F, Brandt A, Goebeler M, Tony HP, Kurze AK, Nollau P, et al. Resistance to anti-CD19/CD3 BiTE in acute lymphoblastic leukemia may be mediated by disrupted CD19 membrane trafficking. *Blood* 2017;**129**:100–4.
63. Mancini SJC, Balabanian K, Corre I, Gavard J, Lazennec G, Le Bousse-Kerdilès MC, et al. Deciphering tumor niches: lessons from solid and hematological malignancies. *Front Immunol* 2021;**12**:766275.
64. Montironi C, Muñoz-Pinedo C, Eldering E. Hematopoietic versus solid cancers and T cell dysfunction: looking for similarities and distinctions. *Cancers* 2021;**13**:284.
65. Nakamura K, Smyth MJ. Myeloid immunosuppression and immune checkpoints in the tumor microenvironment. *Cell Mol Immunol* 2020;**17**:1–12.
66. Tie Y, Tang F, Wei YQ, Wei XW. Immunosuppressive cells in cancer: mechanisms and potential therapeutic targets. *J Hematol Oncol* 2022;**15**:61.
67. Galli F, Aguilera JV, Palermo B, Markovic SN, Nisticò P, Signore A. Relevance of immune cell and tumor microenvironment imaging in the new era of immunotherapy. *J Exp Clin Cancer Res* 2020;**39**:89.
68. Kalaora S, Nagler A, Wargo JA, Samuels Y. Mechanisms of immune activation and regulation: lessons from melanoma. *Nat Rev Cancer* 2022;**22**:195–207.
69. Jiang W, He Y, He W, Wu G, Zhou X, Sheng Q, et al. Exhausted CD8<sup>+</sup> T cells in the tumor immune microenvironment: new pathways to therapy. *Front Immunol* 2021;**11**:622509.
70. Lopez de Rodas M, Schalper KA. Tumour antigen-induced T cell exhaustion—the archenemy of immune-hot malignancies. *Nat Rev Clin Oncol* 2021;**18**:749–50.
71. Brummel K, Eerkens AL, de Bruyn M, Nijman HW. Tumour-infiltrating lymphocytes: from prognosis to treatment selection. *Br J Cancer* 2023;**128**:451–8.
72. Chow A, Perica K, Klebanoff CA, Wolchok JD. Clinical implications of T cell exhaustion for cancer immunotherapy. *Nat Rev Clin Oncol* 2022;**19**:775–90.
73. Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. *Nat Rev Immunol* 2015;**15**:486–99.
74. Haber L, Olson K, Kelly MP, Crawford A, DiLillo DJ, Tavaré R, et al. Generation of T-cell-redireciting bispecific antibodies with differentiated profiles of cytokine release and biodistribution by CD3 affinity tuning. *Sci Rep* 2021;**11**:14397.
75. Voskoboinik I, Whisstock JC, Trapani JA. Perforin and granzymes: function, dysfunction and human pathology. *Nat Rev Immunol* 2015;**15**:388–400.
76. Weigelin B, den Boer AT, Wagena E, Broen K, Dolstra H, de Boer RJ, et al. Cytotoxic T cells are able to efficiently eliminate cancer cells by additive cytotoxicity. *Nat Commun* 2021;**12**:5217.
77. Raskov H, Orhan A, Christensen JP, Gögenur I. Cytotoxic CD8<sup>+</sup> T cells in cancer and cancer immunotherapy. *Br J Cancer* 2021;**124**:359–67.

78. Xie F, Zhang L, Shi S, Zheng A, Di J, Jin S, et al. Liposomal T cell engager and re-director for tumor cell eradication in cancer immunotherapy. *mAbs* 2022;**14**:2115205.
79. Rajé N, Anderson K, Einsele H, Efebera Y, Gay F, Hammond SP, et al. Monitoring, prophylaxis, and treatment of infections in patients with MM receiving bispecific antibody therapy: consensus recommendations from an expert panel. *Blood Cancer J* 2016;**13**:116.
80. Sim BZ, Longhitano A, Er J, Harrison SJ, Slavin MA, Teh BW. Infectious complications of bispecific antibody therapy in patients with multiple myeloma. *Blood Cancer J* 2023;**13**:34.
81. Lamers CH, Sleijfer S, van Steenberghe S, van Elzakker P, van Krimpen B, Groot C, et al. Treatment of metastatic renal cell carcinoma with CAIX CAR-engineered T cells: clinical evaluation and management of on-target toxicity. *Mol Ther* 2013;**21**:904–12.
82. Thistlethwaite FC, Gilham DE, Guest RD, Rothwell DG, Pillai M, Burt DJ, et al. The clinical efficacy of first-generation carcinoembryonic antigen (CEACAM5)-specific CAR T cells is limited by poor persistence and transient pre-conditioning-dependent respiratory toxicity. *Cancer Immunol Immunother* 2017;**66**:1425–36.
83. Viardot A, Goebeler ME, Hess G, Neumann S, Pfreundschuh M, Adrian N, et al. Phase 2 study of the bispecific T-cell engager (BiTE) antibody blinatumomab in relapsed/refractory diffuse large B-cell lymphoma. *Blood* 2016;**127**:1410–6.
84. Budde LE, Sehn LH, Matasar M, Schuster SJ, Assouline S, Giri P, et al. Safety and efficacy of mosunetuzumab, a bispecific antibody, in patients with relapsed or refractory follicular lymphoma: a single-arm, multicentre, phase 2 study. *Lancet Oncol* 2022;**23**:1055–65.
85. Philipp N, Kazerani M, Nicholls A, Vick B, Wulf J, Straub T, et al. T-cell exhaustion induced by continuous bispecific molecule exposure is ameliorated by treatment-free intervals. *Blood* 2022;**140**:1104–18.
86. Howard SC, Jones DP, Pui CH. The tumor lysis syndrome. *N Engl J Med* 2011;**364**:1844–54.
87. Bannerji R, Arnason JE, Advani RH, Brown JR, Allan JN, Ansell SM, et al. Oronextamab, a human CD20×CD3 bispecific antibody in patients with CD20-positive B-cell malignancies (ELM-1): results from the relapsed or refractory non-Hodgkin lymphoma cohort in a single-arm, multicentre, phase 1 trial. *Lancet Haematol* 2022;**9**:e327–39.
88. Hutchings M, Mous R, Clausen MR, Johnson P, Linton KM, Chamuleau MED, et al. Dose escalation of subcutaneous epcoritamab in patients with relapsed or refractory B-cell non-Hodgkin lymphoma: an open-label, phase 1/2 study. *Lancet* 2021;**398**:1157–69.
89. Oates J, Hassan NJ, Jakobsen BK. ImmTACs for targeted cancer therapy: why, what, how, and which. *Mol Immunol* 2015;**67**:67–74.
90. Labrijn AF, Janmaat ML, Reichert JM, Parren PW. Bispecific antibodies: a mechanistic review of the pipeline. *Nat Rev Drug Discov* 2019;**18**:585–608.
91. Budde E, Gopal AK, Kim WS, Flinn IW, Cheah CY, Nastoupil L. A phase 1 dose escalation study of Igm-2323, a novel anti-CD20 × anti-CD3 IgM T cell engager (TCE) in patients with advanced B-cell malignancies. *Blood* 2021;**138**:132.
92. Assouline SE, Kim WS, Sehn LH, Schuster SJ, Cheah CY, Nastoupil LJ, et al. Mosunetuzumab shows promising efficacy in patients with multiply relapsed follicular lymphoma: updated clinical experience from a phase I dose-escalation trial. *Blood* 2020;**136**:42–4.
93. Hutchings M, Carlo-Stella C, Bachy E, Offner FC, Morschhauser F, Crump M, et al. Glofitamab step-up dosing induces high response rates in patients with hard-to-treat refractory or relapsed non-Hodgkin lymphoma. *Blood* 2020;**136**:46–8.
94. Fajgenbaum DC, June CH. Cytokine storm. *N Engl J Med* 2020;**383**:2255–73.
95. Olszewski AJ, Avigdor A, Babu S, Levi I, Abadi U, Holmes H, et al. Single-agent mosunetuzumab is a promising safe and efficacious chemotherapy-free regimen for elderly/unfit patients with previously untreated diffuse large B-cell lymphoma. *Blood* 2020;**136**:43–5.
96. Phillips TJ, Olszewski AJ, Munoz J, Kim TM, Yoon DH, Greil R, et al. Mosunetuzumab, a novel CD20/CD3 bispecific antibody, in combination with CHOP confers high response rates in patients with diffuse large B-cell lymphoma. *Blood* 2020;**136**:37–8.
97. Blanco B, Domínguez-Alonso C, Alvarez-Vallina L. Bispecific immunomodulatory antibodies for cancer immunotherapy. *Clin Cancer Res* 2021;**27**:5457–64.
98. Runcie K, Budman DR, John V, Seetharamu N. Bi-specific and tri-specific antibodies- the next big thing in solid tumor therapeutics. *Mol Med* 2018;**24**:50.
99. Mullard A. Trispecific antibodies take to the clinic. *Nat Rev Drug Discov* 2020;**19**:657–8.
100. Wu L, Seung E, Xu L, Rao E, Lord DM, Wei RR, et al. Trispecific antibodies enhance the therapeutic efficacy of tumor-directed T cells through T cell receptor co-stimulation. *Nature Cancer* 2019;**1**:86–98.
101. Garfall AL, June CH. Trispecific antibodies offer a third way forward for anticancer immunotherapy. *Nature* 2019;**575**:450–1.
102. Sun Y, Yu X, Wang X, Yuan K, Wang G, Hu L, et al. Bispecific antibodies in cancer therapy: target selection and regulatory requirements. *Acta Pharm Sin B* 2023;**13**:3583–97.
103. Seung E, Xing Z, Wu L, Rao E, Cortez-Retamozo V, Ospina B, et al. A trispecific antibody targeting HER2 and T cells inhibits breast cancer growth via CD4 cells. *Nature* 2022;**603**:328–34.
104. Goebeler ME, Bargou R. Blinatumomab: a CD19/CD3 bispecific T cell engager (BiTE) with unique anti-tumor efficacy. *Leuk Lymphoma* 2016;**57**:1021–32.
105. van der Sluis IM, de Lorenzo P, Kotecha RS, Attarbaschi A, Escherich G, Nysom K, et al. Blinatumomab added to chemotherapy in infant lymphoblastic leukemia. *N Engl J Med* 2023;**388**:1572–81.
106. Advani AS, Moseley A, O'Dwyer KM, Wood BL, Fang M, Wieduwilt MJ, et al. Swog 1318: a phase II Trial of blinatumomab followed by POMP maintenance in older patients with newly diagnosed Philadelphia chromosome-negative B-cell acute lymphoblastic leukemia. *J Clin Oncol* 2022;**40**:1574–82.
107. Viardot A, Bargou R. Bispecific antibodies in haematological malignancies. *Cancer Treat Rev* 2018;**65**:87–95.
108. Yu S, Li A, Liu Q, Yuan X, Xu H, Jiao D, et al. Recent advances of bispecific antibodies in solid tumors. *J Hematol Oncol* 2017;**10**:155.
109. Jornada DH, dos Santos Fernandes GF, Chiba DE, de Melo TR, dos Santos JL, Chung MC. The prodrug approach: a successful tool for improving drug solubility. *Molecules* 2015;**21**:42.
110. Autio KA, Boni V, Humphrey RW, Naing A. Probody therapeutics: an emerging class of therapies designed to enhance on-target effects with reduced off-tumor toxicity for use in immuno-oncology. *Clin Cancer Res* 2020;**26**:984–9.
111. Lin WW, Lu YC, Chuang CH, Cheng TL. Ab locks for improving the selectivity and safety of antibody drugs. *J Biomed Sci* 2020;**27**:76.
112. Kessenbrock K, Plaks V, Werb Z. Matrix metalloproteinases: regulators of the tumor microenvironment. *Cell* 2010;**141**:52–67.
113. Liu G, Li Y, Dai Z, Qu J, Shi J, Zhao Q, et al. Abstract 2869: ADG138, a novel HER2×CD3 POWERbody™ integrating bispecific TCE with precision masking to control cytokine release syndrome and on-target off-tumor toxicity for single agent and combination therapies in HER2-expressing solid tumors. *Cancer Res* 2022;**82**:2869.
114. Boustany LM, LaPorte SL, Wong L, White C, Vinod V, Shen J, et al. A probody T cell-engaging bispecific antibody targeting EGFR and CD3 inhibits colon cancer growth with limited toxicity. *Cancer Res* 2022;**82**:4288–98.
115. Shin HG, Yang HR, Yoon A, Lee S. Bispecific antibody-based immune-cell engagers and their emerging therapeutic targets in cancer immunotherapy. *Int J Mol Sci* 2022;**23**:5686.
116. Demaria O, Gauthier L, Debroas G, Vivier E. Natural killer cell engagers in cancer immunotherapy: next generation of immuno-oncology treatments. *Eur J Immunol* 2021;**51**:1934–42.



117. Sivori S, Pende D, Quatrini L, Pietra G, Della Chiesa M, Vacca P, et al. NK cells and ILCs in tumor immunotherapy. *Mol Aspect Med* 2021;**80**:100870.
118. Vacca P, Pietra G, Tumino N, Munari E, Mingari MC, Moretta L. Exploiting human NK cells in tumor therapy. *Front Immunol* 2020; **10**:3013.
119. Wu J, Fu J, Zhang M, Liu D. AFM13: a first-in-class tetravalent bispecific anti-CD30/CD16A antibody for NK cell-mediated immunotherapy. *J Hematol Oncol* 2015;**8**:96.
120. Kerbaay LN, Marin ND, Kaplan M, Banerjee PP, Berrien-Elliott MM, Becker-Hapak M, et al. Combining AFM13, a bispecific CD30/CD16 antibody, with cytokine-activated blood and cord blood-derived NK cells facilitates CAR-like responses against CD30<sup>+</sup> malignancies. *Clin Cancer Res* 2021;**27**:3744–56.
121. Rothe A, Sasse S, Topp MS, Eichenauer DA, Hummel H, Reiners KS, et al. A phase 1 study of the bispecific anti-CD30/CD16A antibody construct AFM13 in patients with relapsed or refractory Hodgkin lymphoma. *Blood* 2015;**125**:4024–31.
122. Bartlett NL, Herrera AF, Domingo-Domenech E, Mehta A, Forero-Torres A, Garcia-Sanz R, et al. A phase 1b study of AFM13 in combination with pembrolizumab in patients with relapsed or refractory Hodgkin lymphoma. *Blood* 2020;**136**:2401–9.
123. Sasse S, Bröckelmann PJ, Momotow J, Plütschow A, Hüttmann A, Basara N, et al. AFM13 in patients with relapsed or refractory classical Hodgkin lymphoma: final results of an open-label, randomized, multicenter phase II trial. *Leuk Lymphoma* 2022;**63**:1871–8.
124. Mandikian D, Takahashi N, Lo AA, Li J, Eastham-Anderson J, Slaga D, et al. Relative target affinities of T cell-dependent bispecific antibodies determine biodistribution in a solid tumor mouse model. *Mol Cancer Therapeut* 2018;**17**:776–85.
125. Liu L, Lam CK, Long V, Widjaja L, Yang Y, Li H, et al. MGD011, A CD19 × CD3 dual-affinity retargeting Bi-specific molecule incorporating extended circulating half-life for the treatment of B-cell malignancies. *Clin Cancer Res* 2017;**23**:1506–18.
126. Dang K, Castello G, Clarke SC, Li Y, Balasubramani A, Boudreau A, et al. Attenuating CD3 affinity in a PSMA×CD3 bispecific antibody enables killing of prostate tumor cells with reduced cytokine release. *J Immunother Cancer* 2021;**19**:e002488.
127. Yu X, Orr CM, Chan HTC, James S, Penfold CA, Kim J, et al. Reducing affinity as a strategy to boost immunomodulatory antibody agonism. *Nature* 2023;**614**:539–47.
128. Wang N, Patel H, Schneider IC, Kai X, Varshney AK, Zhou L. An optimal antitumor response by a novel CEA/CD3 bispecific antibody for colorectal cancers. *Antib Ther* 2021;**4**:90–100.
129. Stafin K, Zuch de Zafra CL, Schutt LK, Clark V, Zhong F, Hristopoulos M, et al. Target arm affinities determine preclinical efficacy and safety of anti-HER2/CD3 bispecific antibody. *JCI Insight* 2020;**5**:e133757.
130. Leong SR, Sukumaran S, Hristopoulos M, Totpal K, Stainton S, Lu E, et al. An anti-CD3/anti-CLL-1 bispecific antibody for the treatment of acute myeloid leukemia. *Blood* 2017;**129**:609–18.
131. Betts A, van der Graaf PH. Mechanistic quantitative pharmacology strategies for the early clinical development of bispecific antibodies in oncology. *Clin Pharmacol Ther* 2020;**108**:528–41.
132. Bacac M, Fauti T, Sam J, Colombetti S, Weinzierl T, Oualet D, et al. A novel carcinoembryonic antigen T-cell bispecific antibody (CEA TCB) for the treatment of solid tumors. *Clin Cancer Res* 2016;**22**:3286–97.
133. Wu X, Sereno AJ, Huang F, Lewis SM, Lieu RL, Weldon C, et al. Fab-based bispecific antibody formats with robust biophysical properties and biological activity. *mAbs* 2015;**7**:470–82.
134. Luo Y, Ye S, Li X, Lu W. Emerging structure–function paradigm of endocrine FGFs in metabolic diseases. *Trends Pharmacol Sci* 2019; **40**:142–53.
135. Sebag M, Raje NS, Bahlis NJ, Costello C, Dholaria B, Solh M, et al. Elranatamab (PF-06863135), a B-cell maturation antigen (BCMA) targeted CD3-engaging bispecific molecule, for patients with relapsed or refractory multiple myeloma: results from magnetism-1. *Blood* 2021;**138**:895.
136. Wu L, Huang Y, Sienkiewicz J, Sun J, Guiang L, Li F, et al. Bispecific BCMA-CD3 antibodies block multiple myeloma tumor growth. *Cancers* 2022;**14**:2518.
137. Santich BH, Cheal SM, Ahmed M, McDevitt MR, Ouerfelli O, Yang G, et al. A self-assembling and disassembling (SADA) bispecific antibody (BsAb) platform for curative two-step pretargeted radioimmunotherapy. *Clin Cancer Res* 2021;**27**:532–41.
138. van de Donk NWCJ, O'Neill C, de Ruijter MEM, Verkleij CPM, Zweegman S. T-cell redirecting bispecific and trispecific antibodies in multiple myeloma beyond BCMA. *Curr Opin Oncol* 2023;**35**:601–11.
139. Ma R, Li Z, Chiocca EA, Caligiuri MA, Yu J. The emerging field of oncolytic virus-based cancer immunotherapy. *Trends Cancer* 2023;**9**:122–39.
140. Yu F, Wang X, Guo ZS, Bartlett DL, Gottschalk SM, Song XT. T-cell engager-armed oncolytic vaccinia virus significantly enhances anti-tumor therapy. *Mol Ther* 2014;**22**:102–11.
141. Crawford A, Chiu D. Targeting solid tumors using CD3 bispecific antibodies. *Mol Cancer Therapeut* 2021;**20**:1350–8.
142. Venugopal S, Sharma V, Mehra A, Singh I, Singh G. DNA intercalators as anticancer agents. *Chem Biol Drug Des* 2022;**100**:580–98.
143. Godziewa M, Ciesielski S. Natural DNA intercalators as promising therapeutics for cancer and infectious diseases. *Curr Cancer Drug Targets* 2020;**20**:19–32.
144. Liu Z, Zou H, Dang Q, Xu H, Liu L, Zhang Y, et al. Biological and pharmacological roles of m6A modifications in cancer drug resistance. *Mol Cancer* 2022;**21**:220.
145. Lajous H, Lelièvre B, Vauléon E, Lecomte P, Garcion E. Rethinking alkylating(-like) agents for solid tumor management. *Trends Pharmacol Sci* 2019;**40**:342–57.
146. Burgess JT, Rose M, Boucher D, Plowman J, Molloy C, Fisher M, et al. The therapeutic potential of DNA damage repair pathways and genomic stability in lung cancer. *Front Oncol* 2020;**10**:1256.
147. Ahlmann M, Hempel G. The effect of cyclophosphamide on the immune system: implications for clinical cancer therapy. *Cancer Chemother Pharmacol* 2016;**78**:661–71.
148. Emadi A, Jones RJ, Brodsky RA. Cyclophosphamide and cancer: golden anniversary. *Nat Rev Clin Oncol* 2009;**6**:638–47.
149. Bracci L, Moschella F, Sestili P, La Sorsa V, Valentini M, Canini I, et al. Cyclophosphamide enhances the antitumor efficacy of adoptively transferred immune cells through the induction of cytokine expression, B-cell and T-cell homeostatic proliferation, and specific tumor infiltration. *Clin Cancer Res* 2007;**13**:644–53.
150. Meermeier EW, Welsh SJ, Sharik ME, Du MT, Garbitt VM, Riggs DL, et al. Tumor burden limits bispecific antibody efficacy through T cell exhaustion averted by concurrent cytotoxic therapy. *Blood Cancer Discov* 2021;**2**:354–69.
151. Oostindie SC, Lazar GA, Schuurman J, Parren PW. Avidity in antibody effector functions and biotherapeutic drug design. *Nat Rev Drug Discov* 2022;**21**:715–35.
152. Birrer MJ, Moore KN, Betella I, Bates RC. Antibody–drug conjugate-based therapeutics: state of the science. *J Natl Cancer Inst* 2019;**111**:538–49.
153. Tsuchikama K, An Z. Antibody–drug conjugates: recent advances in conjugation and linker chemistries. *Protein Cell* 2018; **9**:33–46.
154. Rallis KS, Lai Yau TH, Sideris M. Chemoradiotherapy in cancer treatment: rationale and clinical applications. *Anticancer Res* 2021; **41**:1–7.
155. Sgouros G, Bodei L, McDevitt MR, Nedrow JR. Radiopharmaceutical therapy in cancer: clinical advances and challenges. *Nat Rev Drug Discov* 2020;**19**:589–608.
156. Chan GC, Chan CM. Anti-GD2 directed immunotherapy for high-risk and metastatic neuroblastoma. *Biomolecules* 2022;**12**:358.



157. Zirngibl F, Ivasko SM, Grunewald L, Klaus A, Schwiebert S, Ruf P, et al. GD2-directed bispecific trifunctional antibody outperforms dinutuximab beta in a murine model for aggressive metastasized neuroblastoma. *J Immunother Cancer* 2021;**9**:e002923.
158. Deppisch N, Ruf P, Eissler N, Neff F, Buhmann R, Lindhofer H, et al. Efficacy and tolerability of a GD2-directed trifunctional bispecific antibody in a preclinical model: subcutaneous administration is superior to intravenous delivery. *Mol Cancer Therapeut* 2015;**14**:1877–83.
159. Zhai D, An D, Wan C, Yang K. Radiotherapy: brightness and darkness in the era of immunotherapy. *Transl Oncol* 2022;**19**:101366.
160. Harrington K, Freeman DJ, Kelly B, Harper J, Soria JC. Optimizing oncolytic virotherapy in cancer treatment. *Nat Rev Drug Discov* 2019;**18**:689–706.
161. Kaufman HL, Kohlhapp FJ, Zloza A. Oncolytic viruses: a new class of immunotherapy drugs. *Nat Rev Drug Discov* 2015;**14**:642–62.
162. Hemminki O, Dos Santos JM, Hemminki A. Oncolytic viruses for cancer immunotherapy. *J Hematol Oncol* 2020;**13**:84.
163. Tian Y, Xie D, Yang L. Engineering strategies to enhance oncolytic viruses in cancer immunotherapy. *Signal Transduct Targeted Ther* 2022;**7**:117.
164. Ma R, Li Z, Chiocca EA, Caligiuri MA, Yu J. The emerging field of oncolytic virus-based cancer immunotherapy. *Trends Cancer* 2023;**9**:122–39.
165. Wang Q, Ma X, Wu H, Zhao C, Chen J, Li R, et al. Oncolytic adenovirus with MUC16-BiTE shows enhanced antitumor immune response by reversing the tumor microenvironment in PDX model of ovarian cancer. *Oncol Immunology* 2022;**11**:2096362.
166. Speck T, Heidebuechel JPW, Veinalde R, Jaeger D, von Kalle C, Ball CR, et al. Targeted BiTE expression by an oncolytic vector augments therapeutic efficacy against solid tumors. *Clin Cancer Res* 2018;**24**:2128–37.
167. Leclercq G, Haegel H, Toso A, Zimmermann T, Green L, Steinhoff N, et al. JAK and mTOR inhibitors prevent cytokine release while retaining T cell bispecific antibody *in vivo* efficacy. *J Immunother Cancer* 2022;**10**:e003766.
168. Burger JA. Bruton tyrosine kinase inhibitors: present and future. *Cancer J* 2019;**25**:386–93.
169. Pal Singh S, Dammeijer F, Hendriks RW. Role of Bruton's tyrosine kinase in B cells and malignancies. *Mol Cancer* 2018;**17**:57.
170. Mhibik M, Gaglione EM, Eik D, Kendall EK, Blackburn A, Keyvanfar K, et al. BTK inhibitors, irrespective of ITK inhibition, increase efficacy of a CD19/CD3-bispecific antibody in CLL. *Blood* 2021;**138**:1843–54.
171. Long M, Beckwith K, Do P, Mundy BL, Gordon A, Lehman AM, et al. Ibrutinib treatment improves T cell number and function in CLL patients. *J Clin Invest* 2017;**127**:3052–64.
172. Niemann CU, Herman SE, Maric I, Gomez-Rodriguez J, Biancotto A, Chang BY, et al. Disruption of *in vivo* chronic lymphocytic leukemia tumor-microenvironment interactions by ibrutinib-findings from an investigator-initiated phase II study. *Clin Cancer Res* 2016;**22**:1572–82.
173. Kondo K, Shaim H, Thompson PA, Burger JA, Keating M, Estrov Z, et al. Ibrutinib modulates the immunosuppressive CLL microenvironment through STAT3-mediated suppression of regulatory B-cell function and inhibition of the PD-1/PDL1 pathway. *Leukemia* 2018;**32**:960–70.
174. Zou YX, Zhu HY, Li XT, Xia Y, Miao KR, Zhao SS, et al. The impacts of zanubrutinib on immune cells in patients with chronic lymphocytic leukemia/small lymphocytic lymphoma. *Hematol Oncol* 2019;**37**:392–400.
175. Köhnke T, Krupka C, Tischer J, Knösel T, Subklewe M. Increase of PD-L1 expressing B-precursor ALL cells in a patient resistant to the CD19/CD3-bispecific T cell engager antibody blinatumomab. *J Hematol Oncol* 2015;**8**:111.
176. Kosmaczewska A, Ciszak L, Suwalska K, Wolowicz D, Frydecka I. CTLA-4 overexpression in CD191/CD51 cells correlates with the level of cell cycle regulators and disease progression in B-CLL patients. *Leukemia* 2005;**19**:301–4.
177. Belmontes B, Sawant DV, Zhong W, Tan H, Kaul A, Aeffner F, et al. Immunotherapy combinations overcome resistance to bispecific T cell engager treatment in T cell-cold solid tumors. *Sci Transl Med* 2021;**13**:eabd1524.
178. Martinez-Cannon BA, Castro-Sanchez A, Barragan-Carrillo R, de la Rosa Pacheco S, Platas A, Fonseca A, et al. Adherence to adjuvant tamoxifen in mexican young women with breast cancer. *Patient Prefer Adherence* 2021;**15**:1039–49.
179. Liu D, Bao L, Zhu H, Yue Y, Tian J, Gao X, et al. Microenvironment-responsive anti-PD-L1 × CD3 bispecific T-cell engager for solid tumor immunotherapy. *J Control Release* 2023;**354**:606–14.
180. Cattaruzza F, Nazeer A, To M, Hammond M, Koski C, Liu LY, et al. Precision-activated T-cell engagers targeting HER2 or EGFR and CD3 mitigate on-target, off-tumor toxicity for immunotherapy in solid tumors. *Nat Cancer* 2023;**4**:485–501.
181. Seckinger A, Majocchi S, Moine V, Nouveau L, Ngoc H, Daubeuf B, et al. Development and characterization of NILK-2301, a novel CEACAM5×CD3  $\kappa\lambda$  bispecific antibody for immunotherapy of CEACAM5-expressing cancers. *J Hematol Oncol* 2023;**16**:117.
182. Strohl WR, Naso M. Bispecific T-cell redirection *versus* chimeric antigen receptor (CAR)-T cells as approaches to kill cancer cells. *Antibodies* 2019;**8**:41.
183. Li J, Piskol R, Ybarra R, Chen YJ, Li J, Slaga D, et al. CD3 bispecific antibody-induced cytokine release is dispensable for cytotoxic T cell activity. *Sci Transl Med* 2019;**11**:eaax8861.
184. Norelli M, Camisa B, Barbiera G, Falcone L, Purevdorj A, Genua M, et al. Monocyte-derived IL-1 and IL-6 are differentially required for cytokine-release syndrome and neurotoxicity due to CART cells. *Nat Med* 2018;**24**:739–48.
185. Schildberger A, Rossmann E, Eichhorn T, Strassl K, Weber V. Monocytes, peripheral blood mononuclear cells, and THP-1 cells exhibit different cytokine expression patterns following stimulation with lipopolysaccharide. *Mediat Inflamm* 2013;**2013**:697972.
186. Iwata Y, Sasaki M, Harada A, Taketo J, Hara T, Akai S, et al. Daily ascending dosing in cynomolgus monkeys to mitigate cytokine release syndrome induced by ERY22, surrogate for T-cell redirecting bispecific antibody ERY974 for cancer immunotherapy. *Toxicol Appl Pharmacol* 2019;**379**:114657.
187. Trinklein ND, Pham D, Schellenberger U, Buelow B, Boudreau A, Choudhry P, et al. Efficient tumor killing and minimal cytokine release with novel T-cell agonist bispecific antibodies. *mAbs* 2019;**11**:639–52.
188. Vafa O, Trinklein ND. Perspective: designing T-cell engagers with better therapeutic windows. *Front Oncol* 2020;**10**:446.
189. Wu Z, Cheung NV. T cell engaging bispecific antibody (T-cell redirecting BsAb): from technology to therapeutics. *Pharmacol Ther* 2018;**182**:161–75.
190. Peters IT, Hilders CG, Sier CF, Vahrmeijer AL, Smit VT, Baptist Trimbos J, et al. Identification of cell-surface markers for detecting breast cancer cells in ovarian tissue. *Arch Gynecol Obstet* 2016;**294**:385–93.
191. Sulea T, Rohani N, Baardsnes J, Corbeil CR, Deprez C, Cepero-Donates Y, et al. Structure-based engineering of pH-dependent antibody binding for selective targeting of solid-tumor microenvironment. *mAbs* 2020;**12**:1682866.