DOI: 10.1002/sctm.21-0208

#### PREVIEWS

Previews highlight research articles published in the current issue of STEM CELLS TRANSLATIONAL MEDICINE, putting the results in context for readers.



# Stuart P. Atkinson

Centro de Investigación Príncipe Felipe, Valencia, Spain. Email: satkinson@cipf.es

Mitogen-activated protein kinases (MAPKs or MAP kinases) are serine-/ threonine-specific protein kinases that phosphorylate substrates that control cellular functions such as proliferation, gene expression, differentiation, mitosis, survival, and apoptosis in response to various stimuli. In mammals, MAPKs comprise three subfamilies-the extracellular signalregulated kinase (ERK) MAPKs, which are activated by growth factors and mitogens, and the c-Jun N-terminal kinase (JNK) and p38 (p38) MAPKs, which are activated by cellular stressors and inflammatory cytokines. A large body of work has explored the importance of MAPK signaling to early development and the function of stem cell populations such as embryonic stem cells,<sup>1</sup> skeletal muscle stem cells,<sup>2</sup> and lung stem cells.<sup>3</sup> Given the importance of MAPKs and the specific importance of p38/MAPKs in response to several types of stress, targeting MAPK signaling pathways may represent an interesting means of supporting exogenous stem cell therapies or promoting endogenous stem cell-mediated regeneration and tissue repair in patients suffering from a range of diverse conditions. In the first of our featured articles published this month in STEM CELLS Translational Medicine, Liang et al report that ferulic acid treatment alleviates irradiation-induced impairments in skeletal stem cells (SSCs) and promotes bone regeneration through p38/MAPK and ERK/MAPK signaling pathway reactivation and so may promote bone repair following skeletal tumor treatment.<sup>4</sup> In a related article published recently in STEM CELLS, Shafiguzzaman et al demonstrated, for the first time, the role of the Tak1 (mitogen-activated protein kinase kinase kinase 7)-p38/MAPK pathway in club cell regeneration and bronchiolar epithelial repair and, in doing so, highlighted a potential therapeutic target for bronchiole-related disorders.5

The platelet-derived growth factor (PDGF) family primarily regulates the growth and division of mesenchymal cells such as fibroblasts and smooth muscle cells. The four polypeptide chains encoded by the PDGFA, PDGFB, PDGFC, and PDGFD genes form five different dimeric glycoproteins: PDGF-AA, -BB, -AB, -CC, and -DD. The binding of PDGFs to receptors that possess differential binding affinities induces the dimerization of PDGFR- $\alpha$  and - $\beta$  chain isoforms to form homodimeric and heterodimeric receptors and the activation of receptor tyrosine kinase activity. PDGFR activation stimulates a range of crucial intracellular signaling pathways that promote cell proliferation and migration<sup>6</sup> and plays essential roles during embryogenesis, controlling the development of the lungs, intestines, skin, testis, kidneys, hematopoietic system, and blood vessels. While PDGF also plays a vital role in wound healing in later life,<sup>7</sup> PDGF signaling can contribute to the development of cancer, inflammatory disease, pulmonary fibrosis, atherosclerosis, asthma, and chronic obstructive pulmonary disease.<sup>8</sup> In the second of our featured articles published this month in STEM CELLS Translational Medicine, Shaw et al explore how aging and injury impact the responses of kidney mesenchymal cells through a detailed analysis of marker protein/gene expression, which includes the specific expression of PDGFR- $\alpha$  and PDGFR- $\beta$  by renal myofibroblasts.<sup>9</sup> In a related article published recently in STEM CELLS, Zhou et al reported on how synergistic interactions between mesenchymal stem cells (MSCs) and endothelial cells could improve bone regeneration and how the differentiation of MSCs into pericytes through the PDGF-BB/PDGFR- $\beta$  signaling pathway supported blood vessel formation.<sup>10</sup>

# FEATURED ARTICLES

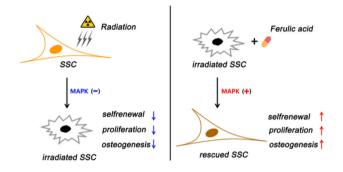
# MAPK Signaling Controls Skeletal Stem Cell-Mediated Tissue Regeneration After Irradiation

Previous research from Heng Zhu (Beijing Institute of Radiation Medicine) and Li Ding (Air Force Medical Center, PLA, Beijing, China) established a role for SSCs in targeting host immune cells to control inflammation<sup>11</sup> and regulating bone remodeling by suppressing inflammatory osteoclastogenesis.<sup>12</sup> Their subsequent research sought to improve bone reconstruction strategies following skeletal tumor treatment by radiotherapy/surgery by exploring the effect of irradiation on SSC biology and bone regeneration and the potential for ferulic acid, a phytochemical with antioxidant and therapeutic activities, to alleviate any potential radiation-induced stem cell damage.<sup>13,14</sup> As reported in their *STEM CELLS Translational Medicine* article,<sup>4</sup> Liang et al discovered that irradiation suppressed proliferation, colony formation, and the osteogenic differentiation of SSCs and induced a general decrease in their number. Furthermore, subsequent proteomic and transcriptomic analyses revealed that irradiation reduced the activation of the p38/MAPK and ERK/MAPK signaling pathways and inhibited the expression of crucial tissue regeneration-associated

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2021 The Author. STEM CELLS TRANSLATIONAL MEDICINE published by Wiley Periodicals LLC on behalf of AlphaMed Press

PREVIEWS

genes. Importantly, ferulic acid treatment of irradiated SSCs induced the reactivation of the p38/MAPK and ERK/MAPK signaling pathways to significantly rescue radiation-induced stem cell impairments; furthermore, the specific activation of these pathways by ferulic acid treatment also improved SSC-mediated bone repair in an irradiated bone defect mouse model. Overall, the authors propose a crucial role for the p38/MAPK and ERK/MAPK signaling pathways in regulating the self-renewal of SSCs and SSC-mediated tissue regeneration following irradiation, thereby underscoring the relevance of these findings to the treatment of skeletal cancer patients.

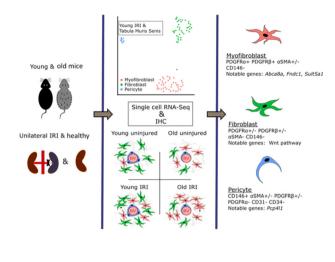


https://doi.org/10.1002/sctm.20-0536

# Analysis of Mesenchymal Cells Helps to Define Age-Related Differences in Kidney Injury Responses

The impact of acute kidney injury<sup>15</sup> on the renal mesenchyme can reduce peritubular capillary density to worsen the loss of nephrons and promote fibrosis, a hallmark of chronic kidney disease.<sup>16</sup> The mesenchymal cells of the kidney significantly influence these disease-associated processes; however, we understand little regarding how aging impacts mesenchymal cells and their response to injury. In the hope of filling this knowledge gap, researchers led by Bruno Péault (University of California, Los Angeles, California, USA) and David A. Ferenbach (University of Edinburgh, UK) characterized renal mesenchymal cell heterogeneity in young and aged mice under normal conditions and after ischemia-reperfusion-injury using multiplexed immunolabeling and single-cell transcriptomics. In their recent STEM CELLS Translational Medicine article,<sup>9</sup> Shaw et al discovered that the expression profiles of a range of perivascular cell marker proteins correlated well with their location within the complex architecture of the renal mesenchyme; of note, PDGFR- $\alpha$  and PDGFR- $\beta$ co-expression efficiently labeled renal myofibroblasts (formed during kidney fibrosis) while CD146 was identified as an optimal renal pericyte marker. Notably, single-cell RNA sequencing of murine kidneys demonstrated that mesenchymal cells segregated into three subtypes with distinct expression patterns with aging and following injury (pericytes, fibroblasts, and myofibroblasts); overall, uninjured aged kidneys resembled injured young kidneys. The authors hope that their integrated description of how marker expression in the

renal mesenchyme changes with age and in response to injury may allow the more accurate definition of renal mesenchymal cell populations and the identification of age-specific pathways and potential therapeutic targets.



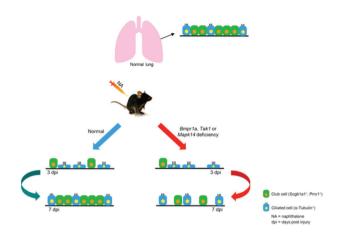
https://doi.org/10.1002/sctm.20-0392

#### **RELATED ARTICLES**

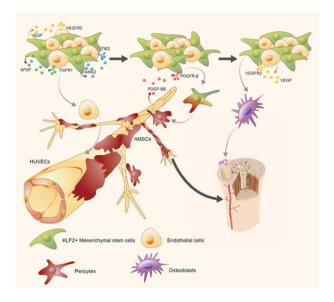
# MAPK Signaling Controls Club Cell Regeneration and Bronchiolar Epithelial Repair

The club cells of the small airways (bronchioles) protect the bronchiolar epithelium and detoxify inhaled harmful substances. Self-renewing progenitor-like club cells differentiate into ciliated epithelial cells and contribute to the repair of the bronchiolar epithelium, which represents a significant site for the development of life-threatening disorders, such as chronic obstructive pulmonary disease and lung adenocarcinoma.<sup>17,18</sup> In a recent STEM CELLS article,<sup>5</sup> researchers led by Huijuan Liu (Shanghai Jiao Tong University, Shanghai, China) explored the identity of progenitor club cells and the mechanisms by which these cells regenerate the bronchiolar epithelium. Shafiguzzaman et al now report that a noncanonical bone morphogenic protein (BMP) pathway involving Tak1 and MAPK<sup>19,20</sup> plays a crucial role in bronchiolar epithelium regeneration by promoting club cell proliferation and expansion. The authors employed lineage tracing and conditional gene knockout mouse studies to establish the homeobox transcription factor Prrx1 as a marker for adult mouse club cells during homeostasis and regeneration. Their analyses also established the importance of a noncanonical signaling pathway comprising BMPs and a BMP receptor type 1A (BMPR1A)-Tak1-p38/MAPK signaling pathway in club cell regeneration; while the loss of Bmpr1a, Tak1, or Mapk14 (encoding  $p38\alpha$ ) expression in Prrx1-expressing club cells had negligible impact under homeostatic conditions, gene deficiencies significantly impacted club cell regeneration and bronchiole repair in adult mice by reducing the proliferation and expansion of regenerating

club cells. Overall, these findings represented the first description of the BMPR1A-mediated Tak1-p38/MAPK pathway as a regulatory mechanism affecting club cell regeneration and bronchiolar epithelium repair and a potential therapeutic target for bronchiole-related disorders.



dimensional cell scaffolds may represent an efficient means of significantly enhancing bone defect regeneration.



https://doi.org/10.1002/stem.3125

#### Exploring the Synergy of MSCs and Endothelial Cells in Bone Regeneration Strategies

As MSC and endothelial cell coculture-based studies suggested a reciprocal increase in MSC-derived osteogenesis<sup>21</sup> and endothelial cellderived angiogenesis,<sup>22</sup> researchers from the laboratories of Mengfei Yu and Huiming Wang (Zhejiang University, Hangzhou, China) explored this synergistic interaction to develop enhanced bone regeneration strategies. As reported in their recent STEM CELLS article,<sup>10</sup> Zhou et al evaluated the interaction of human MSCs expressing Krüppel-like factor 2 (KLF2), which codes for a zinc-finger transcription factor, and human umbilical vein endothelial cells cultivated in conventional twodimensional and advanced three-dimensional gelatin methacrylate hydrogel coculture systems with optimized media formulations. The authors highlighted the crucial role of KLF2 expression in maintaining the stem-like nature of MSCs and enhancing the MSC-mediated induction of endothelial cell-mediated angiogenesis, which itself subsequently stimulated MSC-mediated osteogenesis. Interestingly, the secretion of a range of crucial angiogenic factors by MSCs (including basic fibroblast growth factor, vascular endothelial growth factor, and angiopoietin-2) enhanced endothelial cell-derived angiogenesis; however, this also prompted the differentiation of MSCs into pericytes through the PDGF-BB/PDGFR- $\beta$  signaling pathway to promote the formation of maturing blood vessels. Meanwhile, the increased expression of vascular endothelial growth factor by maturing endothelial cells enhanced the osteogenic potential of the remaining MSCs. Overall, the findings of this exciting study suggested that the implantation of KLF2expressing human MSCs and endothelial cells within advanced three-

#### REFERENCES

https://doi.org/10.1002/stem.3120

- 1. Binétruy B, Heasley L, Bost F, Caron L, Aouadi M. Concise review: regulation of embryonic stem cell lineage commitment by mitogenactivated protein kinases. STEM CELLS. 2007;25:1090-1095.
- 2. Pawlikowski B, Vogler TO, Gadek K, Olwin BB. Regulation of skeletal muscle stem cells by fibroblast growth factors. Dev Dyn. 2017;246:359-367.
- 3. Ventura JJ, Tenbaum S, Perdiguero E, et al. p38 $\alpha$  MAP kinase is essential in lung stem and progenitor cell proliferation and differentiation. Nat Genet. 2007;39:750-758.
- 4. Liang J-W, Li P-L, Wang Q, et al. Ferulic acid promotes bone defect repair after radiation by maintaining the stemness of skeletal stem cells. STEM CELLS TRANSLATIONAL MEDICINE, 2021:10:1217-1231.
- 5. Shafiquzzaman M. Biswas S. Li P. Mishina Y. Li B. Liu H. The noncanonical BMP signaling pathway plays an important role in club cell regeneration. STEM CELLS. 2020;38:437-450.
- 6. Demoulin J-B, Essaghir A. PDGF receptor signaling networks in normal and cancer cells. Cytokine Growth Fact Rev. 2014;25:273-283.
- 7. Andrae J, Gallini R, Betsholtz C. Role of platelet-derived growth factors in physiology and medicine. Genes Dev. 2008;22:1276-1312.
- 8. Kardas G, Daszyńska-Kardas A, Marynowski M, Brzakalska O, Kuna P, Panek M. Role of platelet-derived growth factor (PDGF) in asthma as an immunoregulatory factor mediating airway remodeling and possible pharmacological target. Front Pharmacol. 2020;11:47.
- 9. Shaw IW, O'Sullivan ED, Pisco AO, et al. Aging modulates the effects of ischemic injury upon mesenchymal cells within the renal interstitium and microvasculature. STEM CELLS TRANSLATIONAL MEDICINE. 2021;10:1232-1248.
- 10. Zhou Y, Liu C, He J, et al. KLF2<sup>+</sup> stemness maintains human mesenchymal stem cells in bone regeneration. STEM CELLS. 2020;38:395-409.
- 11. Zhu H, Yang F, Tang B, et al. Mesenchymal stem cells attenuated PLGA-induced inflammatory responses by inhibiting host DC maturation and function. Biomaterials. 2015;53:688-698.
- 12. Li X, Ding L, Wang Y-X, et al. Skeletal stem cell-mediated suppression on inflammatory osteoclastogenesis occurs via concerted action of

- Ma Z-C, Hong Q, Wang Y-G, et al. Effects of ferulic acid on hematopoietic cell recovery in whole-body gamma irradiated mice. *Int J Radiat Biol.* 2011;87:499-505.
- Du K, Li Z, Fang X, et al. Ferulic acid promotes osteogenesis of bone marrow-derived mesenchymal stem cells by inhibiting microRNA-340 to induce β-catenin expression through hypoxia. *Eur J Cell Biol.* 2017; 96:496-503.
- Susantitaphong P, Cruz DN, Cerda J, et al. World incidence of AKI: a meta-analysis. *Clin J Am Soc Nephrol.* 2013;8:1482-1493.
- Chawla LS, Kimmel PL. Acute kidney injury and chronic kidney disease: an integrated clinical syndrome. *Kidney Int.* 2012;82: 516-524.

- 17. Barnes PJ. Senescence in COPD and its comorbidities. *Annu Rev Physiol.* 2017;79:517-539.
- Sutherland KD, Berns A. Cell of origin of lung cancer. Mol Oncol. 2010;4:397-403.
- Shibuya H, Iwata H, Masuyama N, et al. Role of TAK1 and TAB1 in BMP signaling in early Xenopus development. EMBO J. 1998;17:1019-1028.
- 20. Shim J-H, Greenblatt MB, Xie M, et al. TAK1 is an essential regulator of BMP signalling in cartilage. *EMBO J.* 2009;28:2028-2041.
- Xue Y, Xing Z, Hellem S, Arvidson K, Mustafa K. Endothelial cells influence the osteogenic potential of bone marrow stromal cells. *BioMed Eng Online*. 2009;8:34.
- Pedersen TO, Blois AL, Xue Y, et al. Osteogenic stimulatory conditions enhance growth and maturation of endothelial cell microvascular networks in culture with mesenchymal stem cells. J Tissue Eng. 2012;3: 2041731412443236.