

RESEARCH ARTICLE

GeneXpert on patients with human immunodeficiency virus and smear-negative pulmonary tuberculosis

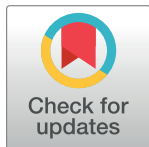
Nguyen Kim Cuong^{1,2}, Nguyen Bao Ngoc³*, Nguyen Binh Hoa⁴‡, Vu Quoc Dat⁵‡, Nguyen Viet Nhung⁴

1 Department of Tuberculosis and Lung Disease, Hanoi Medical University, Hanoi, Vietnam, **2** Department of Respiratory Tuberculosis, National Lung Hospital, Hanoi, Vietnam, **3** Department of Pharmacy, National Lung Hospital, Hanoi, Vietnam, **4** National Tuberculosis Programme, Hanoi, Vietnam, **5** Department of Infectious Diseases, Hanoi Medical University, Hanoi, Vietnam

 These authors contributed equally to this work.

‡ NBH, VQD and NVN also contributed equally to this work.

* baongochup@gmail.com



OPEN ACCESS

Citation: Cuong NK, Ngoc NB, Hoa NB, Dat VQ, Nhung NV (2021) GeneXpert on patients with human immunodeficiency virus and smear-negative pulmonary tuberculosis. PLoS ONE 16(7): e0253961. <https://doi.org/10.1371/journal.pone.0253961>

Editor: Shampa Anupurba, Institute of Medical Sciences, Banaras Hindu University, INDIA

Received: August 27, 2020

Accepted: June 17, 2021

Published: July 6, 2021

Copyright: © 2021 Cuong et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its [Supporting Information](#) files.

Funding: Mr. Cuong got grants from the VIETNAM ADMINISTRATION OF HIV/AIDS CONTROL (grant number: 0095, 2013). URL: <http://vaac.gov.vn/en-us> The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Abstract

Objectives

Vietnam is a high-prevalence country for tuberculosis (TB). Xpert MTB/RIF is a novel PCR-based diagnostic test that is substantially more sensitive for detecting *M. tuberculosis* than traditional smear-based techniques. However, locally-derived evidence of Xpert MTB/RIF in HIV-infected people is limited. This study evaluates the performance of the Xpert MTB/RIF in HIV-infected patients with smear-negative pulmonary TB (SNTB).

Methods

This was a cross-sectional study in 3 hospitals. The performance of Xpert MTB/RIF was compared with the reference standard of liquid culture and phenotypic drug-susceptibility testing for rifampicin (RIF) resistance.

Results

Out of 123 patients, the median age was 37.0 (IQR: 32.0–41.0) and 81.3% were male. The area under the receiver operating characteristic curve, sensitivity (Se) and specificity (Sp) of Xpert MTB/RIF for pulmonary TB diagnosis were 0.72 (95% confidence interval [CI]: 0.63–0.81), 66.7% (95%CI: 54.8–77.1) and 77.1% (95%CI: 62.7–88.0), respectively, while Se and Sp of Xpert MTB/RIF in detecting RIF resistance were 50.0 (11.8–88.2) and 86.4% (95%CI: 72.7–94.8).

Conclusion

The performance of Xpert MTB/RIF in HIV-infected patients with SNTB for the diagnosis of TB and RIF-resistance was low. Further studies are required to evaluate the results of Xpert

MTB/RIF assay in HIV-infected patients with SNTB and the role of Xpert repetition on the same specimens.

Introduction

Tuberculosis (TB) is the globally leading cause of death among people living with human immunodeficiency virus (HIV), accounting for 1.2 million TB-related mortalities among seronegative persons and 208,000 deaths among HIV-infected patients [1]. The World Health Organization (WHO) estimated that among 10.0 million (range, 9.0–11.1 million) TB cases, the proportion of HIV-infected patients were 8.2% [1]. Additionally, TB remains the leading cause of death among HIV-infected patients, accounting for around one in three AIDS-related deaths [2]. HIV is the most important risk factor for TB infection progressing to TB disease. Therefore, it is necessary to prevent, detect and treat TB in HIV co-infected patients. Nowadays, chest X-ray, acid-fast bacilli (AFB) smear, Xpert MTB/RIF, and culture are the most frequent laboratory methods to detect TB. While culture remains the gold standard for TB diagnosis, Xpert MTB/RIF is recommended as the initial test for TB in HIV-infected patients because its sensitivity is substantially higher than that of AFB smear [3, 4].

Among HIV-infected patients, the clinical manifestations are often atypical, particularly in the late stage of HIV infection, non-cavitary disease, lower lobe infiltrates, hilar lymphadenopathy, and pleural effusion [5]. This results in a low proportion of patients with a positive bacteriological test. In December 2010, The WHO first recommended the use of the Xpert MTB/RIF assay (Cepheid, Sunnyvale, CA, USA). The guidelines for using Xpert MTB/RIF in diagnosing pulmonary and extrapulmonary TB in adults and children were reviewed and updated in 2016 [3, 4, 6, 7]. Xpert MTB/RIF is an automated semiquantitative molecular test that allows for the simultaneous detection of mutations in the *rpoB* gene. The test integrates three technologies (gene extraction, amplification and recognition) and provides results within two hours. It is a sensitive test for detecting both *M. tuberculosis* and RIF resistance, allowing early initiation of appropriate antibiotic therapy. WHO endorses Xpert MTB/RIF as the initial test for presumptive pulmonary TB and MDR-TB. In HIV-infected patients, this test is substantially more sensitive than smear and therefore recommended as an initial test for diagnosing with TB [3, 4].

Xpert MTB/RIF plays a key role in diagnosing HIV-infected patients with negative-smear pulmonary TB (SNTB). This measure can shorten turnaround time and provide additional evidence for TB diagnosis when practitioners wait for culture results. Although the first study of the Xpert MTB/RIF showed a sensitivity ranging between 98% in smear-positive TB and 72% in SNTB, subsequent evidence has shown large variability in SNTB, with sensitivity ranging between 26% and 67% [8–14]. This is particularly common in HIV-infected patients, children or patients with SNTB or extrapulmonary TB [11, 14–16].

Since 2011, the Vietnam National Tuberculosis Programme (NTP) has scaled up Xpert MTB/RIF as an initial test to detect TB and RIF resistance for HIV-infected people [17]. There are several studies about applying Xpert MTB/RIF in the diagnosis of TB among HIV-infected patients [13, 18–20]. However, there is a lack of research on the value of this technique in HIV-infected patients with SNTB. Therefore, this study aims to evaluate the performance of the Xpert MTB/RIF in HIV-infected patients with SNTB.

Methods

Study population

This cross-sectional study was conducted from January 2013 to December 2015 in three hospitals in Hanoi, Vietnam, including two hospitals specialized in TB (National Lung Hospital, Hanoi Lung Hospital) and one hospital specialized in HIV (09 Hospital). We consecutively enrolled HIV-infected patients with SNTB \geq 18 years old. The definition of SNTB was that patients had a chest x-ray and clinical symptoms more than two weeks suggesting pulmonary TB (interpreted by two independent trained doctors) and had at least two expectorate negative AFB smears [21]. The exclusion criteria were patients in severe HIV conditions [22].

Sample size

This study aims to determine how sensitive Xpert MTB/RIF is in diagnosing HIV-infected patients with SNTB. The proportion of culture-positive TB cases were 68% in HIV-infected patients with SNTB in Vietnam [23]. Previous studies showed that the sensitivity of Xpert MTB/RIF was about 70% [8–14]. The null hypothesis of this study was that the sensitivity of Xpert MTB/RIF was 70%, while the alternative hypothesis was that sensitivity was not equal to 70%. Therefore, the sample size that would need to have 95% confidence and 80% power to detect a difference of 10% from sensitivity of 70% was 123 patients [24].

Study procedure

All HIV-infected patients with clinically presumptive TB were initially screened for TB by two spontaneous sputum specimens and chest x-ray following national standard practice. Patients who met the inclusion criteria were approached and invited to participate in the study. Patients who consented to participate were enrolled and requested to provide one additional spot sputum sample. When patients could not produce an adequate expectorate, sputum induction was performed by nebulizing sterile 5% saline. This sputum sample was divided into 2 parts for liquid culture and Xpert MTB/RIF. For liquid culture, 5 ml sputum pellets were inoculated on liquid medium (BD BBL Mannula MGIT, Cockeysville, MD, USA) and read using the BD BACTEC Micro MGIT Fluorescence Reader (Cockeysville, MD, USA). The culture-positive samples underwent conventional drug susceptibility testing (DST) for isoniazid (INH), rifampicin (RIF), pyrazinamide (PZA), ethambutol (EMB) and streptomycin (S). The second half of the homogeneous sputum sample was tested with Xpert MTB/RIF by adding the sample reagent to untreated sputum in a 2:1 ratio (1ml of untreated sputum to 2 ml of the sample reagent). After that, 2 ml of this mixture was transferred to the Xpert MTB/RIF cartridge and the cartridge was then loaded into the GeneXpert device. Finally, the results were interpreted and displayed by the GeneXpert system from measured fluorescent signals.

All procedures, including sputum induction, microscopy, Xpert MTB/RIF, liquid culture, and DST, were implemented in each hospital laboratory based on biosafety level III [6]. Sputum specimens from patients were collected in a ventilated sputum collection room and submitted to the laboratory to process within 24 hours [21, 25]. Direct sputum-smear microscopy and Xpert MTB/RIF were performed using good microbiological techniques on an open bench with ventilation [21, 25]. In contrast, procedures related to liquefying specimens for liquid culture and DST were performed in a biological safety cabinet.

We collected data on the DST using the standard conventional drug susceptibility of *M. tuberculosis* isolates to INH, RIF, PZA, EMB, and S. Diagnosis of MDR-TB was based on conventional DST method, BACTEC MGIT 960 culture and Xpert MTB/RIF assay. Demographics and chest x-ray results were extracted from patient records.

Statistical analysis

We provided descriptive statistics on patient demographics and clinical data. We provided medians with interquartile ranges, and conducted the Mann–Whitney U-tests with continuous variables with skewed distribution. Normally distributed continuous variables were presented as mean and standard deviation ranges, and compared by Student's t-tests. Chi-square or Fisher's exact tests were employed for dichotomous variables. We calculated AUC-ROC, sensitivity (Se), specificity (Sp), positive predictive value (PPV), negative predictive value (NPV), positive likelihood ratio (PLR), negative likelihood ratio (NLR) with 95% confidence interval (95%CI) based on a binomial distribution. The measure of agreement was used by Cohen's Kappa statistic. Hypothesis testing was two-sided and p-values less than 0.05 were considered statistically significant. We performed analyses using SPSS 25 (IBM Corp., Armonk, NY, USA).

Ethics statement

This study was given scientific and ethical approval by the Institutional Ethics and Scientific Review Board of the National Lung Hospital under approval letter number 10/2013/NCKHCS. All patients in this study were informed about the risks and benefits of the study and signed an individual written informed consent.

Results

From January 2013 to December 2015, we enrolled 123 HIV-infected patients with SNTB, and the majority of patients were male (81.3%). The median participant age was 37.0 years (IQR: 32.0–41.0). At the time of enrolment, 59 (48.0%) patients were on ART treatment, in which 23 (18.7%) patients were on ART treatment for a median of 7.0 (3.0–20.0) months. The CD4 counts were available for 82 (66.7%) patients with a median of 247.0 (100.8–400.0) cells/ml. There were 6 (5.3%) patients on INH prophylaxis before enrolment. Spontaneous sputum specimens were collected in 76 (61.8%) patients and 47 (38.2%) patients required induced sputum collection. Characteristics of patients in this study are shown in [Table 1](#).

The results of Xpert MTB/RIF assay against BACTEC MGIT 960 culture are in [Table 2](#). Sixty-one patients (49.6%) were diagnosed with TB by Xpert MTB/RIF, while BACTEC MGIT 960 culture detected TB in 75 patients (61.0%). Diagnostic efficacy of Xpert was assessed in all 123 cases with the overall Se, Sp, PPV and NPV of 66.7% (95%CI: 54.8–77.1), 77.1% (95%CI: 62.7–88.0), 82.0% (95%CI: 72.5–88.7) and 59.7% (95%CI: 50.9–68.0), respectively. Compared with the spontaneous expectorate group, Xpert MTB/RIF in the induced sputum group showed similar results in AUC-ROC and sensitivity tests. However, patients in the induced sputum group had the highest PPV and PLR with 85.0% and 4.58, respectively. The kappa of 0.42 in the whole study and 0.50 in the induced sputum group represents a moderate level of agreement between Xpert and BACTEC MGIT 960 culture.

There were 75 culture-positive samples in culture, in which 5 cases happened to errors or contaminations while DST was performed. GeneXpert detected 61 positive cases, in which 3 cases did not be distinguished RIF resistance because of errors. Therefore, only 50 cases had both DST and Xpert MTB/RIF results. The results of GeneXpert MTB/RIF assay against standard conventional DST for MDR-TB are also presented in [Table 3](#). Diagnostic efficacy of Xpert for MDR-TB in culture-positive cases was high with overall Sp and NPV of 86.4% (95% CI: 72.7–94.8) and 92.7% (95%CI: 84.9–96.6), respectively ([Table 3](#)). However, Se and PPV were less than 50%.

Table 1. Baseline characteristics of study participants.

Clinical characteristics	All (N = 123)		Spontaneous sputum (N = 76)		Induced sputum (N = 47)		P- value
	N	Value	N	Value	N	Value	
Age (years) (median, IQR ^a)	123	37.0 (32.0–41.0)	76	36.0 (31.0–40.0)	47	37.0 (34.0–43.0)	0.254
Male gender (%)	100	81.3	65	85.5	35	74.5	0.126
Body mass index (median, IQR)	123	17.6 (16.3–29.5)	76	17.9 (16.2–19.6)	47	17.5 (16.0–18.8)	0.358
Intravenous drug user (%)	66	53.7	44	57.9	22	46.8	0.231
Direct contact with individuals with TB ^b (%)	25	20.3	16	21.1	9	19.1	0.799
ART ^c treatment (%)	59	48.0	30	39.5	29	61.7	0.016
Duration of ART among ART experienced patients (months) (median, IQR)	23	7.0 (3.0–20.0)	10	7.0 (4.3–14.0)	13	7.0 (3.0–25.0)	0.832
INH ^d prophylaxis at any time (%)	6	5.3	2	2.6	4	8.5	0.201
CD4 counts at the time of HIV diagnosis (cells/ml) (median, IQR)	82	247.0 (100.8–400.0)	47	250.0 (114.0–375.0)	35	200.0 (97.0–400.0)	0.409
Pulmonary cavitation on chest radiograph (%)	33	26.8	25	32.9	8	17.0	0.054
Anemia (%)	97	78.9	57	75.0	40	85.1	0.182

IQR^a, Interquartile range;

TB^b, Tuberculosis;

ART^c, Antiretroviral therapy;

INH^d, Isoniazid

<https://doi.org/10.1371/journal.pone.0253961.t001>

Table 2. Performance of Xpert MTB/RIF assay against BACTEC MGIT 960 culture as the reference standard.

Xpert MTB/RIF ^a	BACTEC MGIT 960 culture								
	All n (%)			Induced sputum n (%)			Spontaneous sputum n (%)		
	Negative	Positive	Total	Negative	Positive	Total	Negative	Positive	Total
Negative	37 (30.1)	25 (20.3)	62 (50.4)	18 (38.3)	9 (19.1)	27 (57.4)	19 (25.0)	16 (21.1)	35 (46.1)
Positive	11 (8.9)	50 (40.7)	61 (49.6)	3 (6.4)	17 (36.2)	20 (42.6)	8 (10.5)	33 (43.4)	41 (53.9)
Total	48 (39.0)	75 (61.0)	123 (100.0)	21 (44.7)	26 (55.3)	47 (100.0)	27 (35.5)	49 (64.5)	76 (100.0)
Xpert MTB/RIF	All (95% CI) ⁱ			Induced sputum (95% CI)			Spontaneous sputum (95% CI)		
AUC-ROC ^b	0.72 (0.63–0.81)			0.76 (0.61–0.90)			0.69 (0.56–0.82)		
Kappa	0.42 (0.26–0.57)			0.50 (0.26–0.74)			0.35 (0.15–0.56)		
Se ^c %	66.7 (54.8–77.1)			65.4 (44.3–82.8)			67.4 (52.5–80.0)		
Sp ^d %	77.1 (62.7–88.0)			85.7 (63.7–97.0)			70.4 (49.8–86.3)		
PPV ^e %	82.0 (72.5–88.7)			85.0 (65.7–94.4)			80.5 (69.1–88.4)		
NPV ^f %	59.7 (50.9–68.0)			66.7 (53.4–77.7)			54.3 (42.6–65.5)		
PLR ^g	2.91 (1.69–5.01)			4.58 (1.55–13.54)			2.27 (1.23–4.20)		
NLR ^h	0.43 (0.30–0.62)			0.40 (0.23–0.70)			0.46 (0.29–0.74)		

MTB/RIF^a, Mycobacterium tuberculosis/Rifampicin;

AUC-ROC^b, Area Under the Receiver Operating Characteristic Curve;

Se^c, Sensitivity;

Sp^d, Specificity;

PPV^e, Positive predictive value;

NPV^f, Negative predictive value;

PLR^g, Positive likelihood ratio;

NLR^h, Negative likelihood ratio;

CIⁱ, confidence interval.

<https://doi.org/10.1371/journal.pone.0253961.t002>

Table 3. Performance of Xpert MTB/RIF assay against standard conventional drug susceptibility test for MDR-TB.

GeneXpert MTB/RIF ^a	Drug Susceptibility Test of RIF n (%)		
	Resistant	Sensitive	Total
Resistant	3 (6.0)	6 (12.0)	9 (18.0)
Sensitive	3 (6.0)	38 (76.0)	41 (82.0)
Total	6 (12.0)	44 (88.0)	50 (100.0)
Xpert MTB/RIF (95% CI)¹			
AUC-ROC ^b	0.68 (0.43–0.94)		
Kappa	0.30 (0.01–0.58)		
Se ^c %	50.0 (11.8–88.2)		
Sp ^d %	86.4 (72.7–94.8)		
PPV ^e %	33.3 (14.4–59.9)		
NPV ^f %	92.7 (84.9–96.6)		
PLR ^g	3.67 (1.23–10.93)		
NLR ^h	0.58 (0.26–1.30)		

MTB/RIF^a, Mycobacterium tuberculosis/Rifampicin;

AUC-ROC^b, Area Under the Receiver Operating Characteristic Curve;

Se^c, Sensitivity;

Sp^d, Specificity;

PPV^e, Positive predictive value;

NPV^f, Negative predictive value;

PLR^g, Positive likelihood ratio;

NLR^h, Negative likelihood ratio;

CI¹, confidence interval.

<https://doi.org/10.1371/journal.pone.0253961.t003>

Discussion

Xpert MTB/RIF is a novel PCR-based diagnostic test that is substantially more sensitive for detecting *M. tuberculosis* than traditional smear-based techniques. However, there is limited evidence on the value of Xpert MTB/RIF in HIV-infected patients with SNTB. This study showed that the performance of Xpert MTB/RIF in the diagnosis of TB and MDR-TB was fair and induced sputum collection could detect more TB cases than spontaneously expectorated sputum.

In this study, the Xpert MTB/RIF has moderate sensitivity and specificity with 66.7% (95% CI: 54.8–77.1) and 77.1% (95%CI: 62.7–88.0), respectively. A previous study found that the sensitivity of Xpert MTB/RIF in HIV-infected patients with SNTB was 47.3 (95% CI: 29.2–67.0) [14]. HIV co-infection was associated with a significant reduction of NPV and decreasing trend of sensitivity. A multi-center study in the United States, Brazil, and South Africa found that the sensitivity of Xpert MTB/RIF was low in HIV-infected participants with only 52.1% (95% CI: 38.3%–65.5%) [26]. Results in a meta-analysis assessing the accuracy of Xpert MTB/RIF in HIV-positive individuals by smear status also were comparable to our results [13]. Among HIV-infected patients, the pooled sensitivity of Xpert MTB/RIF was 61% (95%CI: 40%–81%) for SNTB compared with 97% (95%: 90%–99%) for smear-positive pulmonary TB [13]. Overall, studies evaluating performance of Xpert MTB/RIF on HIV-infected patients with SNTB had similar research design and concordant results with our study. The moderate sensitivity and specificity could be due to the inability of Xpert MTB/RIF to detect DNA of *M. tuberculosis* in concentrations below 131 cfu/ml (the lowest threshold of Xpert MTB can detect TB bacteria), while sputum culture could detect TB bacteria at the threshold of 10–100 cfu/ml

[27, 28]. Furthermore, the presence of inhibitors to the gene amplification enzyme (PCR) in the test specimen might present another challenge for the Xpert MTB/RIF [29]. Therefore, the role of Xpert MTB/RIF in diagnosing HIV-infected patients with SNTB should be further optimized as the moderate sensitivity and specificity. These results showed that the final diagnosis should be based on carefully examining clinical symptoms combined with traditional microbiological tests such as sputum smear and culture.

In a previous study on 171 cases of smear-negative and culture-positive sputum, the sensitivity value increased to 72.5%, 85.1%, 90.2% when Xpert MTB/RIF was performed once, twice, and thrice on the same specimen [8]. In Vietnam, Xpert MTB/RIF has been used as the initial test because of the lower sensitivity of AFB smear microscopy and the need for early TB treatment in HIV-infected patients. However, our study showed moderate sensitivity and specificity, so repeating Xpert MTB/RIF test could be a good method to increase its sensitivity. However, the cost of Xpert MTB/RIF is still high compared to smear microscopy, and further studies about the Xpert MTB/RIF performance in HIV-infected patients with SNTB are needed.

The detection of RIF-resistance of Xpert MTB/RIF in this study was lower than in previous studies [10, 30]. However, the sensitivity and specificity of Xpert MTB/RIF for RIF resistant specimens have shown a large variability of 33–100% and 83–100%, respectively [3]. The pooled sensitivity in a meta-analysis was 91% (95%CI: 87–94%); the pooled specificity was 98% (95%CI: 96–99%) [31]. Although this study showed low performance in finding subjects with RIF resistance, it demonstrated an excellent rule-out value for RIF resistance. The discordant results between Xpert MTB/RIF and the conventional gold standard DST had negatively affected treatment decisions in the past [3, 31–34]. Several reasons could explain these differences. Firstly, DST was based on the growth of all *M. tuberculosis*, including both drug-resistant and -susceptible strains, while Xpert MTB/RIF detected genetic mutations. Secondly, the MGIT-DST method was imprecise if the mutation was detected on 511Pro, 516Tyr, 533Pro, 572Phe, and several 526 mutations [35]. A number of recent studies found that some strains of *M. tuberculosis* with Asp 516Tyr mutation in the *rpoB* gene could cause low levels of RIF resistance [33, 36]. Therefore, it was still drug-susceptible on MGIT, whereas the Xpert MTB/RIF result would be classified as RIF resistance. These cases would subsequently be identified as false positives. However, Xpert MTB/RIF could also detect silent mutations (nonfunctional gene) and missense mutations (nonfunctional protein) in the *rpoB* gene, occasionally leading to false-positive RIF resistance [33]. In our study, 3 patients had false-negative results, where RIF sensitive samples yielded resistant isolates on MGIT 960. 2 out of 3 patients were changed to resistance therapy, 1 out of 3 was unable to contact. Causes of false-negative results of Xpert MTB/RIF for RIF resistance have not been well established [37]. Some previous studies showed that although Xpert could detect a larger number of *rpoB* mutations, it was difficult to find all mutations related to rifampin resistance [38, 39]. A recent study on 370 patients with pulmonary TB mentioned that the presence of wild-type sequences detected by the probes, could be one of reasons leading to false-negative results [40]. The false-positive and false-negative cases in this study were not sequenced to determine the mismatch between the results as well as the likelihood of coinfection. This was a limitation of our study and should be addressed in future research. In order to resolve the discrepancy of Xpert MTB/RIF, culture, and DST in detecting *M. tuberculosis* or RIF-resistance, repeat Xpert MTB/RIF, Genotype® MTBDRplus assay, and sequencing of the *rpoB* gene should be implemented [32]. Additionally, stratifying the clinical risk for TB and drug-resistant TB based on patient characteristics and local epidemiology remains critical to optimizing initial TB treatment.

Our study has several strong points. Firstly, the number of HIV-infected patients meeting the criteria of SNTB was a significant sample size for studying this vulnerable population,

which could help to support the accuracy of the results. Secondly, spontaneous sputum and induced sputum were performed to optimize the quality of the sputum samples for testing. Thirdly, Xpert MTB/RIF assay and culture were performed on the identical sample (homogeneous sample), which was the best method for evaluating the accuracy of the diagnostic study. However, this study has some methodological limitations due to financial and logistic constraints. The data about history of HIV treatment, including prophylaxis, duration of ART treatment, CD4 counts at the time of HIV diagnosis, was retrospective and declarative, leading to bias. Sequencing was not performed for patients with discordant RIF resistant results between Xpert MBT/RIF and DST, so it was impossible to determine the exact cause for the discrepancy.

Conclusions

In our study, the diagnostic value of Xpert MTB/RIF in HIV-infected patients with SNTB was moderate compared to BACTEC MGIT 960 and conventional DST. However, Xpert MTB/RIF also showed that it had an excellent rule-out value for RIF resistance. Further studies should be performed to evaluate the performance of Xpert MTB/RIF assay in HIV-infected patients with SNTB and the role of repetition of Xpert MTB/RIF on the same specimens.

Supporting information

S1 Data.
(SAV)

Acknowledgments

The authors were grateful to the study participants and colleagues in Administrative of control and Against HIV/AIDS, National Lung Hospital, National TB Program of Vietnam who supported us in this research.

Author Contributions

Conceptualization: Nguyen Kim Cuong, Nguyen Bao Ngoc, Vu Quoc Dat, Nguyen Viet Nhung.

Data curation: Nguyen Bao Ngoc.

Formal analysis: Nguyen Bao Ngoc.

Funding acquisition: Nguyen Kim Cuong.

Methodology: Nguyen Kim Cuong, Nguyen Bao Ngoc, Nguyen Viet Nhung.

Project administration: Nguyen Kim Cuong, Nguyen Viet Nhung.

Supervision: Nguyen Kim Cuong, Nguyen Viet Nhung.

Visualization: Nguyen Bao Ngoc.

Writing – original draft: Nguyen Bao Ngoc.

Writing – review & editing: Nguyen Kim Cuong, Nguyen Bao Ngoc, Nguyen Binh Hoa, Vu Quoc Dat, Nguyen Viet Nhung.

References

1. World Health Organization. Global tuberculosis report. 2020. https://www.who.int/tb/publications/global_report/en/.
2. UNAIDS. Global HIV & AIDS statistics. 2019. <https://www.unaids.org/en/resources/fact-sheet>.
3. World Health Organization. Xpert MTB/RIF assay for the diagnosis of pulmonary and extrapulmonary TB in adults and children Policy update. 2013. <https://www.who.int/tb/publications/xpert-mtb-rif-assay-diagnosis-policy-update/en/>.
4. World Health Organization. Xpert MTB/RIF assay for the diagnosis TB: meeting report. 2016. <https://apps.who.int/iris/handle/10665/250383>.
5. Raviglione MC, Narain JP, Kochi A. HIV-associated tuberculosis in developing countries: clinical features, diagnosis, and treatment. *Bulletin of the World Health Organization*. 1992; 70(4):515–26. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2393393/>. PMID: 1394786
6. World Health Organization. Companion handbook to the WHO guidelines for the programmatic management of drug-resistant tuberculosis. 2014. https://www.who.int/tb/publications/pmdt_companionhandbook/en/.
7. World Health Organization. Improving the diagnosis and treatment of smear-negative pulmonary and extrapulmonary tuberculosis among adults and adolescents. 2007. <https://www.who.int/hiv/pub/tb/pulmonary/en/>.
8. Boehme CC, Nabeta P, Hillemann D, Nicol MP, Shenai S, Krapp F, et al. Rapid molecular detection of tuberculosis and rifampin resistance. *The New England journal of medicine*. 2010; 363(11):1005–15. <https://doi.org/10.1056/NEJMoa0907847> PMID: 20825313
9. Dorman SE, Schumacher SG, Alland D, Nabeta P, Armstrong DT, King B, et al. Xpert MTB/RIF Ultra for detection of *Mycobacterium tuberculosis* and rifampicin resistance: a prospective multicentre diagnostic accuracy study. *The Lancet Infectious Diseases*. 2018; 18(1):76–84. [https://doi.org/10.1016/S1473-3099\(17\)30691-6](https://doi.org/10.1016/S1473-3099(17)30691-6) PMID: 29198911
10. Kawkitinarong K, Suwanpimolkul G, Kateruttanakul P, Manosuthi W, Ubolyam S, Sophonphan J, et al. Real-Life Clinical Practice of Using the Xpert MTB/RIF Assay in Thailand. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America*. 2017; 64(suppl_2):S171–s8. <https://doi.org/10.1093/cid/cix151> PMID: 28475796
11. Nicol MP, Workman L, Isaacs W, Munro J, Black F, Eley B, et al. Accuracy of the Xpert MTB/RIF test for the diagnosis of pulmonary tuberculosis in children admitted to hospital in Cape Town, South Africa: a descriptive study. *The Lancet Infectious diseases*. 2011; 11(11):819–24. [https://doi.org/10.1016/S1473-3099\(11\)70167-0](https://doi.org/10.1016/S1473-3099(11)70167-0) PMID: 21764384
12. Scott L, David A, Noble L, Nduna M, Da Silva P, Black A, et al. Performance of the Abbott RealTime MTB and MTB RIF/INH Assays in a Setting of High Tuberculosis and HIV Coinfection in South Africa. *Journal of Clinical Microbiology*. 2017; 55(8):2491. <https://doi.org/10.1128/JCM.00289-17> PMID: 28592547
13. Steingart KR, Schiller I, Horne DJ, Pai M, Boehme CC, Dendukuri N. Xpert(R) MTB/RIF assay for pulmonary tuberculosis and rifampicin resistance in adults. *The Cochrane database of systematic reviews*. 2014(1):Cd009593. <https://doi.org/10.1002/14651858.CD009593.pub3> PMID: 24448973
14. Theron G, Peter J, van Zyl-Smit R, Mishra H, Streicher E, Murray S, et al. Evaluation of the Xpert MTB/RIF assay for the diagnosis of pulmonary tuberculosis in a high HIV prevalence setting. *Am J Respir Crit Care Med*. 2011; 184(1):132–40. <https://doi.org/10.1164/rccm.201101-0056OC> PMID: 21493734
15. Sohn H, Aero AD, Menzies D, Behr M, Schwartzman K, Alvarez GG, et al. Xpert MTB/RIF testing in a low tuberculosis incidence, high-resource setting: limitations in accuracy and clinical impact. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America*. 2014; 58(7):970–6. <https://doi.org/10.1093/cid/ciu022> PMID: 24429440
16. Theron G, Zijenah L, Chanda D, Clowes P, Rachow A, Lesosky M, et al. Feasibility, accuracy, and clinical effect of point-of-care Xpert MTB/RIF testing for tuberculosis in primary-care settings in Africa: a multicentre, randomised, controlled trial. *Lancet (London, England)*. 2014; 383(9915):424–35. [https://doi.org/10.1016/s0140-6736\(13\)62073-5](https://doi.org/10.1016/s0140-6736(13)62073-5)
17. Viet Nam Ministry of Health Guideline for application of Gene Xpert MTB/RIF (4921/Q-BYT). 2011. <https://thuvienphapluat.vn/van-ban/The-thao-Y-te/Quy-et-dinh-4921-QD-BYT-nam-2011-huong-dan-Quy-trinh-trien-khai-ky-thuat-Gene-145140.aspx>.
18. Balcells ME, Garcia P, Chanqueo L, Bahamondes L, Lasso M, Gallardo AM, et al. Rapid molecular detection of pulmonary tuberculosis in HIV-infected patients in Santiago, Chile. *The international journal of tuberculosis and lung disease*: the official journal of the International Union against Tuberculosis and Lung Disease. 2012; 16(10):1349–53. <https://doi.org/10.5588/ijtld.12.0156> PMID: 22863872
19. Carriquiry G, Otero L, González-Lagos E, Zamudio C, Sánchez E, Nabeta P, et al. A Diagnostic Accuracy Study of Xpert®MTB/RIF in HIV-Positive Patients with High Clinical Suspicion of Pulmonary

- Tuberculosis in Lima, Peru. *PloS one*. 2012; 7(9):e44626. <https://doi.org/10.1371/journal.pone.0044626> PMID: 22970271
20. Lawn SD, Brooks SV, Kranzer K, Nicol MP, Whitelaw A, Vogt M, et al. Screening for HIV-Associated Tuberculosis and Rifampicin Resistance before Antiretroviral Therapy Using the Xpert MTB/RIF Assay: A Prospective Study. *PLOS Medicine*. 2011; 8(7):e1001067. <https://doi.org/10.1371/journal.pmed.1001067> PMID: 21818180
 21. World Health Organization. Improving the diagnosis and treatment of smear-negative pulmonary and extrapulmonary tuberculosis among adults and adolescents: recommendations for HIV-prevalent and resource-constrained settings / Stop TB Department; Department of HIV/AIDS. 2007. <https://apps.who.int/iris/handle/10665/69463>.
 22. World Health Organization. Guidelines for treatment of tuberculosis, fourth edition. 2010. <https://www.who.int/tb/publications/2010/9789241547833/en/>.
 23. Nguyen Thi Ngoc Lan. Survey about drug-resistant tuberculosis in patients with HIV and non-HIV in Ho Chi Minh City in 1995–1997. *Journal of Practical Medicine*. 2000; 382:92–7.
 24. Hajian-Tilaki K. Sample size estimation in diagnostic test studies of biomedical informatics. *Journal of Biomedical Informatics*. 2014; 48:193–204. <https://doi.org/10.1016/j.jbi.2014.02.013>. <https://www.sciencedirect.com/science/article/pii/S1532046414000501>. PMID: 24582925
 25. World Health Organization. Tuberculosis laboratory biosafety manual. 2012. http://apps.who.int/iris/bitstream/10665/77949/1/978924150438_eng.pdf.
 26. Luetkemeyer AF, A ftACTG, Teams TTCS, Firnhaber C, A ftACTG, Teams TTCS, et al. Evaluation of Xpert MTB/RIF Versus AFB Smear and Culture to Identify Pulmonary Tuberculosis in Patients With Suspected Tuberculosis From Low and Higher Prevalence Settings. *Clinical Infectious Diseases*. 2016; 62(9):1081–8. <https://doi.org/10.1093/cid/ciw035> PMID: 26839383
 27. Helb D, Jones M, Story E, Boehme C, Wallace E, Ho K, et al. Rapid detection of *Mycobacterium tuberculosis* and rifampin resistance by use of on-demand, near-patient technology. *J Clin Microbiol*. 2010; 48(1):229–37. <https://doi.org/10.1128/JCM.01463-09> PMID: 19864480
 28. Lawn SD, Nicol MP. Xpert® MTB/RIF assay: development, evaluation and implementation of a new rapid molecular diagnostic for tuberculosis and rifampicin resistance. *Future Microbiol*. 2011; 6(9):1067–82. <https://doi.org/10.2217/fmb.11.84> PMID: 21958145
 29. Geleta DA, Megerssa YC, Gudeta AN, Akalu GT, Debele MT, Tulu KD. Xpert MTB/RIF assay for diagnosis of pulmonary tuberculosis in sputum specimens in remote health care facility. *BMC Microbiol*. 2015; 15:220-. <https://doi.org/10.1186/s12866-015-0566-6> PMID: 26483194
 30. Horne DJ, Kohli M, Zifodya JS, Schiller I, Dendukuri N, Tollefson D, et al. Xpert MTB/RIF and Xpert MTB/RIF Ultra for pulmonary tuberculosis and rifampicin resistance in adults. *Cochrane Database of Systematic Reviews*. 2019(6). <https://doi.org/10.1002/14651858.CD009593.pub4> PMID: 31173647
 31. Zong K, Luo C, Zhou H, Jiang Y, Li S. Xpert MTB/RIF assay for the diagnosis of rifampicin resistance in different regions: a meta-analysis. *BMC Microbiol*. 2019; 19(1):177-. <https://doi.org/10.1186/s12866-019-1516-5> PMID: 31382894
 32. Mathys V, van de Vyvere M, de Droogh E, Soetaert K, Groenen G. False-positive rifampicin resistance on Xpert(R) MTB/RIF caused by a silent mutation in the *rpoB* gene. *The international journal of tuberculosis and lung disease*. 2014; 18(10):1255–7. <https://doi.org/10.5588/ijtld.14.0297> PMID: 25216843
 33. Mokaddas E, Ahmad S, Eldeen HS, Al-Mutairi N. Discordance between Xpert MTB/RIF assay and Bactec MGIT 960 Culture System for detection of rifampin-resistant *Mycobacterium tuberculosis* isolates in a country with a low tuberculosis (TB) incidence. *Journal of clinical microbiology*. 2015; 53(4):1351–4. <https://doi.org/10.1128/JCM.03412-14> PMID: 25609730
 34. Yakrus MA, Driscoll J, Lentz AJ, Sikes D, Hartline D, Metchock B, et al. Concordance between molecular and phenotypic testing of *Mycobacterium tuberculosis* complex isolates for resistance to rifampin and isoniazid in the United States. *Journal of clinical microbiology*. 2014; 52(6):1932–7. <https://doi.org/10.1128/JCM.00417-14> PMID: 24648563
 35. Rigouts L, Gumusboga M, de Rijk WB, Nduwamahoro E, Uwizeye C, de Jong B, et al. Rifampin resistance missed in automated liquid culture system for *Mycobacterium tuberculosis* isolates with specific *rpoB* mutations. *Journal of clinical microbiology*. 2013; 51(8):2641–5. <https://doi.org/10.1128/JCM.02741-12> PMID: 23761146
 36. Somoskovi A, Deggim V, Ciardo D, Bloemberg GV. Diagnostic implications of inconsistent results obtained with the Xpert MTB/Rif assay in detection of *Mycobacterium tuberculosis* isolates with an *rpoB* mutation associated with low-level rifampin resistance. *J Clin Microbiol*. 2013; 51(9):3127–9. <https://doi.org/10.1128/JCM.01377-13> PMID: 23850949

37. Chakravorty S, Boehme C, Lee J. Tuberculosis Diagnostics in the New Millennium: Role in TB Identification and Control. *Tuberculosis research and treatment*. 2012; 2012:768603. <https://doi.org/10.1155/2012/768603> PMID: 23320162
38. Blakemore R, Story E, Helb D, Kop J, Banada P, Owens MR, et al. Evaluation of the analytical performance of the Xpert MTB/RIF assay. *J Clin Microbiol*. 2010; 48(7):2495–501. <https://doi.org/10.1128/JCM.00128-10> PMID: 20504986
39. Helb D, Jones M, Story E, Boehme C, Wallace E, Ho K, et al. Rapid detection of *Mycobacterium tuberculosis* and rifampin resistance by use of on-demand, near-patient technology. *J Clin Microbiol*. 2010; 48(1):229–37. <https://doi.org/10.1128/JCM.01463-09> PMID: 19864480
40. Zetola NM, Shin SS, Tumedi KA, Moeti K, Ncube R, Nicol M, et al. Mixed *Mycobacterium tuberculosis* complex infections and false-negative results for rifampin resistance by GeneXpert MTB/RIF are associated with poor clinical outcomes. *Journal of clinical microbiology*. 2014; 52(7):2422–9. <https://doi.org/10.1128/JCM.02489-13> PMID: 24789181