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Molecules in Cancer Immunotherapy

Lysine Acetylation/Deacetylation

Modification of Immune-Related

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As major post-translational modifications (PTMs), acetylation and deacetylation are significant factors in signal transmission and cellular metabolism, and are modulated by a dynamic process via two pivotal categories of enzymes, histone acetyltransferases (HATs) and histone deacetylases (HDACs). In previous studies, dysregulation of lysine acetylation and deacetylation has been reported to be associated with the genesis and development of malignancy. Scientists have recently explored acetylation/deacetylation patterns and prospective cancer therapy techniques, and the FDA has approved four HDAC inhibitors (HDACi) to be used in clinical treatment. In the present review, the most recent developments in the area of lysine acetylation/deacetylation alteration in cancer immunotherapy were investigated. Firstly, a brief explanation of the acetylation/ deacetylation process and relevant indispensable enzymes that participate therein is provided. Subsequently, a multitude of specific immune-related molecules involved in the lysine acetylation/deacetylation process are listed in the context of cancer, in addition to several therapeutic strategies associated with lysine acetylation/deacetylation modification in cancer immunotherapy. Finally, a number of prospective research fields related to cancer immunotherapy concepts are offered with detailed analysis. Overall, the present review may provide a reference for researchers in the relevant field of study, with the aim of being instructive and meaningful to further research as well as the selection of potential targets and effective measures for future cancer immunotherapy strategies.

Keywords: acetylation, deacetylation, HAT, HDAC, Cancer, immunotherapy



INTRODUCTION

Post-translational modifications (PTMs) are associated with a large number of cellular processes and regarded as indispensable in maintaining protein function via regulating the activity, stability and localization of proteins (1). As major PTMs, deacetylation and acetylation processes are significant factors in signaling and cellular metabolism (2). The real-life processes of histone acetylation and deacetylation are often regulated by histone deacetylases (HDACs) and histone acetyltransferases (HATs), which together participate in numerous dynamic processes and in a wide range of substrate properties such as protein-DNA interactions, transcriptional and metabolic activities, and cell cycle regulation (3). Notably, since a number of significant processes in biology require histone acetylation and deacetylation, such as diabetes, infectious diseases and cancer, the dysregulation thereof is involved in various diseases (4).

In recent decades, owing to the non-negligible effects of acetylation and deacetylation on tumor cell growth and progress, an increasing amount of scientists have investigated acetylation/deacetylation patterns and potential cancer treatment methods (5, 6). Emerging evidence has indicated that suppression of HDACs can potentially inhibit growth and facilitate the differentiation of tumor cells, thereby producing an anticancer effect in HDACs (7). Four compounds (Romidepsin, Belinostat, Panobinostat and Vorinostat) have already been ratified by the Food and Drug Administration (FDA) as HDAC inhibitors (HDACi) for clinical use, with numerous others currently undergoing clinical assessment (8, 9). In the clinical treatment of tumors, immune checkpoints (for instance, CTLA-4, TIM-3, LAG-3, and most importantly, PD-1/PD-L1) have been demonstrated to be effective targets in cancer immunotherapy due to the ability thereof to regulate immune responses (10). Using HDACi alone or in combination with other therapeutic agents such as immune checkpoint inhibitors (ICIs) has been revealed to be effective in cancer immunotherapy, and the significance of acetylation/deacetylation modulation in cancer has attracted much research attention (11).

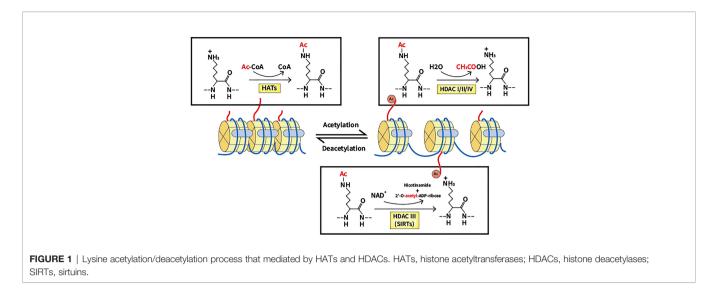
In the present study, relevant achievements in the field of lysine acetylation/deacetylation modification in cancer immunotherapy were analyzed. First, a brief explanation of the acetylation/deacetylation process and relevant indispensable enzymes that participate therein is provided. Subsequently, a multitude of specific immune-related molecules involved in the lysine acetylation/deacetylation process in the context of cancer are listed, in addition to several therapeutic strategies associated with lysine acetylation/deacetylation modification in cancer immunotherapy. Finally, numerous prospective research fields related with cancer immunotherapy concepts are offered with detailed analysis. The present study may serve as a reference for researchers in the relevant field of study, with the aim of being instructive and meaningful to further research as well as the selection of potential targets and effective measures for future cancer immunotherapy strategies.

LYSINE ACETYLATION/DEACETYLATION PROCESS

Acetylation was discovered in 1963 and has since been extensively investigated by scientists, being regarded as a crucial PTM in cells (12). By neutralizing the positive charge of acetylated lysine sites of histones, which regulates the mutual effect between targeted histones and the DNA skeleton with negative charge, acetylation modulates and relaxes the chromatin structure (13). In recent decades, a number of substrates have been reported as substrates of acetylation in addition to histone, reflecting the universality of acetylation in cellular processes (14). At the same time, a myriad of proteins with bromodomain like transcription factors can be used to identify lysine acetylation marks, which activates the next step of the transcriptional process (15). Deacetylation is the reverse process of acetylation, and is able to create a tighter connection between histone and DNA, causing a more condensed structure of chromatin that blocks the subsequent transcriptional process (16). Notably, lysine deacetylation appears to be a significant factor in chromatin reconstruction, further influencing gene expression and cellular processes (refer to Figure 1).

Two groups of essential enzymes are associated with lysine deacetylation and acetylation: histone deacetyltransferases (HDACs) and histone acetyltransferases (HATs). HATs are enzymes that help move the acetyl group from the acetyl coenzyme A to the lysine N-terminal, which removes the basic charge from the unmodified lysine residue, thereby accelerating the resulting active transcription process (17). According to location, HATs can be classified into two types. In the nucleus, Type A HATs are a group of enzymes that facilitate the transcription process by acting as transcriptional activators, which is the common functional feature thereof. The described enzymes can be divided into three separate families depending on the different substrates and functions: the Gcn5-related N-acetyltransferase (GNAT) family, the p300/ CREB-binding protein (CBP) family, and monocytic leukemia zinc finger protein, Ybf2/Sas3, Sas2 and Tip60 (MYST) family (18). The p300/CBP family consists of p300 and CBP, which mainly target H3 and H4 residues. The GNAT family consists of PCAF, Gcn5, Hat1, Hpa2 and Elp3, which mainly act on histones. The MYST family includes Tip60, MOF, Esa1, Sas2, Sas3, Hbo1 and MORF, which mainly target H4 residues (19). Acetylation of newly translated histones is associated with cytoplasmic Type B HATs, and such process is essential for the passage of newly synthesized histones across the nuclear membrane into newly replicated DNA. HatB3.1, Rtt109, HAT1, HAT2, and HAT4 are all types of B-type HATs (20). As well as histones, non-histone proteins such as p53 can also be regulated by HATs. In further studies, many other substrates of nonhistone proteins were verified to be acetylated (21, 22).

Contrastingly, HDACs are a family of 18 enzymes that exhibit opposite functions to HATs. HDACs remove the acetyl group from lysine residues, which function as corepressors of transcription and reduce the acetylation level of histones. Additionally, several non-histone proteins including c-MYC, p53, and STAT3 can also be deacetylated to regulate cellular processes (23, 24). Gene silencing is



caused by lysine deacetylation, which condenses chromatin and limits transcriptional activity (25). HDACs are classified into four types based on the similarity thereof to yeast proteins: Class I, II, III and IV. Class I, II and IV HDACs require the presence of Zn2⁺, while Class III HDACs (also known as sirtuins) are dependent on NAD⁺ (26). Located mainly in the nucleus, a total of four members of Class I HDACs exist: HDAC1, 2, 3, and 8, which are yeast homologs of Rpd3 (27, 28). Class II HDACs can be divided into two groups: Class IIa, which includes HDAC4, 5, 7, and 9, and Class IIb, which includes HDAC6 and 10. The two subclasses are closely related to Hda1 in yeast and can move back and forth between the cytoplasm and the nucleus, allowing Class II HDACs to regulate cytoplasmic substrates (29, 30). Homologous to Sir2 proteins in yeast, Class III HDACs can be categorized into seven elements, including sirtuin (SIRT) 1 to 7. SIRT1, 6, and 7 reside in the nucleus to regulate gene expression by deacetylating substrates (31). SIRT2 has been found to control the cell cycle in both cytoplasm and the nucleus (32, 33). Regarding SIRT3, 4, and 5, said enzymes have been reported to reside in the mitochondria to regulate the activity of oxidative stress (34). A significant characteristic of SIRTs is the ability thereof to not only deacetylate lysine residues but also act as mono-ADP-ribosyltransferases. As such, the enzymes can modulate chromatin, repair DNA and undertake other biological processes (35). In epigenetics, the single member of Class IV HDACs, HDAC11, has emerged as a particular focus of research and is located in the nucleus. Despite preliminary research showing that HDAC11 can negatively modulate IL-10, additional research is required for verification (36).

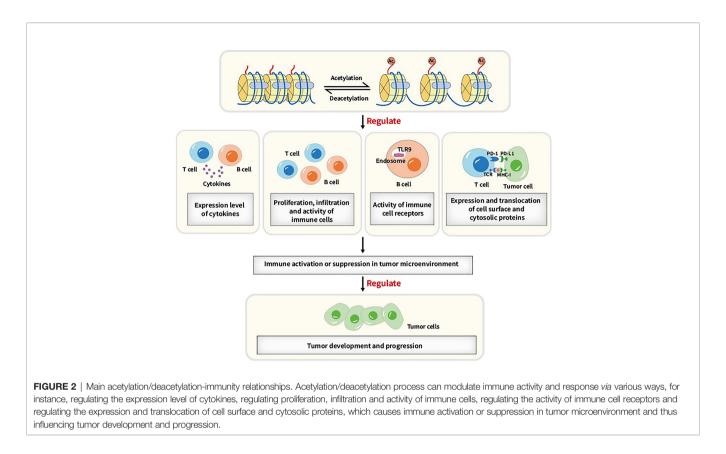
CANCER IMMUNOTHERAPEUTIC MOLECULES AND THE ACETYLATION/ DEACETYLATION MODIFICATION THEREOF

A number of biological processes including DNA damage repair, immunological response and cell cycle arrest require lysine acetylation/deacetylation. Abnormalities in the acetylation/ deacetylation process, such as the dysregulation of the regulating enzymes, have been associated with cancer (37). In previous studies, aberrant expression levels and mutations or translocations of lysine acetylation/deacetylation regulators have been demonstrated in a variety of cancers (38). Acetylation/deacetylation process can modulate immune activity and response *via* various ways, thus influencing tumor development and progression (refer to **Figure 2**). Several specific immune-related targets of acetylation/deacetylation modification are listed as follows (refer to **Table 1**).

PD-L1

The binding of programmed death-1 ligand (PD-L1, also known as B7-H1 or CD274) to programmed death-1 (PD-1) produces a large immune effect in pathological and physiological occurring situations (57). Oncogenic PD-L1 has been found to be overexpressed in many solid tumors, such as melanoma, nonsmall cell lung cancer, and breast cancer (58). PD-L1 is stimulated by PD-1 and signals cancer cells to maintain survival, which leads to cancer cells becoming resistant to Fas signaling pathway-induced apoptosis (59). Further, in the absence of PD-1 signaling, PD-L1 blocks the cytotoxic activity of malignant tumor cells in response to interferons (Type I and II) and CTL cells. Thus, PD-L1 is a significant factor in regulating host immune responses in the context of cancer (60). As one of the most compelling immune checkpoints, PD-L1 has become a target for clinical use, and several therapeutic agents (like PD-L1 inhibitors) have been approved for cancer immunotherapy, such as Durvalumab and Atezolizumab (61). The increased expression level of PD-L1 in immune cells has been reported to be linked with a higher response rate to ICIs and better prognosis in several cancers, including urothelial carcinoma (62), Hence, PD-L1 could be applied in effective cancer immunotherapy strategies.

A large number of PTMs types have been shown to regulate PD-L1, namely ubiquitination, phosphorylation, glycosylation, and acetylation/deacetylation (63). PD-L1 can be acetylated through p300 acetyltransferase at Lys263 in the cytoplasmic region and acetylated PD-L1 is restrained to be translocated



into the nucleus. HDAC2 was also found to be capable of interacting with PD-L1 and inhibiting PD-L1 acetylation mediated by p300, which could increase the nuclear PD-L1 level and facilitate the immune surveillance evasion of tumor cells (39). Such findings provide another theoretical foundation for regulating PD-L1 by acetylation or deacetylation in cancer immunotherapy. Suppressing the shuttle of PD-L1 from cytoplasm to the nucleus by activating p300 or inhibiting the synergy of HDAC2 with PD-1 blockade may recover host immune responses. Such recovery is achieved by facilitating the infiltration of CD8⁺ T cells and downregulating the expression level of TNF- α , which may elevate the immunotherapeutic efficacy for cancer. According to another previous study, the expression of PD-L1 is also relevant to HAT1 expression in pancreatic cancer. HAT1 can acetylate H4K5 and H4K12 residues, further promoting the combination of transcriptional factor BRD4 (bromodomain-containing 4) and the PD-L1 promoter to facilitate PD-L1 transcription. Knockdown of HAT1 can upregulate tumor infiltration of immune effectors such as $\mathrm{CD45^+CD4^+}$ and $\mathrm{CD45^+CD8^+}\ \mathrm{T}$ cells, suppress the infiltration of CD11b⁺Gr1⁺ myeloid cells in tumors and decrease the expression level of PD-L1, which may inhibit tumor proliferation and promote the efficacy of the immune checkpoint blockade (40). As such, targeting HAT1 alone or in combination with ICIs could be a novel strategy in cancer immunotherapy. Moreover, Darvin P et al. observed a consistent overexpression of HDAC1 and p300/CBP enzymes in breast cancer cells, which exhibited a close relationship with

upregulated PD-L1 level and increased breast cancer aggressiveness (41). Another study also reported that HDAC6 can recruit and activate the signal transducer and activator of transcription 3 (STAT3), which increases the expression of PD-L1 through STAT3-mediated activation of the PD-L1 promoter in melanomas. By modulating various mechanisms involved in immune recognition and tumor survival, such as downregulating PD-L1, PD-L2 and B7-H4, the application of HDAC6 inhibitors exhibited an anti-tumor effect, which might provide theoretical basis to apply HDAC6 inhibitors in cancer immunotherapy (42). In cisplatin-resistant non-small cell lung cancer cells, a decreased constitutive photomorphogenesis protein 1 (COP1) level caused the increased accumulation of c-Jun, which suppressed HDAC3 expression and facilitated acetylation of the PD-L1 promoter at the H3 site, thereby upregulating the expression of PD-L1. Increased PD-L1 was found to be crucial for inhibiting the proliferation of CD3⁺ T cells, indicating that the immune checkpoint blockade and HDAC3 activation might be effective therapeutic methods for reversing T cell-related immune suppression in drug-resistant tumors (43). The aforementioned studies reflect the indispensable role of HDACs in PD-L1 regulation, verifying the rationality of the extensive application of HDACi in clinical practice.

However, in the acetylation/deacetylation modification of PD-L1, there remain certain aspects that require further clarification. Although the aforementioned studies provide a macro-level explanation for the link between enzymes and PD-

TABLE 1 | Several specific immune-related targets of acetylation/deacetylation modification.

| Molecules | Enzymes | Modification Sites | Cancer Types | Functions | References |
|-----------|--------------------------------------|-----------------------|-------------------------------|--|--------------|
| PD-L1 | p300 | Lys263 | - | Acetylates PD-L1 and inhibits its translocation into the nucleus. | (39) |
| | HDAC2 | - | - | Inhibits PD-L1 acetylation mediated by p300, which increases the nuclear PD-L1 level. | (39) |
| | HAT1 | H4K5, H4K12 | Pancreatic cancer | Increases PD-L1 expression through BRD4. | (40) |
| | HDAC1, p300, CBP | - | Breast cancer | Modulates chromatin structure and enables transcription factor binding, which increases PD-L1 expression. | (41) |
| | HDAC6 | _ | Melanoma | Recruits and activates STAT3, which increases PD-L1 expression. | (42) |
| | HDAC3 | H3 | Non-small cell lung cancer | Decreased COP1 can cause c-Jun accumulation and suppress HDAC3 expression, which increases PD-L1 expression. | (43) |
| PD-1 | p300, CBP, Tip60 | K120, K164 | Lung cancer | Facilitates PD-1 transcription and expression through acetylated p53. | (44) |
| CTLA-4 | p300, CBP HDACs | – H3K27 | Melanoma | Facilitates the transcriptional activation of CTLA-4 licensed by IFN-γ signaling. Regulates CTLA-4 expression. | (45) (46) |
| TIM-3/ | HDACS HDAC6 | - - | – Melanoma | HDAC6 suppression can decrease the expression level of TIM-3 and LAG-3. | (40) |
| LAG-3 | HDACO | - | IVIEIALIOLITIA | HDAGO Suppression can decrease the expression level of hivi-3 and LAG-3. | (47) |
| B7-H3 | MSL complex (including MOF) | H4K16 | Nasopharyngeal carcinoma | PBK phosphorylates MSL1, promotes enrichment of MSL complex on B7-H3 promoter, leading to increased H4K16 acetylation and the activation of B7-H3 transcription. | (48) |
| | SIRT1 | - | Colorectal carcinoma | MiR-34a can inhibit SIRT1 and consequently cause the acetylation of NF- κ B subunit p65 (a-p65), thereby facilitating its binding with B7-H3 promoter and increasing the transcription and expression of B7-H3. | (49) |
| OX40 | SIRT1, SIRT7 | - | - | OX40 can induce the expression of BATF and BATF3, which bind to Foxp3 promoter and contribute to a closed chromatin configuration to suppress Foxp3 expression through the recruitment of SIRT1 and 7. | (50) |
| | p300, CBP, PCAF, SRC- 1 | H4 | - | NF- κ B can recruit p300, CBP, PCAF and SRC-1 to acetylate OX40 promoter on histone H4 substrate, upregulating the expression of OX40. | (51) |
| | HDAC1 | - | - | NF-κB p50 can recruit HDAC1 to exert its deacetylation effect, which cause the down- regulation of OX40. | (51) |
| OX40L | HDAC1, HDAC3 | - | Ovarian cancer | HDAC1 and HDAC3 can bind at OX40L promoter to downregulate OX40L expression. | (52) |
| 4-1BBL | HDAC1, HDAC3 | - | Ovarian cancer | HDAC1 and HDAC3 can bind at 4-1BBL promoter to downregulate 4-1BBL expression. | (52) |
| | - | - | Cutaneous T- cell lymphoma | GATA6 is epigenetically overexpressed in CTCL cells on account of DNA histone acetylation, which combines with the promoter region of 4-1BBL and thus facilitating the expression of 4-1BBL. | (53) |
| CD70 | HDAC1 | - | - | RFX1 can recruit HDAC1 to the promoter region of CD70 where it deacetylates histone substrate, causing a more condense chromatin structure and decrease expression level of CD70. | (54) |
| | HDAC2, HDAC7 | H3 | - | TSA can suppress the activity of HDAC2 and HDAC7, which causes the increased H3 acetylation at CD70 promoter and thus facilitating the expression of CD70. | (55) |
| | HDAC5 | - | - | Overexpression of HDAC5 is related with upregulated CD70, inducing growth suppression and apoptosis of tumor cells. | (56) |

HATs, histone acetyltransferases; HDACs, histone deacetylases; SIRTs, sirtuins; CBP, CREB-binding protein; CTLA-4, cytotoxic T-lymphocyte-associated antigen 4; PD-L1, programmed death-1 ligand; PD-1, programmed death-1; pSTAT1, phosphorylated STAT1; BRD4, bromodomain-containing 4; OX40, tumor necrosis factor receptor superfamily member 4; OX40L, OX40 ligand; BATF, basic leucine zipper transcription factor ATF-like; 4-1BBL, 4-1BB ligand; RFX1, regulatory factor X 1; STAT3, signal transducer and activator of transcription 3; COP1, constitutive photomorphogenesis protein 1; tim, TIM-3, T-cell immunoglobulin and mucin domain 3; LAG-3, lymphocyte-activation gene 3.

1/PD-L1 expression, the specific signaling pathway and regulatory mechanism of PD-1/PD-L1 modification by HATs/ HDACs are still unclear. Further research concerning the association between HATs/HDACs and PD-1/PD-L1, which can clarify the upstream/downstream molecules in the signaling pathway, may provide new targets and innovative ideas for cancer immunotherapy. Further, most of the regulating pathway and molecular mechanisms of PD-L1 modification by acetylation/deacetylation have been identified at the cellular or animal model stage, and no findings have been made using clinical trials. To expand cancer immunotherapy strategies, further clinical studies that confirm the effect of PD-L1 acetylation/deacetylation modification on tumor development and progression are crucial.

PD-1

PD-1 has been validated to maintain immune homeostasis through PD-1/PD-L1 interactions and is also involved in chronic infections and malignancies. Wherry et al. reported that T cells showed a dysfunctional state and lost the immune function thereof in a mouse model infected with lymphocytic choriomeningitis virus (LCMV). Notably, the expression level of

PD-1 was found to be upregulated, a result that can be considered as one of the characteristics of dysfunctional T cells (64). In human malignancies, T cells with upregulated PD-1 are corelated with tumor reactivity, which is linked with PD-1 expression driven by antigens in both tumors and peripheral blood (65). In hepatocellular carcinoma cells, researchers have demonstrated that blocking the interaction between PD-1 and PD-L1 can suppress the immunosuppressive activity of B cells with high PD-1 level (66). Through reactivating T cell function and restoring host immune responses, several effective antibodies against PD-1 that block PD-1/PD-L1 interaction have been developed and applied in cancer immunotherapy (58).

One study showed that PD-1 can be modified by acetylation. The transcription and expression level of PD-1 can be upregulated by acetylated p53, which is modified by p300, CBP and Tip60 acetyltransferases at K120/K164 sites. After acetylation by the aforementioned HATs, the acetylated nonhistone protein p53 can further recruit the acetyltransferase cofactors to interact with the PD-1 promoter, which facilitates the acetylation level of local chromatin and contributes to a looser chromatin structure, thereby increasing the transcription and expression of PD-1. Upregulated intrinsic PD-1 in lung cancer cells was also found to significantly inhibit tumor growth. Using HDACi upon p53 activation might be considered as a potential strategy for cancer therapy via upregulation of cancer cell-intrinsic PD-1 (44). However, the aforementioned PD-1 regulatory signaling pathway in tumor suppression is regulated in an immunity-independent manner (through inhibiting the AKT/mTOR pathway). Whether tumor growth is inhibited by modulating immunological functions, such as regulating T cell activation and exhaustion, needs further experimental confirmation. Additionally, there is a scarcity of relevant studies on the acetylation/deacetylation modification of PD-1 compared with studies on PD-L1. Further studies on the signaling pathways and mechanisms of acetylation/ deacetylation regulation of PD-1 may provide new therapeutic strategies and acting targets for future cancer immunotherapy.

CTLA-4

Cytotoxic T lymphocyte-associated antigen 4 (CTLA-4, CD152 for short) belongs to the CD28 immunoglobulin subfamily and is generally considered to function as a suppressor receptor. CTLA-4 is involved in a number of suppressive activities such as inhibition of cell cycle progression, T cell proliferation, cytokine expression, and other activities, mainly through the expression of activated effector T cells (67). Further, CTLA-4 directly regulates the homeostasis of Tregs. As a recognized checkpoint in the immune system, CTLA-4 has been extensively investigated as an immunotherapeutic target for a variety of malignancies, including melanoma (68), prostate cancer (69), and hepatocellular cancer (70). Targeting the CTLA-4 signaling pathway by antibodies or fusion proteins has attracted significant attention in the field of cancer therapy. In mouse cancer models, researchers identified that anti-CTLA-4 treatment strategies could cause tumor regression and lead to

resistance to reinoculation of malignant cells (71). Based on the efficacy of blocking CTLA-4 in mouse models, several drug antibodies against CTLA-4 have been used clinically and tested in trials for advanced cancers (72).

The transcription and expression level of CTLA-4 are closely associated with the acetylation process. In prior research, histone acetylation mediated by p300/CBP was found to be critical for the transcriptional activation of CTLA-4 licensed by interferongamma (IFN- γ) signaling in melanoma cells. IFN- γ -regulated STAT1 can be phosphorylated via JAK1/2 dependent pathway and bind to the specific binding site of the CTLA-4 promoter, facilitating the acetylation process regulated by p300/CBP and increasing the expression of CTLA-4 (45). Via inhibiting T cell cytotoxicity and facilitating melanomagenesis and progression in melanocytes and melanoma cells, upregulated CTLA-4 induced by IFN-y was found to contribute to an immunosuppressive microenvironment. Therefore, regulating CTLA-4 acetylation modification in combination with an anti-CTLA-4 strategy might relieve T cell deactivation and produce a curative effect in cancer immunotherapy. A further study demonstrated that low-dose HDAC inhibitor can modulate CTLA-4 expression to facilitate the natural production of Foxp3⁺ Treg cells and restore the suppressive function of Treg cells via the regulation of histone H3K27 acetylation in immune thrombocytopenia (ITP). Such findings indirectly reflect that HDACs are also related to CTLA-4 regulation, and that targeting HDACs to modulate the immune response may have potential application value in cancer immunotherapy (46). Despite the aforementioned findings, the specific types of HDAC that participate in such process need to be identified in further studies. In addition to studies on the acetylation/deacetylation of PD-1/PD-L1, there is also a scarcity of studies on the detailed mechanism and specific signaling pathways of CTLA-4 expression regulated by HATs or HDACs. Further studies will be meaningful for cancer immunotherapy, and carefully designed experiments are needed in the future.

TIM-3/LAG-3

As members of the immune checkpoint family, T-cell immunoglobulin and mucin domain 3 (TIM-3) and lymphocyte activating gene-3 (LAG-3) have negative immunomodulatory functions in tumors. In terms of TIM-3 and LAG-3, the mechanisms of action and potential as therapeutic targets have been validated to a certain extent (73). In particular, TIM-3 expression could be found on functionally exhausted T cells in several mouse tumor models and in a large number of cancer cells such as melanoma and non-small cell lung cancer (74, 75). The levels of TIM-3 have been associated with advanced tumor nodal metastasis (TNM) and poor prognosis and can therefore be considered as a negative prognostic biomarker for different types of cancer (76). Regarding LAG-3, which is upregulated in activated T cells and NK cells, said gene can cause T cell dysfunction or apoptosis. Such result may be a protective mechanism adopted by tumor cells to escape elimination by the immune system (77).

LAG-3 has been reported to be co-expressed with TIM-3 and, most typically, PD-1. Targeting LAG-3/TIM-3 alone or through a combination of drugs acting on different immune checkpoints may be effective (78).

A previous study reported that suppressing HDAC6 can decrease the expression level of LAG-3, TIM-3 and PD-1, relieve the suppressive function of Treg cells and upregulate TIL cytolytic function in melanoma patients, potentially indicating that HDAC6 is involved in the regulatory pathways of LAG-3 and TIM-3. A further indication is that blocking HDAC6 activity may be effective in alleviating T cell suppression. Selective inhibitors of HDAC6 enhance T cell immune properties in melanoma patients, providing a theoretical basis for further studies to evaluate the potential clinical efficacy thereof (47). However, there are limited researches on the acetylation/deacetylation modulation of TIM-3 and LAG-3 compared with researches on PD-L1. Further studies on whether TIM-3 and LAG-3 are regulated by other acetylation/deacetylation molecules and the specific signaling pathways may provide novel targets and therapeutic strategies for future cancer immunotherapy.

B7-H3

As a recently discovered member of the B7 family, B7-H3 (also known as CD276) exhibits a dual role in immune cell responses and has been found to be upregulated in a variety of cancer types, such as colorectal and breast cancers (79, 80). B7-H3 has been reported to have a significant co-stimulatory molecular function in various immune cell responses, such as IFN-y production and T cell activation (81). In a previous study, the expression level of the costimulatory molecule B7-H3 was found to be upregulated in patients with pancreatic cancer, being associated with improved treatment outcome and better postoperative prognosis (82). In patients with colon adenocarcinoma, B7-H3 also exhibited antitumor effects and could be considered as a potential therapeutic molecule in cancer therapy (83). At the same time, B7-H3 also has the ability to inhibit T cell proliferation. Upregulation of B7-H3 is positively associated with poor clinical outcomes in patients with oral squamous cell carcinoma (OSCC). Notably, the expression of B7-H3 promotes tumor growth, while the blockade of B7-H3 inhibits tumor progression (84).

A recent study elucidated a potential upregulation mechanism of B7-H3 in nasopharyngeal carcinoma (NPC). PDZ-binding kinase (PBK) phosphorylates MSL1 and facilitates the enrichment of the MSL complex (consisting of MSL1 and MSL2, MSL3, and MOF) on the B7-H3 promoter region, thereby contributing to the acetylation thereof at H4K16 residue and increasing the transcription of B7-H3. High expression of B7-H3 is positively correlated with CD8-naive, Th2 and nTreg cells but is negatively correlated with Th1, Tfh and CD4 T cells. The cytotoxic T-cell function that attacks NPC cells is also suppressed by increased B7-H3 (48). Thus, targeting both B7-H3 and the acetylation signaling pathway may provide therapeutic benefits for NPC patients. Moreover, another study reported a new signaling pathway through which miR-34a upregulates B7-H3, providing a theoretical basis for targeting B7-H3 in cancer immunotherapy. B7-H3 can be upregulated through the miR-34a/SIRT1/NF-KB/B7-H3 axis in colorectal carcinoma. MiR-34a inhibits SIRT1, which causes the acetylation of the NF-KB subunit p65 (a-p65), thereby promoting the binding thereof to the B7-H3 promoter and increasing the transcription and expression of B7-H3. Upregulated B7-H3 promotes an abundant release of proinflammatory cytokines, including TNF-a, IFN-y, IL-2, IL-6 and IL-10, resulting in the inhibition of the antitumor immune response and the subsequent promotion of tumor growth (49). Therefore, targeting B7-H3 related molecules in acetylation/ deacetylation signaling pathways may affect the expression level of B7-H3, which is a significant factor in regulating the immune response in the tumor microenvironment. However, due to dual effect of B7-H4 in the cellular immune response, further research is needed to explore whether modulating the acetylation/deacetylation process of B7-H4 can produce a specific therapeutic effect in cancer immunotherapy.

OX40/OX40L

Tumor necrosis factor receptor superfamily member 4 (OX40, also known as CD134) and the OX40 ligand (OX40L, also known as CD134L or CD252) thereof are indispensable immune checkpoints that have been reported to be expressed on various cell types, such as activated T cells, B cells, and endothelial cells (85). OX40/OX40L interaction promotes T cell proliferation, upregulates cytokine production, and suppresses the immunosuppressive effects of Treg cells, thereby promoting immune responses (86). In a number of studies, OX40 was found to be expressed on tumor infiltrating lymphocytes (TILs) in many types of cancers, namely colorectal, gastric and ovarian, thereby regulating the development and progression of cancer (87-89). The combination of OX40 agonists with PD-1 blockade has been revealed to be effective in cancer immunotherapy, being able to inhibit the growth of ovarian tumor cells (89). The combination of anti-OX40 therapy with cytokines or radiotherapy can also exert antitumor effects in cancer immunotherapy (90, 91).

OX40 and OX40L have been found to be involved in acetylation/deacetylation regulatory signaling pathways. In a previous study, OX40 could induce the expression of basic leucine zipper transcription factor ATF-like (BATF) and BATF3, both of which bind to the Foxp3 promoter and contribute to a closed chromatin configuration to suppress Foxp3 expression through the recruitment of SIRT1 and 7. In this way, the induction of iTregs was suppressed and immune suppression was relieved (50). Moreover, NF-KB can recruit p300, CBP, PCAF and SRC-1 to acetylate the OX40 promoter on the histone H4 substrate, upregulating the expression of OX40. NF-KB p50 can also recruit HDAC1 to exert a deacetylation effect, which causes the down-regulation of OX40. The differentiation of Tregs could be attributed to the chromatin structure of the OX40 promoter and the OX40 level. Hence, targeting the acetylation/deacetylation process that

regulates OX40 expression might affect the Tregs level and the host immune response (51). Another study demonstrated that HDAC1 and HDAC3 can bind at the OX40L promoter to downregulate OX40L expression in chemoresistant ovarian cancer cells, which could potentially induce the immunosuppression of cancer cells to escape from immune responses (52). Therefore, targeting OX40/OX40L or the upstream/downstream molecules thereof in acetylation/ deacetylation signaling pathways may provide more possibilities and better treatment efficacy for cancer immunotherapy, especially for chemoresistant tumors.

4-1BBL

As a member of the superfamily tumor necrosis factor (TNF), the 4-1BB ligand (known as CD137L) is expressed on activated B cells, T cells, macrophages and dendritic cells (92). By interacting with the high affinity receptor 4-1BB (CD137), 4-1BBL acts as an immunostimulant molecule to supply costimulatory messages to CD4⁺ and CD8⁺ T cells through activating the NF- κ B and c-Jun signaling pathways, which modulates immune response and activity (93). Combined application of vaccine that contains RM-1 cells which expressed 4-1BBL with CTLA-4 blockade can induce the regression of prostate cancer cells, showing the treatment potency of 4-1BBL in cancer immunotherapy (69).

4-1BBL can be modulated by the acetylation/deacetylation process and is related to cancer. In a previous study, HDAC1 and HDAC3 were found to bind at the 4-1BBL promoter to downregulate 4-1BBL expression in chemoresistant ovarian cancer cells, resulting in immune escape and suppression (52). However, the expression level of 4-1BBL has diverse effects on tumor progression in different types of cancers. GATA6 has been reported to be epigenetically overexpressed in cutaneous T-cell lymphoma (CTCL) cells owing to DNA histone acetylation, which combines with the promoter region of 4-1BBL, thereby facilitating the expression of 4-1BBL and promoting tumor progression (53). Blocking 4-1BBL may provide a new immunotherapeutic strategy for the treatment of certain malignancies, despite weakening the antitumor immune response of NK cells and CD8⁺ T cells. Hence, the expression level of 4-1BBL in different types of cancers and the influence thereof on tumor progression need to be further clarified through experiments, so as to provide a theoretical basis for cancer immunotherapeutic strategies. In cancer immunotherapy, 4-1BBL has been utilized according to the immunological characteristics thereof. To date, whole-cell vaccines with 4-1BBL cDNA modified tumor cells have been reported as the most common immunotherapeutic application of 4-1BBL. Researchers have found that the transfer of 4-1BBL cDNA into a mouse squamous cell carcinoma effectively activated primary anti-tumor immune responses of CD8⁺ T cells (94). The combined application of acetylation/deacetylation modulation and tumor cell vaccines may provide a new direction for cancer immunotherapy and needs to be validated by further research.

CD70

CD70 (also referred to as TNFSF7) is a Type II transmembrane glycoprotein and belongs to the TNF superfamily (95). The

interaction between CD70 and the receptor CD27 has been demonstrated to facilitate cytotoxic T cell responses and induce the cytokine production of CD4⁺ and CD8⁺ T cells, thereby regulating the cellular immune response (96). Upregulated CD70 expression has been observed in various types of malignancies, being linked with poor clinical outcomes and prognosis in certain cancers such as breast cancer and B cell lymphoma (97, 98). The aforementioned results may be explained by immune inhibition and escape resulting from an increased amount of iTregs, upregulated lymphocytes apoptosis and induced T cell exhaustion by CD70/ CD27 interaction (99, 100).

In systemic lupus erythematosus (SLE), regulatory factor X 1 (RFX1) was found to be able to recruit HDAC1 to the promoter region of CD70 and deacetylate the histone substrate, resulting in a more condense chromatin structure and a decrease in the expression level of CD70. At the same time, there was a reduction in T cell activation and IgG produced by B cells (54). Moreover, TSA (a histone deacetylase inhibitor) can suppress the activity of HDAC2 and HDAC7, which leads to increased H3 acetylation at the CD70 promoter, and thus, facilitates the expression of CD70 (55). The potential rationality of using HDACi in cancer immunotherapy was also revealed, in addition to a potential direction for further clinical trials. Another study showed that overexpression of HDAC5 is related to upregulated CD70, which induces the growth suppression and apoptosis of tumor cells, indicating that targeting HDAC5 may regulate the immune response and affect tumor progression in cancer immunotherapy (56). However, most of the messages concerning CD70 acetylation/deacetylation modulation have been reported in the context of SLE. Further studies on the acetylation/deacetylation of CD70 in various tumors will be beneficial in establishing a solid theoretical foundation for cancer immunotherapy.

COMBINED APPLICATION OF HDACI AND OTHER IMMUNOTHERAPEUTIC STRATEGIES

In recent decades, many scholars have found that epigenetic abnormalities, especially dysregulated acetylation or deacetylation, are significant features of various cancers (17). As biological inhibitors that target HDACs that are critical in deacetylation modification, HDACi are used as epigenetic regulatory agents in cancer immunotherapy. By interfering with the activity of HDAC, histone acetylation is indirectly induced to regulate gene expression and modulate host antitumor immune responses. Several studies have shown that HDACi can re-express inactivate regulatory genes in cancer cells and reverse malignant phenotypes (101). By decreasing the levels of FoxP3 and Treg, increasing the cytotoxicity of NK and CD8⁺T cells, and increasing the production of proinflammatory cytokines, HDAC inhibitors are conducive to reducing tumor formation and boosting an efficient immune response against cancer cells (102, 103). At the same time, the normal function of the immune system may be further influenced by HDAC

inhibition, which can affect the expression of immune checkpoints and associated ligands. In an SM1 murine melanoma model, the HDAC6 inhibitor greatly reduced PD-L1 and PD-L2 overexpression, which increased the overall survival rate (104). As revealed in another study, HDAC6 inhibition could suppress the T cell expression levels of PD-1, TIM-3 and LAG-3, relieving T cell suppression in melanoma patients (47). The combined application of HDACi and other immunotherapeutic strategies have recently emerged as significant areas of focus in the field of tumor therapy. Several synergistic immunotherapeutic strategies are listed as follows.

COMBINED APPLICATION OF HDACI AND ICIS

Immune checkpoints can effectively suppress immune responses or enable cancers to avoid damage from the entire immune system by activating negative regulatory pathways. Two such immune checkpoints, PD-1/PD-L1 and CTLA-4, have so far received the most attention in cancer immunotherapy. ICIs have made breakthrough progress in many cancer therapeutic fields, bringing new treatment hope to patients. Despite such progress, some patients who use ICIs exhibit no response or even develop drug resistance (105). Studies have shown that HDACi can significantly improve the immune response following the application of ICIs, and can enhance immunity to the effects of treatment. Acetylase inhibitors induce the infiltration of effector T cells, which secrete a variety of immune-related cytokines along with natural killer cells to form a critical line of defense against cancer in the body. HDACi in combination with ICIs have been shown to significantly increase the level of natural killer cells in the tumor microenvironment, resulting in a gradual decrease in tumor cell size (106). Thus, through combined application, HDACi can provide a double guarantee for the efficacy of ICIs by regulating the positive and negative mechanisms of immune response.

Many pre-clinical studies have reported the potential of combining HDACi with ICIs in cancer immunotherapy (refer to Table 2). For instance, HDACi in combination with anti-PD-1 or anti-CTLA-4 antibodies were found to promote the production of granase B in activated CD8⁺ T responder cells, significantly improving the infiltration and function of innate immune cells and enhancing adaptive immune response. Using a hepatocellular carcinoma preclinical model, selective HDAC8 inhibition was found to exert effective responses to anti-PD-L1 treatment by reactivating the production of T cell-trafficking chemokines (107). CG-745, a member of HDACi, has been validated to modulate the immune microenvironment and enhance the therapeutic effect of anti-PD-1 ICIs, which indicates the effectiveness of the combined application of HDACi and ICIs (108). In previous studies, the expression level of PD-L1 was increased by HDAC inhibitors, increasing the susceptibility of the host immune system to anti-PD-L1 and anti-PD-1 therapy, and promoting therapeutic results in patients (109). Combining Nexturastat (HDAC6 inhibitor) with an

anti-PD-1 agent has been reported to suppress tumor growth, decrease the level of pro-tumorigenic M2 macrophages and upregulate effector T cells in melanoma (104). Moreover, the Class I HDAC inhibitor Etinostat facilitated the positive antitumor effect of PD-1 inhibition in syngeneic mouse models of lung and renal cell carcinoma, which could be attributed to the inhibition of the immunosuppressive functions of myeloid-derived suppressor cells (MDSCs) (110). In a recent study, HDACi Romidepsin was found to upregulate PD-L1 expression and the combined application of Romidepsin and anti-PD-1 antibody could enhance the antitumor therapeutic effect and partially reverse the influence of Romidepsin on CD4⁺ and CD8⁺ T cells in colon cancer (111). Further, the combined application of HDACi and antibodies against CTLA-4 could facilitate CD4⁺T cell infiltration and exert a synergistic effect on cancer treatment by intensifying anticancer immune responses (106).

To further investigate the efficacy of the combined application of HDACi and ICIs, researchers have recently conducted several clinical trials and made progress to a certain degree (refer to Table 2). For instance, in metastatic uveal melanoma, the combined application of HDACi Entinostat and PD-1 inhibitor Pembrolizumab has been validated to exert durable responses of tumor regression and prolonged survival in a phase 2 clinical trial, especially for patients with low tumor burden and BAP1 wildtype tumors (124). Further, in a phase 1/1b study, synergizing Vorinostat with Pembrolizumab was well tolerated and exhibited a preliminary anti-tumor effect in advanced/ metastatic non-small cell lung cancer (112). The combined application of Tamoxifen, Vorinostat and Pembrolizumab in a phase 2 trial exhibited clinical benefits in terms of the upregulation of exhausted CD8⁺ T cells and the depletion of Tregs, which was linked with better clinical response and prolonged PFS in ER-positive breast cancer patients. Said combination was also well-tolerated without excess toxicity (114). However, in another phase 2 trial involving the combined application of Pembrolizumab and Vorinostat, significant efficacy was reported with a relatively higher grade of toxicity compared with using anti-PD-1 monoclonal antibodies alone in recurrent metastatic head and neck squamous cell carcinomas and salivary gland cancer (113). As such, immunotherapy involving the coadministration of ICIs and HDACi may be essential in prospective cancer treatment. Such immunotherapy may be particularly beneficial for cancer patients who develop drug resistance to immune checkpoint inhibitors. With the in-depth research on the role of HDACi and ICIs in anti-tumor treatment, the synergistic application of HDACi and ICIs may emerge in the broader field of cancer research and eventually in clinical cancer immunotherapy.

COMBINED APPLICATION OF HDACI AND CANCER VACCINE

Cancer vaccines can be applied in patients with malignant tumors to overcome the immune suppression caused by tumors and activate a

TABLE 2 | Combined application of HDACi and ICIs in pre-clinical studies and clinical trials.

| HADCi | Combined ICIs | Phases | NCT Numbers | Cancer Types | Effects | References |
|--------------|--------------------------|----------------------------|----------------|---|--|------------|
| PCI-34051 | PD-L1 antibody | Pre- clinical | - | Hepatocellular carcinoma | Reactivating T cell-trafficking chemokines production. | (107) |
| CG-745 | PD-1 antibody | Pre- clinical | - | - | Modulating the immune microenvironment and enhancing the therapeutic effect of anti-PD-1 ICIs | (108) |
| Romidepsin | PD-1 antibody | Pre- clinical | - | Lung adenocarcinoma | Increasing the susceptibility of the host immune system to anti-PD-L1 and anti-PD-1 therapy. | (109) |
| Nexturastat | PD-1 antibody | Pre- clinical | - | Melanoma | Suppressing tumor growth, decreasing pro-tumorigenic M2 macrophages level and upregulating effector T cells. | (104) |
| Etinostat | PD-1 inhibition | Pre- clinical | - | Lung and renal cell carcinor | na Facilitating the positive antitumor effect of PD-1 inhibition by inhibiting the immunosuppressive functions of MDSCs. | (110) |
| Romidepsin | PD-1 antibody | Pre- clinical | - | Colon cancer | Enhancing the antitumor therapeutic effect and partially reversing the influence of romidepsin on CD4 ⁺ and CD8 ⁺ T cells. | (111) |
| TSA | CTLA-4 antibody | Pre- clinical | - | Melanoma | Facilitating CD4 ⁺ T cell infiltration and intensifying anticancer immune responses. | (106) |
| Vorinostat | Pembrolizumab | | 02638090 | Non-small cell lung cancer | ORR = 13% | (112) |
| Vorinostat | Pembrolizumab | Clinical phase II | 02538510 | Head and neck squamous carcinomas and salivary gla cancer | | (113) |
| Vorinostat | Pembrolizumab | Clinical phase II | 02395627 | Estrogen receptor-positive l cancer | oreast ORR = 4%, CBR = 19% | (114) |
| Entinostat | Atezolizumab | Clinical phase lb/ll | 03280563 | MORPHEUS receptor-posit breast cancer | ve ORR = 6.7% | (115) |
| Entinostat | Atezolizumab | Clinical phase II | 02708680 | Advanced triple-negative br cancer | east ORR = 10%, CBR = 37.5% | (116) |
| Entinostat | Atezolizumab | Clinical phase I | 03024437 | Renal cell carcinoma | ORR = 20% | (117) |
| Entinostat | Nivolumab and ipilimumab | Clinical phase I | 02453620 | Advanced solid tumors | ORR = 16% | (118) |
| Entinostat | Pembrolizumab | Clinical phase II | 02437136 | Non-small cell lung cancer | ORR = 9.2% | (119) |
| Mocetinostat | Nivolumab and ipilimumab | Clinical phase I | 03565406 | Melanoma | ORR = 70% | (120) |
| Domatinostat | | Clinical phase II | 03812796 | Oesophagogastric and colo cancers | rectal SD = 46.2% | (121) |
| Domatinostat | Pembrolizumab | Clinical phase lb | 03278665 | Melanoma | CBR= 30% | (122) |
| Romidepsin | Nivolumab | Clinical phase I/II | 02393794 | Triple negative breast cance | or ORR = 44% | (123) |

HADCi, histone deacetylase inhibitor; ICIs, immune checkpoint inhibitors; PD-L1, programmed death-1 ligand; PD-1, programmed death-1; MDSCs, myeloid-derived suppressor cells; CTLA-4, cytotoxic T-lymphocyte-associated antigen 4; ORR, overall response rate; CBR, clinical benefit rate; SD, stable disease.

patient's immune system, so as to achieve the purpose of controlling or eliminating tumors (125). As an example, the emergence of the therapeutic HPV DNA vaccine has successfully opened up a new direction for the prevention and treatment of cervical cancer, but the antigenicity of the DNA vaccine itself is weak, and the use of DNA vaccine alone does not have a good anti-tumor effect. The decrease or deletion of MHC Class I molecule expression in cervical cancer cells makes tumor cells unable to recognize and kill CD8⁺T cells, resulting in immune escape. Hence, enhancing the expression of MHC Class I molecules in cervical cancer cells is a significant strategy for cervical cancer immunotherapy. Studies have shown that by increasing the expression of proteins encoded by the DNA vaccine, the combination of HDACi AR-42 and the CRT/E7 DNA vaccine can up-regulate the expression of MHC Class I molecules. Moreover, the expression of MHC Class I molecules can also activate the immune response of $CD8^+T$ cells, thereby producing effective anti-tumor effects (126).

Using MC38-CEA colon and 4T1 triple-negative breast murine carcinoma models, the combined application of Entinostat (HDACi), N-803 (IL-15 superagonist) and cancer vaccine was found to promote an anticancer effect by facilitating the infiltration of activated CD8⁺T cells and activating T cell responses to various tumor-associated antigens in the tumor microenvironment (127). Additionally, the synergistic inhibition of HDAC and proteins of bromodomain and extraterminal (BET) can also enhance the vaccine-induced CD8⁺T cell response and promote the therapeutic effect in melanoma treatment (128). Therefore, the combined application of HDACi and cancer vaccines may have promising prospects in clinical cancer immunotherapy (refer to **Table 3**). However, in addition to HDACi and cancer vaccines, other therapeutic agents were involved in the aforementioned studies. Further studies are needed to confirm whether applying HDACi alone with cancer vaccines can produce the same treatment effect in cancer immunotherapy.

COMBINED APPLICATION OF HDACI AND ANTITUMOR ANTIBODIES

Therapeutic antibodies are laboratory-made antibodies designed to destroy tumor cells in different ways, including antibody-dependent cell-mediated cytotoxicity (ADCC), complement-dependent cytotoxicity (CDC), and direct induction of apoptosis by antibodies (133). Several therapeutic antibodies have been approved for the clinical treatment of cancer. In 1997, Rituximab (Rituxan), the first chimeric antibody against CD20, was approved by the FDA for the treatment of non-Hodgkin's lymphoma. Trastuzumab (Herceptin), the first humanized anti-HER2 monoclonal antibody, was marketed in 1998 for the treatment of breast cancer. Several other directly targeted antitumor antibodies, such as Panitumumab, Cetuximab, and Adesimab, have since received FDA approval for anticancer effects in cancer immunotherapy (134–136).

The combination of HDACi and antitumor antibodies has been reported to enhance antitumor responses and promote the therapeutic efficacy of cancer immunotherapy in numerous studies. By blocking the Akt signaling pathway and increasing DNA breakage, the HDAC Class I inhibitor sNDX-275 can significantly promote Trastuzumab-induced growth inhibition in erbB2-overexpressed breast cancer cells, which may also overcome Trastuzumab resistance in breast cancer immunotherapy (129). Treatment with the HDAC inhibitor Chidamide in synergy with Rituximab was found to significantly upregulate CD20 and inhibit the growth of diffuse large B-cell lymphoma (DLBCL) *in vitro* and *in vivo*, confirming that Chidamide may have application prospects in DLBCB immunotherapy (130). A similar study confirmed the effectiveness of the combination of HDACi and antitumor antibodies, in which Entinostat was reported to promote the anticancer effects of Rituximab and chemotherapeutic agents in B-cell lymphoma (131). Another HDACi, Vorinostat, enhanced cell cycle arrest and restored the sensitivity of Rituximab-resistant lymphoma cells to chemotherapeutic drugs (132). Thus, the synergistic application of HDACi and antitumor antibodies may provide a new direction for improving the effectiveness of immunotherapy in cancer treatment (refer to **Table 3**).

DISCUSSION

As a significant epigenetic regulatory mechanism, acetylation/ deacetylation has been found to well regulate the process of gene expression transcription and modulate the host immune response in various cancers, further aiding tumor development and progression by regulating the expression of oncogenes or tumor suppressor genes and weakening the anti-tumor immune response (137). Numerous studies have reported that targeting HATs and HDACs that are essential in the acetylation/deacetylation process can modulate epigenetic signaling pathways and recover the normal expression level of numerous cancer-related genes and anti-tumor immune responses, thereby offering treatment benefits in cancer immunotherapy (138).

Among the various substances that can target HATs/HDACs directly, or those associated with molecules in acetylation/ deacetylation-relevant signaling pathways, HDAC inhibitors have become a large focus of research and been applied with various degrees of success in different settings to treat cancer (139). Since 2006, four different types of HDAC inhibitors have been authorized by the FDA for clinical use, and over 20 different HDAC inhibitors are currently being tested in clinical studies

TABLE 3 | Combined application of HDACi and cancer vaccines/antitumor antibodies.

| Combined application | HADCi | Combined Drugs | Cancer Types | B Effects | References |
|----------------------------------|--------------------------|-------------------------------|----------------------------------|--|----------------|
| HDACi Cancer vaccines | AR-42 | CRT/ E7 DNA vaccine | Cervical cancer | Up-regulating the expression of MHC class I molecules and activates the immune response of CD8 $^{+}$ T cells. | (126) |
| | Entinostat | Cancer vaccine, N-803 | Colon and breast carcinoma | Promoting the anticancer effect by facilitating infiltration of activated CD8 ⁺ T cells and activating T cell responses to various tumor-associated antigens in tumor microenvironment. | (127) |
| | Romidepsin | Cancer vaccine, IBET151 | Melanoma | Enhancing vaccine-induced CD8 ⁺ T cell response. | (128) |
| HDACi Antitumor antibodies | SNDX-275 | Trastuzumab | Breast cancer | Promoting Trastuzumab-induced growth inhibition by blocking the Akt signaling pathway and increasing DNA breakage. | (129) |
| | Chidamide | Rituximab | Diffuse large B-cell lymphoma | Upregulating CD20 and inhibiting the growth of diffuse large B-cell lymphoma cells. | (130) |
| | Entinostat Vorinostat | Rituximab Rituximab | Lymphoma Lymphoma | Promoting the anticancer effects. Enhancing cell cycle arrest and restoring the sensitivity of Rituximab-resistant lymphoma cells to chemotherapeutic drugs. | (131) (132) |

(140). Notably, as a trending field in oncology research, the therapeutic effect of cancer immunotherapy can be elevated in combination with HDAC inhibitors, showing the significance of combining multiple therapeutic methods in the practice of cancer treatment. The efficacy of HDAC inhibitors can be promoted by means of the synergetic application of numerous immunotherapeutic approaches, including cancer vaccines, immune checkpoint inhibitors and others (141).

Many immune-related molecules are not only modified by acetylation/deacetylation, but also by other PTMs such as phosphorylation, ubiquitination, and methylation to regulate the expression level thereof and further modulate host immune responses. Examples include phosphorylated PD-L1 by glycogen synthase kinase 3β (GSK3 β), which can be further degraded by ubiquitin-proteasome system and Olaparib, a PARP1 inhibitor that maintains and upregulates the expression level of PD-L1 in cancers by suppressing the activity of GSK3B, thereby inhibiting the antitumor immune response and facilitating the tumor immune escape (142). Additionally, acetylation/deacetylation can act synergistically with other types of PTMs on an immune-related molecule to regulate the expression thereof. Consistently Photomorphogenic 1 (COP1) is a ubiquitin E3 ligase, and the reduction thereof can inhibit c-Jun ubiquitination degradation, maintain the stability and facilitate phosphorylation by C-Jun Nterminal kinase (JNK), further promoting the translocation of c-Jun into the nucleus and suppressing the production of HDAC3. Such effects result in a looser chromatin structure by acetylation and increases the expression of PD-L1 (143). Hence, the improvement of anticancer immunotherapeutic efficacy may be achieved through the combination of multiple drugs acting on diverse PTMs or the application of a single chemical compound that targets various substrates. The synergistic application of drugs that modulate acetylation/deacetylation and other PTMs represents a promising treatment approach in future cancer immunotherapy.

Despite a number of researchers having reported anticancer effects in immunotherapy that targets the acetylation/deacetylation process and related molecules, there are still questions that warrant further investigation. For instance, numerous immune-related molecules that regulate antitumor immune responses have not been reported to be altered by acetylation or deacetylation. T-cell immunoglobulin and the ITIM domain (TIGIT) is an emerging immune checkpoint that suppresses innate and adaptive immunity, thereby inhibiting antitumor immune responses. The combined application of drugs that block TIGIT and PD-1 (Atezolizumab/ Tiragolumab) can upregulate CD8⁺ T cell expression and function, which may be considered as a potential cancer immunotherapy strategy (144). Another immune regulatory protein, V-domain Ig suppressor of T cell activation (VISTA), can maintain quiescence on T cells and myeloid-derived cells. In a CT26 colon cancer mouse model, immunotherapy applied in combination with VISTA and PD-L1 ICIs contributed to tumor regression and led to long-term survival in the mouse model, demonstrating the rationale for targeting VISTA in cancer immunotherapy. Further studies of acetylation/deacetylation modifications of immune-related molecules, such as the aforementioned novel immune checkpoints, may facilitate broader ideas and more effective

therapeutic strategies for future cancer immunotherapy. Further, many cytokines have also been found to participate in the regulation of immune homeostasis and response such as transforming growth factor (TGF)- β , IFN- γ and others. TGF- β can enhance the expansion of Tregs and suppress the function of effector T cells, causing immune evasion in tumor microenvironments and poor responses to cancer immunotherapy (145). IFN- γ is crucial for NK, NKT, and T cells trafficking into tumors, which regulates the immune response and tumor progression (146). Targeting the acetylation/deacetylation process to modulate the expression of the aforementioned critical cytokines may relieve immunosuppression and reactivate the immune response in the tumor microenvironment, thereby promoting the efficacy of cancer immunotherapy.

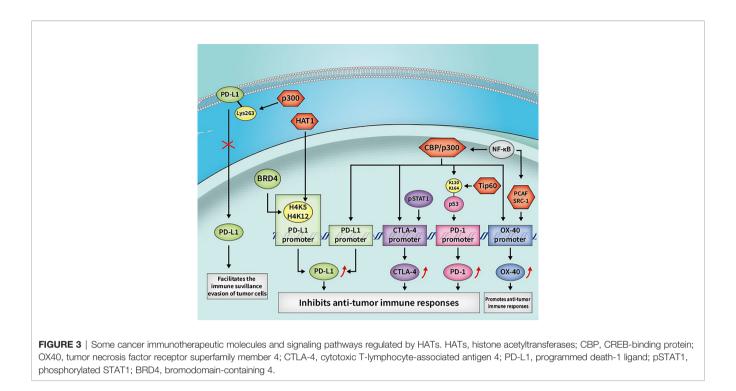
Regarding the existing studies on the regulation of immunerelated molecules by acetylation or deacetylation, the focus has mostly been on the regulatory effect of HDACs, and there is a scarcity of studies on the modification of immune-related molecules by HATs. In the clinical practice of cancer immunotherapy, HDACi have been approved and demonstrated to be valid in cancer treatment. However, unlike HDACi, HAT inhibitors have not been applied in clinical therapy and can be regarded as being stagnant in experimental phases. CCS1477, for example, is the only CBP/p300 inhibitor currently undergoing clinical trials. Even if HAT inhibitors have an essential function in tumor suppression and may exhibit a potential immunotherapeutic effect in cancer treatment, the significance of HAT inhibitors has been well-covered by the in-depth research on HDACi. HAT inhibitors and the modification signaling pathways through which HATs regulate immune-related molecules are of equal significance and deserve the attention of more scientists.

Moreover, a further challenge that inhibits the utilization of HDAC inhibitors is the adverse effects and toxicity observed in early-phase clinical trials. Taking HDACs as an example, only 18 HDACs are responsible for modulating a large number of lysine acetylation residues in over 2000 proteins. Even though HDAC inhibitors can reactivate tumor suppressors, there will be a negative effect on other genes. Future strategies should be directed at developing selective inhibitors to target specific acting sites rather than enzymes, with the aim of overcoming drug toxicity and resistance in cancer immunotherapy.

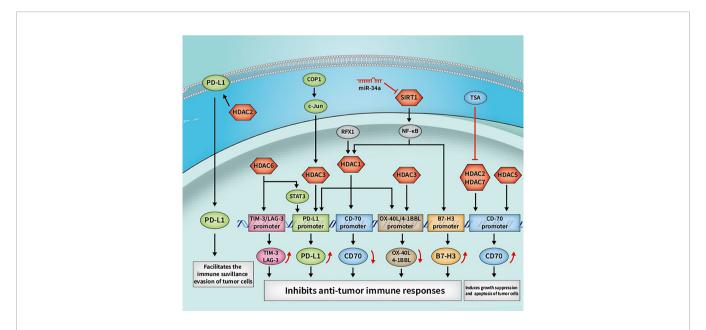
In addition, the focus of most clinical trials has been on the combination of HDACi and ICIs, and there are fewer clinical studies on the anti-tumor efficacy of HDACi combined with cancer vaccines and antitumor antibodies. Future studies on such aspects may provide new strategies for cancer immunotherapy.

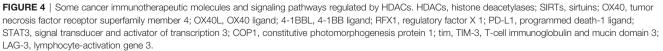
CONCLUSION

In summary, as a significant PTM, lysine acetylation/deacetylation mediated by HATs/HDACs is crucial in regulating various signaling pathways, and also in modulating cancer development and progression by regulating host antitumor immune responses (refer to **Figures 3**, **4**). Owing to the indispensability thereof, lysine



acetylation/deacetylation has received an increasing amount of attention from scientists. The aim of research in the relevant field is to find efficient methods for clinical cancer immunotherapy. Although targeting crucial enzymes in the acetylation/ deacetylation pathway, such as HATs/HDACs, has been shown to recover immune response to exert a cancer therapeutic impact, and certain HDAC inhibitors have even been licensed by the FDA for clinical use, there are still a number of issues to be resolved. The





elucidation of detailed molecules in the acetylation/deacetylation pathway, the discovery of new HAT/HDAC targeted compounds, and the combination of HAT/HDAC inhibitors with other drugs, such as immune checkpoint inhibitors, can all contribute to the finding of new targets and ideas for future cancer immunotherapy.

AUTHOR CONTRIBUTIONS

XY, JH, and JW designed the study. PD, ZM, DLi, MP, and HL searched the literature and wrote the manuscript. YF, YZ, CS, MJ, and DLu searched the literature and made the table, and PD

REFERENCES

- Jayaprakash NG, Surolia A. Role of Glycosylation in Nucleating Protein Folding and Stability. *Biochem J* (2017) 474(14):2333–47. doi: 10.1042/ BCJ20170111
- Choudhary C, Mann M. Decoding Signalling Networks by Mass Spectrometry-Based Proteomics. *Nat Rev Mol Cell Biol* (2010) 11(6):427– 39. doi: 10.1038/nrm2900
- Narita T, Weinert BT, Choudhary C. Functions and Mechanisms of Non-Histone Protein Acetylation. Nat Rev Mol Cell Biol (2019) 20(3):156–74. doi: 10.1038/s41580-018-0081-3
- Drazic A, Myklebust LM, Ree R, Arnesen T. The World of Protein Acetylation. *Biochim Biophys Acta* (2016) 1864(10):1372-401. doi: 10.1016/j.bbapap.2016.06.007
- Gil J, Ramírez-Torres A, Encarnación-Guevara S. Lysine Acetylation and Cancer: A Proteomics Perspective. J Proteomics (2017) 150:297–309. doi: 10.1016/j.jprot.2016.10.003
- Marks P, Rifkind RA, Richon VM, Breslow R, Miller T, Kelly WK. Histone Deacetylases and Cancer: Causes and Therapies. *Nat Rev Cancer* (2001) 1 (3):194–202. doi: 10.1038/35106079
- Liu M, Zhou J, Chen Z, Cheng AS. Understanding the Epigenetic Regulation of Tumours and Their Microenvironments: Opportunities and Problems for Epigenetic Therapy. J Pathol (2017) 241(1):10–24. doi: 10.1002/path.4832
- Singh AK, Bishayee A, Pandey AK. Targeting Histone Deacetylases With Natural and Synthetic Agents: An Emerging Anticancer Strategy. *Nutrients* (2018) 10(6):731. doi: 10.3390/nu10060731
- Zhang H, Shang YP, Chen HY, Li J. Histone Deacetylases Function as Novel Potential Therapeutic Targets for Cancer. *Hepatol Res* (2017) 47(2):149–59. doi: 10.1111/hepr.12757
- Li B, Chan HL, Chen P. Immune Checkpoint Inhibitors: Basics and Challenges. *Curr Med Chem* (2019) 26(17):3009–25. doi: 10.2174/ 0929867324666170804143706
- Gomez S, Tabernacki T, Kobyra J, Roberts P, Chiappinelli KB. Combining Epigenetic and Immune Therapy to Overcome Cancer Resistance. Semin Cancer Biol (2020) 65:99–113. doi: 10.1016/j.semcancer.2019.12.019
- Phillips DM. The Presence of Acetyl Groups of Histones. *Biochem J* (1963) 87(2):258–63. doi: 10.1042/bj0870258
- Glozak MA, Sengupta N, Zhang X, Seto E. Acetylation and Deacetylation of Non-Histone Proteins. *Gene* (2005) 363:15–23. doi: 10.1016/j.gene.2005.09.010
- Norris KL, Lee JY, Yao TP. Acetylation Goes Global: The Emergence of Acetylation Biology. Sci Signaling (2009) 2(97):pe76. doi: 10.1126/ scisignal.297pe76
- Jenuwein T, Allis CD. Translating the Histone Code. Sci (New York NY) (2001) 293(5532):1074–80. doi: 10.1126/science.1063127
- Filippakopoulos P, Knapp S. Targeting Bromodomains: Epigenetic Readers of Lysine Acetylation. Nat Rev Drug Discov (2014) 13(5):337–56. doi: 10.1038/nrd4286
- 17. Choudhary C, Kumar C, Gnad F, Nielsen ML, Rehman M, Walther TC, et al. Lysine Acetylation Targets Protein Complexes and Co-Regulates Major

draw the figures. All authors read and approved the final manuscript.

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Cellular Functions. *Sci (New York NY)* (2009) 325(5942):834–40. doi: 10.1126/science.1175371

- Furdas SD, Kannan S, Sippl W, Jung M. Small Molecule Inhibitors of Histone Acetyltransferases as Epigenetic Tools and Drug Candidates. Archiv Pharm (2012) 345(1):7–21. doi: 10.1002/ardp.201100209
- Roth SY, Denu JM, Allis CD. Histone Acetyltransferases. Annu Rev Biochem (2001) 70:81–120. doi: 10.1146/annurev.biochem.70.1.81
- Yang XJ, Seto E. HATs and HDACs: From Structure, Function and Regulation to Novel Strategies for Therapy and Prevention. Oncogene (2007) 26(37):5310–8. doi: 10.1038/sj.onc.1210599
- Gu W, Roeder RG. Activation of P53 Sequence-Specific DNA Binding by Acetylation of the P53 C-Terminal Domain. *Cell* (1997) 90(4):595–606. doi: 10.1016/s0092-8674(00)80521-8
- Patel JH, Du Y, Ard PG, Phillips C, Carella B, Chen CJ, et al. The C-MYC Oncoprotein Is a Substrate of the Acetyltransferases Hgcn5/PCAF and TIP60. *Mol Cell Biol* (2004) 24(24):10826–34. doi: 10.1128/ mcb.24.24.10826-10834.2004
- Berger SL. The Complex Language of Chromatin Regulation During Transcription. Nature (2007) 447(7143):407–12. doi: 10.1038/nature05915
- Ai T, Cui H, Chen L. Multi-Targeted Histone Deacetylase Inhibitors in Cancer Therapy. *Curr Med Chem* (2012) 19(4):475–87. doi: 10.2174/ 092986712798918842
- Iizuka M, Smith MM. Functional Consequences of Histone Modifications. Curr Opin Genet Dev (2003) 13(2):154–60. doi: 10.1016/s0959-437x(03)00020-0
- Haberland M, Montgomery RL, Olson EN. The Many Roles of Histone Deacetylases in Development and Physiology: Implications for Disease and Therapy. Nat Rev Genet (2009) 10(1):32–42. doi: 10.1038/nrg2485
- Yang XJ, Seto E. The Rpd3/Hda1 Family of Lysine Deacetylases: From Bacteria and Yeast to Mice and Men. *Nat Rev Mol Cell Biol* (2008) 9(3):206– 18. doi: 10.1038/nrm2346
- Moser MA, Hagelkruys A, Seiser C. Transcription and Beyond: The Role of Mammalian Class I Lysine Deacetylases. *Chromosoma* (2014) 123(1-2):67– 78. doi: 10.1007/s00412-013-0441-x
- Gao L, Cueto MA, Asselbergs F, Atadja P. Cloning and Functional Characterization of HDAC11, A Novel Member of the Human Histone Deacetylase Family. J Biol Chem (2002) 277(28):25748–55. doi: 10.1074/ jbc.M111871200
- Ma Z, Liu D, Di S, Zhang Z, Li W, Zhang J, et al. Histone Deacetylase 9 Downregulation Decreases Tumor Growth and Promotes Apoptosis in Non-Small Cell Lung Cancer After Melatonin Treatment. *J Pineal Res* (2019) 67 (2):e12587. doi: 10.1111/jpi.12587
- Guarente L. Calorie Restriction and Sirtuins Revisited. Genes Dev (2013) 27 (19):2072–85. doi: 10.1101/gad.227439.113
- Dryden SC, Nahhas FA, Nowak JE, Goustin AS, Tainsky MA. Role for Human SIRT2 NAD-Dependent Deacetylase Activity in Control of Mitotic Exit in the Cell Cycle. *Mol Cell Biol* (2003) 23(9):3173–85. doi: 10.1128/ mcb.23.9.3173-3185.2003
- Serrano L, Martínez-Redondo P, Marazuela-Duque A, Vazquez BN, Dooley SJ, Voigt P, et al. The Tumor Suppressor SirT2 Regulates Cell Cycle

Progression and Genome Stability by Modulating the Mitotic Deposition of H4K20 Methylation. *Genes Dev* (2013) 27(6):639–53. doi: 10.1101/gad.211342.112

- Verdin E, Hirschey MD, Finley LW, Haigis MC. Sirtuin Regulation of Mitochondria: Energy Production, Apoptosis, and Signaling. Trends Biochem Sci (2010) 35(12):669–75. doi: 10.1016/j.tibs.2010.07.003
- Bosch-Presegué L, Vaquero A. The Dual Role of Sirtuins in Cancer. Genes Cancer (2011) 2(6):648–62. doi: 10.1177/1947601911417862
- Yanginlar C, Logie C. HDAC11 is a Regulator of Diverse Immune Functions. Biochim Biophys Acta Gene Regul Mech (2018) 1861(1):54–9. doi: 10.1016/ j.bbagrm.2017.12.002
- Audia JE, Campbell RM. Histone Modifications and Cancer. Cold Spring Harbor Perspect Biol (2016) 8(4):a019521. doi: 10.1101/cshperspect.a019521
- 38. Sasidharan Nair V, Saleh R, Toor SM, Taha RZ, Ahmed AA, Kurer MA, et al. Transcriptomic Profiling Disclosed the Role of DNA Methylation and Histone Modifications in Tumor-Infiltrating Myeloid-Derived Suppressor Cell Subsets in Colorectal Cancer. *Clin Epigenet* (2020) 12(1):13. doi: 10.1186/s13148-020-0808-9
- Gao Y, Nihira NT, Bu X, Chu C, Zhang J, Kolodziejczyk A, et al. Acetylation-Dependent Regulation of PD-L1 Nuclear Translocation Dictates the Efficacy of Anti-PD-1 Immunotherapy. *Nat Cell Biol* (2020) 22(9):1064–75. doi: 10.1038/s41556-020-0562-4
- Fan P, Zhao J, Meng Z, Wu H, Wang B, Wu H, et al. Overexpressed Histone Acetyltransferase 1 Regulates Cancer Immunity by Increasing Programmed Death-Ligand 1 Expression in Pancreatic Cancer. J Exp Clin Cancer Res CR (2019) 38(1):47. doi: 10.1186/s13046-019-1044-z
- Darvin P, Sasidharan Nair V, Elkord E. PD-L1 Expression in Human Breast Cancer Stem Cells Is Epigenetically Regulated Through Posttranslational Histone Modifications. J Oncol (2019) 2019:3958908. doi: 10.1155/2019/ 3958908
- Leinlaf M, Perez-Villaroel P, Knox T, Pabon M, Sahakian E, Powers J, et al. Essential Role of HDAC6 in the Regulation of PD-L1 in Melanoma. *Mol* Oncol (2016) 10(5):735–50. doi: 10.1016/j.molonc.2015.12.012
- 43. Wang H, Fu C, Du J, Wang H, He R, Yin X, et al. Enhanced Histone H3 Acetylation of the PD-L1 Promoter *via* the COP1/c-Jun/HDAC3 Axis Is Required for PD-L1 Expression in Drug-Resistant Cancer Cells. *J Exp Clin Cancer Res CR* (2020) 39(1):29. doi: 10.1186/s13046-020-1536-x
- 44. Cao Z, Kon N, Liu Y, Xu W, Wen J, Yao H, et al. An Unexpected Role for P53 in Regulating Cancer Cell-Intrinsic PD-1 by Acetylation. *Sci Adv* (2021) 7(14):eabf4148. doi: 10.1126/sciadv.abf4148
- 45. Mo X, Zhang H, Preston S, Martin K, Zhou B, Vadalia N, et al. Interferon-γ Signaling in Melanocytes and Melanoma Cells Regulates Expression of CTLA-4. *Cancer Res* (2018) 78(2):436–50. doi: 10.1158/0008-5472.Can-17-1615
- Zhao HY, Ma YH, Li DQ, Sun T, Li LZ, Li P, et al. Low-Dose Chidamide Restores Immune Tolerance in ITP in Mice and Humans. *Blood* (2019) 133 (7):730–42. doi: 10.1182/blood-2018-05-847624
- Laino AS, Betts BC, Veerapathran A, Dolgalev I, Sarnaik A, Quayle SN, et al. HDAC6 Selective Inhibition of Melanoma Patient T-Cells Augments Anti-Tumor Characteristics. *J Immunother Cancer* (2019) 7(1):33. doi: 10.1186/ s40425-019-0517-0
- Wang MY, Qi B, Wang F, Lin ZR, Li MY, Yin WJ, et al. PBK Phosphorylates MSL1 to Elicit Epigenetic Modulation of CD276 in Nasopharyngeal Carcinoma. Oncogenesis (2021) 10(1):9. doi: 10.1038/s41389-020-00293-9
- Meng F, Yang M, Chen Y, Chen W, Wang W. miR-34a Induces Immunosuppression in Colorectal Carcinoma Through Modulating a SIRT1/NF-κb/B7-H3/TNF-α Axis. *Cancer Immunol Immunother CII* (2021) 70(8):2247–59. doi: 10.1007/s00262-021-02862-2
- Zhang X, Xiao X, Lan P, Li J, Dou Y, Chen W, et al. OX40 Costimulation Inhibits Foxp3 Expression and Treg Induction *via* BATF3-Dependent and Independent Mechanisms. *Cell Rep* (2018) 24(3):607–18. doi: 10.1016/ j.celrep.2018.06.052
- Tone Y, Kojima Y, Furuuchi K, Brady M, Yashiro-Ohtani Y, Tykocinski ML, et al. OX40 Gene Expression is Up-Regulated by Chromatin Remodeling in its Promoter Region Containing Sp1/Sp3, YY1, and NF-Kappa B Binding Sites. J Immunol (Baltimore Md 1950) (2007) 179(3):1760–7. doi: 10.4049/ jimmunol.179.3.1760

- Cacan E. Epigenetic-Mediated Immune Suppression of Positive Co-Stimulatory Molecules in Chemoresistant Ovarian Cancer Cells. Cell Biol Int (2017) 41(3):328–39. doi: 10.1002/cbin.10729
- 53. Kamijo H, Miyagaki T, Shishido-Takahashi N, Nakajima R, Oka T, Suga H, et al. Aberrant CD137 Ligand Expression Induced by GATA6 Overexpression Promotes Tumor Progression in Cutaneous T-Cell Lymphoma. *Blood* (2018) 132(18):1922–35. doi: 10.1182/blood-2018-04-845834
- Zhao M, Sun Y, Gao F, Wu X, Tang J, Yin H, et al. Epigenetics and SLE: RFX1 Downregulation Causes CD11a and CD70 Overexpression by Altering Epigenetic Modifications in Lupus CD4+ T Cells. *J Autoimmun* (2010) 35 (1):58–69. doi: 10.1016/j.jaut.2010.02.002
- Zhou Y, Qiu X, Luo Y, Yuan J, Li Y, Zhong Q, et al. Histone Modifications and Methyl-CpG-Binding Domain Protein Levels at the TNFSF7 (CD70) Promoter in SLE CD4+ T Cells. *Lupus* (2011) 20(13):1365–71. doi: 10.1177/ 0961203311413412
- Huang Y, Tan M, Gosink M, Wang KK, Sun Y. Histone Deacetylase 5 is Not a P53 Target Gene, But its Overexpression Inhibits Tumor Cell Growth and Induces Apoptosis. *Cancer Res* (2002) 62(10):2913–22.
- Ritprajak P, Azuma M. Intrinsic and Extrinsic Control of Expression of the Immunoregulatory Molecule PD-L1 in Epithelial Cells and Squamous Cell Carcinoma. Oral Oncol (2015) 51(3):221-8. doi: 10.1016/ j.oraloncology.2014.11.014
- Ohaegbulam KC, Assal A, Lazar-Molnar E, Yao Y, Zang X. Human Cancer Immunotherapy With Antibodies to the PD-1 and PD-L1 Pathway. *Trends Mol Med* (2015) 21(1):24–33. doi: 10.1016/j.molmed.2014.10.009
- Azuma T, Yao S, Zhu G, Flies AS, Flies SJ, Chen L. B7-H1 is a Ubiquitous Antiapoptotic Receptor on Cancer Cells. *Blood* (2008) 111(7):3635–43. doi: 10.1182/blood-2007-11-123141
- Gato-Cañas M, Zuazo M, Arasanz H, Ibañez-Vea M, Lorenzo L, Fernandez-Hinojal G, et al. PDL1 Signals Through Conserved Sequence Motifs to Overcome Interferon-Mediated Cytotoxicity. *Cell Rep* (2017) 20(8):1818–29. doi: 10.1016/j.celrep.2017.07.075
- 61. Gong J, Chehrazi-Raffle A, Reddi S, Salgia R. Development of PD-1 and PD-L1 Inhibitors as a Form of Cancer Immunotherapy: A Comprehensive Review of Registration Trials and Future Considerations. *J Immunother Cancer* (2018) 6(1):8. doi: 10.1186/s40425-018-0316-z
- 62. Rosenberg JE, Hoffman-Censits J, Powles T, van der Heijden MS, Balar AV, Necchi A, et al. Atezolizumab in Patients With Locally Advanced and Metastatic Urothelial Carcinoma Who Have Progressed Following Treatment With Platinum-Based Chemotherapy: A Single-Arm, Multicentre, Phase 2 Trial. *Lancet (Lond Engl)* (2016) 387(10031):1909– 20. doi: 10.1016/s0140-6736(16)00561-4
- Hsu JM, Li CW, Lai YJ, Hung MC. Posttranslational Modifications of PD-L1 and Their Applications in Cancer Therapy. *Cancer Res* (2018) 78(22):6349– 53. doi: 10.1158/0008-5472.Can-18-1892
- Wherry EJ, Kurachi M. Molecular and Cellular Insights Into T Cell Exhaustion. Nat Rev Immunol (2015) 15(8):486–99. doi: 10.1038/nri3862
- 65. Gros A, Parkhurst MR, Tran E, Pasetto A, Robbins PF, Ilyas S, et al. Prospective Identification of Neoantigen-Specific Lymphocytes in the Peripheral Blood of Melanoma Patients. *Nat Med* (2016) 22(4):433–8. doi: 10.1038/nm.4051
- Zhao L, Yu H, Yi S, Peng X, Su P, Xiao Z, et al. The Tumor Suppressor miR-138-5p Targets PD-L1 in Colorectal Cancer. *Oncotarget* (2016) 7(29):45370– 84. doi: 10.18632/oncotarget.9659
- Krummel MF, Allison JP. CTLA-4 Engagement Inhibits IL-2 Accumulation and Cell Cycle Progression Upon Activation of Resting T Cells. J Exp Med (1996) 183(6):2533–40. doi: 10.1084/jem.183.6.2533
- Wilgenhof S, Neyns B. Anti-CTLA-4 Antibody-Induced Guillain-Barré Syndrome in a Melanoma Patient. Ann Oncol (2011) 22(4):991–3. doi: 10.1093/annonc/mdr028
- Youlin K, Li Z, Xiaodong W, Xiuheng L, Hengchen Z. Combination Immunotherapy With 4-1BBL and CTLA-4 Blockade for the Treatment of Prostate Cancer. *Clin Dev Immunol* (2012) 2012:439235. doi: 10.1155/2012/ 439235
- 70. Sangro B, Gomez-Martin C, de la Mata M, Iñarrairaegui M, Garralda E, Barrera P, et al. A Clinical Trial of CTLA-4 Blockade With Tremelimumab

in Patients With Hepatocellular Carcinoma and Chronic Hepatitis C. J Hepatol (2013) 59(1):81–8. doi: 10.1016/j.jhep.2013.02.022

- Leach DR, Krummel MF, Allison JP. Enhancement of Antitumor Immunity by CTLA-4 Blockade. *Sci (New York NY)* (1996) 271(5256):1734–6. doi: 10.1126/science.271.5256.1734
- Ramagopal UA, Liu W, Garrett-Thomson SC, Bonanno JB, Yan Q, Srinivasan M, et al. Structural Basis for Cancer Immunotherapy by the First-in-Class Checkpoint Inhibitor Ipilimumab. *Proc Natl Acad Sci USA* (2017) 114(21):E4223–e32. doi: 10.1073/pnas.1617941114
- Joller N, Kuchroo VK. Tim-3, Lag-3, and TIGIT. Curr Topics Microbiol Immunol (2017) 410:127–56. doi: 10.1007/82_2017_62
- 74. Gao X, Zhu Y, Li G, Huang H, Zhang G, Wang F, et al. TIM-3 Expression Characterizes Regulatory T Cells in Tumor Tissues and is Associated With Lung Cancer Progression. *PloS One* (2012) 7(2):e30676. doi: 10.1371/ journal.pone.0030676
- 75. Fourcade J, Sun Z, Benallaoua M, Guillaume P, Luescher IF, Sander C, et al. Upregulation of Tim-3 and PD-1 Expression is Associated With Tumor Antigen-Specific CD8+ T Cell Dysfunction in Melanoma Patients. J Exp Med (2010) 207(10):2175–86. doi: 10.1084/jem.20100637
- Zhou E, Huang Q, Wang J, Fang C, Yang L, Zhu M, et al. Up-Regulation of Tim-3 is Associated With Poor Prognosis of Patients With Colon Cancer. *Int J Clin Exp Pathol* (2015) 8(7):8018–27.
- Kuchroo VK, Anderson AC, Petrovas C. Coinhibitory Receptors and CD8 T Cell Exhaustion in Chronic Infections. *Curr Opin HIV AIDS* (2014) 9 (5):439–45. doi: 10.1097/coh.00000000000088
- He Y, Rivard CJ, Rozeboom L, Yu H, Ellison K, Kowalewski A, et al. Lymphocyte-Activation Gene-3, An Important Immune Checkpoint in Cancer. *Cancer Sci* (2016) 107(9):1193–7. doi: 10.1111/cas.12986
- Mao Y, Chen L, Wang F, Zhu D, Ge X, Hua D, et al. Cancer Cell-Expressed B7-H3 Regulates the Differentiation of Tumor-Associated Macrophages in Human Colorectal Carcinoma. *Oncol Lett* (2017) 14(5):6177–83. doi: 10.3892/ol.2017.6935
- Cong F, Yu H, Gao X. Expression of CD24 and B7-H3 in Breast Cancer and the Clinical Significance. Oncol Lett (2017) 14(6):7185–90. doi: 10.3892/ol.2017.7142
- Chapoval AI, Ni J, Lau JS, Wilcox RA, Flies DB, Liu D, et al. B7-H3: A Costimulatory Molecule for T Cell Activation and IFN-Gamma Production. *Nat Immunol* (2001) 2(3):269–74. doi: 10.1038/85339
- Loos M, Hedderich DM, Ottenhausen M, Giese NA, Laschinger M, Esposito I, et al. Expression of the Costimulatory Molecule B7-H3 is Associated With Prolonged Survival in Human Pancreatic Cancer. *BMC Cancer* (2009) 9:463. doi: 10.1186/1471-2407-9-463
- Lupu CM, Eisenbach C, Kuefner MA, Schmidt J, Lupu AD, Stremmel W, et al. An Orthotopic Colon Cancer Model for Studying the B7-H3 Antitumor Effect *In Vivo. J Gastrointest Surg* (2006) 10(5):635–45. doi: 10.1007/bf03239969
- Chen JT, Chen CH, Ku KL, Hsiao M, Chiang CP, Hsu TL, et al. Glycoprotein B7-H3 Overexpression and Aberrant Glycosylation in Oral Cancer and Immune Response. *Proc Natl Acad Sci United States America* (2015) 112 (42):13057–62. doi: 10.1073/pnas.1516991112
- Carboni S, Aboul-Enein F, Waltzinger C, Killeen N, Lassmann H, Peña-Rossi C. CD134 Plays a Crucial Role in the Pathogenesis of EAE and is Upregulated in the CNS of Patients With Multiple Sclerosis. *J Neuroimmunol* (2003) 145(1-2):1–11. doi: 10.1016/j.jneuroim.2003.07.001
- Vu MD, Clarkson MR, Yagita H, Turka LA, Sayegh MH, Li XC. Critical, But Conditional, Role of OX40 in Memory T Cell-Mediated Rejection. *J Immunol (Baltimore Md 1950)* (2006) 176(3):1394–401. doi: 10.4049/ jimmunol.176.3.1394
- Weixler B, Cremonesi E, Sorge R, Muraro MG, Delko T, Nebiker CA, et al. OX40 Expression Enhances the Prognostic Significance of CD8 Positive Lymphocyte Infiltration in Colorectal Cancer. *Oncotarget* (2015) 6 (35):37588–99. doi: 10.18632/oncotarget.5940
- Martins MR, Santos RLD, Jatahy KDN, Matta MCD, Batista TP, Júnior JIC, et al. Could OX40 Agonist Antibody Promote Activation of the Anti-Tumor Immune Response in Gastric Cancer? J Surg Oncol (2018) 117(5):840–4. doi: 10.1002/jso.25001
- Guo Z, Wang X, Cheng D, Xia Z, Luan M, Zhang S. PD-1 Blockade and OX40 Triggering Synergistically Protects Against Tumor Growth in a

Murine Model of Ovarian Cancer. *PloS One* (2014) 9(2):e89350. doi: 10.1371/journal.pone.0089350

- Redmond WL, Triplett T, Floyd K, Weinberg AD. Dual Anti-OX40/IL-2 Therapy Augments Tumor Immunotherapy via IL-2R-Mediated Regulation of OX40 Expression. PloS One (2012) 7(4):e34467. doi: 10.1371/ journal.pone.0034467
- Yokouchi H, Yamazaki K, Chamoto K, Kikuchi E, Shinagawa N, Oizumi S, et al. Anti-OX40 Monoclonal Antibody Therapy in Combination With Radiotherapy Results in Therapeutic Antitumor Immunity to Murine Lung Cancer. *Cancer Sci* (2008) 99(2):361–7. doi: 10.1111/j.1349-7006.2007.00664.x
- Li SY, Liu Y. Immunotherapy of Melanoma With the Immune Costimulatory Monoclonal Antibodies Targeting CD137. *Clin Pharmacol Adv Appl* (2013) 5(Suppl 1):47–53. doi: 10.2147/cpaa.S46199
- Cannons JL, Choi Y, Watts TH. Role of TNF Receptor-Associated Factor 2 and P38 Mitogen-Activated Protein Kinase Activation During 4-1BB-Dependent Immune Response. J Immunol (Baltimore Md 1950) (2000) 165(11):6193–204. doi: 10.4049/jimmunol.165.11.6193
- 94. Mogi S, Sakurai J, Kohsaka T, Enomoto S, Yagita H, Okumura K, et al. Tumour Rejection by Gene Transfer of 4-1BB Ligand Into a CD80(+) Murine Squamous Cell Carcinoma and the Requirements of Co-Stimulatory Molecules on Tumour and Host Cells. *Immunology* (2000) 101(4):541–7. doi: 10.1046/j.1365-2567.2000.t01-1-00138.x
- Boursalian TE, McEarchern JA, Law CL, Grewal IS. Targeting CD70 for Human Therapeutic Use. Adv Exp Med Biol (2009) 647:108–19. doi: 10.1007/978-0-387-89520-8_7
- Lens SM, Tesselaar K, van Oers MH, van Lier RA. Control of Lymphocyte Function Through CD27-CD70 Interactions. *Semin Immunol* (1998) 10 (6):491–9. doi: 10.1006/smim.1998.0154
- Bertrand P, Maingonnat C, Penther D, Guney S, Ruminy P, Picquenot JM, et al. The Costimulatory Molecule CD70 is Regulated by Distinct Molecular Mechanisms and is Associated With Overall Survival in Diffuse Large B-Cell Lymphoma. *Genes Chromosomes Cancer* (2013) 52(8):764–74. doi: 10.1002/ gcc.22072
- Petrau C, Cornic M, Bertrand P, Maingonnat C, Marchand V, Picquenot JM, et al. CD70: A Potential Target in Breast Cancer? J Cancer (2014) 5(9):761–4. doi: 10.7150/jca.10360
- 99. Lee WW, Yang ZZ, Li G, Weyand CM, Goronzy JJ. Unchecked CD70 Expression on T Cells Lowers Threshold for T Cell Activation in Rheumatoid Arthritis. J Immunol (Baltimore Md 1950) (2007) 179 (4):2609–15. doi: 10.4049/jimmunol.179.4.2609
- 100. Wang QJ, Hanada K, Robbins PF, Li YF, Yang JC. Distinctive Features of the Differentiated Phenotype and Infiltration of Tumor-Reactive Lymphocytes in Clear Cell Renal Cell Carcinoma. *Cancer Res* (2012) 72(23):6119–29. doi: 10.1158/0008-5472.Can-12-0588
- 101. Ho TCS, Chan AHY, Ganesan A. Thirty Years of HDAC Inhibitors: 2020 Insight and Hindsight. J Med Chem (2020) 63(21):12460–84. doi: 10.1021/ acs.jmedchem.0c00830
- 102. Shen L, Ciesielski M, Ramakrishnan S, Miles KM, Ellis L, Sotomayor P, et al. Class I Histone Deacetylase Inhibitor Entinostat Suppresses Regulatory T Cells and Enhances Immunotherapies in Renal and Prostate Cancer Models. *PloS One* (2012) 7(1):e30815. doi: 10.1371/journal.pone.0030815
- West AC, Smyth MJ, Johnstone RW. The Anticancer Effects of HDAC Inhibitors Require the Immune System. Oncoimmunology (2014) 3(1): e27414. doi: 10.4161/onci.27414
- 104. Knox T, Sahakian E, Banik D, Hadley M, Palmer E, Noonepalle S, et al. Selective HDAC6 Inhibitors Improve Anti-PD-1 Immune Checkpoint Blockade Therapy by Decreasing the Anti-Inflammatory Phenotype of Macrophages and Down-Regulation of Immunosuppressive Proteins in Tumor Cells. Sci Rep (2019) 9(1):6136. doi: 10.1038/s41598-019-42237-3
- 105. He M, Yang T, Wang Y, Wang M, Chen X, Ding D, et al. Immune Checkpoint Inhibitor-Based Strategies for Synergistic Cancer Therapy. Adv Healthcare Mater (2021) 10(9):e2002104. doi: 10.1002/adhm.202002104
- 106. Cao K, Wang G, Li W, Zhang L, Wang R, Huang Y, et al. Histone Deacetylase Inhibitors Prevent Activation-Induced Cell Death and Promote Anti-Tumor Immunity. *Oncogene* (2015) 34(49):5960-70. doi: 10.1038/onc.2015.46

- 107. Yang W, Feng Y, Zhou J, Cheung OK, Cao J, Wang J, et al. A Selective HDAC8 Inhibitor Potentiates Antitumor Immunity and Efficacy of Immune Checkpoint Blockade in Hepatocellular Carcinoma. *Sci Trans Med* (2021) 13 (588):eaaz6804. doi: 10.1126/scitranslmed.aaz6804
- 108. Kim YD, Park SM, Ha HC, Lee AR, Won H, Cha H, et al. HDAC Inhibitor, CG-745, Enhances the Anti-Cancer Effect of Anti-PD-1 Immune Checkpoint Inhibitor by Modulation of the Immune Microenvironment. J Cancer (2020) 11(14):4059–72. doi: 10.7150/jca.44622
- 109. Zheng H, Zhao W, Yan C, Watson CC, Massengill M, Xie M, et al. HDAC Inhibitors Enhance T-Cell Chemokine Expression and Augment Response to PD-1 Immunotherapy in Lung Adenocarcinoma. *Clin Cancer Res* (2016) 22 (16):4119–32. doi: 10.1158/1078-0432.Ccr-15-2584
- 110. Orillion A, Hashimoto A, Damayanti N, Shen L, Adelaiye-Ogala R, Arisa S, et al. Entinostat Neutralizes Myeloid-Derived Suppressor Cells and Enhances the Antitumor Effect of PD-1 Inhibition in Murine Models of Lung and Renal Cell Carcinoma. *Clin Cancer Res* (2017) 23(17):5187–201. doi: 10.1158/1078-0432.Ccr-17-0741
- 111. Shi Y, Fu Y, Zhang X, Zhao G, Yao Y, Guo Y, et al. Romidepsin (FK228) Regulates the Expression of the Immune Checkpoint Ligand PD-L1 and Suppresses Cellular Immune Functions in Colon Cancer. *Cancer Immunol Immunother CII* (2021) 70(1):61–73. doi: 10.1007/s00262-020-02653-1
- 112. Gray JE, Saltos A, Tanvetyanon T, Haura EB, Creelan B, Antonia SJ, et al. Phase I/Ib Study of Pembrolizumab Plus Vorinostat in Advanced/Metastatic Non-Small Cell Lung Cancer. *Clin Cancer Res* (2019) 25(22):6623–32. doi: 10.1158/1078-0432.Ccr-19-1305
- 113. Rodriguez CP, Wu QV, Voutsinas J, Fromm JR, Jiang X, Pillarisetty VG, et al. A Phase II Trial of Pembrolizumab and Vorinostat in Recurrent Metastatic Head and Neck Squamous Cell Carcinomas and Salivary Gland Cancer. *Clin Cancer Res* (2020) 26(4):837–45. doi: 10.1158/1078-0432.Ccr-19-2214
- 114. Terranova-Barberio M, Pawlowska N, Dhawan M, Moasser M, Chien AJ, Melisko ME, et al. Exhausted T Cell Signature Predicts Immunotherapy Response in ER-Positive Breast Cancer. Nat Commun (2020) 11(1):3584. doi: 10.1038/s41467-020-17414-y
- 115. Sonnenblick A, Im SA, Lee KS, Tan A, Telli M, Strulov Shachar S, et al. 267p Phase Ib/II Open-Label, Randomized Evaluation of Second- or Third-Line (2L/3L) Atezolizumab (Atezo) + Entinostat (Entino) in MORPHEUS-HR+ Breast Cancer (M-HR+BC). Ann Oncol (2021) 32:S479. doi: 10.1016/ j.annonc.2021.08.550
- 116. O'Shaughnessy J, Moroose RL, Babu S, Baramidze K, Chan D, Leitner SP, et al. Results of ENCORE 602 (TRIO025), A Phase II, Randomized, Placebo-Controlled, Double-Blinded, Multicenter Study of Atezolizumab With or Without Entinostat in Patients With Advanced Triple-Negative Breast Cancer (aTNBC). J Clin Oncol (2020) 38(15_suppl):1014. doi: 10.1200/ JCO.2020.38.15_suppl.1014
- 117. Pili R, Adra N, Damayanti N, Logan TF, Narayan V, Monk P, et al. Immunomodulation by HDAC Inhibition: Results From a Phase I Study With Entinostat in Combination With Atezolizumab and Bevacizumab in Metastatic Renal Cell Carcinoma Patients. J Clin Oncol (2020) 38 (15_suppl):5064. doi: 10.1200/JCO.2020.38.15_suppl.5064
- 118. Roussos Torres ET, Rafie C, Wang C, Lim D, Brufsky A, LoRusso P, et al. Phase I Study of Entinostat and Nivolumab With or Without Ipilimumab in Advanced Solid Tumors (ETCTN-9844). *Clin Cancer Res* (2021) 27 (21):5828–37. doi: 10.1158/1078-0432.Ccr-20-5017
- 119. Hellmann MD, Jänne PA, Opyrchal M, Hafez N, Raez LE, Gabrilovich DI, et al. Entinostat Plus Pembrolizumab in Patients With Metastatic NSCLC Previously Treated With Anti-PD-(L)1 Therapy. *Clin Cancer Res* (2021) 27 (4):1019–28. doi: 10.1158/1078-0432.Ccr-20-3305
- 120. Weber JS, Laino AS, Vassallo M, Pavlick A, Malatyali S, Krishnarajapet S, et al. Preclinical and Clinical Studies of a Class I/IV HDAC Inhibitor, Mocetinostat, in Melanoma. J Clin Oncol (2020) 38(15_suppl):10052. doi: 10.1200/JCO.2020.38.15_suppl.10052
- 121. Cartwright E, Turkes F, Saffery C, Tran A, Smith G, Moreno SE, et al. 443p EMERGE: A Phase II Trial Assessing the Efficacy of Domatinostat Plus Avelumab in Patients With Previously Treated Advanced Mismatch Repair Proficient Oesophagogastric and Colorectal Cancers – Phase IIA Dose Finding. Ann Oncol (2021) 32:S555–S6. doi: 10.1016/j.annonc.2021.08.964
- 122. Hassel JC, Berking C, Schlaak M, Eigentler T, Gutzmer R, Ascierto PA, et al. Results From the Phase Ib of the SENSITIZE Trial Combining Domatinostat

With Pembrolizumab in Advanced Melanoma Patients Refractory to Prior Checkpoint Inhibitor Therapy. *J Clin Oncol* (2021) 39(15_suppl):9545. doi: 10.1200/JCO.2021.39.15_suppl.9545

- 123. Sharma P, Abramson VG, O'Dea A, Nye LE, Mayer IA, Crane GJ, et al. Romidepsin (HDACi) Plus Cisplatin and Nivolumab Triplet Combination in Patients With Metastatic Triple Negative Breast Cancer (mTNBC). J Clin Oncol (2021) 39(15_suppl):1076. doi: 10.1200/JCO.2021.39.15_suppl.1076
- 124. Ny L, Jespersen H, Karlsson J, Alsén S, Filges S, All-Eriksson C, et al. The PEMDAC Phase 2 Study of Pembrolizumab and Entinostat in Patients With Metastatic Uveal Melanoma. *Nat Commun* (2021) 12(1):5155. doi: 10.1038/ s41467-021-25332-w
- DeMaria PJ, Bilusic M. Cancer Vaccines. Hematol oncol Clinics North America (2019) 33(2):199–214. doi: 10.1016/j.hoc.2018.12.001
- 126. Lee SY, Huang Z, Kang TH, Soong RS, Knoff J, Axenfeld E, et al. Histone Deacetylase Inhibitor AR-42 Enhances E7-Specific CD8⁺ T Cell-Mediated Antitumor Immunity Induced by Therapeutic HPV DNA Vaccination. J Mol Med (Berlin Germany) (2013) 91(10):1221–31. doi: 10.1007/s00109-013-1054-9
- 127. Hicks KC, Knudson KM, Lee KL, Hamilton DH, Hodge JW, Figg WD, et al. Cooperative Immune-Mediated Mechanisms of the HDAC Inhibitor Entinostat, an IL15 Superagonist, and a Cancer Vaccine Effectively Synergize as a Novel Cancer Therapy. *Clin Cancer Res* (2020) 26(3):704– 16. doi: 10.1158/1078-0432.Ccr-19-0727
- 128. Badamchi-Zadeh A, Moynihan KD, Larocca RA, Aid M, Provine NM, Iampietro MJ, et al. Combined HDAC and BET Inhibition Enhances Melanoma Vaccine Immunogenicity and Efficacy. J Immunol (Baltimore Md 1950) (2018) 201(9):2744–52. doi: 10.4049/jimmunol.1800885
- 129. Huang X, Wang S, Lee CK, Yang X, Liu B. HDAC Inhibitor SNDX-275 Enhances Efficacy of Trastuzumab in Erbb2-Overexpressing Breast Cancer Cells and Exhibits Potential to Overcome Trastuzumab Resistance. *Cancer Lett* (2011) 307(1):72–9. doi: 10.1016/j.canlet.2011.03.019
- 130. Guan XW, Wang HQ, Ban WW, Chang Z, Chen HZ, Jia L, et al. Novel HDAC Inhibitor Chidamide Synergizes With Rituximab to Inhibit Diffuse Large B-Cell Lymphoma Tumour Growth by Upregulating CD20. *Cell Death Dis* (2020) 11(1):20. doi: 10.1038/s41419-019-2210-0
- 131. Frys S, Simons Z, Hu Q, Barth MJ, Gu JJ, Mavis C, et al. Entinostat, a Novel Histone Deacetylase Inhibitor is Active in B-Cell Lymphoma and Enhances the Anti-Tumour Activity of Rituximab and Chemotherapy Agents. Br J Haematol (2015) 169(4):506–19. doi: 10.1111/bjh.13318
- 132. Xue K, Gu JJ, Zhang Q, Mavis C, Hernandez-Ilizaliturri FJ, Czuczman MS, et al. Vorinostat, A Histone Deacetylase (HDAC) Inhibitor, Promotes Cell Cycle Arrest and Re-Sensitizes Rituximab- and Chemo-Resistant Lymphoma Cells to Chemotherapy Agents. J Cancer Res Clin Oncol (2016) 142(2):379–87. doi: 10.1007/s00432-015-2026-y
- Hussain K, Cragg MS, Beers SA. Remodeling the Tumor Myeloid Landscape to Enhance Antitumor Antibody Immunotherapies. *Cancers* (2021) 13 (19):4904. doi: 10.3390/cancers13194904
- 134. Gradishar WJ. HER2 Therapy-an Abundance of Riches. *N Engl J Med* (2012) 366(2):176–8. doi: 10.1056/NEJMe1113641
- 135. Feugier P. A Review of Rituximab, the First Anti-CD20 Monoclonal Antibody Used in the Treatment of B Non-Hodgkin's Lymphomas. *Future* Oncol (London England) (2015) 11(9):1327–42. doi: 10.2217/fon.15.57
- 136. Petrelli F, Ardito R, Ghidini A, Zaniboni A, Ghidini M, Barni S, et al. Different Toxicity of Cetuximab and Panitumumab in Metastatic Colorectal Cancer Treatment: A Systematic Review and Meta-Analysis. Oncology (2018) 94(4):191–9. doi: 10.1159/000486338
- 137. Park JW, Han JW. Targeting Epigenetics for Cancer Therapy. Arch Pharmacal Res (2019) 42(2):159-70. doi: 10.1007/s12272-019-01126-z
- 138. Cheng Y, He C, Wang M, Ma X, Mo F, Yang S, et al. Targeting Epigenetic Regulators for Cancer Therapy: Mechanisms and Advances in Clinical Trials. Signal Transduct Target Ther (2019) 4:62. doi: 10.1038/s41392-019-0095-0
- 139. Hull EE, Montgomery MR, Leyva KJ. HDAC Inhibitors as Epigenetic Regulators of the Immune System: Impacts on Cancer Therapy and Inflammatory Diseases. *BioMed Res Int* (2016) 2016:8797206. doi: 10.1155/ 2016/8797206
- 140. Li Y, Seto E. HDACs and HDAC Inhibitors in Cancer Development and Therapy. Cold Spring Harbor Perspect Med (2016) 6(10):a026831. doi: 10.1101/cshperspect.a026831

- 141. Park J, Thomas S, Munster PN. Epigenetic Modulation With Histone Deacetylase Inhibitors in Combination With Immunotherapy. *Epigenomics* (2015) 7(4):641–52. doi: 10.2217/epi.15.16
- 142. Jiao S, Xia W, Yamaguchi H, Wei Y, Chen MK, Hsu JM, et al. PARP Inhibitor Upregulates PD-L1 Expression and Enhances Cancer-Associated Immunosuppression. *Clin Cancer Res* (2017) 23(14):3711–20. doi: 10.1158/ 1078-0432.Ccr-16-3215
- 143. Migliorini D, Bogaerts S, Defever D, Vyas R, Denecker G, Radaelli E, et al. Cop1 Constitutively Regulates C-Jun Protein Stability and Functions as a Tumor Suppressor in Mice. J Clin Invest (2011) 121(4):1329–43. doi: 10.1172/jci45784
- 144. Chauvin JM, Zarour HM. TIGIT in Cancer Immunotherapy. J Immunother Cancer (2020) 8(2):e000957. doi: 10.1136/jitc-2020-000957
- Batlle E, Massagué J. Transforming Growth Factor-β Signaling in Immunity and Cancer. Immunity (2019) 50(4):924–40. doi: 10.1016/j.immuni.2019.03.024
- 146. Castro F, Cardoso AP, Gonçalves RM, Serre K, Oliveira MJ. Interferon-Gamma at the Crossroads of Tumor Immune Surveillance or Evasion. Front Immunol (2018) 9:847. doi: 10.3389/fimmu.2018.00847

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